

# Jesse McNichol

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## Summary of Experience

I am an Assistant Professor in the department of Biology at St. Francis Xavier University, where I teach microbiology and run a research program in marine microbial ecology. My research is currently primarily funded by the Simons Foundation International as part of the [CBIOMES collaboration](#).

My academic background is in oceanography and microbial ecology, and I have experience in both wet-lab and *in silico* approaches for generating and analyzing high-throughput sequencing data. This includes 3-domain metabarcoding, shotgun metagenomics, as well as related approaches for quantifying microbial activity using fluorescence *in situ* hybridization.

## Education

**Massachusetts Institute of Technology - Woods Hole Oceanographic Institution**, Cambridge/Woods Hole, Massachusetts, USA

*Ph.D., Biological Oceanography*

2011-2016

Thesis: [Productivity, Metabolism and Physiology of Free-Living Chemoautotrophic \*Epsilonproteobacteria\*](#) (Advisor: Dr. Stefan Sievert)

**Mount Allison University**, Sackville, New Brunswick, Canada

*B.Sc., First Class Honours with Distinction, Biology*

2003-2008

Thesis: Endophytic Fungi of Liverworts (*Bryophyta*) in a Copper-Contaminated Environment (Advisors: Drs. Robert Thompson and Felix Baerlocher)

## Professional Experience

**St. Francis Xavier University**; Antigonish, NS, Canada

*Assistant Professor*

2023 - present

**University of Southern California, Fuhrman Lab**; Los Angeles, CA, USA

*Postdoctoral Scholar*

2018 - 2023

- Compiled an atlas of microbial biogeography ([GRUMP](#)) based on existing meta-'omics datasets and newly generated amplicon sequence libraries
- Developed software pipelines for the analysis of amplicon sequences, intercomparison with metagenomes, and *in-silico* evaluation of primer performance

**Chinese University of Hong Kong, Luo Lab;** Shatin, New Territories, HK

*Research Assistant* 2016 - 2017

- Developed plate-based isolation methods for high-throughput isolation of fastidious chemoautotrophic *Campylobacteria*
- Isolated sulfur-oxidizing microorganisms from hydrothermal vents and coastal sediments for pangenome and population genetic analysis
- Developed a high-through put isolation system for selective enrichment of roseobacter group bacteria from local sediments

**Woods Hole Oceanographic Institution, Sievert Lab;** Woods Hole, MA, USA

*Graduate Research Assistant* 2011-2016

- Incubated deep-sea hydrothermal vent chemoautotrophic communities at *in-situ* pressure and temperature to infer their ecophysiology and biogeochemical impact
- Cultivated *Sulfurimonas denitrificans* in a custom-built chemostat to validate a theoretical model of energy conservation for this organism

**National Research Council of Canada;** Halifax, NS, Canada

*Technical Officer* 2009-2011

**Environment Canada;** Moncton, NB, Canada

*Environmental Technician* 2009

**Atlantic Canada Conservation Data Center;** Sackville, NB, Canada

*Assistant Field Botanist* 2008

**Marine Macroecology and Biogeochemistry Lab;** Sackville, NB, Canada

*Summer Research Student* 2007-2008

**Grants &  
Fellowships**

**2024-2026: NRC Ocean Cluster Support Program (\$88,384 CAD)**

*Effects of nano/microplastic exposure on zebrafish (Danio rerio) gut microbiome*

**2024-2027: CBIOMES Scholars Program, Simons Foundation  
International (\$359,396 USD)**

*Predicting Community Resilience to Environmental Perturbations by  
Quantifying in situ Physiology of Microdiverse Subclusters of Uncultured  
Marine Prokaryotes*

**2023: StFX Startup Funding (\$27,000 CAD)**

**2016: JGI Small-Scale Microbial / Metagenome Program**

*Investigating the Genetic Basis of Differential Oxygen Tolerance in Sulfurimonas Ecotypes from the Seafloor Biosphere Using Single-Cell Genomics*

**2014-2016: NASA Earth Systems Science Fellowship**

*Quantifying Energy Metabolism and Associating Function with Taxonomy for Chemosynthetic Microbial Communities at Deep-Sea Hydrothermal Vents*

**2013-2016: Natural Sciences and Engineering Research Council of Canada, Post-Graduate Scholarship (Doctoral level)**

*Quantifying Energy Metabolism and Associating Function with Taxonomy for Chemolithoautotrophic Microbial Communities at Deep-Sea Hydrothermal Vents*

**2013-2014: Canadian Meteorological and Oceanographic Society Scholarship Supplement**

**2011-2012: Natural Sciences and Engineering Research Council of Canada, Post-Graduate Scholarship (Master's level)**

*Psychrophilic bacteria in the Canadian Arctic*

**Research Impact** Total citations: 817, 499 citations since 2020  
h-index: 13 (source: [Google Scholar](#))

**Preprints & in-review publications** **McNichol, J.\***, Sievert, S.M.\*, 2019. [Reconciling a Model of Core Metabolism with Growth Yield Predicts Biochemical Mechanisms and Efficiency for a Versatile Chemoautotroph](#). bioRxiv. \*Co-corresponding authors.

**McNichol, J.\***, Williams L.R.\*, Raut, Y., Fuhrman, J.A. *et al.* [Characterizing organisms from three domains of life with universal primers from throughout the global ocean](#). bioRxiv (in review for *Scientific Data*). \*Co-first authors.

**Publications in Prep** Ser-Giacomi, E.\*, **McNichol, J.\***, Raut, Y., Yeh, Y.-C., Ribalet, F., Hassle., C., Rees, A., Tarran, G., Gradoville, M., Turk-Kubo, K., Zehr, J., Dutkiewicz, S., Follows, M., Fuhrman, J.A. Estimating Absolute Microbial Abundances By Combining Metabarcoding and Cell-Count Data. \*Co-first authors.

**Peer-  
Reviewed  
Publications**

Zakem, E.J., **McNichol, J.**, Weissman, J.L., Raut, Y., Xu, L., Halewood, E.R., Carlson, C.A., Dutkiewicz, S., Fuhrman, J.A., Levine, N.M., 2025. Functional biogeography of marine microbial heterotrophs. *Science* (accepted March 6th 2025).

Mishra, A., **McNichol, J.**, Fuhrman, J., Blei, D., Müller, C.L., 2024. Variational inference for microbiome survey data with application to global ocean data. *ISME Communications* (accepted March 21, 2025).

Jones-Kellett, A.E., **McNichol, J.**, Raut, Y., Cain, K.R., Ribalet, F., Armbrust, E.V., Follows, M.J., Fuhrman, J.A., 2024. [Amplicon Sequencing with Internal Standards Yields Accurate Absolute Picocyanobacteria Cell Abundances as Validated with Flow Cytometry](#). *ISME Communications*. 4(1), ycae115.

Mino, S., Fukazawa, S., Tsuchiya, J., **McNichol, J.**, Sievert, S.M., Yamaki, S., Ando, Y., Sawabe, T., 2023. [Hydrogenimonas cancrithermarum](#) sp. nov., a hydrogen- and thiosulfate-oxidizing mesophilic chemolithoautotroph isolated from diffuse-flow fluids on the East Pacific Rise, and an emended description of the genus *Hydrogenimonas*. *IJSEM*. 73, 006132.

Milke, F., Sanchez-Garcia, S., Dlugosch, L., **McNichol, J.**, Fuhrman J.A., Simon, M., Wagner-Döbler I., 2022. [Composition and Biogeography of Pro- and Eukaryotic Communities in the Atlantic Ocean: Primer Choice Matters](#). *Frontiers in Microbiology*. 13, 895875.

**McNichol, J.**<sup>1,2</sup>, Dykma S.<sup>1</sup>, Mußmann M., Seewald, J., Sylva, S., Sievert S.<sup>2</sup>, 2022. [Genus-Specific Carbon Fixation Activity Measurements Reveal Distinct Responses to Oxygen Among Hydrothermal Vent \*Campylobacteria\*](#). *Applied and Environmental Microbiology*. 88(2), e02083-21. <sup>1</sup>Co-first authors. <sup>2</sup>Co-corresponding authors.

Lin, X.Q., Chu, X., **McNichol, J.**, Qian, Y., Luo, H.W., 2022. [Cryptic Niche Differentiation of Novel Sediment Ecotypes of \*Ruegeria pomeroyi\* Correlates with Nitrate Respiration](#). *Environmental Microbiology*. 24(1), 390-403.

**McNichol, J.\***, Berube, P., Biller, S., Fuhrman, J., 2021. [Evaluating and Improving SSU rRNA PCR Primer Coverage for Bacteria, Archaea, and Eukaryotes Using Metagenomes from Global Ocean Surveys](#). *mSystems*. 6(3), e00565-21. \*Corresponding author.

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- Yeh, Y.C., **McNichol, J.**, Needham, D., Fichot, E., Berdjeb, L., Fuhrman, J., 2021. [Comprehensive single-PCR 16S and 18S rRNA community analysis validated with mock communities, and estimation of sequencing bias against 18S](#). *Environmental Microbiology*. doi: 10.1111/1462-2920.15553.
- Götz, F., Pjevac, P., Markert, S., **McNichol, J.**, Becher, D., Schweder, T., Mußmann, M., Sievert, S.M., 2019. [Transcriptomic and proteomic insight into the mechanism of cyclooctasulfur- versus thiosulfate-oxidation by the chemolithoautotroph \*Sulfurimonas denitrificans\*](#). *Environmental Microbiology* 21, 244–258.
- Labonté, J.M., Pachiadaki, M., Fergusson, E., **McNichol, J.**, Grosche, A., Gulmann, L.K., Vetriani, C., Sievert, S.M., Stepanauskas, R., 2019. [Single Cell Genomics-Based Analysis of Gene Content and Expression of Prophages in a Diffuse-Flow Deep-Sea Hydrothermal System](#). *Frontiers in Microbiology*. 10.
- McNichol, J.\***, Stryhanyuk, H., Sylva, S.P., Thomas, F., Musat, N., Seewald, J.S., Sievert, S.M.\*, 2018. [Primary Productivity Below the Seafloor at Deep-Sea Hot Springs](#). *Proceedings of the National Academy of Sciences*. 115, 6756–6761. \*Co-corresponding authors.
- Götz, F., Longnecker, K., Soule, M.C.K., Becker, K.W., **McNichol, J.**, Kujawinski, E.B., Sievert, S.M., 2018. [Targeted metabolomics reveals proline as a major osmolyte in the chemolithoautotroph \*Sulfurimonas denitrificans\*](#). *Microbiology Open*. e586.
- McNichol, J.**, Sylva, S.P., Thomas, F., Taylor, C.D., Sievert, S.M., Seewald, J.S., 2016. [Assessing microbial processes in deep-sea hydrothermal systems by incubation at \*in situ\* temperature and pressure](#). *Deep Sea Research Part I: Oceanographic Research Papers*. 115, 221–232.
- McNichol, J.**, MacDougall, K.M., Melanson, J.E., McGinn, P.J., 2012. [Suitability of soxhlet extraction to quantify microalgal fatty acids as determined by comparison with \*in situ\* transesterification](#). *Lipids*. 47, 1–13.
- MacDougall, K.M., **McNichol, J.**, McGinn, P.J., O’Leary, S.J.B., Melanson, J.E., 2011. [Triacylglycerol profiling of microalgae strains for biofuel feedstock by liquid chromatography–high-resolution mass spectrometry](#). *Analytical and Bioanalytical Chemistry*. 401, 2609–2616.
- Park, K.C., Whitney, C., **McNichol, J.**, Dickinson, K.E., MacQuarrie, S., Skrupski, B.P., Zou, J., Wilson, K.E., O’Leary, S.J.B., McGinn, P.J., 2011. [Mixotrophic and photoautotrophic cultivation of 14 microalgae isolates](#)

from Saskatchewan, Canada: potential applications for wastewater remediation for biofuel production. *Journal of Applied Phycology*. 24, 339–348.

**Book  
Chapters &  
Other  
Scientific  
Writing**

**McNichol, J.**, 2024. [Mineral-eating microorganisms at extinct hydrothermal vents](#). "News and Views" for Nature Microbiology. 9, 589-590.

Sievert, S.M., **McNichol, J.**, Thomas, F., Panayotova, E., Watson R., 2018. [How Do Deep-Sea Hot Spring Ecosystems Work?](#) *Environmental Science Journal for Teens*.

**McNichol, J.**, 2017. [Breaking the Oxygen Barrier in Microbial Cultivation](#), Small Things Considered Blog, American Society for Microbiology.

**McNichol, J.**, 2014. [A First-Time Diver's Experience](#). Blog for research cruise AT2623 (R/V *Atlantis*).

**McNichol, J.**, 2014. "Mail Buoy" responses to gradeschool student questions about deep-sea hydrothermal vent ecosystems: [Jan 12th](#), [Jan 13th](#), [Jan 20th](#), [Jan 30th](#). Research cruise AT26-10 (R/V *Atlantis*).

**McNichol, J.**, McGinn, P.J., 2012. [Adapting Mass Algalculture for a Northern Climate](#), in: Gordon, R., Seckbach, J. (Eds.), *The Science of Algal Fuels, Cellular Origin, Life in Extreme Habitats and Astrobiology*. Springer Netherlands, pp. 131–146.

**McNichol, J.**, Gordon, R., 2012. [Are We from Outer Space?: A Critical Review of the Panspermia Hypothesis](#), in: Seckbach, J. (Ed.), *Genesis - In The Beginning, Cellular Origin, Life in Extreme Habitats and Astrobiology*. Springer Netherlands, pp. 591– 619.

**McNichol, J.**, 2008. [Primordial soup, fool's gold, and spontaneous generation](#). *Biochemistry and Molecular Biology Education* 36, 255–261.

**Scientific Workflows & Tools**     **McNichol, J.**, Yeh Y.C., Aleman, M., Fuhrman J., 2019. A custom bash pipeline to process 515Y/926R mixed 16S/18S amplicons into ASVs ([protocol link](#)).

**McNichol, J.**, 2021. *MGPrimerEval*: A reproducible snakemake pipeline to compare amplicon primer sequences with metagenomes to quantify their *in-silico* coverage and real-world performance by comparing metagenomes with amplicons ([pipeline link](#)).

**Teaching Experience**     **St. Francis Xavier University**; Antigonish, NS, Canada  
    *Biology 398: Exploring Native Microbiota* (~8 students, taught yearly)  
    *Biology 315: Introduction to Microbiology* (~40 students, taught yearly)  
    *Biology 115: Introduction to Microbiology for Nursing* (~90 students, taught yearly)  
    *Biology 215: Introduction to Microbiology for Nutrition* (~20 students, taught yearly)

**Woods Hole Oceanographic Institution**; Woods Hole, MA, USA  
    *Biological Oceanography* (TA, Spring 2015)  
    *Marine Microbiology and Biogeochemistry* (TA, Fall 2013)

**Mount Allison University**; Sackville, NB, Canada  
    *Native Flora* (TA, Fall 2007)

**Research Mentoring & Supervisory Experience**     **2023 - present:** Mentoring undergraduate student Jillian Davies in microbial metabarcoding techniques for analyzing zebrafish microbiomes in collaboration with NRC colleagues and Dr. Tammy Rodela.

**2021 - present:** Mentoring PhD student Laiza Faria in laboratory techniques for metabarcoding, metagenomics, and associated data analysis. Also assisted with successful application for funding to the Scientific Committee on Antarctic Research (SCAR) to travel to Antigonish, NS for research collaboration (July-August 2024).

**2021 - 2023, USC:** Mentored postdoctoral scholar Yubin Raut in a project that aims to use bioinformatic data from global ocean basins to validate and improve with the global-scale biogeochemical model DARWIN.

**2019 - 2021, USC:** Mentored undergraduate Bruce Yanpui Chan in bioinformatic analyses and assisted with a successful funding application



to the program Student Opportunities for Academic Research (SOAR).

**2018 - 2022, USC:** Assisted graduate students Selene Sanchez-Garcia (Technical University Braunschweig) and Felix Milke (University of Oldenburg) with amplicon analysis, data interpretation, and conference presentations.

**2018 - 2023, USC:** Assisted Fuhrman lab graduate students in implementing a qiime2 workflow for the analysis of PCR amplicon data.

**2017 - 2018, CUHK:** Guided the final-year undergraduate project\* of Annie Wing-Yi Lo, which tested the effect of microoxic conditions on the isolation of sulfur-oxidizing microbes from shallow-water hydrothermal vents and local sediments. \*Equivalent of Honours thesis.

**2015 - 2016, WHOI:** Supervised the independent research project of volunteer Dali Smolsky to isolate novel autotrophic *Campylobacteria* from salt marsh and hydrothermal vent environments.

**2010, NRC:** Taught students techniques for lipid extraction and transesterification of microalgal biomass with bio-compatible solvents.

## **Pedagogical and Research Training** **Maple League of Universities**

*The Maple League Micro-Certificate in Teaching and Learning (2023-2024)*

**edX Online Learning Platform**

*Becoming a More Equitable Educator: Mindsets and Practices (Spring 2021)*

**CyVerse Learning Network**

*Foundational Open Science Skills Online (Spring 2021)*

**University of Southern California, Center for Excellence in Teaching**

*Future Faculty Teaching Institute (Spring 2020)*

**Science Outreach & Other** X-CHEM, Introduction to Environmental Microbiology (July 2024, grades 5-8, 2 hour class)

**Educational Experience** Fun Interactive Science Hour ([FISH](#)) Outreach Program (USC) / Los Angeles Public Library (Dec 2020 - 2023)

Chinese Summer Language Village, Mount Allison University (2008)

Planet Performers environmental drama, Mount Allison University (2007-2008)



**University and Department Service** Faculty of Science Secretary (2023 - present)  
Science Atlantic Representative (Biology, 2024 - present)  
Joint Occupational Health and Safety Committee (starting January 2025)  
University Open House Day (Oct 2023, 2024)  
3 Minute Thesis judge (2024)  
Woods Hole Oceanographic Institution Joint Program Career Panel (2024)  
Honours thesis second reader: 2 (2023-2024: 2)

**Selected Data Products** McNichol, J., 2023. 3-Domain Amplicon Sequencing of Unfractionated (> 0.2 µm) DNA Collected During GO-SHIP P16N and P16S Lines (2005/2006).

McNichol, J., 2019. Exact Amplicon Sequence Variant Abundances from the ANT28-5 Latitudinal Transect of the Atlantic Ocean.

McNichol, J., Dyksma S., Mußmann M., and Sievert S., 2020. CARD-FISH Images from Incubations of Deep-Sea Hydrothermal Vent Fluid and Testing of Novel Probes for *Arcobacter*, *Sulfurimonas*, and *Sulfurovum*.

**Technical Comments** McNichol, J., Sievert, S.M. Comment on PMID 26929299: Carbon Fixation Driven by Molecular Hydrogen Results in Chemolithoautotrophically Enhanced Growth of *Helicobacter pylori*. In: PubMed Commons [Internet]. Bethesda (MD): National Library of Medicine; 2017 Feb 16. [Permalink](#).

**Invited Talks** Institute for Chemistry and Biology of the Marine Environment (ICBM); Oldenburg, Germany (COVID e-seminar, Nov 25th, 2020)

*Using Global Metagenomes to Evaluate and Improve PCR Primer Coverage and the Application of 3-Domain Amplicon Data to Trait-Based Models*

**Scripps Institution of Oceanography**; San Diego, CA (COVID e-seminar, June 17th, 2020).

*Using Global Metagenomes to Quantify PCR Primer Coverage and Integrating 3-Domain Amplicon Data with Models*

**CBIOMES e-meeting**; (online seminar, Dec 12th, 2018).

*Biogeography of Exact Amplicon Sequence Variants*

**The Swire Institute of Marine Science;** Hong Kong S.A.R., China (Mar 20th, 2016)

*Brimstone Bacteria: Primary Productivity and Microbial Ecology of Deep-Sea Hydrothermal Vents*

**University of Hong Kong, School of Biological Sciences;** Hong Kong S.A.R., China (Jan 29th, 2016)

*Brimstone Bacteria: Primary Productivity and Microbial Ecology of Deep-Sea Hydrothermal Vents*

**Max Planck Institute for Marine Microbiology;** Bremen, Germany (July 8th, 2014)

*Insights into chemolithoautotrophy at deep-sea hydrothermal vents from in-situ experiments and metabolic modeling*

**Conference &  
Workshop  
Activity**

**Scientific Committee on Antarctic Research (2024),** Pucón  
Punta Arenas, Chile

Co-author on 2 presentations, representing active collaborations:

*Unraveling the Antarctic Krill microbiome present in Krill internal and external body parts* (Mireia Miestre, National Museum of Natural Sciences, Madrid, Spain)

*Latitudinal Gradients and Vertical Connectivity in the Scotia Sea Microbiome* (Mireia Miestre, National Museum of Natural Sciences, Madrid, Spain)

**Ocean Sciences Meeting (2024)** New Orleans, LA, USA

Co-author on 5 presentations, representing active collaborations:

*Characterizing and comparing distributions of organisms within all 3-domains of life with universal primers from 1577 samples throughout the global ocean* (Nathan Williams, collaborator at USC)

*Using global rRNA metabarcoding datasets to assess a trait-based ecosystem model reveals the importance of missing and under-surveyed phytoplankton* (Yubin Raut, collaborator at MIT)

*The Fluid Dynamical Structuring of Microbial Communities Along an Eastern North Pacific Transect* (Lexi Jones-Kellett, collaborator at MIT)

*A mechanistic species coexistence theory explains the observed increase in marine microbial biodiversity with depth* (Liang Xu, collaborator at Carnegie Institute)

*Emergent structure and function of microbial heterotrophic communities in a model ocean* (Emily Zakem, collaborator at Carnegie Institute)

**Ocean Sciences Meeting (2020)** San Diego, CA, USA

*Testing model predictions and revealing basin-scale biogeography with whole-community PCR amplicons from GEOTRACES* (Oral Presentation)

*Atlantic and Pacific Ocean bacterioplankton diversity* (Co-author on student presentation).

**Ocean Nucleic Acids ‘omics Intercalibration and Standardization Workshop (2020)** UNC Chapel Hill, NC, USA (Invited Participant)

**Gordon Research Conference/Seminar in Marine Molecular Ecology (2017)** Hong Kong S.A.R., China

*Primary Productivity and Ecophysiology of Chemosynthetic Campylobacteria* (Poster Presentation)

**International Society for Microbial Ecology (2016)** Montreal, QC, Canada

*Primary Productivity and Ecology of the Subseafloor Biosphere at Deep-Sea Hydrothermal Vents, 9 °N East Pacific Rise* (Oral Presentation)

**Gordon Research Conference in Marine Molecular Ecology (2015)** Hong Kong S.A.R., China

*Bacterial Chemosynthesis at Deep-Sea Hydrothermal Vents Quantified by Cultivation at in-situ Pressure and NanoSIMS Analysis* (Poster Presentation)

**Gordon Research Seminar in Marine Molecular Ecology (2015)** Hong Kong S.A.R., China

*Incubations of Hydrothermal Vent Communities at In-situ Pressure and Temperature Quantify Community Primary Productivity of the Subseafloor*

*Biosphere* (Oral Presentation)

**American Society for Microbiology General Meeting (2015)** New Orleans, LA, USA *Simulated Seafloor Conditions Reveal Epsilonproteobacteria as Dominant Chemoautotrophs in Fluids from the Subseafloor Biosphere at Deep-Sea Vents* (Young Investigator Oral Presentation)

**American Society for Microbiology General Meeting (2014)** Boston, MA, USA

*A Genome-Scale Metabolic Model of Sulfurimonas denitrificans Provides Insight into the Process of Autotrophic Denitrification* (Young Investigator Oral Presentation)

**Department Seminars** **Woods Hole Oceanographic Institution** (Aug 1st, 2016)  
*Productivity, Metabolism and Physiology of Free-Living Chemoautotrophic Epsilonproteobacteria* (Thesis defense)

**Woods Hole Oceanographic Institution** (Feb 19th, 2015)  
*Simulated Seafloor Conditions Reveal Epsilonproteobacteria as Dominant Chemoautotrophs in Fluids from the Subseafloor Biosphere at Deep-Sea Vents*

**Professional Service** **Reviewer for:** Scientific Reports, PLoS One, Frontiers in Microbiology, International Journal of Systematic and Evolutionary Microbiology, Journal of Open-Source Science, Nature Communications, PeerJ, Molecular Ecology Resources, Environmental Microbiology, ISME Journal, Journal of Geophysical Research - Biogeosciences.

**Professional Affiliations** American Society for Microbiology (ASM), Canadian Society for Microbiology (CSM), The Oceanography Society (TOS)

**Research Cruises** **November 2014**, East Pacific Rise 9°N, R/V *Atlantis*: AT26-23, Chemoautotrophic Carbon Production at Deep-Sea Hydrothermal Vents

**January 2014**, East Pacific Rise 9°N, R/V *Atlantis*: AT26-10, Dimensions of Biodiversity, An Integrated Study of Energy Metabolism, Carbon Fixation, and Colonization Mechanisms in Chemosynthetic Microbial Communities at Deep-Sea Vents

**July-Aug 2012**, Rhode Island Continental Shelf, R/V *Endeavor*: Deep Ocean

## Benthic Sampler Cruise

### Technical Research Skills & Experience

- Aquatic microbial field sampling, preservation, and DNA extraction
- SSU rRNA amplicon / metagenomic / -transcriptomic library preparation
- Microbial pure culture isolation, including for oxygen-sensitive microorganisms (Hungate technique)
- Physiological growth experiments, including both batch and chemostat
- Incubations for measuring biogeochemical rates and microbial growth, including stable/radioisotope methods
- Microscopic enumeration and identification of environmental microorganisms, including Fluorescence *In Situ* Hybridization methods (e.g. CARD-FISH) and subsequent single-cell activity measurements (e.g. NanoSIMS)

### Software / Bioinform- atic skills

**Scripting and Plotting Languages:** Python, bash, R.

**Selected Bioinformatics Software Experience:** qiime2, jupyter, conda/mamba, snakemake, anvi'o, (meta)SPAdes, look@NanoSIMS, GTDB-tk, phyloFlash, bbtools.

### Languages

Written Chinese (functional with traditional and simplified characters), Mandarin Chinese (conversational), Cantonese (basic spoken), French (conversational), Spanish (functional)