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## TOWARDS A COMPREHENSIVE SURVEY OF $C_3$ AND $C_4$ PHOTOSYNTHETIC PATHWAYS IN CYPERACEAE

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#### ABSTRACT

Members of the family Cyperaceae were surveyed by original observation and from the literature to assess the distribution of  $C_3$  and  $C_4$  photosynthetic pathways in the family. All 107 genera were included in the current sample, with 91 genera assessed as consistently  $C_3$  and 11 genera as  $C_4$ . The genera Abildgaardia, Cyperus, Eleocharis, Fimbristylis, and Rhynchospora are variable for this trait. Of the total number (1474) of specific (1406) and infraspecific (68) taxa sampled, 938 taxa (63%) are  $C_3$ , 527 taxa (36%) are  $C_4$ , and nine species of Eleocharis are debatably intermediate or variable in pathway. Some data suggesting further infrageneric variation in photosynthetic pathways are discussed. The "one cell distant criterion" accurately predicts  $C_4$  pathway in sedges, except in Eleocharis. Distribution and variability of photosynthetic pathways in Eleocharis are discussed. Photosynthetic pathway was found to be a useful taxonomic marker in Cyperaceae, despite variability in this trait at various taxonomic levels and the apparently multiple origin of  $C_4$  photosynthesis within the family. A checklist of 3395 records of  $C_3$  and  $C_4$  sedges is presented.

Key words:  $C_3$ ,  $C_4$ ,  $C_3$ - $C_4$  intermediate, Cyperaceae,  $δ^{13}$ C values, *Eleocharis*, Γ, photosynthetic pathway, sedge.

#### INTRODUCTION

Two distinct patterns of vegetative anatomy in sedges have long been recognized (Haberlandt 1884: 281). One, with "radiate" chlorenchyma and a green (chlorenchymatous) sheath situated within the vascular bundles (Kranz anatomy), "is seen in certain species of Cyperus" (Haberlandt 1884: 284). The other, exemplified by Carex and many other Cyperus species, involves non-radiate chlorenchyma and vascular bundles enclosed by a "sheath of large colorless cells." Botanists were quick to incorporate these discontinuities into the taxonomic framework of Cyperaceae, e.g., "Chlorocyperaceen" and "Eucyperaceen" of Rikli (1895: 560), see also Clarke (1908). More recent authors (e.g., Druyts-Voets 1970; Metcalfe 1971), in extending these anatomical studies and recognizing further variants (e.g., Sharma and Mehra 1972; Carolin et al. 1977; Gilliland and Gordon-Gray 1978; Bruhl et al. 1987; Bruhl 1995), have extended the taxonomic utility of vegetative anatomy in this

Subsequent to the discovery of the  $C_4$  photosynthetic pathway, the correlations between chlorocyperoid (Kranz) anatomy and  $C_4$  photosynthesis, and eucyperoid anatomy and  $C_3$  (Calvin cycle) photosynthesis became apparent. Further correlations were detected within Cyperaceae, as with other families, of photosynthetic pathways with characteristic carbon isotope ratio ( $\delta^{13}$ C value) ranges (Bender 1971), with  $CO_2$  compensation point ( $\Gamma$ ) values (Krenzer et al. 1975), and with geographical distributions, there being a concentration of  $C_4$  sedge species and genera in the tropics and  $C_3$  taxa in the temperate regions (Raynal 1972). Biochemistry has also been used to assess pathways (Bruhl et al. 1987; Sage et al. 1999), by measuring the ratio of phosphoenol-

pyruvate carboxylase (PEPCase) to ribulose-1,5-bisphosphate carboxylase (Rubisco) activity and by measuring the initial products of photosynthesis. Use of these correlates, particularly anatomical and  $\delta^{13}$ C values, has allowed extensive prediction of C<sub>3</sub> and C<sub>4</sub> photosynthetic pathways in sedges (Bruhl et al. 1992; Bruhl 1993, 1995), and has led to a reassessment, along structural/functional and evolutionary lines (Soros and Bruhl 2000), of earlier taxonomic decisions based on purely anatomical discontinuities. Thus Raynal (1973) positioned his predominantly C<sub>4</sub> Cypereae and Fimbristylideae as terminal assemblages in a scheme of phylogenetic relationships of the Cyperoideae, with the C<sub>4</sub> genera uppermost, indicating their assumed derived states. Molecular studies by Muasya et al. (1998, 2002) found C3 taxa sister to C<sub>4</sub> taxa, indicating that rbcL, a photosynthetic gene, reflects phylogeny.

There is need for some caution when using anatomical observations or  $\delta^{13}C$  values alone as predictors of photosynthetic pathway, since  $C_3$ – $C_4$  intermediates may be overlooked (Bruhl et al. 1987; Hattersley 1987; Bruhl and Perry 1995; Sage 2002). The "maximum cells distant count" (Hattersley and Watson 1975) has proved to be a very reliable anatomical criterion in relation to grasses (Hattersley 1987). This explicit anatomical criterion for  $C_3$ / $C_4$  assignment, though seemingly applicable to sedges (Hattersley et al. 1977), had not previously been tested on them on a large scale. Instead, the relatively vague concepts of "radiate" chlorenchyma and "Kranz" anatomy (e.g., Ueno and Koyama 1987) have continued in use (Bruhl et al. 1987).

Cyperaceae have been covered in a number of surveys of photosynthetic pathway variation (e.g., Black 1976; Raghavendra and Das 1976; Takeda et al. 1985; Wang 2003), and

various taxonomic conclusions regarding the family have been drawn (Lerman and Raynal 1972; Raynal 1973; Takeda et al. 1985). Major contributions to  $C_3/C_4$  assessments, in terms of the numbers of species and genera sampled, have been made by Lerman and Raynal (1972, see below) and Takeda et al. (1985). No genus remains unknown in this respect, though there is conflicting information about the photosynthetic pathways of some.

We have obtained additional anatomical,  $\delta^{13}$ C and  $\Gamma$  data to examine critically the correlation between physiological, biochemical, and anatomical data pertaining to photosynthetic pathways, and to fill significant gaps in the taxonomic coverage, to locate new variation within genera, and to identify taxa where C<sub>3</sub>-C<sub>4</sub> intermediates may occur. In what follows, those new observations are presented and current knowledge of C<sub>3</sub>/C<sub>4</sub> photosynthetic pathway variation in Cyperaceae is summarized. Appendix 1 contains the first extensive, up-to-date compilation of photosynthetic pathway determinations for Cyperaceae, with sources, and presents, also for the first time, the valuable, original data underlying the publications by Lerman and Raynal (1972), Raynal (1972, 1973), and Stock et al. (2004). All new, previously unpublished data and previously published data are compiled in Appendix 1. The compilation is believed to be comprehensive with respect to the physiological and biochemical data.

#### MATERIALS AND METHODS

#### Plant Material

For study of living material, plants collected from across Australia were grown under half-shade in glasshouses maintained between 35°C (day maximum) and 15°C (night minimum), and regularly fertilized with Ruakura nutrient solution (Smith et al. 1983). Identities were checked and vouchers have been lodged at CANB or NE (by JJB) or NSW (by KLW). Where samples were taken from herbarium material, voucher labels were attached to the sheets.

#### Anatomy and "One Cell Distant Criterion"

Plant material was selected and prepared as stated in Bruhl et al. (1987), Wilson (1991), and Bruhl and Perry (1995). Hand-cut sections of rehydrated herbarium material or fresh material, temporarily mounted, are generally adequate for ascertaining the "maximum cells distant count" and applying the "one cell distant criterion." The latter states that "in C<sub>4</sub> species no chlorenchymatous mesophyll cell is separated from the nearest parenchymatous bundle sheath (PBS) cell by more than one other chlorenchymatous mesophyll cell" (Hattersley and Watson 1975: 325). In Cyperaceae, application of this criterion involves counting the numbers of primary carbon assimilation (PCA) cells distant from the photosynthetic carbon reduction (PCR) cells, ignoring non-PCR mestome sheath cells, PBS cells and any non-chlorenchymatous cells.

#### CO2 Compensation Point Analyses

A pulse flow system for CO<sub>2</sub> compensation point analysis (Brown et al. 1985; Brown and Hattersley 1989) was used. Fresh healthy leaves or culms (photosynthetic material only)

of glasshouse-grown plants were placed in a 50 mL clear glass syringe. The syringe was fitted with a needle and the needle tip sealed with a rubber plug. The plunger was made airtight by lubrication with liquid paraffin. The sealed loaded syringe was placed in a growth cabinet under a photosynthetic photon flux density of 500 µmol quanta m<sup>-2</sup>s<sup>-1</sup> at 30°C, and was incubated for at least 20 minutes. A 30 mL gas sample from the syringe was then passed through a calcium chloride H<sub>2</sub>O trap, to an infrared CO<sub>2</sub> gas analyzer (model ZAR, Fuji Electric, Tokyo, Japan). High grade nitrogen, at a flow rate of 4 liters min<sup>-1</sup>, was used as a carrier gas. The output from the analyzer was recorded on an RDK Rikadenki chart recorder (RDK, Tokyo, Japan). The system was calibrated with 1, 2, and 3 mL samples of pure CO<sub>2</sub> (delivered with an SGE microlitre syringe) equivalent to 33.3, 66.6, and 100 parts per million (ppm) of CO<sub>2</sub> in 30 mL volumes, respectively. The CO<sub>2</sub> concentration of the sample gas was calculated from the peak height of the CO2 pulse. Controls were used to ensure that the sole source of CO<sub>2</sub> was that derived from the sample, and constituted the delivery of 30 mL of CO<sub>2</sub>-free air, which resulted in no pen movement beyond the base line. The  $\Gamma$  values of JJB presented in Appendix 1 represent means based on four replicates, except for the controls, where there were two replicates.

#### $\delta^{13}C$ Values

- (a) This study (see Bruhl 1990).—For  $\delta^{13}$ C value determinations by JJB, mature healthy leaf or culm samples from cultivated plants oven-dried at 70°C, or from herbarium specimens, were ground finely in liquid nitrogen with a mortar and pestle, or chopped finely with a razor. Samples of 0.2 to 3 mg were combusted using a modification of the classical Dumar method in a Carlo Erba 1106 Elemental Analyzer (CE Instruments, Milan, Italy). The CO<sub>2</sub> produced was trapped automatically at liquid nitrogen temperature, then distilled from the cold finger and passed to a VG Isogas Sira-24 mass spectrometer (Thermo Electron Corporation, Boston, Massachusetts, USA) for analysis. Standards used were the laboratory internal CO<sub>2</sub> standard gas and a standardized sucrose calibrated against international carbonate standards. The  $^{13}$ C/ $^{12}$ C ratios are reported as  $\delta^{13}$ C values in  $^{96}$ C.
- (b) Source of values used in Lerman and Raynal (1972).—Before his untimely death, Jean Raynal gave K. L. Wilson (NSW) a copy of the data sheets for the  $\delta^{13}$ C value determinations associated with their landmark papers (Lerman and Raynal 1972; Raynal 1972, 1973). After receiving a copy and seeking permission from Dr. Lerman, J. J. Bruhl incorporated these data in a survey of photosynthetic pathways (Bruhl 1990). Lerman and Raynal's data are presented here, in full, in a refereed publication for the first time.
- (c) Source of values used in Stock et al. (2004).—While the present paper was in review, a paper by Stock et al. (2004) appeared in Austral Ecology with data in summary form only. Professor Will Stock and Dr. Tony Verboom kindly agreed to make their data available to us, and we present them here, in full, for the first time.

#### Evaluation of Literature; Nomenclature

Photosynthetic pathway determinations were collated from original publications, rather than from reviews, and anatomical data from publications preceding the discovery of C<sub>4</sub> photosynthesis have been used only where they permit unambiguous interpretation. Description of leaf or stem chlorenchyma as radiate is not a reliable criterion for assigning photosynthetic pathway (Bruhl et al. 1987) and was ignored. For example, consider "chlorenchyma slightly radiating" (Govindarajalu 1969a: 28) for species of *Fuirena* that are C<sub>3</sub> and "chlorenchyma not radiating" (Govindarajalu 1974: 245–246) for *Cyperus clarkei* T. Cook, which is C<sub>4</sub>. For methods used to assess photosynthetic pathways see the original papers.

Nomenclature and generic and subgeneric circumscriptions used here are informed by the relatively recent classifications and studies of Cyperaceae (especially Bruhl 1995; Goetghebeur 1998; Muasya et al. 2000) and the arrangement of Cyperaceae at the National Herbarium of New South Wales (NSW) by KLW, as well as the draft World Checklist of Cyperaceae being coordinated by D. Simpson and R. Govaerts from Royal Botanic Gardens, Kew, with contributions from other cyperologists such as the present authors.

#### RESULTS AND DISCUSSION

The draft World Checklist of Cyperaceae (see above) lists 103 genera and about 5400 species in Cyperaceae. For the current study we recognize 107 genera of which all genera and 1406 species and an additional 68 subspecies and varieties have been investigated for photosynthetic pathway (Appendix 1) anatomically (2350 records), or physiologically and/or biochemically (1045 records), totaling 3395 records (Appendix 1). The compilation includes new  $\Gamma$  values for seven genera and 29 species, new δ<sup>13</sup>C value determinations for 15 genera and 50 species and 1047 new anatomical records obtained in this study, along with 1305 anatomical records based on our assessment of published literature. In addition, we present the 246 original δ<sup>13</sup>C value determinations summarized by, but not presented in, Lerman and Raynal (1972), as well as the 68 original  $\delta^{13}$ C value records of Stock et al. (2004).

The determination of photosynthetic pathways, particularly at the generic level, is comprehensive for this reasonably large family, and affords a sound basis from which to generalize about the likelihood of finding further variation, predict the photosynthetic pathway of the unassessed taxa, and discuss taxonomic implications of the available data. All genera that we recognize here can be assigned to a photosynthetic pathway (Table 1) with confidence on the basis of biochemical, physiological, and anatomical evidence (Appendix 1). The present state of knowledge shows that Abildgaardia, Cyperus s.l., Eleocharis, Fimbristylis, and Rhynchospora s.l. are variable, comprising both C<sub>3</sub> and C<sub>4</sub> species, while the remaining genera are consistently either C<sub>3</sub> (91 genera) or C<sub>4</sub> (11 genera) (Table 1; Appendix 1) (see also Bruhl et al. 1987; Bruhl and Perry 1995; Soros and Bruhl 2000). Of the total number (1474) of specific (1406) and infraspecific (68) taxa sampled, 938 taxa (63%) are C<sub>3</sub>, 527 taxa (36%) are C<sub>4</sub>, and nine species of *Eleocharis* are debatably intermediate or variable in pathway.

Sage et al. (1999), based on data and generalizations from Bruhl et al. (1992) and Bruhl (1995), gave the percentage of genera and species of Cyperaceae that are C<sub>4</sub> as 21% and 27%, respectively. Here, not recognizing as many segregate genera, and including only the species actually examined (Appendix 1), we find that 15% of genera (including the five genera that are variable for photosynthetic pathway) and 34% of the species sampled are C<sub>4</sub>. Given the different bases of the calculations, their similarity provides some confidence in them. The differences in numbers, however, indicate the need to continue to improve the estimate of phylogeny for the family and hence a better basis for its classification, and the need to complete the survey of photosynthetic pathways of species of Cyperaceae.

The anatomical sample is rather patchy at the species level, in that most of the smaller genera, along with some large ones (e.g., *Cyperus, Eleocharis*, and *Rhynchospora*) have been extensively sampled, while other large genera such as *Lagenocarpus* and *Pleurostachys* have been examined for only one or two species. Nonetheless, the sample size compares favorably with those used for many other micromorphological or anatomical features. For example, recent taxonomic treatments (Raynal 1973; Goetghebeur 1986; Bruhl 1995; Goetghebeur 1998) of Cyperaceae placed a great deal of reliance on embryo morphology, where the available data are much less comprehensive.

#### "One Cell Distant Criterion"

Cross-referencing between the different kinds of evidence presented in Appendix 1 shows excellent correspondence between the different methods of assessing photosynthetic pathways in this family. It also shows that, perhaps with the exception of some species of *Eleocharis* (see below), all the photosynthetic pathways are correctly predicted using anatomical criteria. The few conflicting records are considered below. The data tabulated in Appendix 1 allow evaluation of the "maximum cells distant count" as a predictor of photosynthetic pathway in Cyperaceae. Of the 105 genera investigated anatomically for this criterion by us (i.e., except for Pleurostachys and Trichoschoenus), 84 genera and many of the species sampled have also been analyzed for their  $\delta^{13}$ C values, and 14 genera and 91 species for  $\Gamma$ . The photosynthetic pathway of 14 of the genera including 63 species has been determined biochemically. Congruence of the data shows, with the exception of *Eleocharis*, that the "one cell distant criterion" (Hattersley and Watson 1975) is an accurate predictor of C<sub>4</sub> in Cyperaceae, while counts of greater than one cell accurately predict C<sub>3</sub>. Given the simplicity and ease with which anatomical preparations can be made to determine photosynthetic pathway type, it is reasonable to suggest that such evidence and determinations should accompany the descriptions of new species and genera as a matter of routine.

In *Eleocharis*, the unequivocally  $C_4$  taxa (Appendix 1; Bruhl et al. 1987) yield counts of one to four cells, i.e., often exceeding a count of one, even if the chlorophyllous layer of cells adjacent to the mestome sheath is considered to constitute a PBS and ignored (Bruhl et al. 1987). However, the chloroplast abundance in the PCA ( $C_4$  mesophyll) cells is relatively low and the more distant cells are equivocally

Table 1. Genera and generic sample for distribution of photosynthetic pathway in Cyperaceae. (^ = World Checklist of Cyperaceae; www.kew.org/wcsp/home.do) (Simpson et al. 2005); incl. = including; PP = photosynthetic pathway; \* = includes two additional species published since first version of Checklist was distributed to cyperologists in 2004).

Genera sampled (unless indicated otherwise)	PP	World Checklist <sup>^</sup>	No. of species^
Abildgaardia Vahl	C <sub>3</sub> /C <sub>4</sub>	(= Fimbristylis)	
Actinoschoenus Benth.	$C_3$	Actinoschoenus	3
Actinoscirpus (Ohwi) R. Haines & K. Lye	$C_3$	Actinoscirpus	1
Afrotrilepis (Gilg) J. Raynal	$C_3$	Afrotrilepis	2
Alinula J. Raynal	$C_4$	Alinula	4
Amphiscirpus Oteng-Yeboah	$\mathbf{C}_3$	Amphiscirpus	1
Androtrichum (Brongn.) Brongn.	$\mathbb{C}_3$	Androtrichum	2
Arthrostylis R. Br.	$C_3$	Arthrostylis	1
Ascolepis Nees ex Steud.	$\mathbb{C}_4$	Ascolepis	22
Asterochaete Nees	$C_3$	(= Carpha)	
Baumea Gaudich.	$C_3$	(= Machaerina)	
Becquerelia Brongn.	$\mathbf{C}_3$	Becquerelia	8
Bisboeckelera Kuntze	$C_3$	Bisboeckelera	4
Blysmus Panz. ex Schult.	$C_3$	Blysmus	3
Bolboschoenus (Aschers.) Palla	$\mathbb{C}_3$	Bolboschoenus	10
Bulbostylis Kunth	$C_4$	Bulbostylis	206
Calyptrocarya Nees	$C_3$	Calyptrocarya	8
Capeobolus J. Browning	$\mathbb{C}_3$	Capeobolus	1
Capitularina Kern	$\mathbb{C}_3$	Capitularina	1
Carex L. (incl. Cymophyllus Mack. ex Britton & A. Br. and Vesicarex Steyerm.)	$C_3$	Carex	1757
Carpha Banks & Sol. ex R. Br.	$\mathbb{C}_3$	Carpha	16
Caustis R. Br.	$\mathbb{C}_3$	Caustis	5
Cephalocarpus Nees	$C_3$	Cephalocarpus	4
(= Eleocharis)		Chillania Roiv.	1
Chorizandra R. Br.	$\mathbb{C}_3$	Chorizandra	5
Chrysitrix L.	$C_3$	Chrysitrix	4
Cladium P. Browne	$C_3$	Cladium	3
Coleochloa Gilly	$\mathbb{C}_3$	Coleochloa	7
Costularia C. B. Clarke (incl. Lophoschoenus Stapf)	$C_3$	Costularia	24
Courtoisina Soják	$C_3$	Courtoisina	2
Crosslandia W. V. Fitzg.	$\mathbb{C}_4$	Crosslandia	1
Cyathochaeta Nees	$C_3$	Cyathochaeta	6
Cyathocoma Nees	$C_3$	(= Tetraria)	
Cyperus L. (incl. Anosporum Nees, Juncellus (Griseb.) C. B. Clarke, Mariscus Vahl, Remirea Aubl., Torulinium Desv.)	C <sub>3</sub> /C <sub>4</sub>	Cyperus	686
(= Carex)	$C_3$	Cymophyllus	1
Cypringlea M. Strong	$C_3$	Cypringlea	2
Desmoschoenus Hook. f.	$C_3$	Desmoschoenus Didymiandrum	1
Didymiandrum Gilly	$C_3$	•	1
Diplacrum R. Br. Diplasia Rich. ex Pers.	$C_3$	Diplacrum Diplacia	9 1
Dulichium Pers.	$C_3$	Diplasia Duli alaina	
Egleria L. T. Eiten	$C_3$	Dulichium	1
Eleocharis R. Br.	$C_3$	Egleria Eleocharis	1 252
Epischoenus C. B. Clarke	$C_3/C_4$	Epischoenus	8
*	$C_3$	*	
Eriophorum L. (incl. Eriophoropsis Palla & Erioscirpus Palla)  Evandra R. Br.	$C_3$	Eriophorum Evandra	18 2
Evanara R. Bl. Everardia Ridl. ex Oliver	$C_3$	Evanara Everardia	12
	$C_3$	Exocarya	
Exochogyne C. B. Clarke	$C_3$	•	1 3
Ficinia Schrad. (incl. Sickmannia Nees)	$C_3$	Exochogyne	72
	$C_3$	Ficinia	
Finderistylis Vahl (incl. Tylocarya Nelmes)	$C_3/C_4$	Fimbristylis	306
Fuirena Rottb.	$C_3$	Fuirena Gahnia	58 41
Gahnia J. R. Forst. & G. Forst.	$C_3$		41
Gymnoschoenus Nees	$C_3$	Gymnoschoenus	2
Hellmuthia Steud.	$C_3$	Hellmuthia	1
Hypolytrum Rich.	$C_3$	Hypolytrum	57
Isolepis R. Br. (incl. Eleogiton Link)	$C_3$	Isolepis	74
Karinia A. Reznicek & R. McVaugh	$C_3$	Karinia	1
Khaosokia D. A. Simpson, Chayam. & J. Parn.	$C_3$	(Not in first version of World Checklist)	1

Table 1. Continued.

Genera sampled (unless indicated otherwise)	PP	World Checklist <sup>^</sup>	No. of species^
Kobresia Willd.	$C_3$	Kobresia	71
Koyamaea W. W. Thomas & G. Davidse	$C_3$	Koyamaea	1
Kyllinga Rottb. (∼ Cyperus)	$C_4$	Kyllinga	73
Kyllingiella R. Haines & K. Lye	$\mathbf{C}_3$	Kyllingiella	3
Lagenocarpus Nees	$C_3$	Lagenocarpus	61
Lepidosperma Labill.	$C_3$	Lepidosperma	56
Lepironia Rich.	$C_3$	Lepironia	1
Lipocarpha R. Br.	$C_4$	Lipocarpha	36
Machaerina Vahl	$\mathbf{C}_3$	Machaerina	51
Mapania Aubl. (incl. Thoracostachyum Kurz)	$C_3$	Mapania	84
Mesomelaena Nees	$C_3$	Mesomelaena	5
Microdracoides Hua	$C_3$	Microdracoides	1
Morelotia Gaudich.	$C_3$	Morelotia	2
Neesenbeckia Levyns	C <sub>3</sub>	Neesenbeckia	1
Nelmesia Van der Veken	$C_4$	Nelmesia	1
Nemum Desv.	$C_4$	Nemum	5
Oreobolopsis T. Koyama & E. R. Guaglianone	$C_3$	Oreobolopsis	3
Oreobolus R. Br. (incl. Chillania Roiv.)	$C_3$	Oreobolus	17
Oxycaryum Nees	$C_3$ $C_3$	Oxycaryum	1
Paramapania Uittien		• •	7
	$C_3$	Paramapania Plantia a simus	3*
Phylloscirpus C. B. Clarke	$C_3$	Phylloscirpus	50
Pleurostachys Brongn.	$C_3$	Pleurostachys	
Principina Uittien	$C_3$	Principina	1
Pseudoschoenus (C. B. Clarke) Oteng-Yeboah	C <sub>3</sub>	Pseudoschoenus	1
Ptilothrix K. L. Wilson	$C_3$	Ptilothrix	1
Pycreus Beauv. (~ Cyperus)	$C_4$	Pycreus	118
Queenslandiella Domin (~ Cyperus)	$C_4$	Queenslandiella	1
Reedia F. Muell.	$C_3$	Reedia	1
(= Cyperus)		Remirea Aubl.	1
Rhynchocladium T. Koyama	$C_3$	Rhynchocladium	1
Rhynchospora Vahl	$C_3/C_4$	Rhynchospora	341
Schoenoplectus (Rchb.) Palla	$C_3$	Schoenoplectus	64
Schoenoxiphium Nees	$C_3$	Schoenoxiphium	20
Schoenus L.	$C_3$	Schoenus	108
Scirpodendron Zipp. ex Kurz	$C_3$	Scirpodendron	2
Scirpoides Ség.	$C_3$	Scirpoides	3
Scirpus L.	$C_3$	Scirpus	67
Scleria Berg.	$C_3$	Scleria	264
Sphaerocyperus K. Lye	$\mathrm{C}_4$	Sphaerocyperus	1
Sumatroscirpus Oteng-Yeboah	$C_3$	Sumatroscirpus	1
Tetraria Beauv.	C <sub>3</sub>	Tetraria	57
Trachystylis S. T. Blake	$C_3$	Trachystylis	1
Trianoptiles Fenzl	$C_3$	Trianoptiles	3
Trichophorum Pers.	$C_3$	Trichophorum	9
Trichoschoenus J. Raynal	C <sub>3</sub>	Trichoschoenus	1
Tricostularia Nees	$C_3$	Tricostularia	5
Trilepis Nees	$C_3$	Trilepis	7
Uncinia Pers.	$C_3$	Uncinia	66
Volkiella Merxm. & Czech.	$C_3$	Volkiella	1
Websteria S. H. Wright	$C_3$	Websteria	1
Zameioscirpus Dhooge & Goetgh.	$C_3$	(Not in first version of World Checklist)	1
107 genera sampled		104 genera	5401

chlorophyllous. Even where the criterion can be applied with confidence, the PBS may be chlorophyllous or more or less non-chlorophyllous (e.g., in *Fimbristylis*), with variation apparent within and between species. The stoichiometric and physiological significance of such variation is not clear, and warrants further investigation.

 $C_4$  sedges are generally NADP-ME, whereas the  $C_4$  species of *Eleocharis* are NAD-ME type—the only occurrence of this biochemical type in Cyperaceae. So variation in biochemical type coincides with breakdown of the  $C_4$  anatomical predictor (Ueno et al. 1986; Bruhl et al. 1987). The breakdown does not seriously impair use of the "one cell

distant criterion" in predicting photosynthetic pathway, because of the apparent rarity of the NAD-ME type in this family. Although  $C_4$  *Eleocharis* species would be incorrectly assigned to photosynthetic pathway using this criterion, they are accurately predicted by ultrastructural features of the mitochondria and chloroplast grana in PCR cells (Bruhl and Perry 1995). Exceptions to the "one cell distant criterion" highlight the advantage of using at least two means to assess photosynthetic pathway in Cyperaceae. Anatomy,  $\delta^{13}C$  value, and ultrastructure have the advantage that they can be assessed from fresh or herbarium samples.

#### Conflicting Reports

There are conflicting reports about the photosynthetic pathway status of some species (marked with "+" in Appendix 1). We suggest that most, if not all, of these conflicts result from misidentification of the material studied (see Appendix 2 for discussion). Some vouchers have been reexamined and redetermined by the current authors. These redeterminations together with our assessment of the conflicts are indicated in Appendix 1.

One case is presented here as it highlights the need for publication of data (= results) and for vouchers to be lodged in recognized herbaria to underpin discussion and conclusions in scientific papers. In their recent publication, Stock et al. (2004) submitted δ13C data linked to vouchers deposited in a recognized herbarium but were not allowed by the journal to include them in the paper. The summary  $\delta^{13}$ C data (Stock et al. 2004: Table 1) contained two surprises. One species of Lipocarpha is listed as C3 and one of Schoenoplectus is listed as C<sub>4</sub>. The specific data obtained from the authors reveal these species to be L. rehmannii and S. pulchellus (Appendix 1). Their datum for L. rehmannii is at odds with five other records for the species based on anatomical observations and  $\delta^{13}$ C values. All the pieces of this specimen appear to match other material of L. rehmannii at BOL (A. Verboom pers. comm., Aug 2004). We hope to examine this specimen anatomically. The C<sub>4</sub> δ<sup>13</sup>C value for S. pulchellus is the first report for this species. We do not yet have access to material of this species to check it. The specimen in question is not mixed and appears to have been correctly identified by cyperologist Jane Browning (A. Verboom pers. comm., Aug 2004). There are 52 records of C<sub>3</sub> pathway for the genus (Appendix 1), plus two apparently anomalous C<sub>4</sub> reports for S. lateriflorus (Hofstra et al. 1972). We plan to survey all the species in this genus for photosynthetic pathway and clarify the photosynthetic pathway status of S. pulchellus. To minimize the chance of erroneous reports, to support novel findings, and to allow the authentication of names used in botanical studies it is crucial that journals and book editors insist on the publication at the same time of supporting data linked to voucher specimens lodged in a recognized herbarium (see also Goldblatt et al. 1992; Hosking et al. 1996).

## Significance of Photosynthetic Pathways for Solving Taxonomic Problems

Two contrasting and independent examples indicate the value of photosynthetic pathway data in posing or solving taxonomic problems. Firstly, the monotypic *Syntrinema* H.

Pfeiff. is variously regarded, largely on the basis of floral morphology, as belonging to *Rhynchospora* (Ballard 1934; Goetghebeur 1986; Wayt Thomas pers. comm.) or as a genus belonging to a separate tribe (Eiten 1976). It is C<sub>4</sub> (Appendix 1), and its vegetative anatomy (Bruhl et al. 1987; Ueno and Koyama 1987) is typical of the C<sub>4</sub> *Rhynchospora* species with rhynchosporoid anatomy (Bruhl et al. 1987). Rhynchosporoid anatomy is found only in these two genera and, therefore, strongly supports the former taxonomic affiliation. We have included *Syntrinema* here under *Rhynchospora*.

Secondly, Abildgaardia and Fimbristylis, two closely related and often synonymized genera (Bruhl et al. 1992; Bruhl 1995; Ghamkhar et al. 2007), have previously been considered to be exclusively C<sub>4</sub> (Appendix 1). Indeed, Raynal (1973) and Goetghebeur (1986) place these genera in a tribe in part characterized by C<sub>4</sub> photosynthesis and fimbristyloid anatomy (see Bruhl et al. 1987). Our anatomical observations and  $\delta^{13}$ C value data (Appendix 1), however, indicate that Abildgaardia hygrophila and Fimbristylis variegata are C<sub>3</sub>. Furthermore, Gordon-Gray's (1971: 562) observations for F. variegata ("even the smaller bundles of the outer ring lie not within the mesophyll but merely in contact with its inner margin . . . The mesophyll in this species is especially well organized, the cells being palisadelike"), considered in retrospect, also hint at C<sub>3</sub> anatomy and Metcalfe's (1971: 276) description of the chlorenchyma in A. hygrophila (treated under Fimbristylis; "up to 6 layers of palisade cells") clearly indicates C<sub>3</sub> anatomy.

Photosynthetic pathway and vegetative anatomy are valuable in substantiating the inclusion of *Syntrinema* in *Rhynchospora*. In most cases, photosynthetic pathway is a valuable taxonomic criterion (as seen by its consistency within most genera), but the *Abildgaardia* example illustrates the need for caution when generalizing from small samples of species.

#### Eleocharis

Rikli's (1895) "Chlorocyperaceen" genera have generally been found to be C<sub>4</sub> (Lerman and Raynal 1972), e.g., Ascolepis, Cyperus subgen. Cyperus, Fimbristylis, Kyllinga, Lipocarpha, and Monandrus ined. Rikli (1895) suggested division of the essentially helophytic genus Eleocharis (as Heleocharis) into two genera: Eleocharis with "eucyperoid" anatomy, and Chlorocharis with "chlorocyperoid" anatomy (i.e., with an inner chlorophyllous parenchyma sheath, or boundary layer cells). This seemed to be misleading in the context of photosynthetic pathways, in that subsequent literature on Eleocharis anatomy and photosynthetic pathway indicated a solidly C<sub>3</sub> genus, i.e., including some of his species of Chlorocharis.

More recently it has been shown that at least some species of *Eleocharis* (including one "*Chlorocharis*" species) are  $C_4$  (Appendix 1) (Bruhl et al. 1987; Ueno et al. 1988b; see also Bruhl and Perry 1995). Of the eight species included in Rikli's (1895) *Chlorocharis*, he listed *C. palustris* (L.) Rikli (= *E. palustris*), *C. tuberculosa* (Michx.) Rikli (= *E. tuberculosa*) and *C. vivipara* (Link) Rikli (= *E. vivipara*) as having an "inner and outer parenchymatous sheath". Terrestrial forms of *Eleocharis vivipara* have recently been found to be  $C_4$  (and NAD-ME) (Bruhl et al. 1987; Ueno et al. 1988b, etc.), but  $\delta^{13}$ C values for *E. palustris*, and *E. tuberculosa* are

typical of C<sub>3</sub> species (Appendix 1). Rikli (1895) listed five other species—C. balansaeana (Boeck.) Rikli (= E. filiculmis), C. emarginata (Nees) Rikli (= E. quinquangularis), C. capitata (L.) Rikli (= E. geniculata), C. geniculata (L.) Rikli (= E. geniculata), and C. subprolifera Rikli (= E. pellucida)—as having only an "inner parenchymatous sheath" (i.e., possessing prominent chlorophyllous border parenchyma), implying that a typical PBS is absent. However, one of these species, Eleocharis geniculata, has been examined in the present study, and it possesses an obvious non-chlorenchymatous C3-like PBS outside the mestome sheath (Bruhl et al. 1987); while  $\delta^{13}$ C and  $\Gamma$  values (Appendix 1) and biochemical assays (Bruhl et al. 1987) all confirm its C<sub>3</sub> status. This is despite the border parenchyma cells being somewhat more prominent and chlorophyllous than in most other C<sub>3</sub> species (cf. Bruhl and Perry 1995).

The essentially terrestrial species *E. filiculmis*, *E. pellucida*, and *E. quinquangularis* were studied by Ueno et al. (1989). They are members of the series *Sulcatae* and *Multicaules* (with spirally disposed bracts; Svenson 1939), and are not closely related to the C<sub>4</sub> species of *Eleocharis* that constitute part of the series *Tenuissimae* possessing distichous floral bracts (Svenson 1937). These three species, therefore, were predicted to be C<sub>3</sub> (Bruhl 1990). Ueno et al. (1989) found the first two species to be C<sub>3</sub>, while *E. quinquangularis* was reported to be "C<sub>3</sub>–C<sub>4</sub>?". Of Rikli's *Chlorocharis*, only *E. tuberculosa* and *E. vivipara* were assigned to series *Tenuissimae* by Svenson (1937); the former is also C<sub>3</sub> (Ueno et al. 1989), while the dimorphic *E. vivipara* can be C<sub>4</sub> (Ueno et al. 1988a) (Appendix 1).

Ueno et al. (1988b) provided convincing evidence in the form of  $\delta^{13}$ C values, pulse-chase experiments and  $C_4$  acid decarboxylation enzyme assays that the terrestrial form of *E. vivipara* is  $C_4$  and the submerged aquatic form is  $C_3$ . Subsequent studies by Ueno and colleagues (e.g., Ueno 2001; Ueno and Ishimaru 2002) have focused on variation between and within individuals. *Eleocharis baldwinii* displays much of the intraspecific variation in photosynthetic pathway seen in *E. vivipara* (Ueno et al. 1989; Uchino et al. 1995; Ueno and Ishimaru 2002; Ueno 2004) (Appendix 1).

The variability correlates with the breakdown in the "one cell distant criterion" amongst the previously known C<sub>4</sub> sedges (see above; Bruhl et al. 1987), suggesting that these variabilities may have a common basis. Both the C<sub>4</sub> forms of *E. vivipara* and *E. baldwinii*, and the apparently consistently C<sub>4</sub> species of *Eleocharis* are NAD-ME type (Bruhl et al. 1987; Ueno and Samejima 1989; Bruhl and Perry 1995; Ueno 2004) and are members of ser. *Tenuissimae* (Bruhl and Smith 2002).

Eleocharis is home to further photosynthetic pathway variations that highlight its importance in understanding photosynthetic pathway evolution and development. On the basis of intermediate anatomy (Bruhl and Perry 1995), low or undetectable  $C_4$  enzyme values (Bruhl et al. 1987),  $C_3$   $\delta^{13}C$  values and intermediate G values (Appendix 1), supported by some ultrastructural evidence (Bruhl and Perry 1995), *E. pusilla* is interpretable as a  $C_3$ -like  $C_3$ - $C_4$  intermediate. Similarly, on the basis of intermediate anatomy and mostly  $C_3$ -like  $\delta^{13}C$  values (Ueno et al. 1989; Guaglianone and Ueno 1990) *E. cylindrica, E. quinquangularis,* and *E. reverchonii* are  $C_3$ -like  $C_3$ - $C_4$  intermediates or variable and in need of further study.

The variability, especially infraspecific, of photosynthetic pathways in *Eleocharis* is interesting in the context of the mechanism of development of  $C_4$  photosynthesis (see Ueno 2001; Agarie et al. 2002; Ueno and Ishimaru 2002; Ueno and Kobayashi 2002; Ueno 2004). There is evidence that *E. baldwinii* responds to different  $CO_2$  environments in water by shifting the relative proportion of  $C_3$  and  $C_4$  photosynthesis;  $C_4$ -like intermediacy in this species is considered to be a response to  $CO_2$  depletion (Ueno 2004).

These findings further highlight *Eleocharis* in general and specifically series *Tenuissimae* (see Bruhl and Smith 2002) as a singularly appropriate group to study the evolution and expression of the C<sub>4</sub> syndrome. Studies should be extended to grow a wider sample of species of *Eleocharis* that exhibit a range of photosynthetic pathway characteristics (Appendix 1) and the related aquatic monotypics *Egleria* and *Websteria* (Bruhl 1995; Roalson and Friar 2000) under terrestrial conditions to test the stability of their C<sub>3</sub> status.

#### CONCLUSIONS

The taxonomic sample for photosynthetic pathways in Cyperaceae is particularly broad, covering all genera recognized here. Some large genera remain poorly sampled, notably *Carex* and *Scleria*, but these are considered likely not to be variable. On the other hand, more biochemical typing is necessary across the family, particularly with regard to C<sub>4</sub> anatomical variation in *Eleocharis* and *Rhynchospora*. Usefulness of the "one cell distant criterion" is confirmed for all groups of Cyperaceae except *Eleocharis*. The most promising and interesting area for discovery of further C<sub>4</sub> species or further infrageneric variation is the predominantly C<sub>3</sub> Scirpeae (sensu Bruhl 1995), within and around *Eleocharis*. Similarly, more study is needed of *Rhynchospora* s.l. and Abildgaardieae.

Information on photosynthetic pathway variation, especially with regard to anatomical aspects, has had a significant impact on taxonomy particularly at the species and generic levels (see Raynal 1973; Haines and Lye 1983).

The helophytic habit, typically associated with high light and high temperature and, by definition, availability of water, coupled with cold sensitivity, and tolerance of salinity and low nitrogen levels (Wilson 1991: 391) of many (perhaps most) C<sub>4</sub> sedges, particularly the C<sub>4</sub> species of *Eleocharis*, offers an attractive model to investigate the functional significance of C<sub>4</sub> photosynthesis in terms of nitrogen-use efficiency, as well as in terms of the traditional, but seemingly inappropriate, hypothesis which relates C<sub>4</sub> photosynthesis to water-use efficiency (see also Bruhl 1990). "CO2-use efficiency" seems important for C4 species when submerged (Ueno 2004). Future investigations of the mechanisms of C<sub>4</sub> photosynthesis regulation in sedges should also address questions of particular agronomic interest (e.g., control of the "world's most troublesome weeds" (Wills 1987), the C<sub>4</sub> Cyperus rotundus and C. esculentus), as well as addressing fundamental questions of differentiation and development.

This paper is part of our ongoing study of photosynthetic pathways in Cyperaceae. We intend to follow up with an update of phylogenetic aspects of photosynthetic pathway (cf. Soros and Bruhl 2000). Finally, given the significance and application of a knowledge of photosynthetic pathway, we will present and maintain the survey data for Cyperaceae

via the Internet (see www.une.edu.au/botany/jjbres.htm), and encourage those who make and publish observations to contribute these new records to the ongoing database, where of course they will be fully acknowledged.

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#### LITERATURE CITED

- AGARIE, S., M. KAI, H. TAKATSUJI, AND O. UENO. 2002. Environmental and hormonal regulation of gene expression of C<sub>4</sub> photosynthetic enzymes in the amphibious sedge *Eleocharis vivipara*. *Pl. Sci. (Elsevier)* **163**: 571–580.
- AKITA, S., A. MIYASAKA, AND Y. MURATA. 1969. Studies on the differences of photosynthesis among species. 1. Differences in the response of photosynthesis among species in normal oxygen concentration as influenced by some environmental factors. *Proc. Crop Sci. Soc. Japan* 38: 507–523.
- ALVES, M. V., M. E. M. ESTELITA, M. G. J. WANDERLEY, AND W. W. THOMAS. 2002. Aplicações taxonômicas da anatomia foliar das espécies brasileiras de *Hypolytrum* Rich. (Cyperaceae). *Revista Brasil. Bot.* 25: 1–9.
- AUCOUR, A., C. HILLAIRE-MARCEL, AND R. BONNEFILLE. 1994. Late Quaternary biomass changes from <sup>13</sup>C measurements in a highland peatbog from equatorial Africa (Burundi). *Quatern. Res.* 41: 225–233
- Ballard, F. 1934. Rhynchospora confusa Ballard. Hooker's Icon. Pl. 2: 3.
- BASKIN, J. M., AND C. C. BASKIN. 1981. Photosynthetic pathways indicated by leaf anatomy in fourteen summer annuals of Cedar Glades. *Photosynthetica* 15: 205–209.
- BENDER, M. M. 1971. Variations in the <sup>13</sup>C/<sup>12</sup>C ratios of plants in relation to the pathway of photosynthetic carbon dioxide fixation. *Phytochemistry* **10**: 1239–1244.
- BETTS, M. W. 1920. Notes on the autecology of certain plants of the Peridotite Belt, Nelson. Part I—Structure of some of the plants (No. 3). *Trans. New Zealand Inst.* **52**: 276–314.
- BLACK, C. C. 1976. Checklists of C<sub>4</sub>-dicarboxylic acid photosynthesis plant species, pp. 425–426. *In R. H. Burris and C. C. Black [eds.]*, CO<sub>2</sub> metabolism and plant productivity. University of Park Press, Baltimore, Maryland, U.S.A.
- —, W. H. CAMPBELL, T. M. CHEN, AND P. DITTRICH. 1973. The monocotyledons: their evolution and comparative biology. III. Pathways of carbon metabolism related to net carbon dioxide assimilation by monocotyledons. *Quart. Rev. Biol.* 48: 298–313.
- BLACK, C. C. J., AND H. H. MOLLENHAUER. 1971. Structure and distribution of chloroplasts and other organelles in leaves with various rates of photosynthesis. *Pl. Physiol. (Lancaster)* 47: 15–23.
- Borchers, E., W. Frey, and H. H. Hilger. 1982. C<sub>4</sub> species in the highlands of Iran and Afghanistan. *Notes Roy. Bot. Gard. Edinburgh* 40: 99–113.
- BOUTTON, T. W., A. TYRONE HARRISON, AND B. N. SMITH. 1980. Distribution of biomass of species differing in photosynthetic pathway along an altitudinal transect in Southeastern Wyoming grassland. *Oecologia* **45**: 287–298.
- Brown, R. H., J. H. Bouton, P. T. Evans, H. E. Malter, and L. L. RIGSBY. 1985. Photosynthesis, leaf anatomy, and cytogenetics of hybrids between C<sub>3</sub> and C<sub>3</sub>/C<sub>4</sub> *Panicum* species. *Pl. Physiol. (Lancaster)* 77: 653–658.
- —, AND P. W. HATTERSLEY. 1989. Leaf anatomy of C<sub>4</sub> species as related to evolution of C<sub>4</sub> photosynthesis. *Pl. Physiol. (Lan-caster)* 91: 1543–1550.
- Brown, W. V. 1975. Variations in anatomy, associations and origins of Kranz tissue. Amer. J. Bot. 62: 395–402.
- BRUHL, J. J. 1990. Taxonomic relationships and photosynthetic pathways in the Cyperaceae. Ph.D. thesis, Australian National University, Canberra, Australia. 196 pp.
- \_\_\_\_\_. 1993. Sedge genera of the world. Delta Newslett. 8: 19.

- . 1995. Sedge genera of the world: relationships and a new classification of the Cyperaceae. *Austral. Syst. Bot.* **8**: 125–305.
- —, AND S. PERRY. 1995. Photosynthetic pathway-related ultrastructure of C<sub>3</sub>, C<sub>4</sub> and C<sub>3</sub>-like C<sub>3</sub>-C<sub>4</sub> intermediate sedges (Cyperaceae), with special reference to *Eleocharis*. Austral. J. Pl. Physiol. 22: 1–10.
- , AND G. S. SMITH. 2002. *Eleocharis* R. Brown (subg. *Eleocharis* sect. *Eleocharis*) ser. *Tenuissimae* Svenson, pp. 90–99. *In* Flora of North America Editorial Committee [eds.], Flora of North America, Vol. 23. Oxford University Press, New York, USA.
- N. E. STONE, AND P. W. HATTERSLEY. 1987. C<sub>4</sub> acid decarboxylation enzymes and anatomy in sedges (Cyperaceae): first record of NAD-malic enzyme species. *Austral. J. Pl. Physiol.* 14: 719–728.
- , L. WATSON, AND M. J. DALLWITZ. 1992. Genera of Cyperaceae: interactive identification and information retrieval. *Taxon* 41: 225–234.
- CAROLIN, R. C., S. W. L. JACOBS, AND M. VESK. 1977. The ultrastructure of Kranz cells in the family Cyperaceae. *Bot. Gaz.* 138: 413–419
- CHEN, T. M., R. H. BROWN, AND C. C. BLACK. 1970. CO<sub>2</sub> compensation concentration, rate of photosynthesis, and carbonic anhydrase activity of plants. Weed Sci. 18: 399–403.
- ——, P. DITTRICH, W. H. CAMPBELL, AND C. C. BLACK. 1974. Metabolism of epidermal tissues, mesophyll cells, and bundle sheath strands resolved from mature nutsedge leaves. *Arch. Biochem. Biophys.* 163: 246–262.
- CHERMEZON, H. 1926. Sur la feuille de certaines espèces de *Scleria*. *Rev. Gén. Bot.* **38**: 337–353.
- . 1930. Structure de la feuille et de la tige chez Eriospora setifera. Arch. Bot. 4: 41–59.
- —. 1933. Observations sur le genre *Microdracoides. Bull. Soc. Bot. France* **80**: 90–97.
- CLARKE, C. B. 1908. New genera and species of Cyperaceae. Bull. Misc. Inform. Kew 8: 1–196.
- COLLINS, R. P., AND M. B. JONES. 1985. The influence of climatic factors on the distribution of C<sub>4</sub> species in Europe. *Vegetatio* **64**: 121–129
- Denton, M. F. 1983. Anatomical studies of the *Luzulae* group of *Cyperus* (Cyperaceae). *Syst. Bot.* 8: 250–262.
- DHOOGE, S., AND P. GOETGHEBEUR. 2002. A new Andean species and a new combination in *Oreobolopsis* (Cyperaceae). *Novon* **12**: 338–342
- genus of Cyperaceae from South America. *Pl. Syst. Evol.* **243**: 73–84.
- Downton, W. J. S., and E. B. Tregunna. 1968. Carbon dioxide compensation—its relation to photosynthetic carboxylation reactions, systematics of the Gramineae, and leaf anatomy. *Canad. J. Bot.* **46**: 207–215.
- DRUYTS-VOETS, E. 1970. Types van stengel: en bladstrukturen in het genus *Cyperus* L. *Natuurw. Tijdschr. Ned.-Indië* **52**: 28–49.
- EHLERINGER, J. R., Z. F. LIN, C. B. FIELD, G. C. SUN, AND C. Y. KUO. 1987. Leaf carbon isotope ratios of plants from a subtropical monsoon forest. *Oecologia* 72: 109–114.
- EITEN, L. T. 1969. The vegetative anatomy of *Eleocharis interstincta* (Vahl) Roem. et Schult. *Arq. Bot. Estado São Paulo* **4**: 187–228.
- ——. 1976. The morphology of some critical Brazilian species of Cyperaceae. *Ann. Missouri Bot. Gard.* **63**: 113–199.
- ELLERY, K., W. N. ELLERY, AND B. T. VERHAGEN. 1992. The distribution of C<sub>3</sub> and C<sub>4</sub> plants in a successional sequence in the Okavango Delta. *S. African J. Bot.* **58**: 400–402.
- Frey, W., and H. Kürschner. 1983. Photosyntheseweg und Zonierung von Halophyten an Salzseen in der Türkei, in Jordanien und im Iran. *Flora* **173**: 293–310.
- Getliffe Norris, F. M. 1983. Anatomy of the genus *Kyllinga* in South Africa. *Bothalia* 14: 809–814.

- GHAMKHAR, K., A. MARCHANT, K. L. WILSON, AND J. J. BRUHL. 2007.
  Phylogeny of Abildgaardieae (Cyperaceae) inferred from ITS and trnL–F data, pp. 149–164. In J. T. Columbus, E. A. Friar, J. M. Porter, L. M. Prince, and M. G. Simpson [eds.], Monocots: comparative biology and evolution—Poales. Rancho Santa Ana Botanic Garden, Claremont, California, USA.
- GILLILAND, M. G., AND K. D. GORDON-GRAY. 1978. Kranz and non-Kranz cells in Cyperaceae. Proceedings of the Electron Microscopy Society of Southern Africa 8: 85–86.
- GOETGHEBEUR, P. 1986. Genera Cyperacearum. Een bijdrage tot de kennis van de morfologie, systematiek en fylogenese van de Cyperaceae-genera. Ph.D. thesis, Rijksuniversiteit Gent, Belgium. 1164 pp. [In Flemish.]
- . 1998. Cyperaceae, pp. 141–190. In K. Kubitzki, H. Huber, P. J. Rudall, P. S. Stevens, and T. Stuetzel [eds.], The families and genera of vascular plants, Vol. 4. Springer-Verlag, Berlin, Germany.
- GOLDBLATT, P., P. C. HOCH, AND L. M. McCOOK. 1992. Documenting scientific data: the need for voucher specimens. *Ann. Missouri Bot. Gard.* **79**: 969–970.
- GORDON-GRAY, K. D. 1971. *Fimbristylis* and *Bulbostylis*: generic limits as seen by a student of southern African species. *Mitt. Bot. Staatssamml. München* 10: 549–574.
- GOVINDARAJALU, E. 1966. The systematic anatomy of South Indian Cyperaceae: *Bulbostylis* Kunth. *Bot. J. Linn. Soc.* **59**: 289–304.
- \_\_\_\_\_\_. 1969a. The systematic anatomy of South Indian Cyperaceae: Fuirena Rottb. Bot. J. Linn. Soc. 62: 27–40.
- . 1969b. The systematic anatomy of South Indian Cyperaceae: *Cyperus* L. subgen. *Kyllinga* (Rottb.) Suringar. *Bot. J. Linn. Soc.* **62**: 41–58.
- . 1974. The systematic anatomy of South Indian Cyperaceae: Cyperus L. subgen. Juncellus, Cyperus subgen. Mariscus and Lipocarpha R. Br. Bot. J. Linn. Soc. 68: 235–266.
- 1975a. The systematic anatomy of South Indian Cyperaceae: Eleocharis R. Br., Rhynchospora Vahl and Scleria Bergius. Adansonia 14: 581–632.
- ——. 1975b. Studies in Cyperaceae. XIV. Endomorphic evidence for placing *Cyperus hyalinus* under the new subgenus *Queenslan-diella*. Reinwardtia 9: 187–195.
- . 1976. The systematic anatomy of South Indian Cyperaceae: *Scirpus L. s. lat. Adansonia* **16**: 13–38.
- . 1981. Studies in Cyperaceae. 19. Novelty in *Lipocarpha*: L. raynaleana and its vegetative anatomy. Adansonia 20: 369–375
- . 1985. Studies in Cyperaceae. 19. Species novarum Cyperacearum in memoriam optimi Professori Swamy. *Proc. Indian Acad. Sci.*, *Pl. Sci.* 94: 11–31.
- . 1990a. Cyperaceae Indiae Australis Precursores—nova species in *Fimbristyle* (L.) Vahl and their vegetative anatomy. *Proc. Indian Acad. Sci., Pl. Sci.* 100: 77–84.
- . 1990b. Cyperaceae Indiae Australis Precursores: a novelty in *Cyperus* Linn. and its vegetative anatomy. *Proc. Indian Acad. Sci.*, *Pl. Sci.* 100: 409–413.
- ——, AND J. RAYNAL. 1976. Notes cypérologiques. 28. Compléments sur le genre *Rikliella* J. Rayn. *Adansonia* 16: 219–228.
- GUAGLIANONE, E. R. 1990. Una especie nueva de *Cyperus* (Cyperaceae) de Misiones Argentina. *Darwiniana* 30: 233–236.
- —, AND O. UENO. 1990. A disjunct species in *Eleocharis* (Cyperaceae). *Darwiniana* 30: 223–229.
- ——, AND B. RUTHSATZ. 1998. Note on *Eleocharis tuc-umanensis* (Cyperaceae). *Darwiniana* **35**: 169–173.
- HABERLANDT, G. 1882. Vergleichende Anatomie des assimilatorischen Gewebesystems der Pflanzen. Jahrb. Wiss. Bot. 13: 74–188.
- . 1884. Physiological plant anatomy. Today and Tomorrow's Book Agency, New Delhi, India. 777 pp.

- HAINES, R. W., AND K. A. LYE. 1983. The sedges and rushes of East Africa. East African Natural History Society, Nairobi, Kenya. 404 pp.
- HATTERSLEY, P. W. 1987. Variations in photosynthetic pathway, pp.
  49–64. *In* T. R. Soderstrom, K. W. Hilu, C. S. Campbell, and M. E. Barkworth [eds.], Grass systematics and evolution. Smithsonian Institution Press, Washington, D.C., USA.
- ——, AND L. WATSON. 1975. Anatomical parameters for predicting photosynthetic pathways of grass leaves: The "maximum lateral cell count" and the "maximum cells distant count". *Phytomorphology* 25: 325–333.
- ——, AND C. B. OSMOND. 1977. *In situ* immunofluorescent labelling of ribulose-1,5-bisphosphate carboxylase in leaves of C<sub>3</sub> and C<sub>4</sub> plants. *Austral. J. Pl. Physiol.* **4**: 523–539.
- —, S.-C. Wong, S. Perry, and Z. Roksandic. 1986. Comparative ultrastructure and gas exchange characteristics of the C<sub>3</sub>-C<sub>4</sub> intermediate *Neurachne minor S. T. Blake* (Poaceae). *Pl. Cell Environ.* 9: 217–233.
- HAYASAKA, E. 2002. Taxonomic revision of the genus *Schoenoplectus* (Cyperaceae), with special reference to nutlet micromorphology. Ph.D. thesis, Tohoku University, Sendai, Japan. 325 pp.
- Hesla, B. L., L. L. Teiszen, and S. K. Imbamba. 1982. A systematic survey of C<sub>3</sub> and C<sub>4</sub> photosynthesis in the Cyperaceae of Kenya, East Africa. *Photosynthetica* **16**: 196–205.
- HNATIUK, R. J. 1980. C<sub>4</sub> photosynthesis in the vegetation of Aldabra Atoll. *Oecologia* **44**: 327–334.
- HOFSTRA, J. J., S. AKSORNKOAE, S. ATMOWIDJOJO, J. F. BANAAG, SANTOSA, R. A. SASTROHOETOMO, AND L. T. N. THU. 1972. A study on the occurrence of plants with a low CO<sub>2</sub> compensation point in different habitats in the tropics. *Ann. Bogor.* 5: 143–157.
- HOSKING, J. R., G. R. SAINTY, AND S. W. L. JACOBS. 1996. Certainty and uncertainty in plant identification, pp. 464–467. *In* R. C. H. Shepherd [ed.], Proceedings of the Eleventh Australian Weeds Conference. Weed Science Society, Frankston, Victoria, Australia.
- IMAI, K., AND Y. MURATA. 1979. Changes in apparent photosynthesis, CO<sub>2</sub> compensation point and dark respiration of leaves of some Poaceae and Cyperaceae species with senescence. *Pl. Cell Physiol.* 20: 1653–1658.
- JONES, M. B., G. E. HANNON, AND M. D. COFFEY. 1981. C<sub>4</sub> photosynthesis in *Cyperus longus* L., a species occurring in temperate climates. *Pl. Cell Environ.* 4: 161–168.
- ——, AND T. R. MILBURN. 1978. Photosynthesis in papyrus (*Cyperus papyrus* L.). *Photosynthetica* **12**: 197–199.
- KALAPOS, T., A. BALOGHNE-NYAKAS, AND P. CSONTOS. 1997. Occurrence and ecological characteristics of C<sub>4</sub> dicot and Cyperaceae species in the Hungarian flora. *Photosynthetica* **33**: 227–240.
- Keeley, J. E., L. Sternberg, and M. J. Deniro. 1986. The use of stable isotopes in the study of photosynthesis in freshwater plants. *Aquatic Bot.* **26**: 213–223.
- KÖRNER, C., G. D. FARQUHAR, AND Z. ROKSANDIC. 1988. A global survey of carbon isotope discrimination in plants from high altitude. *Oecologia* **74**: 623–632.
- KOYAMA, T. 1966. The systematic significance of leaf structure in the Cyperaceae–Mapanieae. *Mem. New York Bot. Gard.* **15**: 136–150
- . 1967. The systematic significance of leaf structure in the tribe Sclerieae (Cyperaceae). *Mem. New York Bot. Gard.* 16: 46– 70
- , AND E. R. GUAGLIANONE. 1987. Oreobolopsis: a new genus of Cyperaceae (Scirpeae) from Bolivia, South America. Darwiniana 28: 79–85.
- KRENZER, E. G. J., D. N. Moss, AND R. K. CROOKSTON. 1975. Carbon dioxide compensation points of flowering plants. *Pl. Physiol.* (*Lancaster*) 56: 194–206.
- KÜKENTHAL, G. 1935. Cyperaceae–Scirpoideae–Cypereae 1, pp. 1– 160. In A. Engler [ed.], Das Pflanzenreich, Heft 101. W. Engelmann, Berlin, Germany.

- . 1936. Cyperaceae-Scirpoideae-Cypereae 2, pp. 161-671. In A. Engler [ed.], Das Pflanzenreich, Heft 101. W. Engelmann, Berlin, Germany.
- KUKKONEN, I. 1967. Vegetative anatomy of *Uncinia* (Cyperaceae). Ann. Bot. (Oxford) 31: 523–544.
- . 1970. Vegetative anatomy of Carex microglochin Wahlenb. and Carex camptoglochin Krech. Bot. J. Linn. Soc. 63 (suppl. 1): 137–145.
- . 1995. (1192–1193) Two proposals to conserve species names in Cyperaceae. *Taxon* 44: 625–627.
- ——, AND K. A. LYE. 1996. Cyperus medusaeus (Cyperaceae) redescribed. Ann. Bot. Fenn. 33: 21–27.
- KUOH, C.-S., AND S.-H. T. CHIANG. 1984. The comparative leaf anatomy of the genus Fimbristylis (Cyperaceae). Yushania 1: 29–43.
- LERMAN, J. C., AND J. RAYNAL. 1972. La teneur en isotopes stables du carbone chez les Cypéracées: sa valeur taxonomique. Compt. Rend. Hebd. Séances Acad. Sci., Sér. D 275: 1391–1394.
- LI, M. 1993. Distribution of C<sub>3</sub> and C<sub>4</sub> species of *Cyperus* in Europe. *Photosynthetica* 28: 119–126.
- —, AND M. B. JONES. 1994. Kranzkette, a unique C<sub>4</sub> anatomy occurring in *Cyperus japonicus* leaves. *Photosynthetica* 30: 117–131.
- , D. A. Wedin, And L. L. Tieszen. 1999. C<sub>3</sub> and C<sub>3</sub> photosynthesis in *Cyperus* (Cyperaceae) in temperate eastern North America. *Canad. J. Bot.* 77: 209–218.
- LIN, C. H., Y. S. TAI, D. J. LIU, AND M. S. B. KU. 1993. Photosynthetic mechanisms of weeds in Taiwan. *Austral. J. Pl. Physiol.* 20: 757–769.
- MANI, A. P. 1963. Leaf anatomy in the identification of *Cyperus* species. *Trans. Bose Res. Inst.*, *Calcutta* 26: 119–128.
- MATEU ANDRES, I. 1991. Leaf anatomy of plants from coastal Mediterranean salt marshes. Monocotyledons. Candollea 46: 345–358.
- McNaughton, S. J., L. L. Wallace, and M. B. Coughenour. 1983. Plant adaptation in an ecosystem context: effects of defoliation, nitrogen and water on growth of an African C<sub>4</sub> sedge. *Ecology* **64**: 307–318.
- MEINZER, F. C. 1978. Observation on the taxonomic and ecologic distribution of C<sub>4</sub> carbon photosynthesis in vegetation of northwest Central America. *Revista Biol. Trop.* 26: 359–369.
- METCALFE, C. R. 1971. Cyperaceae, pp. 260–261. *In C. R.* Metcalfe [ed.], Anatomy of the monocotyledons, Vol. 5. Clarendon Press, Oxford, UK. 597 pp.
- Moss, D. N., E. G. J. Krenzer, and W. A. Brun. 1969. Carbon dioxide compensation points in related plant species. *Science* 164: 187–188.
- MUASYA, A., J. J. BRUHL, D. A. SIMPSON, A. CULHAM, AND M. W. CHASE. 2000. Suprageneric phylogeny of Cyperaceae, pp. 593–601. *In* K. L. Wilson and D. A. Morrison [eds.], Monocots: systematics and evolution. CSIRO Publishing, Collingwood, Victoria, Australia.
- , D. A. SIMPSON, AND M. W. CHASE. 2002. Phylogenetic relationships in *Cyperus* L. s.l. (Cyperaceae) inferred from plastid DNA sequence data. *Bot. J. Linn. Soc.* 138: 145–153.
- , , , , , , AND A. CULHAM. 1998. An assessment of suprageneric phylogeny in Cyperaceae using *rbc*L DNA sequences. *Pl. Syst. Evol.* **211**: 257–271.
- MULROY, T. W., AND P. W. RUNDEL. 1977. Annual plants: adaptations to desert environments. *Bioscience* 27: 109–114.
- NAUTIYAL, D. D., AND B. K. DAS. 1982. Contributions to the anatomy of Cyperaceae. I: Cyperus tenuispica Steud. Phyta, Studies on Living & Fossil Plants, Pant Commem. Vol., pp. 173–197.
- PONESSA, G., M. TORRES, AND L. BENÍTEZ. 1997. Estudio taxonómico de *Scirpus* sección *Monocephales* (Cyperaceae) en Tucumán, República Argentina. *Lilloa* 39: 5–30.
- Prakash, N., T. C. Shen, K. C. Yap, and K. M. Yim. 1976. A survey of the leaf structure and its relationship to the photosynthetic pathways in certain Malaysian plants. *Malaysian J. Sci.* **4**: 67–73.

- RAGHAVENDRA, A. S., AND S. R. DAS. 1976. Distribution of the C<sub>4</sub> dicarboxylic acid pathway of photosynthesis in local monocotyledonous plants and its taxonomic significance. *New Phytol.* 76: 301–305.
- RAYNAL, J. 1972. La teneur en isotopes stables du carbone chez les Cypéracées: sa valeur taxonomique. Compt. Rend. Hebd. Séances Acad. Sci., Sér. D 275: 1391–1394.
- . 1973. Notes cypérologiques. 19. Contribution à la classification de la sous-famille des Cyperoideae. Adansonia 13: 145– 171.
- RIKLI, M. 1895. Beiträge zur vergleichenden Anatomie der Cyperaceen mit besonderer Berücksichtigung der inneren Parenchymscheide. *Jahrb. Wissensch. Bot.* 27: 485.
- ROALSON, E. H., AND E. A. FRIAR. 2000. Infrageneric classification of *Eleocharis* (Cyperaceae) revisited: evidence from the internal transcribed spacer (ITS) region of nuclear ribosomal DNA. *Syst. Bot.* 25: 323–336.
- RODRIGUES, A. C., AND M. E. M. ESTELITA. 2003. Origin and structure of the Kranz tissue in bracts of *Cyperus giganteus* Vahl (Cyperaceae). *Revista Brasil. Bot.* 26: 445–452.
- SABNIS, T. S. 1921. The physiological anatomy of the plants of the Indian Desert. *J. Indian Bot.* 2: 163–299.
- SAGE, R. F. 2002. C<sub>4</sub> photosynthesis in terrestrial plants does not require Kranz anatomy. *Trends Pl. Sci.* 7: 283–285.
- , M. LI, AND R. K. MONSON. 1999. The taxonomic distribution of C<sub>4</sub> photosynthesis, pp. 551–584. *In R. F. Sage and R. K. Monson [eds.]*, C<sub>4</sub> plant biology. Academic Press, San Diego, California, USA.
- SAXENA, K. G., AND P. S. RAMAKRISHNAN. 1984. C<sub>3</sub>/C<sub>4</sub> species distribution among successional herbs following slash and burn in north-eastern India. *Acta Oecologica* **5**: 335–346.
- SHARMA, O. P. 1972. Anatomy of *Scirpus squarrosus* L. *Curr. Sci.* 41: 494–497.
- . 1973. Anatomy of Eriophorum comosum. Phytomorphology 23: 17–24.
- ——, AND P. N. MEHRA. 1970. Comparative anatomy of *Kobresia* Willd. (Cyperaceae). *Res. Bull. Punjab Univ., New Ser., Sci.* 21: 119–128.
- —, AND —. 1972. Systematic anatomy of *Fimbristylis* Vahl (Cyperaceae). *Bot. Gaz.* **133**: 87–95.
- SHEPHERD, G. J. 1976. The use of anatomical characters in the intrageneric classification of *Carex* (Cyperaceae). *Hoehnea* **6**: 33–54.
- SIMPSON, D. 1990. A revision of *Cyperus* sect. *Leucocephali. Kew Bull.* **45**: 485–501.
- SIMPSON, D. A., A. M. MUASYA, K. CHAYAMARIT, J. A. N. PARNELL, S. SUDDEE, B. D. E. WILDE, M. B. JONES, J. J. BRUHL, AND R. POOMA. 2005. Khaosokia caricoides, a new genus and species of Cyperaceae from Thailand. Bot. J. Linn. Soc. 149: 357–364.
- SMITH, B. N., AND S. EPSTEIN. 1971. Two categories of <sup>13</sup>C/<sup>12</sup>C ratios for higher plants. *Pl. Physiol. (Lancaster)* **47**: 380–384.
- SMITH, G. S., C. M. JOHNSTON, AND I. S. CORNFORTH. 1983. Comparison of nutrient solutions for growth of plants in sand culture. New Phytol. 94: 537–548.
- SONNENBERG, B. J., AND C. E. J. BOTHA. 1992. An investigation of leaf-blade anatomy and photosynthetic characteristics of four Cyperaceae species from the Albany and Bathurst districts in the eastern Cape. S. Afr. J. Bot. 58: 297–303.
- Soros, C. L., AND J. J. BRUHL. 2000. Multiple evolutionary origins of C<sub>4</sub> photosynthesis in the Cyperaceae, pp. 629–636. *In* K. L. Wilson and D. A. Morrison [eds.], Monocots: systematics and evolution. CSIRO Publishing, Collingwood, Victoria, Australia.
- , AND N. G. DENGLER. 2001. Ontogenetic derivation and cell differentiation in photosynthetic tissues of C<sub>3</sub> and C<sub>4</sub> Cyperaceae. *Amer. J. Bot.* 88: 992–1005.
- STANDLEY, L. A. 1987a. Anatomical and chromosomal studies of

- Carex section Phacocystis in eastern North America. Bot. Gaz. 148: 507-518.
- . 1987b. Anatomical studies of *Carex cuchumatanensis*, *C. decidua* and *C. hermannii* (Cyperaceae) and comparisons with North American taxa of the *C. acuta* complex. *Brittonia* **39**: 11–19
- ——. 1990. Anatomical aspects of the taxonomy of sedges (*Carex*, Cyperaceae). *Canad. J. Bot.* **68**: 1449–1456.
- STERNBERG, L., M. J. DENIRO, AND J. E. KEELEY. 1984. Hydrogen, oxygen, and carbon isotope ratios of cellulose from submerged aquatic crassulacean acid metabolism and non-crassulacean acid metabolism plants. *Pl. Physiol. (Lancaster)* 76: 68–70.
- STOCK, W. D., D. K. CHUBA, AND G. A. VERBOOM. 2004. Distribution of South African C<sub>3</sub> and C<sub>4</sub> species of Cyperaceae in relation to climate and phylogeny. *Austral Ecol.* **29**: 313–319.
- SVENSON, H. K. 1937. Monographic studies in the genus *Eleocharis*. IV. *Rhodora* **39**: 236–273.
- . 1939. Monographic studies in the genus *Eleocharis*. V. *Rhodora* 41: 73–77.
- SYVERTSEN, J. P., G. L. NICKELL, R. W. SPELLENBERG, AND G. L. CUNNINGHAM. 1976. Carbon reduction pathways and standing crop in three Chihuahuan desert plant communities. *Southwestern Naturalist* 21: 311–320.
- TAKEDA, T., O. UENO, AND W. AGATA. 1980. The occurrence of C<sub>4</sub> species in the genus *Rhynchospora* and its significance in Kranz anatomy of the Cyperaceae. *Bot. Mag. (Tokyo)* 93: 55–65.
- —, —, M. SAMEJIMA, AND T. OHTANI. 1985. An investigation for the occurrence of C<sub>4</sub> photosynthesis in the Cyperaceae from Australia. *Bot. Mag. (Tokyo)* 98: 393–411.
- THOMAS, W. W. 1984. The systematics of *Rhynchospora* sect. *Dichromena. Mem. New York Bot. Gard.* 37: 1–116.
- TREGUNNA, E. B., B. N. SMITH, J. A. BERRY, AND W. J. S. DOWNTON. 1970. Some methods for studying the photosynthetic taxonomy of the angiosperms. *Canad. J. Bot.* 48: 1209–1214.
- TROUGHTON, J. H., K. A. CARD, AND C. H. HENDY. 1974. Photosynthetic pathways and carbon isotope discrimination by plants. *Yearb. Carnegie Inst. Washington* **73**: 768–780.
- UCHINO, A., M. SAMEJIMA, R. ISHII, AND O. UENO. 1995. Photosynthetic carbon metabolism in an amphibious sedge, *Eleocharis baldwinii* (Torr.) Chapm.: modified expression of C<sub>4</sub> characteristics under submerged aquatic conditions. *Pl. Cell Physiol.* 36: 229–238.
- UENO, O. 1996. Structural characterization of photosynthetic cells in an amphibious sedge, *Eleocharis vivipara*, in relation to C<sub>3</sub> and C<sub>4</sub> metabolism. *Planta* 199: 382–393.
- . 1998a. Immunogold localization of photosynthetic enzymes in leaves of various C<sub>4</sub> plants, with particular reference to pyruvate orthophosphate dikinase. J. Exper. Bot. 49: 1637–1646.
- . 1998b. Induction of Kranz anatomy and C<sub>4</sub>-like biochemical characteristics in a submerged amphibious plant by abscisic acid. Pl. Cell 10: 571–583.
- . 2001. Environmental regulation of C<sub>3</sub> and C<sub>4</sub> differentiation in the amphibious sedge *Eleocharis vivipara*. *Pl. Physiol. (Lan-caster)* 127: 1524–1532.
- 2004. Environmental regulation of photosynthetic metabolism in the amphibious sedge *Eleocharis baldwinii* and comparisons with related species. *Pl. Cell Environm.* 27: 627–639.
- , AND K. ISHIMARU. 2002. Effects of an inhibitor of phosphoenolpyruvate carboxylase on photosynthesis of the terrestrial forms of amphibious *Eleocharis* species. *Photosyn. Res.* 71: 265–272.
- —, AND Y. KOBAYASHI. 2002. Cellular regulation of glycine decarboxylase in the amphibious C<sub>3</sub>/C<sub>4</sub> sedge, *Eleocharis vivipa*ra. Pl. Cell Physiol. 43: S173–S174.
- —, AND T. KOYAMA. 1987. Distribution and evolution of C<sub>4</sub> syndrome in *Rhynchospora* (Rhynchosporeae: Cyperaceae). *Bot. Mag. (Tokyo)* 100: 63–85.

- , AND M. SAMEJIMA. 1989. Structural features of NAD-Malic Enzyme Type C<sub>4</sub> *Eleocharis*: an additional report of C<sub>4</sub> acid decarboxylation types of the Cyperaceae. *Bot. Mag. (Tokyo)* **102**: 393–402.
- , AND ——. 1990. Immunogold localization of ribulose 1,5-bisphosphate carboxylase in amphibious *Eleocharis* species in relation to C<sub>3</sub> and C<sub>4</sub> photosynthesis. *Current Research in Photosynthesis* **4**: 867–870.
- of C<sub>4</sub> syndrome in *Eleocharis*, a sedge group inhabiting wet and aquatic environments, based on culm anatomy and carbon isotope ratios. *Ann. Bot.* **64**.
- , —, S. MUTO, AND S. MIYACHI. 1988a. Photosynthetic characteristics of an amphibious plant, *Eleocharis vivipara*: expression of C<sub>4</sub> and C<sub>3</sub> modes in contrasting environments. *Proc. Natl. Acad. Sci. U.S.A.* **85**: 6733–6737.
- —, AND T. TAKEDA. 1992. Photosynthetic pathways, ecological characteristics, and the geographical distribution of the Cyperaceae in Japan. *Oecologia* 89: 195–203.
- ———, AND E. MAEDA. 1988b. Leaf ultrastructure of C<sub>4</sub> species possessing different Kranz anatomical types in the Cyperaceae. *Bot. Mag. (Tokyo)* 101: 141–152.
- ———, AND T. MURATA. 1986. C<sub>4</sub> acid decarboxylating enzyme activities of C<sub>4</sub> species possessing different Kranz anatomical types in the Cyperaceae. *Photosynthetica* 20: 111–116.
- Vorster, P. 1990. Anatomy of the South African species of *Mariscus* (Cyperaceae), and its relation to environmental conditions. *Mitt. Inst. Allg. Bot. Hamburg* 23a: 367–386.
- ——. 1996. Justification for the generic status of *Courtoisina* (Cyperaceae). *Bot. J. Linn. Soc.* 121: 271–280.
- WANG, R. Z. 2003. C<sub>4</sub> plants in the vegetation of Tibet, China: their natural occurrence and altitude distribution pattern. *Photosynthetica* **41**: 21–26.
- WILLIAMS, G. J., AND R. K. MONSON. 1981. Gas exchange in *Carex stenophylla* and its relationship to leaf anatomy, pp. 11–21. *In G.* Akoyunoglou [ed.], Photosynthesis VI. Photosynthesis and productivity, photosynthesis and environment. Balaban International Science Services, Philadelphia, USA.
- WILLS, G. D. 1987. Description of purple and yellow nutsedge (Cyperus rotundus and C. esculentus). Weed Technol. 1: 2–9.
- WILSON, K. L. 1981. Revision of the genus Mesomelaena (Cyperaceae). Telopea 2: 181–195.
- ——. 1991. Systematic studies in *Cyperus* section *Pinnati* (Cyperaceae). *Telopea* 4: 361–496.
- WINTER, K., AND J. H. TROUGHTON. 1978. Photosynthetic pathways in plants of coastal and inland habitats of Israel and the Sinai. Flora. Abteilung A, Physiologie und Biochemie. Jena 167: 1–34.

### APPENDIX 1: Published and Inferred Records of Photosynthetic Pathway Type in Cyperaceae

This list includes both previously published records of photosynthetic pathway type in species of Cyperaceae and newly published records.

Column 1: Species.—A taxon is listed by what the present authors believe is its currently accepted name. For previously published records, the name used in the original publication is indicated if different from the current name. The present authors have examined and confirmed or re-determined the vouchers concerned in some cases. For example, one of the vouchers for Mesomelaena stygia (Coveny 8296, NSW) cited by Takeda et al. (1985) is actually M. preissii (Wilson 1981). Similarly, Takeda et al. (1985) cited NT 42319 (Henshall 249) as Cyperus angustatus, but that specimen belongs to C. fucosus (voucher re-determined by KLW at NSW; Wilson 1991). For other records, the authors have updated the nomenclature from recent monographic and floristic works, as well as the draft World Checklist being coordinated by D. Simpson and R. Go-

vaerts, and by consulting other cyperologists (see Acknowledgments). This does not guarantee, of course, that the vouchers were correctly identified by the original researchers, so the vouchers should be reexamined in any critical cases.

Column 2: Photosynthetic pathway.—Pathway type is indicated as being  $C_3$ ,  $C_4$ , or apparently intermediate or variable in some species. These types are inferred by the present authors from the analyses either quoted here for new records or given in previously published works. + = an anomalous record that is discussed in Appendix 2 or in the text.

Column 3: References.—References are to previously published and new records, which are treated differently.

Previously published records.—For these, the publication is cited, followed by the method by which the pathway type was assessed, using the abbreviations below, and as discussed in the main text. In some older publications that pre-date recognition of the significance of anatomical arrangements, we have inferred the pathway type from the anatomical sections illustrated—indicated by "[A]". Actual values are not given here. For these, the reader should consult the original publications or the current authors' database, which will be available on-line by the second half of 2006 (or by contacting one of the two authors).

A = anatomy

[A] = anatomy deduced from older publications

c = chlorocyperoid anatomy

e, [e] = eleocharoid anatomy

f = fimbristyloid anatomy

r = rhynchosporoid anatomy

B = biochemistry

 $\Gamma = CO_2$  compensation point (ml/liter or ppm  $CO_2$ )

 $\delta^{13}$ C =  $\delta^{13}$ C values in %0

IL = immunofluorescence labelling of Rubisco

ASP = aspartate as the initial product of photosynthesis

MAL = malate as the initial product of photosynthesis

NAD = NAD-malic enzyme (ME)

NADP = NADP-ME

PHOS = sugar phosphates as the initial products of photosynthesis

PIB = post-illumination CO<sub>2</sub> burst effect

US = ultrastructure

S = submerged form of an *Eleocharis* species

T = terrestrial form of an Eleocharis species

New records.—Records newly published here come from several sources. Many are from the current authors' separate or joint research, indicated by "JB", "KW" or "BW" (see abbreviations listed below). Records for Phylloscirpus are based on leaf sections cut by Sandra Dhooge (GENT) and interpreted by JB and KW. Records labelled as "LR" are the original analyses by J. C. Lerman and the late Jean Raynal in the early 1970s that were the basis of several publications (Lerman and Raynal 1972; Raynal 1972, 1973). Similarly, those labelled as "SCV" come from the work of W. D. Stock, D. K. Chuba, and G. A. Verboom and underlie their recent publication on South African species (Stock et al. 2004). See text for further discussion. All vouchers for new LR records are in P; all vouchers for SCV records are in BOL. The default herbarium is NSW for new records from Bruhl and Wilson; all other herbaria are indicated using the abbreviations in Index Herbariorum (current version accessible on the New York Botanical Garden website, www. nybg.org).

JB = J. J. Bruhl, this study, including records listed in Bruhl (1990)

KW = K. L. Wilson, this study

BW = J. Bruhl and K. Wilson, this study

BDW = J. Bruhl, S. Dhooge, and K. Wilson, this study

LR = the records from J. C. Lerman and J. Raynal that underlaid Lerman and Raynal (1972) and Raynal (1972, 1973)

SCV = the records from W. D. Stock, D. K. Chuba, and G. A. Verboom that underlay Stock et al. (2004).

Appendix 1. Published and inferred records of photosynthetic pathway type in Cyperaceae.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
Abildgaardia hygrophila (Gordon-Gray) K. Lye	$C_3$	JB (A: Ward 2794 BRI, 5519 BRI. δ <sup>13</sup> C: -27.9, Ward 2794, -28.2, Ward 5519)
A. macrantha (Boeck.) ined., as Fimbristylis macrantha Boeck.	$C_4$	Takeda et al. 1985 (Af)
A. ovata (Burm. f.) Kral, as Fimbristylis monostachyos (L.) Hassk.a, as F. ova- ta (Burm. f.) Kernb	$\mathrm{C}_4$	Gordon-Gray 1971 ([A]) <sup>b</sup> ; Sharma and Mehra 1972 ([A]) <sup>a</sup> ; Raghavendra and Das 1976 (A. Γ) <sup>a</sup> ; Carolin et al. 1977 (USf) <sup>b</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C) <sup>b</sup> ; Kuoh and Chiang 1984 (A) <sup>b</sup> ; Bruhl et al. 1987 (Af. B) <sup>b</sup> ; LR (δ <sup>13</sup> C: –11.3, <i>Chevalier</i> 22172) <sup>b</sup>
A. oxystachya (F. Muell.) ined., as Fimbristylis oxystachya F. Muell.	$\mathrm{C}_4$	Takeda et al. 1985 (Af)
A. schoenoides R. Br., as Fimbristylis squarrulosa F. Muell.	$C_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C)
A. triflora (L.) Abeyw., as Fimbristylis tri- flora (L.) K. Schum. <sup>a</sup>	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; JB (Af: <i>Davidse 8228</i> BRI. δ <sup>13</sup> C: -10.6, <i>Davidse 8228</i> ); LR (δ <sup>13</sup> C: -12.0, <i>Boivin s. n.</i> ) <sup>a</sup>
A. vaginata R. Br., as Fimbristylis brownii Benth.	$C_4$	Bruhl et al. 1987 (Af. B); JB (Af: s. coll. CANB 114505; Bruhl 233, 236 CANB)
Actinoschoenus filiformis (Thwaites) Benth.	$C_3$	JB (A: Ramos NSW 181450); LR (δ <sup>13</sup> C: -27.4, Poilane 23085)
Actinoscirpus grossus (L. f.) Goetgh. & D. A. Simpson, as Hymenochaeta grossa (L. f.) Nees <sup>a</sup> , as Scirpus grossus L. f. <sup>b</sup>	$C_3$	Govindarajalu 1976 ([A]) <sup>b</sup> ; Takeda et al. 1985 (A) <sup>a</sup> ; JB (A: <i>Specht 1243</i> CANB)
Afrotrilepis jaegeri J. Raynal	$C_3$	JB (A: Jaeger 7869 NSW 181677)
A. pilosa (Boeck.) J. Raynal, as Eriospora pilosa (Boeck.) Benth. <sup>a</sup>	$C_3$	Chermezon 1930 ([A]) <sup>a</sup> ; JB (A: <i>Letouzey 13915</i> ); LR (δ <sup>13</sup> C: -32.3, <i>Hallé &amp; Villiers 4978</i> , -33.1, Serre Orsay cult., 1972)
Alinula lipocarphoides (Kük.) J. Raynal	$\mathbb{C}_4$	JB (Ac: Robinson 5018 EA)
A. malawica (J. Raynal) Goetgh. & Vorster, as Mariscus malawicus J. Raynal	$\mathrm{C}_4$	Raynal 1973 (A)
A. paradoxa (Cherm.) Goetgh. & Vorster	$\mathbf{C}_3$	SCV ( $\delta^{13}$ C: $-25.08$ , Burtt-Davy 1749)
A. peteri (Kük.) Goetgh. & Vorster, as As- colepis peteri Kük. <sup>a</sup>	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; JB (Ac: <i>Greenway 13488</i> EA); LR (δ <sup>13</sup> C: -14.1, <i>Robinson 4438</i> ) <sup>a</sup>
Amphiscirpus nevadensis (S. Watson) Oteng-Yeboah	$C_3$	JB (A: Peck 15386 K)
Androtrichum trigynum (Spreng.) H. Pfeiff. Arthrostylis aphylla R. Br.	$C_3$ $C_3$	JB (A: Rosengurttx B3904 U); LR (δ <sup>13</sup> C: -27.5, Hatschbach 15198) Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: Reeve 127 CANB); LR (δ <sup>13</sup> C: -27.4, Leichhardt 33)
Ascolepis capensis (Kunth) Ridl.	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C); JB (Ac: <i>Cooper</i> , Mar 1873, MEL 1543822); LR (δ <sup>13</sup> C: -12.4, <i>Meurillon CNAD 688</i> ); SCV (δ <sup>13</sup> C: -10.31, <i>Bolus 3944</i> )
A. dipsacoides (K. Schum.) J. Raynal	$\mathbf{C}_4$	LR (δ <sup>13</sup> C: -10.3, Annet 53)
A. pinguis C. B. Clarke	$\mathbf{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
A. protea Welwitsch	$\mathbf{C}_4$	LR (δ <sup>13</sup> C: -10.4, <i>Letouzey 7567</i> )
A. pusilla Ridl.	$\mathbf{C}_4$	LR (δ <sup>13</sup> C: -11.4, Demange 3276)
Asterochaete capitellata Nees (Carpha capitellata (Nees) Boeck.)	$C_3$	JB (A: MEL 1543862)
Baumea acuta (Labill.) Palla	$\mathbf{C}_3$	JB (A: Newbey 4625 CANB)
B. articulata (R. Br.) S. T. Blake, as Ma-	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C)
chaerina articulata (R. Br.) Koyama B. deplanchei Boeck.	C	LR (δ¹³C: −30.4, Raynal & Jaffré 16490)
B. glomerata Gaud.	$C_3$ $C_3$	JB (A: van Royen 5218 CANB)
B. juncea (Nees) Boeck., as Machaerina juncea (R. Br.) Koyama	$C_3$	Takeda et al. 1985 (A)
B. rubiginosa (Spreng.) Boeck., as Ma- chaerina nipponensis (Ohwi) Ohwi & Koyama <sup>a</sup> , as Machaerina rubiginosa (Spreng.) Koyama <sup>b</sup>	C <sub>3</sub>	Takeda et al. 1985 (A. $\delta^{13}C)^b$ ; Ueno et al. 1986 (A. B) <sup>a</sup> ; JB (A: Bruhl 518 CANB)
B. teretifolia (R. Br.) Palla, as Machaerina teretifolia (R. Br.) Koyama	$C_3$	Takeda et al. 1985 (A)
B. tetragona (Labill.) S. T. Blake	$C_3$	KW (A: Beauglehole 36048; Sharpe 1365)
Becquerelia cymosa Brongn. subsp. cymosa	$C_3$	JB (A: Harley 20171 K); LR (δ <sup>13</sup> C: -32.0, Sastre 97)
Bisboeckelera irrigua (Nees) Kuntze	$C_3$	LR (δ <sup>13</sup> C: -36.6, Smith 2785)
B. microcephala (Boeck.) Koyama	$C_3$	JB (A: Florschuetz 1819 U)
Blysmus compressus (L.) Panz. ex Link	$\mathbf{C}_3$	JB (A: Manchester 1387); LR (δ <sup>13</sup> C: -27.5, Duclos s. n., 1924)
B. rufus (Huds.) Link (Blysmopsis rufa (Huds.) Oteng-Yeboah)	$C_3$	JB (A: Stafleu 338; Laurer NSW 181496)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
Bolboschoenus cf. medianus (V. J. Cook) Soják, as Scirpus maritimus L.	$C_3$	Takeda et al. 1985 (A)
B. fluviatilis (Torr.) Soják, as Scirpus fluviatilis (Torr.) A. Gray <sup>a</sup>	$C_3$	Takeda et al. 1985 (A) <sup>a</sup> ; Ueno et al. 1986 (A. B) <sup>a</sup> ; JB (A: <i>Gray 3921</i> CANB)
B. maritimus (L.) Palla, as Scirpus maritimus L. <sup>a</sup>	$C_3$	Sabnis 1921 ([A]) <sup>a</sup> ; Govindarajalu 1976 ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); Mateu Andres 1991 ([A]) <sup>a</sup> ; LR (δ <sup>13</sup> C: −29.7, <i>Perrottet 818</i> )
B. robustus (Pursh) Soják, as Scirpus ro- bustus Pursh	$C_3$	Bender 1971 ( $\delta^{13}$ C)
Bulbostylis abortiva (Steud.) C. B. Clarke	$C_4$	LR (\delta^{13}C: -13.3, Tisserant 3161)
B. argenteobrunnea C. B. Clarke	$\mathbf{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
B. atrosanguinea (Boeck.) C. B. Clarke	$\mathbb{C}_4$	Hesla et al. 1982 (A. $\delta^{13}$ C)
B. barbata (Rottb.) C. B. Clarke	$\mathrm{C}_4$	Govindarajalu 1966 ([A]); Hesla et al. 1982 (δ¹³C); Takeda et al. 1985 (Af); Ueno et al. 1986 (Af. B); Ueno et al. 1988b (USf); Ueno and Takeda 1992 (A. Γ); JB (Af: <i>McKee 10720</i> CANB; <i>Bruhl 540</i> CANB)
B. basalis Fosberg	$\mathbb{C}_4$	Hnatiuk 1980 (A)
B. boeckeleriana (Schweinf.) Beetle	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C: $-10.7$ )
B. bozumensis Cherm.	$\mathbf{C}_4$	LR (δ <sup>13</sup> C: -13.8, <i>Badré 40</i> )
B. coleotricha (Hochst. ex A. Rich.) C. B. Clarke	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C); LR ( $\delta^{13}$ C: $-12.5$ , <i>Pobéguin 408</i> )
B. contexta (Nees) Bodard	$C_4$	Gordon-Gray 1971 ([A])
<ul> <li>B. densa (Wall. ex Roxb.) HandMazz., as</li> <li>B. capillaris (L.) C. B. Clarke subsp.</li> <li>trifida (Nees) Koyama<sup>a</sup>, as B. capillaris var. trifida (Nees) C. B. Clarke<sup>b</sup></li> </ul>	$\mathrm{C}_4$	Govindarajalu 1966 ([A]) <sup>b</sup> ; Hesla et al. 1982 ( $\delta^{13}$ C); Takeda et al. 1985 (Af. $\delta^{13}$ C); Ueno and Takeda 1992 (A) <sup>a</sup>
subsp. <i>afrimontana</i> (K. Lye) R. Haines, as <i>B. capillaris</i>	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
B. filamentosa (Vahl) C. B. Clarke, as B. cardiocarpa (Ridl.) C. B. Clarke	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
<ul><li>B. hispidula subsp. filiformis (C. B. Clarke)</li><li>R. Haines, as B. filiformis C. B.</li><li>Clarke</li></ul>	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
B. glaberrima Kük.	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
B. hispidula (Vahl) R. W. Haines, as Fimbristylis hispidula (Vahl) Kunth	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C); LR ( $\delta^{13}$ C: $-13.4$ , Chevalier 9243)
B. humilis (Kunth) C. B. Clarke	$\mathbf{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
B. lanata (Kunth) Lindm.	$\mathbb{C}_4$	JB (Af: <i>McKee 10720</i> CANB)
B. laniceps C. B. Clarke	$C_4$	LR (δ <sup>13</sup> C: -14.1, Chevalier 27599)
B. lanifera (Boeck.) Kük.	$C_4$	LR (δ <sup>13</sup> C: -12.6, Berhaut 2617)
B. mucronata C. B. Clarke	$\mathbb{C}_4$	LR ( $\delta^{13}$ C: $-10.6$ , Humbert 15943)
B. oligostachys (Hochst. ex A. Rich.) K. Lye, as Fimbristylis oligostachys	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
Hochst. ex A. Rich.	C	I.D. (\$13C) 12.0 Letour at (\$722)
B. oritrephes (Ridl.) K. Lye B. paradoxa (Spreng.) Lindm.	$C_4$	LR (8 <sup>13</sup> C: -12.0, <i>Letouzey 8733</i> ) JB (Af <sup>2</sup> , <i>McKee 11199</i> CANB)
B. pilosa (Willd.) Cherm.	$egin{array}{c} C_4 \ C_4 \end{array}$	Hesla et al. 1982 ( $\delta^{13}$ C); LR ( $\delta^{13}$ C: $-12.0$ , Boivin s. n., 1850)
B. puberula (Poir.) C. B. Clarke var. gracilis (Nees) Fisch.	$C_4$	Govindarajalu 1966 ([A])
var. puberula, as B. puberula	$C_4$	Govindarajalu 1966 ([A])
B. pusilla (Hochst. ex A. Rich.) C. B. Clarke	$\mathbf{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C); LR (δ <sup>13</sup> C: -16.0, <i>Le Testu 2868</i> )
B. schimperiana (A. Rich.) C. B. Clarke, as Fimbristylis humilis A. Peter	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
B. subspinescens C. B. Clarke	$\mathbb{C}_4$	Govindarajalu 1966 ([A])
B. swamyi Govindarajalu	$\mathbf{C}_4$	Govindarajalu 1985 ([A])
B. trichobasis (Baker) C. B. Clarke	$\mathbf{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
B. turbinata S. T. Blake	$C_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C)
B. vanderystii Cherm.	$\mathbb{C}_4$	LR (δ <sup>13</sup> C: -11.3, Chevalier 27314)
Calyptrocarya glomerulata (Brongn.) Urb.	$C_3$	JB (Af: Campbell MEL 1543844; Harris 438 K); LR (δ <sup>13</sup> C: -37.2, Pinto & Sastre 971)
Capeobolus brevicaulis (C. B. Clarke) J. Browning	$C_3$	JB (A: Bruhl 1720, 1736 NE; Moss 7612 K)
Capitularina involucrata (Valck. Sur.) Kern	$\mathbb{C}_3$	JB (A: van Royen 4005 CANB); LR (δ <sup>13</sup> C: -33.8, Hombron s. n., 1841)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
Carex alligata Boott, as C. sandwichensis [sic] Boeck.	$C_3$	Standley 1990 ([A])
C. appressa R. Br.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: <i>Bruhl 119</i> CANB; <i>Bruhl 15</i> CANB); LR (δ <sup>13</sup> C: -24.6, <i>Drummond 272</i> )
C. baccans Nees	$C_3$	Hofstra et al. 1972 (A. $\Gamma$ )
C. bequaertii De Wild.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. bohemica Schreb., as C. cyperoides L.	$C_3$	LR ( $\delta^{13}$ C: $-27.7$ , Duclos s. n., 1929)
C. breviculmis R. Br.	$C_3$	Takeda et al. 1985 (A)
C. camptoglochin Krech.	$C_3$	Kukkonen 1970 ([A])
C. capillacea Boott	$C_3$	JB (A: Adams 2619 CANB)
C. castanostachya K. Schum.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. cephalotes F. Muell.	$C_3$	JB (A: <i>Doing</i> , 8 Jan 1964 CANB; <i>Gray</i> 4785 CANB)
C. chlorosaccus C. B. Clarke		Hesla et al. 1982 ( $\delta^{13}$ C)
	$C_3$	
C. collumanthus (Steyerm.) L. E. Mora (Vesicarex collumanthus Steyerm.)	$C_3$	JB (A: Cleef 5611 U)
C. conferta Hochst. ex A. Rich.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
var. <i>lycurus</i> (K. Schum.) K. Lye, as C.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
lycurus K. Schum.	_	
C. cruciata Vahl	$C_3$	Saxena and Ramakrishnan 1984 (A)
C. cuchumatanensis Standley & Steyerm.	$\mathbb{C}_3$	Standley 1987b ([A]); Standley 1990 ([A])
C. curvula Allioni	$C_3$	Körner et al. 1988 ( $\delta^{13}$ C)
C. decidua Boott	$C_3$	Standley 1987 <i>b</i> ([A])
C. declinata Boott	$C_3$	Takeda et al. 1985 (A)
C. dietrichiae Boeck.	$C_3$	LR ( $\delta^{13}$ C: $-31.7$ , Schlechter 18277)
C. doniana Spreng., as C. japonica Thunb. subsp. chlorostachys (D. Don) Koya- ma	$C_3$	Ueno et al. 1986 (A. B)
C. duriuscula C. A. Mey., as C. eleocharis L. H. Bailey	$C_3$	Boutton et al. 1980 (A)
C. echinochloe Kunze	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. elgonensis Nelmes	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. emoryi Dewey	$C_3$	Standley 1987 <i>a</i> ([A])
C. ericetorum Pollich	$\mathbf{C}_3$	LR ( $\delta^{13}$ C: $-25.9$ , Lerman s. n., 1970)
C. erythrorrhiza Boeck.	$\mathbf{C}_3$	Hesla et al. 1982 (δ <sup>13</sup> C)
C. extensa Good.	$C_3$	Mateu Andres 1991 ([A])
C. fascicularis Solander ex Boott	$\mathbf{C}_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: <i>Gray 5825</i> CANB)
C. filifolia Nutt.	$C_3$	Boutton et al. 1980 (A)
C. flava L.	$C_3$	LR ( $\delta^{13}$ C: $-27.4$ , Lerman s. n., 1971)
C. fraserianus (Ker Gawl.) Kartesz & Gan- dhi (Cymophyllus fraseri (Andrews) Mack.)	$C_3$	JB (A: s. coll. MEL 154850)
C. grayi J. Carey	$C_3$	Hofstra et al. 1972 (A. $\Gamma$ )
C. haydenii Dewey	$C_3$	Standley 1987 <i>a</i> ([A])
C. hermannii Cochrane	$C_3$	Standley 1987 <i>b</i> ([A])
C. indica L.	$C_3$	LR (8 <sup>13</sup> C: -27.8, Zollinger 313)
C. inversa R. Br.	$C_3$	JB (A: <i>Moore 8135</i> CANB)
C. johnstonii Boeck.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. kobomugi Ohwi	$C_3$	Akita et al. 1969 ([A]); Takeda et al. 1980 (A. $\Gamma$ )
C. lacustris Willd.		Bender 1971 ( $\delta^{13}$ C)
	$C_3$	
C. lenticularis Michx. var. lenticularis	$C_3$	Standley 1987a ([A]); Standley 1990 ([A])
C. liparocarpos Gaudin	$C_3$	Kalapos et al. 1997 (A. $\Gamma$ )
C. maculata Boott	$C_3$	KW (A: Wilson 10201)
C. maritima Gunnerus, as C. incurva Lightfoot	$C_3$	LR ( $\delta^{13}$ C: $-26.9$ , Duclos s. n., 1938)
C. microglochin Wahlenb.	$C_3$	Kukkonen 1970 ([A])
C. monostachya A. Rich.	$C_3$	Hesla et al. 1982 (A. $\delta^{13}$ C); Aucour et al. 1994 ( $\delta^{13}$ C)
C. neoguinensis C. B. Clarke	$C_3$	Hofstra et al. 1972 (A. Γ); JB (A: Smith ANU 15115 CANB)
C. nigra (L.) Reichard	$C_3$	Standley 1987 <i>a</i> ([A])
C. obnupta L. H. Bailey	$\mathbf{C}_3$	Standley 1990 ([A])
C. pachystylis J. Gay	$C_3$	Winter and Troughton 1978 (δ <sup>13</sup> C); LR (δ <sup>13</sup> C: -33.1, Phytotron Gif cult., 1972)
	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
C. petitiana A. Rich., as C. cuprea (Kük.)  Nelmes <sup>a</sup> , as C. fischeri K. Schum. <sup>b</sup> , as  C. longipedunculata K. Schum. <sup>c</sup> , as C.	C <sub>3</sub>	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a,b,c,d</sup>
ninagongensis (Kük.) Robyns <sup>d</sup>	C	I.D. (\$13C). 20.2 Council a. r. 1012)
C. pilulifera L.	$C_3$	LR (8 <sup>13</sup> C: -30.3, Seyrat s. n., 1912)
C. polyantha F. Muell.	$C_3$	JB (A: Bruhl, 17 Nov 1985 CANB)
C. rafflesiana Boott	$C_3$	JB (A: <i>Bruhl 551</i> CANB); Hofstra et al. 1972 (A. Γ)
C. runssoroensis K. Schum.	$C_3$	Aucour et al. 1994 (δ <sup>13</sup> C); Hesla et al. 1982 (δ <sup>13</sup> C)
C. setifolia Kunze ex Kunth	$C_3$	LR ( $\delta^{13}$ C: $-31.9$ , Phytotron Gif cult., 1972)
C. simensis A. Rich.	$C_3$	Hesla et al. 1982 (8 <sup>13</sup> C)
C. sp.	$C_4^+$	Smith and Epstein 1971 ( $\delta^{13}$ C)
C. sp.	$C_3$ $C_3$	Troughton et al. 1974 ( $\delta^{13}$ C) Boutton et al. 1980 (A)
C. sp.		SCV (δ <sup>13</sup> C: -27.72, Schlechter 4759)
C. spicatopaniculata C. B. Clarke	$C_3$	
C. stenophylla Wahlenb.	$C_3$	Williams and Monson 1981 ([A], Γ)  Pender 1071 (\$13C): Standley 1087α ([A])
C. stricta Lam.	$C_3$	Bender 1971 ( $\delta^{13}$ C); Standley 1987 <i>a</i> ([A])
C. strigosa Huds.	$C_3$	LR (δ <sup>13</sup> C: -30.1, <i>De Vergnes s. n.</i> , 1902) SCV (δ <sup>13</sup> C: -25.18, <i>Dod 3467</i> )
C. subinflata Nelmes	$C_3$	
C. torta Boott	$C_3$	Standley 1987 <i>a</i> ([A]); Standley 1990 ([A])
C. vallis-rosetto K. Schum., as C. green- wayi Nelmes	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. verticillata Zoll. & Moritzi	$C_3$	Hofstra et al. 1972 (A. Γ)
C. vesicaria L.	$C_3$	Shepherd 1976 ([A])
Carpha alpina R. Br.	C <sub>3</sub>	Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: <i>Craven 1770</i> CANB; <i>NGF 10208</i> CANB); LR (δ <sup>13</sup> C: -26.6, <i>Le Guillou s. n.</i> , 1840)
C. nivicola F. Muell.	$C_3$	JB (A: Totterdell 373 CANB)
Caustis blakei Kük.	$C_3$	Takeda et al. 1985 (A)
C. dioica R. Br.	$C_3$	Takeda et al. 1985 (A)
C. flexuosa R. Br.	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C); JB (A: <i>Bruhl 130</i> CANB)
C. pentandra R. Br.	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C); LR ( $\delta^{13}$ C: $-30.7$ , <i>Rodway 1558</i> )
C. recurvata Spreng.	$C_3$	Takeda et al. 1985 (A)
Cephalocarpus rigidus Gilly	$C_3$	JB (A: Maguire 32831U); LR (δ <sup>13</sup> C: -27.7, Maguire et al. 30159)
C. australis K. L. Wilson C. cymbaria R. Br.	C <sub>3</sub> C <sub>3</sub>	<ul> <li>KW (A: Bates 4022; Beauglehole 68256; Williamson NSW 122681)</li> <li>Takeda et al. 1985 (A); JB (A: Bruhl 101 CANB; Prober 243 CANB); KW</li> <li>(A: Beauglehole 24875; Constable 6204, 4443; Coveny 3832, 5050, 6236, 6693, 9093; Jacobs 3229; Johnson NSW 241350; McBarron 9276; McKee 30542, 43984 NOU; Olsen 2017; Rodway 268; Schmid 3080 NOU; Wilson 3053, 3086, 3094); LR (δ¹³C: -27.4, Raynal &amp; Jaffré 16459)</li> </ul>
C. enodis Nees	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); KW (A: <i>Blake 20736</i> ; <i>Coveny 8102</i> ; <i>Gunn 1401</i> ; <i>Maiden</i> NSW 22493; <i>Melville 1612</i> ; <i>Walter</i> NSW 242326; <i>Wilson 2773</i> )
C. multiarticulata Nees	$C_3$	KW (A: Conn 3545; P.G. Wilson 7069 PERTH)
C. sphaerocephala R. Br.	$\mathbf{C}_3$	KW (A: Beauglehole 32890; Brooks NSW 242163; Camfield NSW 2249, NSW 22496; Coveny 831, 3755, 5025, 6283, 6704; Constable NSW 46244; Maiden NSW 242388; McBarron 8208; Rodway NSW 242331, NSW 242387)
Chorizandra sp. G (Wilson 7192)	$C_3$	KW (A)
Chrysitrix capensis L.	$\mathbf{C}_3$	JB (A: Williams 3240 PRE); LR (δ <sup>13</sup> C: -27.9, Humbert 9600)
C. dodii C. B. Clarke	$\mathbf{C}_{3}^{J}$	JB (A: Esterhuysen 2917 PRE)
C. junciformis C. B. Clarke	$\mathbf{C}_3$	JB (A: Taylor 3888 PRE); SCV (δ <sup>13</sup> C: -23.33, Stokoe s. n.)
Cladium jamaicense Crantz, as C. mariscus (L.) Pohl subsp. jamaicense (Crantz) Kük.a, as C. mariscus var. jamaicense (Crantz) ined.b	C <sub>3</sub>	Bender 1971 ( $\delta^{13}$ C); Hesla et al. 1982 ( $\delta^{13}$ C); Aucour et al. 1994 ( $\delta^{13}$ C) <sup>b</sup> ; LR ( $\delta^{13}$ C: $-26.0$ , Degener 27989) <sup>a</sup>
C. mariscus (L.) Pohl	$C_3$	Aucour et al. 1994 ( $\delta^{13}$ C)
C. procerum S. T. Blake <sup>a</sup> , as C. mariscus <sup>b</sup>	$\mathbf{C}_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C) <sup>a,b</sup> ; JB (A: Crisp 6878 CBG)
Coleochloa abyssinica (A. Rich.) Gilly, as Eriospora abyssinica Hochst. ex A. Rich. <sup>a</sup>	$C_3$	Chermezon 1930 ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); LR (δ <sup>13</sup> C: -26.1, <i>Dillon &amp; Pettit s. n.</i> , 1842)
C. schweinfurthiana (Boeck.) Nelmes C. setifera (Ridl.) Gilly, as Eriospora seti- fera (Ridl.) C. B. Clarke <sup>s</sup>	C <sub>3</sub> C <sub>3</sub>	JB (A: s. coll. MEL 1543821) Chermezon 1930 ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); SCV (δ <sup>13</sup> C: -22.20, Esterhuysen 21464)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
Costularia arundinacea (Solander ex Vahl) Kük. (Lophoschoenus arundinaceus (Solander ex Vahl) Stapf)	$C_3$	LR (\delta^{13}C: -27.7, Raynal & Jaffré 16508)
C. chamaedendron (Guill.) Kük.	$C_3$	JB (A: <i>McPherson 3055</i> ); LR (δ <sup>13</sup> C: -29.6, <i>Raynal &amp; Jaffré 16514</i> , -30.4, Serre Orsay cult., 1972)
C. comosa (C. B. Clarke) Kük. (Lophos- choenus comosus (C. B. Clarke)	$C_3$	JB (A: <i>McKee</i> 7726 BRI)
Stapf)	C	ID (A. Handlan 15075 CAND)
C. fragilis (Däniker) Kük.	$C_3$	JB (A: <i>Hartley 15075</i> CANB)
C. leucocarpa (Ridl.) H. Pfeiff. C. pantopoda (Baker) C. B. Clarke	$C_3$ $C_3$	JB (A: Bossa 7773 K) LR (δ¹³C: -25.1, Humbert 22674)
C. pilisepala (Steud.) Kern, as C. urvilleana (Gaud. ex Boeck.) Kük.	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C)
C. pubescens J. Raynal (Lophoschoenus pubescens (J. Raynal) ined.)	$C_3$	JB (A: McKee 1051A BRI)
C. stagnalis (Däniker) Kük.	$C_3$	JB (A: Hartley 15072 CANB)
Courtoisina assimilis (Steud.) Maquet, as Mariscus assimilis (Steud.) Podlech	$C_3$	Hesla et al. 1982 (δ <sup>13</sup> C); Vorster 1996 (A)
C. cyperoides (Roxb.) Soják	$C_3$	Druyts-Voets 1970 ([A]); Vorster 1996 (A); JB (A: MEL 1543845, MEL 1543847); LR (δ <sup>13</sup> C: -28.2, <i>Fotius 931</i> )
Crosslandia setifolia W. V. Fitzg., as Crosslandia <sup>a</sup>	$\mathrm{C}_4$	Lerman and Raynal 1972 (A) <sup>a</sup> ; Takeda et al. 1985 (Af. δ <sup>13</sup> C); JB (Af: <i>van Rijn 19</i> CANB; <i>Latz 2774</i> CANB; <i>Leutert 74</i> CANB); LR (δ <sup>13</sup> C: –11.5, <i>MacKee 8432</i> )
Cyathochaeta avenacea (R. Br.) Benth.	$C_3$	Takeda et al. 1985 (A); JB (A: <i>Morrison</i> , 3 Dec 1903 BRI. δ <sup>13</sup> C: -26.3, <i>Morrison</i> , 3 Dec 1903); LR (δ <sup>13</sup> C: -25.9, <i>Morrison s. n.</i> , 1915)
C. diandra (R. Br.) Nees	$C_3$	Takeda et al. 1985 (A)
Cyathocoma hexandra (Nees) J. Browning, as Macrochaetium hexandrum (Nees) H. Pfeiff. <sup>a</sup>	$C_3$	JB (A: Garside, 10 Oct 1920 K); LR (δ <sup>13</sup> C: -29.3, Drège 3944) <sup>a</sup>
Cyperus acuminatus Torr. & Hook.	$C_3$	Denton 1983 (A); Li et al. 1999 (A)
C. aggregatus (Willd.) Endl.	$C_4$	KW (Ac: Jacobs NSW 144403)
	$C_3$ +	Li et al. 1999 (A)
C. ajax C. B. Clarke	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. albopilosus (C. B. Clarke) Kük., as  Mariscus albopilosus C. B. Clarke	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. albosanguineus Kük., as Mariscus albosanguineus (Kük.) Napper	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. albostriatus Schrad.	$C_3$	KW (A: Schlechter 2569; Wilson 4383)
C. alopecuroides Rottb.	$\begin{array}{c} \mathrm{C_4} + \\ \mathrm{C_4} \end{array}$	Sonnenberg and Botha 1992 (A. PIB: NADP) Druyts-Voets 1970 ([Ac]); Hesla et al. 1982 (δ <sup>13</sup> C); Li and Jones 1994 (A)
C. alterniflorus R. Br.	$C_4$	KW (Ac: <i>Martensz</i> 263)
C. alternifolius L.	C <sub>3</sub>	Haberlandt 1884 ([A]); Rikli 1895 ([A]); Brown 1975 (A); Takeda et al. 1980 (A. Γ); Hesla et al. 1982 (δ¹³C); Krenzer et al. 1975 (Γ); Li 1993 (A. Γ. δ¹³C); Li and Jones 1994 (A); LR (δ¹³C: -30.3, <i>Perrier 14816</i> , 35.5, Serre Orsay cult., 1972)
C. amabilis Vahl	$C_4$	Druyts-Voets 1970 ([A]); Meinzer 1978 (A); Hesla et al. 1982 (δ <sup>13</sup> C); KW (Ac: <i>Adam 2258</i> P; <i>de la Bâthie 13097</i> ); LR (δ <sup>13</sup> C: -13.0, <i>Boivin s. n.</i> ); SCV (δ <sup>13</sup> C: -12.31, <i>Schupers 893</i> )
C. amauropus Steud., as Mariscus amauropus (Steud.) Cufod. <sup>a</sup>	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: s. coll. 1301, Kenya P); LR (δ <sup>13</sup> C: –14.0, Humbert 8502 bis) <sup>a</sup>
C. amuricus Maxim.	$C_4$	Ueno and Takeda 1992 (A); KW (Ac: Inamasu 425)
C. anderssonii Boeck.	$\mathbf{C}_4$	KW (Ac: Schimpff 13 P)
C. andinus Palla	$C_3$	LR (\delta^{13}C: -26.5, Mandon 1396)
C. angolensis Boeck.	$\mathbf{C}_4$	Druyts-Voets 1970 ([A]); KW (Ac: Robinson 2683 P)
C. angustatus R. Br.	$\mathbf{C}_4$	KW (Ac: <i>Latz 3651</i> )
C. aquatilis R. Br.	$C_3$	Carolin et al. 1977 (US); Takeda et al. 1985 (A. $\delta^{13}$ C)
C. articulatus L.	$C_4$	Druyts-Voets 1970 ([Ac]); Hesla et al. 1982 (δ <sup>13</sup> C); Meinzer 1978 (A)
C. astartodes K. L. Wilson	$C_4$	KW (Ac: Specht 649; Wilson 5151)
C. atractocarpus Ridl. C. atroviridis C. B. Clarke, as C. aterrimus	$egin{array}{c} C_4 \ C_4 \end{array}$	Druyts-Voets 1970 ([A]) Druyts-Voets 1970 ([A]); Hesla et al. 1982 ( $\delta^{13}$ C)
Steud.  C. aucheri Jaub. & Spach	$C_4$	KW (Ac: H. Wilson 4, Arabia P)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
C. baoulensis (Chevalier) Kük.	$C_4$	KW (Ac: Hall 306 P)
C. bellus Kunth [may be referable to C.	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
tanganyicanus (Kük.) K. Lye] C. bernieri Cherm.	C	KW (Ac: Boivin Voyage 1847–52, 2320 P)
C. betchei (Kük.) S. T. Blake subsp. betchei	$egin{array}{c} { m C}_4 \\ { m C}_4 \end{array}$	KW (Ac. Bolvin Voyage 1647–22, 2520 1) KW (Ac: Payne 4; Wilson 746)
subsp. commiscens K. L. Wilson	$C_4$ $C_4$	KW (Ac: Latz 7090)
C. bifax C. B. Clarke, as Cyperus rotundus	$C_4$	Carolin et al. 1977 (USc); Druyts-Voets 1970 ([A]) <sup>a</sup> ; Takeda et al. 1985 (Ac.
subsp. <i>retzii</i> <sup>a</sup>	<b>C</b> <sub>4</sub>	$\delta^{13}$ C)
C. blakeanus K. L. Wilson	$\mathbf{C}_4$	KW (Ac: Latz 2200; Wilson 5373)
C. blysmoides C. B. Clarke	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. bowmanii F. Muell. ex Benth.	$\mathbf{C}_4$	Carolin et al. 1977 (USc); Takeda et al. 1985 (Ac. $\delta^{13}$ C)
C. bulbosus Vahl	$C_4$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 (δ <sup>13</sup> C); Takeda et al. 1985 (Ac); KW (Ac: <i>O'Connell</i> NSW 121327); LR (δ <sup>13</sup> C: -14.0, <i>Audru 2536</i> )
C. burkartii Guaglianone	$C_3$	Guaglianone 1990 ([A])
C. callistus Ridl.	$C_4$	Druyts-Voets 1970 ([A]); KW (Ac: Gossweiler 209 P)
C. cancrorum Cherm.	$\mathbf{C}_4$	KW (Ac: Decary 1686 P)
C. capensis (Steud.) Endl. var. capensis	$C_4$	KW (Ac: Schlechter 3779)
C. capitatus Vandelli	$C_4$	Collins and Jones 1985 (δ <sup>13</sup> C); Li 1993 (A. δ <sup>13</sup> C); JB (Ac: <i>s. coll.</i> MEL 1543828); LR (δ <sup>13</sup> C: -11.1, <i>Mabille Hb Cors.</i> 96)
C. carinatus R. Br.	$C_4$	KW (Ac: Latz 5176; Wilson 4669)
C. castaneus Willd.	$C_4$	Carolin et al. 1977 (USc); Takeda et al. 1985 (Ac. $\delta^{13}$ C)
C. centralis K. L. Wilson	$\mathbf{C}_4$	KW (Ac: Latz 1911, 2012)
C. cephalotes Vahl (Anosporum cephalotes (Vahl) Kurz)	$C_3$	Hesla et al. 1982 (8 <sup>13</sup> C); JB (A: <i>Dharmawardhana 25</i> CANB; <i>Pullen 7523</i> CANB); KW (A: <i>Schmid s. n.</i> , Indochina P)
C. chalaranthus J. Presl & C. Presl	$C_3$	KW (A: Lescure 604 P)
C. chordorrhizus Chiovenda	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. chrysocephalus (K. Schum.) Kük.	$C_4$	KW (Ac: Baum 311a BRI; de Witte 7185 PRE)
C. circumclusus (C. B. Clarke) Kük., as Mariscus circumclusus C. B. Clarke <sup>a</sup> ,	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a,b</sup> ; KW (Ac: <i>Lewalle s. n.</i> , Burundi P); LR (δ <sup>13</sup> C: –11.1 <i>Tisserant 1877</i> ) <sup>b</sup>
as M. macropus C. B. Clarke <sup>b</sup>	-	TWY (4 - DI 1 - 515 ( TW) - 55 (0)
C. clarus S. T. Blake	$C_4$	KW (Ac: Blake 5174; Wilson 5749)
C. clavinux C. B. Clarke	$C_4$	KW (Ac: Fotius 1603)
C. colymbetes Kotschy & Peyr.	$C_3$	Druyts-Voets 1970 ([A]) Takada et al. 1985 (Ae): Hang and Takada 1992 (A)
C. compactus Retz., as Mariscus compac- tus (Retz.) Boldingh <sup>a</sup>	$C_4$	Takeda et al. 1985 (Ac); Ueno and Takeda 1992 (A) <sup>a</sup>
C. compressus L.	$\mathrm{C}_4$	Druyts-Voets 1970 ([Ac]); Hofstra et al. 1972 (A. Γ); Prakash et al. 1976 (A); Hesla et al. 1982 (δ¹³C); Takeda et al. 1985 (Ac); Ueno et al. 1986 (Ac. B); Ueno and Takeda 1992 (A. Γ); Lin et al. 1993 (Ac); KW (Ac: Henty NGF9870); LR (δ¹³C: -9.2, Boivin s. n., 1847); SCV (δ¹³C: -9.85, de Winter and Giess 6889)
C. concinnus R. Br.	$C_3$	KW (A: Wilson 1463)
C. confertus Sw.	$\mathbf{C}_4$	Rikli 1895 ([A])
C. congensis C. B. Clarke	$\mathbf{C}_4$	Druyts-Voets 1970 ([A])
C. congestus Vahl, as Mariscus congestus (Vahl) C. B. Clarke <sup>a</sup>	$\mathrm{C}_4$	Takeda et al. 1985 (Ac. δ <sup>13</sup> C); Li 1993 (A. Γ. δ <sup>13</sup> C); Li and Jones 1994 (A); Sonnenberg and Botha 1992 (A. PIB: NADP) <sup>a</sup> ; KW (Ac: <i>Wilson 1442</i> ); SCV (δ <sup>13</sup> C: -9.56, <i>Mogg 11683</i> ) <sup>a</sup>
C. conglomeratus Rottb.	$\mathbf{C}_4$	Sabnis 1921 ([A]); Druyts-Voets 1970 ([A]); Hnatiuk 1980 (A); KW (Ac: <i>de Fabrègues 2759</i> P); LR (δ <sup>13</sup> C: -11.6, <i>Jamin s. n.</i> , 1852)
C. conicus (R. Br.) Boeck.	$C_4$	Carolin et al. 1977 (USc); Takeda et al. 1985 (Ac); KW (Ac: <i>Wilson 1502</i> , 3529)
C. constanzae Urban	$C_3$	KW (A: Ekman 6879 K; Harris 12350 K)
C. cornelii-ostenii Kük.	$\mathbb{C}_4$	KW (Ac: Krapovickas 24323 P)
C. corymbosus Rottb., as C. corymbosus var. longispiculatus (O. Kuntze) Kük. <sup>a</sup>	$C_4$	Mani 1963 ([A]) <sup>a</sup> ; SCV (δ <sup>13</sup> C: -10.33, <i>Maputaland Expedition 14319</i> )
C. cracens K. L. Wilson	$\mathbb{C}_4$	KW (Ac: Benson 2088b; Craven 5826)
C. crassipes Vahl, as C. maritimus Poir.a	$C_4$	Druyts-Voets 1970 ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ1 <sup>3</sup> C) <sup>a</sup> ; KW (Ac: <i>Adam 1538</i> P); LR (δ1 <sup>3</sup> C: -12.0, <i>Mahoux SF6764</i> ) <sup>a</sup>
C. crispulus K. L. Wilson	$C_4$	KW (Ac: Blake 17673; Dunlop 5240; Wilson 5303)
C. croceus Vahl (C. globulosus Aublet)	$C_4$	KW (Ac: Schallert 28257 P)
C. cunninghamii (C. B. Clarke) C. Gardner subsp. cheradicus K. L. Wilson	$C_4$	KW (Ac: Wilson 5454)
subsp. cunninghamii, as C. cunninghamii <sup>a</sup>	$C_4$	Takeda et al. 1985 (Ac. δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Coveny 517</i> ; <i>Latz 9499</i> ; <i>Mitchell 453</i> ; <i>Tate</i> NSW 22742; <i>Wilson 5379</i> )

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
subsp. uniflorus K. L. Wilson	$C_4$	KW (Ac: Wilson 5302)
C. cuspidatus Kunth	$C_4$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 (δ¹³C); Takeda et al. 1985 (Ac); KW (Ac: <i>de la Bâthie 16099</i> )
C. cyperinus (Retz.) Valck. Sur.	$\mathbf{C}_4$	Mani 1963 ([A]); Hofstra et al. 1972 (A. Γ)
C. cyperoides (L.) Kuntze, as Mariscus sie-	$\mathbf{C}_4$	Hofstra et al. 1972 (A. $\Gamma$ ); Hesla et al. 1982 ( $\delta^{13}C$ ) <sup>a</sup> ; Ueno and Takeda 1992
berianus Nees ex C. B. Clarke <sup>a</sup> , as M. sumatrensis (Retz.) J. Raynal <sup>b</sup>		$(A. \Gamma)^b$
C. dactylotes Benth.	$C_4$	Takeda et al. 1985 (Ac. δ <sup>13</sup> C); KW (Ac: <i>Latz 5126</i> ; <i>Martensz 4134</i> )
C. deciduus Boeck.	$C_3$ +	Druyts-Voets 1970 ([A]); Vorster 1990 ([A]); KW (A: Angus 2987 P; Miller 5634; Robinson 4020 P)
C. decompositus (R. Br.) F. Muell.	$\mathbf{C}_4$	Takeda et al. 1985 (Ac. δ <sup>13</sup> C); KW (Ac: <i>Tryon</i> NSW 608854)
C. dentatus Torr.	$C_3$	Li et al. 1999 (A. δ <sup>13</sup> C); KW (A: Fernald 16273)
C. denudatus L. f.a, as C. phaeorrhizus K. Schum.b, as C. phaeorrhizus var. filifolia ined.c	$C_3$	Druyts-Voets 1970 ([A]) <sup>a,b,c</sup> ; Hesla et al. 1982 (A. $\delta^{13}$ C) <sup>a,b</sup> ; Aucour et al. 1994 ( $\delta^{13}$ C) <sup>a</sup>
C. dereilema Steud.a, as C. dereilema var. deckenii (Boeck.) ined.b	$C_3$	Druyts-Voets 1970 ([A]) <sup>a,b</sup> ; Hesla et al. 1982 ( $\delta^{13}$ C) <sup>a</sup>
C. dichroostachyus A. Rich.	$C_3$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 (δ <sup>13</sup> C)
C. dietrichiae Boeck.	$\mathbf{C}_4$	KW (Ac: Flecker NSW 608855)
C. difformis L.	$C_3$	Akita et al. 1969 ([A]); Druyts-Voets 1970 ([A]); Hofstra et al. 1972 (A. Γ); Imai and Murata 1979 (Γ); Takeda et al. 1980 (A. Γ); Hesla et al. 1982 (δ <sup>13</sup> C); Takeda et al. 1985 (A); Li 1993 (A. Γ. δ <sup>13</sup> C); Li et al. 1999 (A. δ <sup>13</sup> C); KW (A: <i>Wilson 1464</i> )
C. digitatus Roxb.	$\mathbf{C}_4$	Ueno and Takeda 1992 (A); LR (δ <sup>13</sup> C: -13.4, <i>Drummond 6576</i> )
subsp. <i>auricomus</i> (Sieb. ex Spreng.) Kük., as <i>C. auricomus</i> Sieb. ex Spreng.	$\mathrm{C}_4$	Druyts-Voets 1970 ([A]); Li 1993 (δ¹³C)
C. disjunctus C. B. Clarke	$C_3$	KW (A: Wilson 9907)
C. distans L. f.a, as C. distans var. densiflorus Kük.b	$C_4$	Druyts-Voets 1970 ([A]) <sup>a,b</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; Ueno et al. 1986 (Ac. B) <sup>a</sup> ; Ueno et al. 1988 <i>b</i> (USc) <sup>a</sup> ; Ueno and Takeda 1992 (A) <sup>a</sup> ; KW (Ac: <i>Berhaut 3611</i> P; <i>Flecker</i> NSW 65486; <i>Wilson 3805</i> ) <sup>a</sup>
C. distinctus Steud.	$C_3$	Denton 1983 (A)
C. diurensis Boeck., as Mariscus diurensis (Boeck.) C. B. Clarke <sup>a</sup>	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Humbert 7234</i> P)
C. dives Del. <sup>a</sup> , as C. immensus C. B. Clar-ke <sup>b</sup>	$\mathrm{C}_4$	Druyts-Voets 1970 ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a,b</sup> ; KW (Ac: <i>Berhaut 5100</i> P) <sup>a</sup> ; LR (δ <sup>13</sup> C: −13.0, <i>Schlieben 1274</i> ) <sup>b</sup>
C. drummondii Torr. & Hook., as C. virens var. drummondii (Torr. & Hook.) Kük.	$C_3$	Denton 1983 (A)
C. dubius Rottb., as Mariscus dubius (Rottb.) Kük. ex Fisch. <sup>a</sup>	$C_4$	Hnatiuk 1980 (A): Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Vorster 2566</i> ); LR (δ <sup>13</sup> C: -12.7, <i>Chevalier 21665</i> P) <sup>a</sup>
C. duripes I. M. Johnst.	$C_4$	KW (Ac: Anthony 400 US)
C. durus Kunth, as Mariscus durus (Kunth) C. B. Clarke	$C_4$	Vorster 1990 (A)
C. echinatus (L.) Alph. Wood, as C. ovu- laris (Michx.) Torr.	$C_4$	Li 1993 (δ <sup>13</sup> C)
C. ekmannii Kük.	$C_4$	KW (Ac: Ekman 14980 US)
C. elatus L. C. elegans L.	$egin{array}{c} C_4 \ C_4 \end{array}$	Mani 1960 ([A]) Druyts-Voets 1970 ([A]); KW (Ac: <i>Pringle 5946</i> ; <i>Gentle 801</i> ); LR (δ <sup>13</sup> C: –11.3, <i>Rodriguez 3200</i> )
C. entrerianus Boeck.	$C_3$	-11.5, <i>Roanigue</i> 2.200) LR (δ <sup>13</sup> C: -26.6, <i>Hassler 7866</i> )
C. eragrostis Vahl, as C. vegetus Willd. <sup>a</sup>	C <sub>3</sub>	Druyts-Voets 1970 ([A]); Denton 1983 (A); Bruhl et al. 1987 (A. B); Li 1993 (A. Γ. δ <sup>13</sup> C); Li and Jones 1994 (A) <sup>a</sup> ; Soros and Dengler 2001 (A); JB (A: <i>Bruhl 658</i> CANB; <i>Ferreira 63</i> BRI; <i>Tryon</i> BRI 186483. Γ: 42, 46, <i>Bruhl 658</i> . δ <sup>13</sup> C: -26.6, <i>Tryon</i> BRI 186483); KW (A: <i>Wilson</i>
	$C_4+$	633); LR ( $\delta^{13}$ C: $-26.8$ , Duffort SEFFH 1649, $-27.6$ , Brown 119) Downton and Tregunna 1968 ( $\Gamma$ ); Troughton et al. 1974 ( $\delta^{13}$ C)
C. erythrorhizos Muhl.	$C_4$ $C_4$	Li et al. 1999 (A)
C. esculentus L.	C <sub>4</sub>	Moss et al. 1969 (Γ); Chen et al. 1970 (Γ); Druyts-Voets 1970 ([A]); Syvertsen et al. 1976 (A); Krenzer et al. 1975 (Γ); Hesla et al. 1982 (δ <sup>13</sup> C); L 1993 (Γ. δ <sup>13</sup> C); Li et al. 1999 (A. δ <sup>13</sup> C); JB (Ac: <i>Everist 6052</i> BRI); KW (Ac: <i>Johnson 7692</i> ); LR (δ <sup>13</sup> C: -12.1, <i>Sellier Sté Roch. 4501</i> P)
C. exaltatus Retz.	$C_4$	Mani 1963 ([A]); Hesla et al. 1982 (8 <sup>13</sup> C); Takeda et al. 1985 (Ac); JB (Ac: <i>s. coll.</i> CANB 112270); KW (Ac: <i>Solling 496</i> )

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
subsp. iwasakii (Makino) T. Koyama	$C_4$	Ueno and Takeda 1992 (A)
C. exilis Willd. ex Kunth	$C_4$	KW (Ac: Bosser 15824 P)
C. fastigiatus Rottb.	$C_4$	Sonnenberg and Botha 1992 (A. PIB: NAD/PCK); KW (Ac: Burchell 1773)
C. fertilis Boeck.	$C_3$	Druyts-Voets 1970 ([A]); LR (δ <sup>13</sup> C: -33.7, Serre Orsay cult., 1972 P, -36.6, <i>de Witte 7660</i> )
C. filiculmis Vahl	$C_4$	Bender 1971 ( $\delta^{13}$ C); Li et al. 1999 (A. $\delta^{13}$ C)
C. fischerianus A. Rich.	$C_3$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 (δ <sup>13</sup> C)
C. fissus Steud.	$C_4$	Druyts-Voets 1970 ([A])
C. flaccidus R. Br.	$C_3$	Ueno and Takeda 1992 (A. Γ); KW (A: Coveny 4861)
C. flexuosus Vahl (Torulinium flexuosum (Vahl) T. Koyama)	$\mathrm{C}_4$	KW (Ac: Wilson 2399)
C. foliaceus C. B. Clarke	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. frerei C. B. Clarke	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. fucosus K. L. Wilson, as C. angustatus [voucher re-determined at NSW by KLW]	$C_4$	Takeda et al. 1985 (A. δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: Wilson 5501)
C. fulgens C. B. Clarke, as C. fulgens var. fulgens <sup>a</sup>	$C_4$	Druyts-Voets 1970 ([A]); SCV (δ <sup>13</sup> C: -11.41, Rev. Lawson s. n.) <sup>a</sup>
C. fuligineus Chapm.	$C_4$	KW (Ac: Shafer 2488 P)
C. fulvus R. Br.	$C_4$	Carolin et al. 1977 (USc); KW (Ac: Coveny 3916; Wilson 5820, 5826)
C. fuscus L.	$C_3$	Li 1993 (A. $\delta^{13}$ C) Kalapos et al. 1997 (A. Γ. $\delta^{13}$ C); LR ( $\delta^{13}$ C: $-28.1$ , Bec in Arènes 3742)
C. gardneri Nees var. gardneri	$C_3$	KW (A: van Hermann 107 P)
var. vegetior Kük.	$C_3$	KW (A: Wilson 1344 P)
C. giganteus Vahl	$C_4$	Rodrigues and Estelita 2003 (A); LR (δ <sup>13</sup> C: -12.1, <i>Glaziou s. n.</i> , 1880)
C. gilesii Benth., as C. aff. gilesii <sup>a</sup>	$C_4$	Carolin et al. 1977 (USc) <sup>a</sup> ; Takeda et al. 1985 (Ac. δ <sup>13</sup> C); KW (Ac: <i>Mills &amp; Cox 23; Milthorpe &amp; Cunningham 1725</i> )
C. glaber L., as Chlorocyperus glaber (L.) Palla <sup>a</sup>	$\mathrm{C}_4$	Collins and Jones 1985 ( $\delta^{13}$ C); Li 1993 (A. $\delta^{13}$ C); Kalapos et al. 1997 (A. $\delta^{13}$ C) <sup>a</sup>
	$C_3$ +	Li 1993 (A. $\delta^{13}$ C)
C. glaucophyllus Boeck.	$C_3$	Druyts-Voets 1970 ([A])
C. glomeratus L., as Chlorocyperus glomeratus (L.) Palla <sup>a</sup>	$C_4$	Collins and Jones 1985 (δ <sup>13</sup> C); Ueno and Takeda 1992 (A); Li 1993 (A); Kalapos et al. 1997 (A. δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Licent 1671</i> P)
a	$C_3$ +	Li 1993 (A)
C. gracilis R. Br.	C <sub>3</sub>	JB (A: <i>Bruhl 519</i> CANB. Γ: 40, <i>Bruhl 519</i> ); LR (δ <sup>13</sup> C: -29.2, <i>Schmid 3472</i> , -31.2, Serre Orsay cult., 1972)
C. grandibulbosus C. B. Clarke (C. giolii Chiovenda)	$C_4$	KW (Ac: Mwangangi 600 P)
C. grandis C. B. Clarke	$\mathbf{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. grayi Torr.	$\mathbb{C}_4$	Li et al. 1999 (A. $\delta^{13}$ C3); KW (Ac: <i>Dowell 6014</i> )
C. grayoides Mohl., as C. "grayioides" [sic]	$C_4$	Li et al. 1999 (A. δ <sup>13</sup> C)
C. gunnii Hook. f. subsp. gunnii, as Mariscus gunnii (Hook. f.) C. B. Clarke <sup>a</sup>	$C_4$	Bruhl et al. 1987 (Ac. B) <sup>a</sup> ; JB (Ac: <i>Bruhl 29 CANB</i> ); KW (Ac: <i>Salasoo 2996</i> ; <i>Wilson 4422, 4439</i> )
subsp. <i>novaehollandiae</i> (Boeck.) K. L. Wilson	$\mathrm{C}_4$	KW (Ac: Wilson 3789)
C. gymnocaulos Steud.	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C); JB (A: Martensz 3249 CANB)
C. hamulosus M. Bieb. ("Monandrus hamulosus" (M. Bieb.) ined.)	$C_4$	Li 1993 (A. δ <sup>13</sup> C); JB (Ac: <i>Paun</i> MEL 11543826; <i>Smith 1138</i> PRE); KW (Ac: <i>Ramsay 8</i> MEL)
C. haspan L.	$C_3$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 ( $\delta^{13}$ C); Ueno et al. 1986 (A. B); Ueno and Takeda 1992 (A. $\Gamma$ )
C. hemisphaericus Boeck., as Mariscus hemisphaericus (Boeck.) C. B. Clarke <sup>a</sup>	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Pawek 6514</i> P; <i>Wilson 2014</i> )
C. hensii C. B. Clarke	$C_4$	Druyts-Voets 1970 ([A]); KW (Ac: Chevalier 11242 P)
C. hermaphroditus (Jacq.) Standley, as C. incompletus (Jacq.) Link	$C_4$	Rikli 1895 ([Ac])
C. hesperius K. L. Wilson	$\mathbb{C}_4$	KW (Ac: Beauglehole 11352, 48650)
C. hieronymi Boeck.	$\mathbf{C}_{3}$	KW (A: Venturi 6841 US)
C. hillebrandii Boeck.	$C_4$	KW (Ac: Hillebrand NSW 608853)
C. holoschoenus R. Br.	$C_4$	Carolin et al. 1977 (USc); Takeda et al. 1985 (Ac. δ <sup>13</sup> C); KW (Ac: <i>Blake</i> 12556, 16301)
C. holostigma C. B. Clarke ex Schweinf.	$C_4$	KW (Ac: Kelly 129 P)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
C. holstii Kük.	$\mathbf{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C); LR (δ <sup>13</sup> C: -10.5, Sacleux 2310)
C. houghtonii Torr.	$C_4$	Li et al. 1999 (A. δ <sup>13</sup> C); KW (Ac: <i>Umbach 2470</i> )
C. humilis Kunth	$C_3$	KW (A: von Turkheim NSW 608978); LR (δ <sup>13</sup> C: -28.0)
C. hystricinus Fernald	$C_4$	Li et al. 1999 (A)
C. imbricatus Retz.	$\mathbf{C}_4$	Mani 1963 ([A]); Druyts-Voets 1970 ([A]); Ueno and Takeda 1992 (A); LR (δ <sup>13</sup> C: -13.1, <i>Le Testu 2847</i> )
C. impubes Steud., as Mariscus impubes (Steud.) Napper	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
var. fallax (Cherm.) Kük. (C. fallax Cherm.)	$\mathbf{C}_4$	KW (Ac: de la Bâthie 13098 P)
C. incompressus C. B. Clarke	$C_4$	KW (Ac: Jacques-Félix 7216 P); LR (δ <sup>13</sup> C: -11.0, Jacques-Félix 7216)
C. incomtus Kunth	$C_3$	KW (A: Venturi 5633)
C. indecorus Kunth	$C_4$	KW (Ac: O'Connor 16)
C. indecorus var. decurvatus (C. B. Clarke) Kük.	$\mathbf{C}_4$	KW (Ac: Vorster 2504)
C. intricatus Schrad. ex Schultes	$C_3$	Denton 1983 (A)
C. involucratus Rottb., as C. alternifolius L. subsp. flabelliformis (Rottb.) Kük.a, as C. flabelliformis Rottb.b	C <sub>3</sub>	Druyts-Voets 1970 ([A]) <sup>b</sup> ; Hofstra et al. 1972 (A. $\Gamma$ ) <sup>b</sup> ; Ehleringer et al. 1987 ( $\delta^{13}$ C) <sup>a</sup> ; Li and Jones 1994 (A); KW (A: <i>Wilson 4384</i> )
C. iria L.	$C_4$	Akita et al. 1969 ([A]); Druyts-Voets 1970 ([A]); Hofstra et al. 1972 (A. Γ); Carolin et al. 1977 (USc); Takeda et al. 1980 (Ac. Γ); Hesla et al. 1982 (δ <sup>13</sup> C); Takeda et al. 1985 (Ac); Ueno et al. 1986 (Ac. B); Ueno et al. 1988b (USc); Lin et al. 1993 (Ac); Ueno 1998a (US); Ueno 2004 (B [kinetics]); JB (Ac: Bruhl 207 CANB; Latz 1527 CANB); KW (Ac: Coveny 3499; Streimann & Kairo NGF 27568); LR (δ <sup>13</sup> C: −11.9, Chevalier 24693)
var. flavescens Benth.	$C_4$	Druyts-Voets 1970 ([A])
C. isabellinus K. L. Wilson	$C_4$	KW (Ac: Wilson 3348)
C. ixiocarpus F. Muell.	$C_4$	KW (Ac: Chippendale 2078; Latz 1255, 6698)
C. javanicus Houtt., as M. javanicus (Houtt.) Merr. & Metc. <sup>a</sup>	$C_4$	Hofstra et al. 1972 (A. $\Gamma$ ); Ueno and Takeda 1992 (A) <sup>a</sup>
C. jeminicus Rottb.	$C_4$	LR (δ <sup>13</sup> C: -12.7, Chevalier 1235)
C. kaessneri C. B. Clarke	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. kappleri Hochst. ex Steud.	$C_4$	KW (Ac: Holt 260 NY)
C. karlschumanii C. B. Clarke	$C_4$	KW (Ac: Aké Assi 6513 P)
C. kerstenii Boeck., as Mariscus kerstenii (Boeck.) C. B. Clarke	$\mathbf{C}_4$	Hesla et al. 1982 (A. $\delta^{13}$ C)
C. kilimandscharicus Kük.	$C_4$	Hesla et al. 1982 (A. $\delta^{13}$ C)
C. kipasensis Cherm., as C. platycaulis var. kipasensis (Cherm.) A. Peter & Kük. <sup>a</sup>	$C_3$	Druyts-Voets 1970 ([A]) <sup>a</sup> ; KW (A: de Witte 3339 NSW ex P)
C. kirkii C. B. Clarke	$C_4$	KW (Ac: Biegel 2065 P)
C. koyaliensis Cherm.	$C_4$	LR (δ <sup>13</sup> C: -12.6, <i>Chevalier 20553</i> P)
C. lacunosus Griseb.	$C_4$	KW (Ac: Leon 8199 P)
C. laeteflorens (C. B. Clarke) Kük.	$C_4$	KW (Ac: McKee 7956)
C. laevigatus L., as Juncellus laevigata [sic] (L.) C. B. Clarke <sup>a</sup>	$C_4$	Borchers et al. 1982 (A); Druyts-Voets 1970 ([A]); Hesla et al. 1982 (δ <sup>13</sup> C); Frey and Kürschner 1983 (A. δ <sup>13</sup> C); Takeda et al. 1985 (Ac. δ <sup>13</sup> C); Li 1993 (A. δ <sup>13</sup> C); Bruhl et al. 1987 (Ac. B) <sup>a</sup> ; JB (Ac: <i>Bruhl 65</i> CANB; <i>Paijmans 2762</i> CANB; <i>Symon 13169</i> CANB); KW (Ac: <i>Barry 65</i> ); LR (δ <sup>13</sup> C: -12.1, <i>Balansa 736</i> P); SCV (δ <sup>13</sup> C: -9.31, <i>Bolus 715</i> )
C. laevis R. Br.	$C_3$	KW (A: Rodd 2262)
C. lancastriensis Porter	$C_4$	Li et al. 1999 (A. $\delta^{13}$ C)
C. latifolius Poir.	$C_4$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 (δ <sup>13</sup> C); Aucour et al. 1994 (δ <sup>13</sup> C) LR (δ <sup>13</sup> C: -13.0, <i>Humbert 7778</i> )
C. latzii K. L. Wilson	$C_4$	KW (Ac: Latz 5270)
C. laxus Lam., as C. diffusus Vahla	$C_3$	Prakash et al. 1976 (A); LR (8 <sup>13</sup> C: -32.7, Serre Orsay cult., -36.5, <i>Gilles</i> 180)
subsp. <i>buchholzii</i> (Boeck.) K. Lye, as <i>C. diffusus</i> subsp. <i>buchholzii</i> (Boeck.) Kük.	C <sub>3</sub>	Druyts-Voets 1970 ([A])
subsp. <i>sylvestris</i> (Ridl.) K. Lye, as <i>Cype-rus diffusus</i> subsp. <i>sylvestris</i> (Ridl.) Kük.	C <sub>3</sub>	Druyts-Voets 1970 ([A])

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
C. lecontei Torr. ex Steud.	$C_3$	KW (A: Curtiss 5714)
C. leiocaulon Benth.	$C_4$	Carolin et al. 1977 (USc); KW (Ac: Payne 16)
C. leptocladus Kunth	$C_3$	SCV (δ <sup>13</sup> C: -27.37, Maputaland Expedition 14316)
C. leucocephalus Retz.	$C_3$	Simpson 1990 (A); KW (A: Nat. Collector no. D1210, Thailand P)
C. lhotskyanus Boeck. (Mariscus rutilans	$C_4$	Hattersley et al. 1977 (Ac. B [IL]) <sup>a</sup> ; Takeda et al. 1985 (Ac. $\delta^{13}$ C) <sup>a</sup> ; JB (Ac:
C. B. Clarke), as C. rutilans (C. B.	- 4	Hattersley, 10 Dec 1979 voucher at RSBS); KW (Ac: Beauglehole
Clarke) Maiden & Betche <sup>a</sup>		49697; Rupp NSW 65130; Wilson 4441, 4442, 5877, 5878)
C. ligularis L., as Mariscus ligularis (L.) Urb. <sup>a</sup>	$\mathrm{C}_4$	Hnatiuk 1980 (A); LR (δ <sup>13</sup> C: -11.1, Leprieur s. n., 1824) <sup>a</sup>
C. limosus Maxim.	$C_3$	LR (δ¹³C: −26.0, <i>Maximowicz s. n.</i> , 1859, R. Amur)
C. longibracteatus Cherm., as Mariscus longibracteatus Cherm. <sup>a</sup>	$C_4$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Bosser 7220</i> P); LR (δ <sup>13</sup> C: -10.8, <i>Le Testu 8875</i> ) <sup>a</sup>
var. niger (C. B. Clarke) K. Lye, as Mar- iscus keniensis (Kük.) S. S. Hooper	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C); Vorster 1990 (A); SCV (δ <sup>13</sup> C: -9.99, <i>Burtt-Davy s. n.</i> )
var. rubrotinctus (Cherm.) Kük., as Mariscus rubrotinctus Cherm.	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
C. longus L., as C. longus var. longus <sup>a</sup>	$\mathrm{C}_4$	Haberlandt 1882 ([Ac]); Lerman and Raynal 1972 (A); Jones et al. 1981 (Ac. B. Γ. USc); Hesla et al. 1982 (δ <sup>13</sup> C); Li 1993 (A. Γ. δ <sup>13</sup> C); Li and Jones 1994 (A); LR (δ <sup>13</sup> C: -9.1, <i>Lejeune 713</i> ); SCV (δ <sup>13</sup> C: -8.16, <i>Schlechter 3925</i> ) <sup>a</sup>
subsp. <i>tenuiflorus</i> (Rottb.) Kük., as <i>C. longus</i> var. <i>tenuiflorus</i> (Rottb.) Boeck. <sup>a</sup>	$C_4$	Druyts-Voets 1970 ([A]); SCV (δ <sup>13</sup> C: -10.90, <i>Bolus 3926</i> ) <sup>a</sup>
var. pallidus Boeck.	$C_4$	Druyts-Voets 1970 ([Ac]); Borchers et al. 1982 (A)
C. lucidus R. Br. (Mariscus lucidus (R. Br.) C. B. Clarke)	$C_4$	JB (Ac: Bruhl 75 CANB); KW (Ac: Constable 6217; Corrick 7936)
C. lupulinus (Spreng.) Marcks	$C_4$	Li et al. 1999 (A. δ <sup>13</sup> C); KW (Ac: <i>Tolstead 41551</i> MO)
subsp. <i>macilentus</i> (Fernald) Marcks	$C_4$	KW (Ac: Kneucker 91)
C. luteus Boeck., as Mariscus luteus	$C_4$ $C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
(Boeck.) C. B. Clarke	•	
C. luzulae (L.) Retz.	$C_3$	Druyts-Voets 1970 ([A]); Denton 1983 (A); KW (Ac: Wilson 1586); LR (δ <sup>13</sup> C: -30.0, Rodriguez 4853)
C. macrocarpus (Kunth) Boeck., as Mariscus macrocarpus C. B. Clarke <sup>a</sup>	$C_4$	Hesla et al. 1982 (A. δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Troupin 7178</i> P; <i>Vorster 2637</i> )
var. humbertii (Cherm.) Kük.	$\mathbf{C}_4$	KW (Ac: <i>Bosser 18951</i> P)
var. pseudoflavus (Kük.) K. Lye, as Mariscus macer Kunth	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
C. macrocephalus Liebm. (Torulinium macrocephalum (Liebm.) C. B. Clarke, C. eggersii Boeck.)	$\mathrm{C}_4$	KW (Ac: Croat 23370 P; Leon 9120 P)
C. maculatus Boeck.	$C_4$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 ( $\delta^{13}$ C)
C. malaccensis Lam.	$C_4$	Mani 1963 ([A]; Hofstra et al. 1972 (A. Γ); KW (Ac: <i>Floyd</i> NGF 8041)
C. manimae Kunth var. asperrimus (Liebm.) Kük.	$C_4$	KW (Ac: Pringle 13237)
var. manimae (C. phaeocephalus Griseb.)	$C_4$	KW (Ac: Benoist 2646 P)
C. mannii C. B. Clarke	$C_3$	LR (\delta^{13}C: -33.0, Letouzey 7957)
C. mapanioides C. B. Clarke	$C_3$	Druyts-Voets 1970 ([A]); LR ( $\delta^{13}$ C: $-31.9$ , Le Testu 2763)
var. major Boeck.	$C_3$	Druyts-Voets 1970 ([A])
C. maranguensis K. Schum.	$C_4$	Hesla et al. 1982 (A. $\delta^{13}$ C)
C. margaritaceus Vahla, as C. margarita-	$C_4$	Druyts-Voets 1970 ([A]) <sup>a,b,c</sup> ; KW (Ac: <i>Schlechter 11591</i> ); LR ( $\delta^{13}$ C: $-10.6$ ,
ceus var. prorepens Kük. <sup>b</sup> , as C. margaritaceus var. pseudoniveus (Boeck.) C. B. Clarke <sup>c</sup>	-4	Sacleux 873) <sup>a</sup>
C. marginatus Thunb.	$C_3$	SCV (δ¹³C: −26.27, Ward 12292)
C. medusaeus Chiovenda	$C_4$	Kukkonen and Lye 1996 (A)
C. meeboldii Kük.	$C_4$	LR (\delta^{13}C: -12.5, Audru 2215)
C. megalanthus (Kük.) G. C. Tucker, as C.	$C_3$	Denton 1983 (A)
pseudovegetus var. megalanthus Kük.	-5	( )
C. meyenianus Kunth	$C_4$	KW (Ac: Montes 1507; Orth 708; Riedel 904 US)
C. michelianus (L.) Delile, as Dichostylis micheliana (L.) Nees <sup>a</sup> , as C. micheli-	$C_4$	Druyts-Voets 1970 ([A]); Li 1993 (A. δ <sup>13</sup> C); Kalapos et al. 1997 (A. δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Kneucker 33a</i> ); LR (δ <sup>13</sup> C: -12.3, <i>Duclos s. n.</i> , 1933) <sup>b</sup>
anus subsp. michelianus <sup>b</sup> C. michoacanensis Britton	$C_3$	Simpson 1990 (A); KW (A: Pringle 4269 P; Purpus 267 p.p. US)
c. menoucunensis Billion	$\sim_3$	5 mpson 1770 (11), 11 (11. 1 mgt 7207 1, 1 mps 207 p.p. 03)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
C. microcephalus R. Br. [identity unclear: voucher number is for a dicot]	$C_4$	Takeda et al. 1985 (A)
subsp. chersophilus K. L. Wilson	$C_4$	KW (Ac: Fitzgerald NSW 153243; Wilson 4891)
subsp. microcephalus	$C_4$	KW (Ac: Black 14; Latz 7377; Wilson 4874, 5202, 5556)
subsp. saxicola K. L. Wilson	$C_4$	KW (Ac: Jacobs 1535; Latz 3102; Wilson 5191, 5223)
C. microglumis D. A. Simpson	C <sub>3</sub>	Simpson 1990 (A)
C. microiria Steud.	$C_4$	Imai and Murata 1979 (Γ); Ueno et al. 1986 (Ac. B); Ueno and Takeda 1992 (A. Γ); Li et al. 1999 (A); KW (Ac: <i>Ohwi NSM 308; Hashimoto TNS 1269</i> )
C. miliifolius Poepp. & Kunth ex Kunth	$C_3$	KW (A: Croat 19806 P)
C. mitis Steud.	$C_4$	Mani 1963 ([A])
C. mollipes (C. B. Clarke) K. Schum. (Ascopholis gamblei C. E. C. Fisch.), as Mariscus mollipes C. B. Clarke <sup>a</sup>	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; JB (Ac: <i>Nijalingappa</i> NSW 709497); KW (Ac: <i>Lewalle 2052</i> P)
C. mutisii (Kunth) Griseb.	$\mathbf{C}_4$	KW (Ac: Barkley 14092 NY; Pringle 4476)
C. nanus Willd.	$\mathbf{C}_4$	KW (Ac: <i>Harris 12359</i> P)
C. natalensis Hochst.	$\mathbf{C}_4$	SCV ( $\delta^{13}$ C: $-9.95$ , $Hood\ 860$ )
C. nayaritensis Tucker	$C_3$	Simpson 1990 (A)
C. nduru Cherm., as C. margaritaceus var. nduru (Cherm.) Kük.	$\mathrm{C}_4$	Druyts-Voets 1970 ([A])
C. nipponicus Franch. & Sav.	$C_4$	Ueno et al. 1986 (Ac. B); Ueno et al. 1988 $b$ (USc); Ueno and Takeda 1992 (A. $\Gamma$ )
C. niveus Retz.	$\mathbf{C}_4$	Sabnis 1921 ([A]); Hnatiuk 1980 (A)
var. flavissimus (Schrad.) K. Lye, as C. obtusiflorus var. flavissimus (Schrad.) Boeck.	$\mathrm{C}_4$	Druyts-Voets 1970 ([A])
var. leucocephalus (Kunth) Fosberg, as C. obtusiflorus Vahl <sup>a</sup> , as C. obtusiflo- rus var. tenerior C. B. Clarke <sup>b</sup> , as C. obtusiflorus var. macrostachys ined. <sup>c</sup>	$\mathrm{C}_4$	Druyts-Voets 1970 ([A]) <sup>a,b,c</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); KW (Ac: <i>Decary 12835</i> ); LR (δ <sup>13</sup> C: -12.4, <i>Bachmann 78</i> )
var. tisserantii (Cherm.) K. Lye	$C_4$	KW (Ac: Boudet 2420 P)
C. nutans Vahl	$C_4$	Ueno et al. 1986 (Ac. B); Ueno and Takeda 1992 (A)
subsp. eleusinoides (Kunth) Haines	$C_4$	KW (Ac: Blake 7718)
C. oblongo-incrassatus Kük., as Mariscus taylori C. B. Clarke	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. obsoletinervosus A. Peter & Kük., as  Mariscus obsoletinervosus (A. Peter & Kük.) Greenway <sup>a</sup>	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Polhill &amp; Paulo 964</i> P)
C. ochraceus Vahl	$C_3$	Denton 1983 (A)
C. odoratus L., as Torulinium odoratum (L.) S. S. Hooper <sup>a</sup> , as T. ferax (Rich.) Ham. <sup>b</sup>	$\mathbf{C}_4$	Bender 1971 (δ <sup>13</sup> C); Ueno et al. 1988 <i>b</i> (USc) <sup>b</sup> ; Ueno and Takeda 1992 (A) <sup>a</sup> ; Ueno et al. 1986 (Ac. B) <sup>b</sup> ; Li et al. 1999 (A. δ <sup>13</sup> C); JB (Ac: <i>Darbyshire 708</i> ); KW (Ac: <i>Heller 2466</i> P); LR (δ <sup>13</sup> C: -10.5, <i>Mocquerys s. n.</i> , Venezuela) <sup>a</sup>
C. ohwii Kük.	$C_4$	Ueno et al. 1986 (Ac. B); Ueno and Takeda 1992 (A. Γ)
C. orgadophilus K. L. Wilson	$C_4$	KW (Ac: Latz 7144)
C. orthostachys Franch. & Sav., as C. truncatus C. A. Mey. ex Turcz. <sup>a</sup>	$C_4$	Lerman and Raynal 1972 (A)*; Ueno et al. 1986 (Ac. B); Ueno and Takeda 1992 (A. Γ); KW (Ac: <i>Fox</i> NSW 618282); LR (δ <sup>13</sup> C: -15.0, <i>Karo</i> 90)*
C. oxycarpus S. T. Blake	$C_4$	KW (Ac: Blake 9209; Latz 7298)
C. oxylepis Nees ex Steud.	$C_4$	KW (Ac: Pedersen 9611)
C. palianparaiensis Govindarajalu	$C_3$	Govindarajalu 1990b ([A])
C. panamensis (C. B. Clarke) Britton	$C_4$	KW (Ac: Standley 29144 US)
C. pangorei Rottb.	$C_4$	KW (Ac: Wight NSW 608850)
C. pannonicus Jacq., as Acorellus pannonicus (Jacq.) Palla <sup>a</sup>	$C_4$	Li 1993 (A. δ <sup>13</sup> C); Kalapos et al. 1997 (A. δ <sup>13</sup> C) <sup>a</sup> ; JB (Ac: <i>s. coll.</i> MEL 1543854)
C. papyrus L.a, as C. papyrus subsp. ugandensis Chiov.b	$\mathrm{C}_4$	Haberlandt 1884 ([Ac]) <sup>a</sup> ; Rikli 1895 ([A]) <sup>a</sup> ; Druyts-Voets 1970 ([A]) <sup>a,b</sup> ; Krenzer et al. 1975 (Γ) <sup>a</sup> ; Jones and Milburn 1978 (A. Γ) <sup>a</sup> ; Hesla et al. 1982 (A. δ <sup>13</sup> C) <sup>a</sup> ; Li 1993 (A. Γ. δ <sup>13</sup> C) <sup>a</sup> ; Aucour et al. 1994 (δ <sup>13</sup> C) <sup>a</sup> ; Li and Jones 1994 (A) <sup>a</sup> ; JB (Ac: <i>Lau 2112</i> NE; <i>Lepschi 1505</i> NE) <sup>a</sup> ; LR (δ <sup>13</sup> C: –11.0, –11.9, <i>Dang 178</i> , –12.0, –12.8, <i>Killick 3419</i> ) <sup>a</sup>
as Papyrus [sic] <sup>b</sup> , as Papyrus cicuta <sup>a</sup>	$C_3+$	Moss et al. 1969 ( $\Gamma$ ) <sup>a</sup> ; Tregunna et al. 1970 (A. B. $\Gamma$ . $\delta^{13}$ C) <sup>b</sup> ; Hofstra et al. 1972 (A. $\Gamma$ )
C. pectinatus Vahl, as C. nudicaulis Poir. <sup>a</sup>	$C_3$	Druyts-Voets 1970 ([A]) <sup>a</sup> ; Ellery et al. 1992 ( $\delta^{13}$ C); LR ( $\delta^{13}$ C: $-27.8$ , Leprieur s. n., Senegal) <sup>a</sup>

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
C. pedunculatus (R. Br.) Kern, as Remi-	$C_4$	Hofstra et al. 1972 (A. Γ); Lerman and Raynal 1972 (A)a; Ueno and Takeda
rea <sup>a</sup> , as Remirea maritima Aubl. <sup>b</sup> , as		1992 (A)c; JB (Ac: Blake 8261 BRI; Bruhl 496 CANB; Lazarides 563
Mariscus pedunculatus (R. Br.) Koya- ma <sup>c</sup>		CANB; Pullen 1181 CANB); LR (δ <sup>13</sup> C: -13.5, Chevalier 23474) <sup>b</sup>
C. perangustus S. T. Blake	$\mathbf{C}_4$	KW (Ac: Blake 11304)
C. perennis (M. E. Jones) O'Neill	$C_3$	KW (A: Gentry 14432 US; Purpus 267 p.p. US)
C. phillipsiae (C. B. Clarke) Kük., as Mariscus phillipsiae C. B. Clarke	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. phleoides (Nees ex Kunth) Hillebr. var. hawaiiensis (H. Mann) Kük.	$\mathrm{C}_4$	KW (Ac: Alexander 5234)
C. picardae Boeck.	$C_4$	KW (Ac: <i>Howard 8842</i> P)
C. pilosus Vahl	$C_4$	Ehleringer et al. 1987 ( $\delta^{13}$ C); Ueno et al. 1986 (Ac. B); Ueno et al. 1988 (USc); Ueno and Takeda 1992 (A. $\Gamma$ ); KW (Ac: <i>McKee 1547</i> ; <i>Murata T-15902</i> P)
C. plateilema (Steud.) Kük., as Mariscus plateilema Steud.	$\mathrm{C}_4$	Hesla et al. 1982 (A. $\delta^{13}$ C)
C. platycaulis Baker <sup>a</sup> , as C. platycaulis var. lucenti-nigricans (K. Schum.) Kük. <sup>b</sup> , as C. platycaulis var. serpens (Cherm.) Kük. <sup>c</sup>	$C_3$	Druyts-Voets 1970 ([A]) <sup>a,b,c</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup>
C. platystylis R. Br.	$C_3$	Mani 1960 ([A]); Druyts-Voets 1970 ([A]); Takeda et al. 1985 (A); KW (A: <i>Specht 1159</i> ); LR (δ <sup>13</sup> C: -27.7, <i>Poilane 21428</i> )
C. plukenetii Fernald	$\mathbf{C}_4$	Li et al. 1999 (A)
C. podocarpus Boeck.	$\mathbf{C}_4$	KW (Ac: Adam 14957 P)
C. portae-tartari K. L. Wilson, as C. ixi- ocarpus F. Muell. <sup>a</sup>	$C_4$	Carolin et al. 1977 (USc) <sup>a</sup> ; KW (Ac: <i>Dunlop 4455</i> ; <i>Jacobs 1527</i> )
C. procerus Rottb.	$\mathbf{C}_4$	Mani 1960 ([A]); KW (Ac: Auld NSW 84668; Salasoo NSW 91196)
var. <i>vanderystii</i>	$\mathbf{C}_4$	Druyts-Voets 1970 ([A])
C. prolifer Lam.	$\mathbf{C}_3$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 (δ <sup>13</sup> C); KW (A: Wilson 4382)
C. prolixus Kunth	$\mathbf{C}_4$	KW (Ac: Pedersen 9601 P); LR (δ <sup>13</sup> C: -12.7, Bourgeau s. n., 1866)
C. pseudoleptocladus Kük.a, as C. pseudo- leptocladus var. polycarpus Kük.b	$C_3$	Druyts-Voets 1970 ([A]) <sup>a,b</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; KW (A: <i>Pawek 6484</i> P) <sup>a</sup>
C. pseudovegetus Steud., as C. pseudovegetus var. pseudovegetus <sup>s</sup>	$C_3$	Denton 1983 (A) <sup>a</sup> ; Li et al. 1999 (A)
C. pseudovestitus (C. B. Clarke) Kük. (Mariscus goniobolbus Cherm.), as Mariscus pseudovestitus C. B. Clarke <sup>a</sup>	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; Vorster 1990 (A) <sup>a</sup> ; KW (Ac: <i>Bosser 13474</i> P; <i>Vorster 2497</i> )
C. pulchellus R. Br., as C. leucocephalus Retz. var. pulchellus (R. Br.) ined. <sup>a</sup>	$C_3$	Druyts-Voets 1970 ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); Takeda et al. 1985 (A. δ <sup>13</sup> C); Simpson 1990 (A); KW (A: <i>Bosser 4654</i> P; <i>McKee 9189</i> P)
C. pulchellus	$\mathbf{C}_4 +$	LR (δ <sup>13</sup> C: -13.2, Gillet 1698)
C. pulcher Thunb.	$C_3$	Sonnenberg and Botha 1992 (A. PIB)
C. pustulatus Vahl	$C_4$	Druyts-Voets 1970 ([A])
C. pycnostachyus (Kunth) Kunth	$C_4$	KW (Ac: Pringle 6313 P)
C. pygmaeus Rottb., as C. michelianus subsp. pygmaeus (Rottb.) Aschers. & Graebn. <sup>a</sup>	$C_4$	Carolin et al. 1977 (USc); Hesla et al. 1982 (δ¹³C); Ueno and Takeda 1992 (A. Γ)²; JB (Ac: <i>H. Eichler 1818</i> 2 CANB); KW (Ac: <i>Beauglehole 46512</i> ; <i>McGillivray 2943</i> ; <i>Solling 486</i> ; <i>Wilson 1465</i> ); LR (δ¹³C: −13.3, <i>Kotschy 329</i> )²
C. radians Nees & Meyen	$C_4$	KW (Ac: Petelot 5480, 5602 P)
C. reduncus Boeck.	$C_3$	Druyts-Voets 1970 ([A])
C. reflexus Vahl, as C. reflexus var. reflexus sa, as C. reflexus var. fraternus (Kunth) Kuntzeb	$C_3$	Denton 1983 (A) <sup>a,b</sup> ; KW (A: Wilson 1441)
C. retroflexus var. pumilus (Britton) R. Carter & S. D. Jones (C. subuniflorus Britton)	$\mathrm{C}_4$	KW (Ac: Pringle 807 P)
C. refractus Engelm. ex Boeck.	$C_4$	Li et al. 1999 (A)
C. remotus (C. B. Clarke) Kük., as Maris- cus remotus C. B. Clarke	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
C. renschii Boeck.	$C_3$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 (δ¹³C)
C. retrofractus (L.) Torr.	$C_4$	Li et al. 1999 (A); KW (Ac: Radford & Bozeman 11380 P)
C. retrorsus Chapm.	$C_4$	Li et al. 1999 (A. $\delta^{13}$ C)
C. rhynchosporoides Kük.	$C_4$	KW (Ac: Richards 16898 P)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
C. rigens J. Presl & C. Presl	$C_4$	Takeda et al. 1985 (Ac. δ <sup>13</sup> C); KW (Ac: Wilson 1445)
C. rigidellus (Benth.) J. Black, as C. sub- pinnatus Kük. <sup>a</sup>	$C_4$	Takeda et al. 1985 (Ac) <sup>a</sup> ; KW (Ac: <i>Beauglehole 23111</i> ; Wilson 742, 1466)
C. rigidifolius Steud.	$\mathrm{C}_4$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 (A. δ <sup>13</sup> C); LR (δ <sup>13</sup> C: -12.9, <i>Pap-pi 2155</i> )
C. rohlfsii Boeck., as Mariscus rohlfsii (Boeck.) C. B. Clarke	$\mathrm{C}_4$	Hesla et al. $1982 (\delta^{13}C)$
C. rotundus L., as C. rotundus subsp. rotundus <sup>a</sup>	$C_4$	Sabnis 1921 ([A]); Mani 1963 ([A]); Akita et al. 1969 ([A]); Chen et al. 1970 (Γ); Druyts-Voets 1970 ([Ac]); Black and Mollenhauer 1971 (Ac); Black et al. 1973 (B); Hofstra et al. 1972 (A. Γ) <sup>a</sup> ; Chen et al. 1974 (A. B); Troughton et al. 1974 (δ <sup>13</sup> C); Prakash et al. 1976 (A); Meinzer 1978 (A); Takeda et al. 1980 (Ac. Γ); Borchers et al. 1982 (A); Hesla et al. 1982 (δ <sup>13</sup> C); Ueno et al. 1986 (Ac. B); Bruhl et al. 1987 (Ac. B); Li 1993 (A. Γ. δ <sup>13</sup> C); Lin et al. 1993 (Ac); Li and Jones 1994 (A); Li et al. 1999 (A. δ <sup>13</sup> C); JB (Ac: <i>I. B. Wilson 197</i> CANB); KW (Ac: <i>Wilson 902</i> )
subsp. <i>merkeri</i> (C. B. Clarke) Kük., as <i>C. merkeri</i> C. B. Clarke <sup>a</sup>	$C_4$	Druyts-Voets 1970 ([A]); Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup>
C. rubicundus Vahl, as C. teneriffae Poir.a	$C_4$	Druyts-Voets 1970 ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Subko 104</i> P); LR (δ <sup>13</sup> C: -11.8, <i>Schimper 1323</i> ) <sup>a</sup>
C. rubiginosus Hook. f.	$\mathrm{C}_4$	KW (Ac: Hooker s. n. P; Schimpff 14 US; Wheeler et al. 5 US; Wiggins & Porter 568 NY)
C. rupestris Kunth	$\mathbf{C}_4$	KW (Ac: Biegel 2065 P; Pienaar 271 PRE; Wood 8546)
C. sandwicensis Kük.	$\mathbf{C}_4$	KW (Ac: Degener 8460 NY)
C. sanguineo-ater Boeck.	$\mathbf{C}_4$	KW (Ac: Pringle 3844)
C. scaber R. Br., as Mariscus scaber (R. Br.) Boeck. <sup>a</sup>	$\mathrm{C}_4$	Bruhl et al. 1987 (Ac. B) <sup>a</sup> ; JB (Ac: <i>Bruhl 234, 497</i> CANB); KW (Ac: <i>Mc-Kee 9040</i> P)
C. scariosus R. Br.	$\mathbf{C}_4$	Takeda et al. 1985 (Ac. $\delta^{13}$ C)
C. schimperianus Steud., as C. schimperianus var. minor Boeck. <sup>a</sup>	$\mathrm{C}_4$	Druyts-Voets 1970 ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C)
C. schomburgkianus Nees	$C_3$	Simpson 1990 (A)
C. schweinitzii Torr.	$C_4$	Li et al. 1999 (A. $\delta^{13}$ C)
C. secubans K. L. Wilson	$C_4$	KW (Ac: Coveny 8812)
C. seemanianus Boeck.	$C_4$	KW (Ac: Campbell NSW 22740)
C. semitrifidus Schrad.	$C_4$	KW (Ac: Schlechter 2513)
C. serotinus Rottb., as Juncellus serotinus (Rottb.) C. B. Clarke <sup>a</sup>	$\mathrm{C}_4$	Akita et al. 1969 ([A]); Takeda et al. 1980 (Ac. Γ); Collins and Jones 1985 (δ <sup>13</sup> C); Ueno et al. 1986 (Ac. B); Li 1993 (A. Γ. δ <sup>13</sup> C); Kalapos et al. 1997 (δ <sup>13</sup> C) <sup>a</sup> ; JB (Ac: <i>s. coll.</i> MEL 1543840); KW (Ac: <i>Naito</i> NSW 608949); LR (δ <sup>13</sup> C: -13.1, <i>Bourgeau 32</i> )
C. seslerioides Kunth	$C_3$	LR (\delta^{13}C: -30.3, Pringle 806)
C. setigerus Torr. & Hook.	$\mathbf{C}_4$	Li et al. 1999 (A)
C. sexflorus R. Br.	$C_4$	KW (Ac: Blake 17534; de Lestang 338; Wilson 5341)
C. socialis C. B. Clarke	$C_4$	KW (Ac: <i>Halle 3059</i> P)
C. solidus Kunth, as Mariscus solidus (Kunth) Vorster var. involutus (C. B. Clarke) Vorster ined. <sup>a</sup>	$C_4$	Vorster 1990 (A) <sup>a</sup> ; KW (Ac: Guillaumin s. n., cult. P; Medley Wood 12023)
C. sordidus J. Presl & C. Presl (C. howellii O'Neill & Ben. Ayers)	$\mathrm{C}_4$	KW (Ac: Mason & Hanna 14594 US)
C. soyauxii Boeck., as Mariscus soyauxii (Boeck.) C. B. Clarke <sup>a</sup>	$\mathrm{C}_4$	KW (Ac: Adam 1918); LR (δ <sup>13</sup> C: -12.8, Adam 1918) <sup>a</sup>
C. sp. (Isolepis humillima (Benth.) K. L. Wilson)	$C_4$	KW (Ac: Clarke 24)
<i>C.</i> sp.	$C_4$	Smith and Epstein 1971 ( $\delta^{13}$ C)
C. sp. as Cyperus japonicus Makino	$C_4+$	Li and Jones 1994 (A)
C. sp. as Mariscus psilostachys (C. B. Clarke) Kük. (non C. psilostachys Steud.) <sup>a</sup>	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Gillett 12841</i> P)
C. sp. nov. aff. pedunculosus F. Muell.	$C_3$	KW (A: Sharpe 1455)
C. sp. aff. sexflorus	$C_4$	Carolin et al. 1977 (USc)
C. speciosus Vahl	$\mathbf{C}_4$	KW (Ac: Kotov s. n., Transcaucasia P)
C. spectabilis Spreng.	$\mathbf{C}_4$	KW (Ac: Arsène 5914 P; Pringle 13233)
C. sphacelatus Rottb.	$\mathbf{C}_4$	Druyts-Voets 1970 ([A])

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
C. sphaerospermus Schrad.	$C_3$	KW (A: O'Connor 71 PRE)
C. sporobolus R. Br.	$\mathbf{C}_4$	KW (Ac: Dunlop 4096)
C. squarrosus L. (Monandrus squarrosus	$\mathbb{C}_4$	Mani 1960 ([A]) <sup>b</sup> ; Druyts-Voets 1970 ([A]) <sup>a,b</sup> ; Mulroy and Rundel 1977 (A) <sup>c</sup> ;
(L.) ined.) <sup>a</sup> , as C. aristatus Rottb. <sup>b</sup> , as		Baskin and Baskin 1981 (A) <sup>c</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C) <sup>d</sup> ; Takeda et al.
C. aristatus var. inflexusc, as Mariscus		1985 (Ac) <sup>a</sup> ; Li 1993 (A. $\delta^{13}$ C) <sup>a</sup> ; Li et al. 1999 (A. $\delta^{13}$ C) <sup>a</sup> ; JB (Ac: <i>Da</i> -
squarrosus (L.) C. B. Clarke <sup>d</sup>		<i>vidson 347</i> BRI; <i>Steele</i> , 6 Aug 1909 BRI); KW (Ac: <i>Wilson 1501</i> ); LR (δ <sup>13</sup> C: -13.1, <i>Gillet 3272</i> ) <sup>d</sup>
C. stenophyllus Valck. Sur.	$C_4$	KW (Ac: Schlechter 16700 P)
C. stolonifer Retz.	$C_4$	Ueno and Takeda 1992 (A)
C. stradbrokensis Domin	$\mathbb{C}_3$	KW (A: Johnson 7593)
C. strigosus L., as Mariscus strigosus (L.) C. B. Clarke <sup>a</sup>	$C_4$	Lerman and Raynal 1972 (A) <sup>a</sup> ; Li 1993 (A. $\delta^{13}$ C); Li et al. 1999 (A. $\delta^{13}$ C); LR ( $\delta^{13}$ C: $-12.0$ , Louis Marie s. n., 1927) <sup>a</sup>
C. subbadius Kük.	$C_4$	KW (Ac: Leroy s. n., Madagascar P; Decary 644G)
C. sublimis (C. B. Clarke) Dandy, as M.	$\mathbb{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C); LR (δ <sup>13</sup> C: -12.6, Waterlot 1265)
sublimis C. B. Clarke		
C. submicrolepis Kük.	$C_3$	Druyts-Voets 1970 ([A])
C. subparadoxus Kük., as Mariscus paradoxus (Cherm.) Cherm.ª	$C_4$	KW (Ac: <i>Haines 4138</i> P); LR (δ <sup>13</sup> C: -13.5, <i>Haines 4138</i> ) <sup>a</sup>
C. subulatus R. Br.	$C_4$	KW (Ac: Whittet NSW 65182; Wilson 1493)
C. subumbellatus Kük., as Mariscus alter- nifolius Vahl	$C_4$	Hesla et al. 1982 $(\delta^{13}C)^a$
C. subxerophilus Kük.	$C_3$	Druyts-Voets 1970 ([A])
C. surinamensis Rottb.	$C_3$	Denton 1983 (A); LR (δ <sup>13</sup> C: -28.3, Leblond 40)
C. tabularis Schrad.	$C_4$	KW (Ac: Schlechter 10675)
C. tanganyicanus (Kük.) K. Lye, as C. bellus var. tanganyicanus Kük.	$C_4$	Druyts-Voets 1970 ([A])
C. tenax Boeck. <sup>a</sup> , as C. tenax var. actinos- tachys (Welw. ex Ridl.) Kük. <sup>b</sup>	$C_4$	Druyts-Voets 1970 ([A]) <sup>a,b</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Stolz 1046</i> P) <sup>a</sup>
C. tenellus L. f.	$C_3$	Druyts-Voets 1970 ([A]); Takeda et al. 1985 (A. $\delta^{13}$ C)
C. tenerrimus J. Presl & C. Presl	$C_3$	Simpson 1990 (A)
C. tenuiculmis Boeck.a, as C. zollingeri var. parvus C. B. Clarkeb, as C. tenuiculmis f. compactus ined.c, as C. tenui-	$\mathrm{C}_4$	Druyts-Voets 1970 ([A]) <sup>a,b,c,d</sup> ; KW (Ac: <i>Audru 3760</i> P; <i>Wilson 3819</i> ) <sup>a</sup> ; LR (δ <sup>13</sup> C: -11.9, <i>Hooker s. n.</i> , India 1853) <sup>a</sup>
culmis var. densior ined.d var. tenuiculmis, as C. tenuiculmis var.	$\mathrm{C}_4$	Druyts-Voets 1970 ([A])
longiramulosus Kük.	_	
var. schweinfurthianus (Boeck.) S. S. Hooper, as C. schweinfurthianus Boeck.	$C_4$	Druyts-Voets 1970 ([A])
C. tenuis var. luridus (C. B. Clarke) Kük.,	C	LR (8 <sup>13</sup> C: -12.3, <i>Chevalier 23601</i> )
as <i>Mariscus luridus</i> C. B. Clarke (non <i>C. luridus</i> Govindarajalu)	$C_4$	EK (0 °C. –12.3, Chevaner 23001)
C. tenuispica Steud.	$C_3$	Mani 1960 ([A]); Druyts-Voets 1970 ([A]); Nautiyal and Das 1982 ([A]); Ueno and Takeda 1992 (A); LR (δ <sup>13</sup> C: -28.1, <i>Jacques-Félix 7241</i> )
C. tenuispiculatus Boeck.	$C_4$	KW (Ac: de la Bâthie, Madagascar Apr 1928 P)
C. tetracarpus Boeck.	$C_4$	KW (Ac: Blake 15581)
C. textilis Thunb.	$C_3$	Druyts-Voets 1970 ([A])
	$C_4$ +	Li 1993 (A)
C. thomsonii Boeck.	$C_4$	KW (Ac: <i>Balansa</i> 2836 P)
C. thunbergii Vahl	$C_4$	KW (Ac: <i>Humbert 10215</i> P)
C. thyrsiflorus Junghuhn	$C_4$	KW (Ac: Fisher NSW 608977)
C. tomaiophyllus K. Schum., as Mariscus tomaiophyllus (K. Schum.) C. B. Clarke	$ extstyle{C_4}$	Hesla et al. 1982 (δ <sup>13</sup> C); LR (δ <sup>13</sup> C: -11.9, <i>Humbert 8639</i> )
C. tonkinensis C. B. Clarke	$C_4$	KW (Ac: Tixier 11 P)
var. <i>baikei</i> (C. B. Clarke ex Kük.) S. S. Hooper, as <i>C. baikiei</i> C. B. Clarke ex	$C_4$	Druyts-Voets 1970 ([Ac])
Kük.		
C. trachysanthos Hook. & Arn.	$\mathbf{C}_4$	KW (Ac: Degener 11059 BRI; Forbes 2359.0 P)
C. trichodes Griseb.	$\mathbf{C}_4$	KW (Ac: <i>Harris 12882</i> NY; <i>Proctor 34283</i> NY)
C. tuberosus Rottb., as C. rotundus subsp. tuberosus (Rottb.) Kük. <sup>a</sup>	$\mathrm{C}_4$	Druyts-Voets 1970 ([Ac]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); Takeda et al. 1985 (Ac)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
C. turrillii Kük., as Mariscus laxiflorus Turrilla	$C_4$	KW (Ac: Koechlin 5313 P); SCV (δ <sup>13</sup> C: -10.55, Rogers 7006) <sup>a</sup>
C. uncinulatus Schrad. ex Nees	$C_3$	LR (δ <sup>13</sup> C: -27.4, Weddell 3056)
C. undulatus Kük.	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
C. usitatus Burch.	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
var. stuhlmannii (C. B. Clarke) K. Lye, as Cyperus stuhlmannii C. B. Clarke	$C_4$	Hesla et al. 1982 $(\delta^{13}C)$
C. ustulatus A. Rich.	$\mathbf{C}_4$	Troughton et al. 1974 ( $\delta^{13}$ C)
C. vaginatus R. Br.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); KW (A: Payne 20; Wilson 1354)
C. varicus (C. B. Clarke) Kük.	$\mathbb{C}_4$	KW (Ac: Decary 10649 P)
C. vestitus Krauss, as Mariscus vestitus (Krauss) C. B. Clarke	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C); SCV ( $\delta^{13}$ C: $-9.99$ , Acocks 21042)
C. victoriensis C. B. Clarke	$\mathbf{C}_4$	Carolin et al. 1977 (USc); Takeda et al. 1985 (Ac); KW (Ac: <i>Payne 19</i> )
C. virens Michx., as C. virens var. virens <sup>a</sup>	$C_3$	Denton 1983 (A) <sup>a</sup> ; LR (δ <sup>13</sup> C: -29.4, <i>Curtis 5238</i> )
var. minarum (Boeck.) Denton	$C_3$	Denton 1983 (A)
var. montanus (Boeck.) Denton	$C_3$	Denton 1983 (A)
C. viscidulus K. L. Wilson	$\mathbb{C}_4$	KW (Ac: Beauglehole 47687; Dunlop 5231)
C. vorsteri K. L. Wilson, as Mariscus grantii C. B. Clarke	$\mathrm{C}_4$	Vorster 1990 (A)
C. zollingeri Steud., as C. rubroviridis Cherm.	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
Cypringlea analecta (Beetle) M. T. Strong Desmoschoenus spiralis (A. Rich.) Hook. f.	$C_3$ $C_3$	BW (A: Lyonet 1318 US; Pringle 3175 US; Purpus 2889 US)  JB (A: Clifford, 17 Nov 1973 BRI. δ <sup>13</sup> C: -24.6, Clifford, 17 Nov 1973); LI (δ <sup>13</sup> C: -24.7, Hombron s. n., ca. 1840)
Didymiandrum stellatum (Boeck.) Gilly	$C_3$	JB (A: <i>Prance</i> 9789 K)
Diplacrum africanum C. B. Clarke	$C_3$	JB (A: Schweinfurth 2573 BRI); LR (δ¹³C: -29.1, Jacques-Félix 7328)
D. caricinum R. Br., as Scleria caricina (R. Br.) Benth. <sup>a</sup>	$C_3$	Govindarajalu 1975a ([A]) <sup>a</sup> ; JB (A: <i>Brass 19343</i> CANB; <i>Schweinfurth 2573</i> BRI)
D. pygmaeum (R. Br.) Nees ex Boeck., as Scleria pygmaea R. Br. <sup>a</sup>	$C_3$	Takeda et al. 1985 (A) <sup>a</sup> ; JB (A: <i>Bruhl 304</i> CANB)
Diplasia karataefolia Rich. ex Pers.	$C_3$	Koyama 1967 ([A]); Metcalfe 1971 ([A]); JB (A: Croat 17547)
Dulichium arundinaceum (L.) Britton, as Dulichium <sup>a</sup>	$C_3$	Lerman and Raynal 1972 (A) <sup>a</sup> ; JB (A: <i>Smith</i> , 25 Aug 1946 BRI); LR (δ <sup>13</sup> C: -30.7, <i>Barkley Bogdan 380028</i> )
Egleria fluctuans L. T. Eiten	$C_3$	JB (A: <i>Ducke</i> , 20 July 1912 BRI. δ <sup>13</sup> C: -25.6, -27.6, <i>Ducke</i> , 20 July 1912 BRI)
Eleocharis acicularis (L.) Roem. & Schult.	$C_3$	Akita et al. 1969 ([A]); Sternberg et al. 1984 (δ <sup>13</sup> C); Keeley et al. 1986 (δ <sup>13</sup> C); Ueno et al. 1989 (A. δ <sup>13</sup> C); Ueno and Takeda 1992 (A); Lin et al. 1993 (A); JB (A: MEL 1543839; MEL 1543860)
E. acuta R. Br.	C <sub>3</sub>	Takeda et al. 1985 (A. δ <sup>13</sup> C); Bruhl et al. 1987 (A. B); Ueno et al. 1989 (A) Bruhl and Perry 1995 (US); JB (A: <i>Phillips 2841761</i> CANB; <i>Bruhl 33</i> , 74, 125 CANB. Γ: 44, <i>Bruhl 74</i> ; 47, <i>Bruhl 33</i> ; 47, <i>Bruhl 125</i> . δ <sup>13</sup> C: -28.2, <i>Bruhl 125</i> ; -28.4, <i>Bruhl 125</i> )
E. acutangula (Roxb.) Roem. & Schult., as E. fistulosa (Poir.) Link <sup>a</sup>	$C_3$	Govindarajalu 1975 <i>a</i> ([A]); Hesla et al. 1982 (δ <sup>13</sup> C); Ueno et al. 1989 (A) <sup>a</sup> ; Ueno and Takeda 1992 (A) <sup>a</sup> ; KW (A: <i>Blake 9371</i> ); LR (δ <sup>13</sup> C: -27.3, <i>Smith 6710</i> )
E. acutisquamata Buckley	$C_3$	Ueno et al. 1989 (A)
E. albibractea Nees & Meyen	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. albida Torr.	$C_3$	Ueno et al. 1989 (A)
E. alveolata Svenson	$C_4$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. amazonica C. B. Clarke	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. ambigens Fernald	$C_3$	Ueno et al. 1989 (A)
E. atricha R. Br.	$C_3$	Takeda et al. 1985 (A); JB (A: Beauglehole 6525 CANB)
E. atropurpurea (Retz.) Presl	C <sub>3</sub>	Govindarajalu 1975a ([A]); Hesla et al. 1982 (δ¹³C); Takeda et al. 1985 (A) Ueno et al. 1989 (A); Ueno and Takeda 1992 (A); JB (A: <i>Latz</i> 2226 CANB); SCV (δ¹³C: -26.90, <i>Schlieben</i> 6398)
E. attenuata (Franch. & Sav.) Palla	$\mathbb{C}_3$	Ueno and Takeda 1992 (A); JB (A: Flennley ANU 2536 CANB)
E. bahamensis Boeck.	$\mathbf{C}_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. baldwinii (Torr.) Chapm.	$C_4$	Uchino et al. 1995 (A (T): [e]. B(T). B $^{14}$ C pulse- $^{12}$ C chase (T)); Ueno and Samejima 1989 (B); Ueno et al. 1989 (A. $\delta^{13}$ C); Ueno and Samejima 1990 ( $\delta^{13}$ C)
	$C_4$ & $C_4$ -like $C_3$	Uchino et al. 1995 (A [S]: [e]. B [S]. B <sup>14</sup> C pulse- <sup>12</sup> C chase [S]); Ueno 2004 (A. B. B [antiserum]. B [IL]. B [kinetics]. US: NAD)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
	$C_3 - C_4$	Ueno and Samejima 1990 (δ¹³C)
	$C_3$	Ueno and Samejima 1990 (δ <sup>13</sup> C)
E. bella (Piper) Svenson	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. bolanderi A. Gray	$C_3$	Ueno et al. 1989 (A)
E. bonariensis Nees	$C_3$	Ueno et al. 1989 (A)
E. brassii S. T. Blake	$\mathbb{C}_3$	Takeda et al. 1985 (A); KW (A: Wilson 5318, 5375)
E. caespitosissima J. G. Baker	$C_4$	Bruhl et al. 1987 (Ae. B); Bruhl and Perry 1995 (USe); JB (Ae, <i>Dunlop</i> 4212 CANB; <i>Bruhl 356, 357, 365, 399, 409</i> CANB. Γ: 1, <i>Bruhl 356</i> ; 1, <i>Bruhl 375</i> ; 1, <i>Bruhl 375</i> . δ <sup>13</sup> C: -13.0, -13.6, <i>Bruhl 356</i> )
E. cancellata S. Wats.	$C_3$	Ueno et al. 1989 (A. δ <sup>13</sup> C); JB (A: <i>Pringle 3269</i> MEL)
E. capillacea Kunth	$C_3$	Ueno et al. 1989 (A)
E. cellulosa Torr.	$C_3$	Ueno et al. 1989 (A)
E. compressa Sull.	$C_3$	Ueno et al. 1989 (A)
E. congesta D. Don	$C_3$	Govindarajalu 1975 <i>a</i> ([A]); Ueno et al. 1989 (A. δ <sup>13</sup> C); Ueno 2004 (B [kinetics]); JB (A: <i>Uva</i> , Sep 1890 PDA)
subsp. japonica (Miq.) Koyama	$C_3$	Ueno and Takeda 1992 (A. Γ); JB (A: ?Koniegalle 1867 PDA)
E. crinalis (Griseb.) C. B. Clarke	$C_3$	Ueno et al. 1989 (A)
E. cylindrica Buckley	$C_3$ – $C_4$	Guaglianone and Ueno 1990 (A)
as E. spegazzinii Barros	$C_3 - C_4$ ?	Ueno et al. 1989 (A. δ <sup>13</sup> C)
E. cylindrostachys Boeck.	$C_3$	Ueno et al. 1989 (A)
E. debilis Kunth	$C_3$	Ueno et al. 1989 (A)
E. decumbens C. B. Clarke	$C_3$	Ueno et al. 1989 (A)
E. densa Benth.	$C_3$	Ueno et al. 1989 (A)
E. dietrichiana Boeck.	$C_3$	Ueno et al. 1989 (A)
E. dombeyana Kunth	$C_3$	Ueno et al. 1989 (A)
E. dulcis (Burm. f.) Hensch.	$C_3$	Takeda et al. 1985 (A); Ueno and Takeda 1992 (A); Ueno et al. 1989 (A. δ <sup>13</sup> C); Ellery et al. 1992 (δ <sup>13</sup> C); JB (A: <i>Dharmawardhana 14</i> CANB. Γ: 49, <i>Dharmawardhana 14</i> ); KW (A: <i>Wilson 5005, 5097a</i> )
E. dunensis Kük.	$C_3$	Ueno et al. 1989 (A)
E. elegans (Kunth) Roem. & Schult.	$C_3$	Ueno et al. 1989 (A)
E. elliptica Kunth	$C_3$	Ueno et al. 1989 (A)
E. elongata Chapm.	C <sub>3</sub>	Ueno et al. 1989 (A)
E. engelmannii Steud.	C <sub>3</sub>	Ueno et al. 1989 (A)
E. equisetina J. Presl & C. Presl	C <sub>3</sub>	Takeda et al. 1985 (A); KW (A: Constable 6535; Wilson 3825)
E. equisetoides (Elliott) Torr.	C <sub>3</sub>	Ueno et al. 1989 (A)
E. erythropoda Steud., as E. calva Torr. [nom. inval.]	$C_3$	Ueno et al. 1989 (A)
E. exigua (Kunth) Roem. & Schult.	$C_3$ ?	Ueno et al. 1989 (A. δ <sup>13</sup> C)
E. filiculmis Kunth	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. flavescens (Poir.) Urban	$C_3$	Ueno et al. 1989 (A)
var. <i>olivacea</i> (Torr.) Gleason, as <i>E. olivacea</i> Torr.	$C_3$	Ueno et al. 1989 (A)
E. geniculata (L.) Roem. & Schult. <sup>a</sup> , as E. caribaea (Rottb.) S. F. Blake <sup>b</sup>	$C_3$	Govindarajalu 1975 <i>a</i> ([A]); Takeda et al. 1985 (A) <sup>a,b</sup> ; Bruhl et al. 1987 (A. B); Ueno et al. 1989 (A); Ueno and Takeda 1992 (A); Bruhl and Perry 1995 (US); JB (A: <i>Bruhl 231 CANB.</i> Γ: 51, <i>Bruhl 231.</i> δ <sup>13</sup> C: -28.1, <i>Bruhl 231</i> ); LR (δ <sup>13</sup> C: -29.8, <i>Sintenis 1968</i> )
E. glauca Boeck.	$C_4$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. gracilis R. Br., as E. cunninghamii Boeck. <sup>a</sup>	$C_3$	Takeda et al. 1985 (A); Ueno et al. 1989 (A) <sup>a</sup>
E. grisea Kük.	$C_3$	Ueno et al. 1989 (A)
E. haumaniana Barros	$C_3$	Ueno et al. 1989 (A)
E. intermedia Schult.	$C_3$	Ueno et al. 1989 (A)
E. interstincta (Vahl) Roem. & Schult.	$C_3$	Eiten 1969 ([A]); Ueno et al. 1989 (A)
E. intricata Kük.	$C_3$	Ueno et al. 1989 (A); JB (A: Arnold 470 PRE)
E. kamtschatica (C. A. Mey.) Komarov	$C_3$	Ueno et al. 1989 (A); Ueno and Takeda 1992 (A)
E. kuroguwai Ohwi	$C_3$	Akita et al. 1969 ([A]); Ueno and Takeda 1992 (A. $\Gamma$ )
E. lanceolata Fernald	$\mathbf{C}_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. limosa (Schrad.) Schult.	$\mathbf{C}_3$	Ueno et al. 1989 (A); SCV (δ <sup>13</sup> C: -26.17, Flanagan 903)
× E. macounii Fernald (= E. intermedia Schult. × E. obtusa (Willd.) Schult.)	$C_3$	Ueno et al. 1989 (A)
E. macrostachya Britton	$C_3$	Keeley et al. 1986 (δ¹³C); Ueno et al. 1989 (A)
E. maculosa (Vahl) Roem. & Schult.	$C_3$	Ueno et al. 1989 (A)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
E. mamillata (Lindb.) Lindb.	$C_3$	Ueno et al. 1989 (A); Ueno and Takeda 1992 (A)
E. margaritacea (Hulten) Miyabe & Kudo	$C_3$	Ueno and Takeda 1992 (A)
E. marginulata Steud.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
E. melanocarpa Torr.	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. melanostachys (Urville) C. B. Clarke	$C_3$	Ueno et al. 1989 (A)
E. microcarpa Torr.	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. minarum Boeck.	$C_3$	Ueno et al. 1989 (A)
E. minima Kunth	$C_4$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. minuta Boeck.	$C_3$	Ueno et al. 1989 (A); JB (A: <i>Bruhl 201</i> CANB. Γ: 47, <i>Bruhl 201</i> . δ <sup>13</sup> C: –28.0, <i>Bruhl 201</i> )
E. minutissima Britton	$C_3$	Ueno et al. 1989 (A)
E. mitracarpa Steud.	$C_3$	Ueno et al. 1989 (A)
E. mitrata (Griseb.) C. B. Clarke	$C_3$	Ueno et al. 1989 (A. δ <sup>13</sup> C)
E. montana (Kunth) Roem. & Schult., as E. nodulosa (Roth) Schult.	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. montevidensis Kunth <sup>a</sup> , as E. palmeri Svenson <sup>b</sup>	$C_3$	Ueno et al. 1989 (A) <sup>a,b</sup>
E. multicaulis Sm.	$C_3$	Ueno et al. 1989 (A)
E. mutata (L.) Roem. & Schult.	$C_3$	Ueno et al. 1989 (A); LR (δ <sup>13</sup> C: -27.4, <i>Mélinon 13</i> )
E. nana Kunth	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. naumanniana Boeck.	$C_3$	JB (A: Smith 2988 PRE)
E. nervata Svenson	$C_3$	Ueno et al. 1989 (A)
E. nigrescens (Nees) Steud.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C); Ueno et al. 1989 (A. $\delta^{13}$ C)
E. nitida Fernald	$C_3$	Ueno et al. 1989 (A); JB (A: Fernald 328 MEL)
E. nuda C. B. Clarke	$C_3$	Ueno et al. 1989 (A)
E. nudipes (Kunth) Palla	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. nupeensis Hutch. & Dalziel	$C_3$	Ueno et al. 1989 (A)
E. obicis L. Johnson & O. Evans	$C_3$	KW (A: Wilson 5655)
E. obtusa (Willd.) Schult.	$C_3$	Ueno et al. 1989 (A)
E. ochrostachys Steud.	$C_3$	JB (A: Dharmawardhana 17 CANB. Γ: 47, Dharmawardhana 17)
E. oligantha C. B. Clarke	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. ovata (Roth) Roem. & Schult.	$C_3$	Ueno et al. 1989 (A); Ueno and Takeda 1992 (A)
E. pachycarpa E. Desv.	$C_3$ ?	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. pachystyla (C. Wright) C. B. Clarke	$C_3$ ?	Ueno et al. 1989 (A. δ <sup>13</sup> C)
E. pallens S. T. Blake	$C_3$	Takeda et al. 1985 (A); JB (A: Bruhl 33, 246 CANB. Γ: 45, Bruhl 246)
E. palustris (L.) Roem. & Schult. <sup>a</sup> , as E. smallii Britton <sup>b</sup>	$C_3$	Ueno et al. 1989 (A <sup>a,b</sup> , $\delta^{13}$ C <sup>b</sup> ); LR ( $\delta^{13}$ C: $-27.3$ , Buchet s. n., 1853)
E. palustris subsp. parvinux (Ohwi) T. Koyama	$C_3$	Ueno and Takeda 1992 (A)
E. parishii Britton	$C_3$	Ueno et al. 1989 (A)
E. <i>parodii</i> Barros	$C_3$	Ueno et al. 1989 (A)
E. parvula (Roem. & Schult.) Link ex Bluff	$C_3$	Bender 1971 (δ <sup>13</sup> C); Ueno et al. 1989 (A); LR (δ <sup>13</sup> C: -24.5, <i>Bourgeau 453</i> )
E. pellucida J. Presl & C. Presl	$C_3$	Ueno et al. 1989 (A)
E. philippinensis Svenson	$C_3$	KW (A: Wilson 3587)
E. plana S. T. Blake	$C_3$	Takeda et al. 1985 (A)
E. plicarhachis (Griseb.) Svenson	$C_3$	Ueno et al. 1989 (A)
E. pusilla R. Br.	$C_3 - C_4$ ?	Bruhl et al. 1987 (A. B)
,	$C_3$ -like $C_3$ - $C_4$	Bruhl and Perry 1995 (US); JB (A: Anderson 45, 384, 1667, 1678 CANB; Canning 3543A CANB; Eichler 15636 CANB; Gauba 347, 2422 CANB; Moore 799 CANB. Γ: 29, Bruhl 179 CANB; 29, 30, 31, Bruhl 682 CANB. δ <sup>13</sup> C: -26.7, Bruhl 179)
E. quadrangulata (Michx.) Roem. & Schult.	$C_3$	Ueno et al. 1989 (A)
E. quinquangularis Boeck. E. quinqueflora (Hartmann) O. Schwarz, as	$C_3-C_4$ ? $C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C) Boutton et al. 1980 (A) <sup>a</sup> ; Ueno et al. 1989 (A) <sup>a</sup>
E. pauciflora (Lightf.) Link <sup>a</sup>	~	II 1 1000 (1)
E. radicans (Poir.) Kunth	$C_3$	Ueno et al. 1989 (A)
E. retroflexa (Poir.) Urb.	$C_3+$	Govindarajalu 1975a ([A])
subsp. <i>chaetaria</i> (Roem. & Schult.) T. Koyama	$\mathrm{C}_4$	Ueno and Samejima 1989 (B); Ueno et al. 1989 (A. δ <sup>13</sup> C); Ueno 2004 (B. E [antiserum]); JB (Ae, <i>Sonder 187</i> MEL 1543842. δ <sup>13</sup> C: -10.4, -10.9, <i>Sonder 187</i> )

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
subsp. retroflexa, as E. retroflexa <sup>a</sup>	$C_4$	Bruhl et al. 1987 (Ae. B) <sup>a</sup> ; Ueno et al. 1989 (A. δ <sup>13</sup> C); Bruhl and Perry 1995 (USe); Soros and Dengler 2001 (Ae); JB (Ae, <i>Blake 14421</i> CANB; <i>Godwin C.2967</i> CANB. δ <sup>13</sup> C: -12.9, -12.9, <i>Godwin C.2967</i> )
subsp. subtilissima (Nelmes) K. Lye	$C_4$	JB (Ae, Ellery 15 PRE. δ <sup>13</sup> C: -14.4, -14.7, Ellery 15)
E. reverchonii Svenson	$C_3 - C_4$ ?	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. robbinsii Oakes	$\mathbf{C}_3$	Ueno et al. 1989 (A)
E. rostellata (Torr.) Torr.	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. schaffneri Boeck.	$\mathbf{C}_3$	Ueno et al. 1989 (A)
E. schlechteri C. B. Clarke	$\mathbf{C}_3$	JB (A: Schlechter 3829 PRE)
E. schweinfurthiana Boeck.	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. sellowiana Kunth	$C_3$	Ueno et al. 1989 (A)
E. setifolia (A. Rich.) J. Raynal	$\mathbf{C}_3$	JB (A: <i>Latz 2751</i> CANB)
E. sintenisii Boeck.	$C_3$	Ueno et al. 1989 (A)
E. sp. (?cf. acuta), as E. carniolica C. Koch	$\mathbf{C}_3$	Ueno et al. 1989 (A)
E. sp. aff. nuda (Wilson 5245)	$C_3$	KW (A: Wilson 5012)
E. sp. aff. ochrostachys (Wilson 5166)	$C_3$	KW (A)
E. sp. aff. variegata (Wilson 5248)	$C_3$	KW (A: Wilson 5160)
E. sphacelata R. Br.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); Bruhl et al. 1987 (A. B); Ueno et al. 1989 (A JB (A: <i>Bruhl 124, 579</i> CANB. Γ: 42, <i>Bruhl 124, 51, Bruhl 579</i> ); KW (A: <i>Coveny 5031</i> ; <i>Wilson 2075</i> )
E. spiralis (Rottb.) Roem. & Schult.	$C_3$	Govindarajalu 1975 <i>a</i> ([A]); Takeda et al. 1985 (A. δ <sup>13</sup> C); Ueno et al. 1989 (A); KW (A: <i>Wilson 3666, 5097</i> )
E. squamigera Svenson	$C_3$	Ueno et al. 1989 (A)
E. stenocarpa Svenson	$C_3$	Ueno et al. 1989 (A)
E. subarticulata (Nees) Boeck.	$C_3$	Ueno et al. 1989 (A)
E. subcancellata C. B. Clarke	$\mathbf{C}_4$	JB (Ae, <i>Pringle 4339</i> MEL 1543837, 1543838. δ <sup>13</sup> C: -11.3, -13.0, <i>Pringle 4339</i> MEL 1543837, -11.4, -11.9, <i>Pringle 4339</i> MEL 1543838)
	$C_3$ +	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. sundaica Kern	$C_3$	KW (A: Wilson 5011, 5165)
E. tenuis (Willd.) Schult.	$C_3$	Ueno et al. 1989 (A)
E. tetraquetra Nees	$\mathbf{C}_3$	Govindarajalu 1975 <i>a</i> ([A]); Ueno and Takeda 1992 (A); JB (A: <i>Bruhl 672</i> CANB. Γ: 41, <i>Bruhl 672</i> )
E. tortilis (Link) Schult.	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. tricostata Torr.	$C_3$	Ueno et al. 1989 (A)
E. tuberculosa (Michx.) Roem. & Schult.	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C); LR ( $\delta^{13}$ C: $-30.1$ , [collector's name obscured] 1919 Louisiana)
E. tucumanensis Barros	$C_3$	Guaglianone et al. 1998 (A)
E. uniglumis (Link) Schult.a, as E. halophi- la (Fernald & Brackett) Fernaldb	$C_3$	Ueno et al. 1989 (A) <sup>a,b</sup>
E. variegata (Poir.) C. Presl	$C_3$	Ueno et al. 1989 (A)
E. viridans Kük.	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
E. vivipara Link.	$\mathrm{C}_4$	Ueno et al. 1988 <i>a</i> (A. B. $\delta^{13}$ C); Ueno et al. 1989 (A. $\delta^{13}$ C); Ueno and Sam jima 1990 ( $\delta^{13}$ C); Soros and Dengler 2001 (Ae)
	$C_3 \& C_4 +$	Ueno 1996 (A. US: NAD); Ueno 1998b (A. B. US); Ueno 2004 (B. B [ant serum]. B [kinetics])
	$C_3$ +	Ueno et al. 1988 $a$ (A. B); Ueno and Samejima 1990 ( $\delta^{13}$ C)
E. wichurai Boeck.	$C_3$	Ueno and Takeda 1992 (A. $\Gamma$ )
E. wolfii (A. Gray) A. Gray ex Britton	$C_3$	Ueno et al. 1989 (A. $\delta^{13}$ C)
Epischoenus adnatus Levyns	$\mathbf{C}_3$	LR (δ <sup>13</sup> C: -27.3, Schlechter 7402)
E. cernuus Levyns	$C_3$	SCV (δ <sup>13</sup> C: -24.73, <i>Levyns 8873</i> )
E. complanatus Levyns	$C_3$	JB (A: Esterhuysen 17776 PRE; Stokoe 2162 PRE); SCV (δ <sup>13</sup> C: -26.21, Eterhuysen 11575)
E. dregeanus (Boeck.) Levyns	$\mathbf{C}_3$	SCV ( $\delta^{13}$ C: $-24.45$ , Levyns 9379)
E. gracilis Levyns	$C_3$	JB (A: Esterhuyen 27597 PRE)
E. lucidus (C. B. Clarke) Levyns	$C_3$	SCV (δ <sup>13</sup> C: -25.20, Esterhuysen 11312)
E. villosus Levyns	$C_3$	SCV (δ <sup>13</sup> C: -22.07, Esterhuysen 16927)
Eriophorum angustifolium Honckeny	$C_3$	Bender 1971 (\delta^{13}C); JB (A: <i>Watson</i> , 9 July 1987 pickled fragments at RSBS, ANU)
E. comosum (Wall.) Wall. ex Nees (Erios- cirpus comosus (Wall.) Palla)	$C_3$	Sharma 1973 ([A]); JB (A: Singh 189)
E. latifolium Hoppe	$C_3$	LR ( $\delta^{13}$ C: $-29.6$ , Bec in Arènes 1222)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
E. microstachyum Boeck. (Erioscirpus mi- crostachyus (Boeck.) Palla)	$C_3$	JB (A: <i>Parker 2785</i> )
E. virginicum L. (Eriophoropsis virginica (L.) Palla)	$C_3$	JB (A: Knowlton, NSW 709552; Roy 3950)
Evandra aristata R. Br.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); LR (δ <sup>13</sup> C: -28.5, <i>Drummond 397</i> )
E. pauciflora R. Br.	C <sub>3</sub>	JB (A: Royce 2683 BRI. 8 <sup>13</sup> C: -28.0, Royce 2683)
Everardia montana Ridl.	$C_3$	LR (δ <sup>13</sup> C: -28.2, Wurdeck 1380)
E. montana subsp. duidae (Gilly) T. Koyama & Maguire	$C_3$	JB (A: Steyermark 93322 K)
Exocarya sclerioides (F. Muell.) Benth.	$C_3$	Koyama 1967 ([A]); Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: <i>Brass 18277</i> CANB; <i>Jones 3437</i> CANB)
Exochogyne amazonica C. B. Clarke	$C_3$	JB (A: <i>Steyermark</i> , 1 Sep 1961 BRI. δ <sup>13</sup> C: -26.9, <i>Steyermark</i> , 1 Sep 1961)
Ficinia acuminata (Nees) Nees	$C_3$	SCV (δ <sup>13</sup> C: -25.40, <i>Levyns 11212</i> )
F. angustifolia C. B. Clarke	$C_3$	JB (A: Esterhuysen 90877 K); SCV (δ <sup>13</sup> C: -24.20, Esterhuysen 26427)
F. capillifolia C. B. Clarke	C <sub>3</sub>	JB (A: Fourcade 3017 K)
F. elongata Boeck.	C <sub>3</sub>	JB (A: Stirton 6382 K)
F. fascicularis Nees	$C_3$	JB (A: Acocks 9090 K)
F. filiformis (Lam.) Schrad.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
F. gracilis (Poir.) Schrad.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
F. gydomontana Arnold	C <sub>3</sub>	SCV ( $\delta^{13}$ C: $-21.20$ , Esterhuysen 27706)
F. indica (Lam.) H. Pfeiff.	$C_3$	LR (δ <sup>13</sup> C: -26.8, Schlechter 8402)
F. nodosa (Rottb.) Goetgh., A. M. Muasya	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C)
& D. A. Simpson, as <i>Scirpus nodosus</i> Rottb.	,	· /
F. pallens (Schrad.) Nees var. lithosperma (Boeck.) Arnold	$C_3$	JB (A: Arnold 1007 K)
F. radiata (L.) Kunth (Sickmannia radiata (L. f.) Nees)	$C_3$	JB (A: Arnold 965 PRE)
F. stolonifera Boeck.	$C_3$	SCV (δ <sup>13</sup> C: -25.31, <i>Levyns 6863</i> )
Fimbristylis acicularis R. Br.	$C_4$	Takeda et al. 1985 (Af)
F. acuminata Vahl, as F. rhyticarya F. Muell.	$C_4$	Takeda et al. 1985 ( $\delta^{13}$ C)
F. aestivalis (Retz.) Vahl	$\mathrm{C}_4$	Akita et al. 1969 ([A]); Kuoh and Chiang 1984 (A); Takeda et al. 1985 (Af); Bruhl et al. 1987 (Af. B); Ehleringer et al. 1987 (δ <sup>13</sup> C)
F. annua (Allioni) Roem. & Schult.	$C_4$	Ehleringer et al. 1987 ( $\delta^{13}$ C)
F. aphylla Steud.	$C_4$	LR (δ <sup>13</sup> C: -12.7, Tisserant 3330)
F. autumnalis (L.) Roem. & Schult.	$C_4$	Ueno and Takeda 1992 (A. Γ)
F. bisumbellata (Forsk.) Bubani	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C); Takeda et al. 1985 (Af. $\delta^{13}$ C); Ueno and Takeda 1992 (A)
F. bivalvis (Lam.) K. Lye (F. longiculmis Steud.)	$C_4$	SCV ( $\delta^{13}$ C: $-10.05$ , Rogers 4563)
F. caespitosa R. Br.	$\mathbf{C}_4$	Takeda et al. 1985 (Af)
F. caroliniana (Lam.) Fernald	$\mathbf{C}_4$	Brown 1975 (Af)
F. cephalophora F. Muell.	$\mathbb{C}_4$	Takeda et al. 1985 (Af)
F. compacta Turrill	$C_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C)
F. complanata (Retz.) Link	$\mathrm{C}_4$	Sharma and Mehra 1972 ([A]); Hesla et al. 1982 (A. $\delta^{13}$ C); Ehleringer et al. 1987 ( $\delta^{13}$ C); Ueno et al. 1988 <i>b</i> (USf); Ueno and Takeda 1992 (A. Γ); Ueno et al. 1986 (Af. B)
subsp. keniaeensis (Kük.) K. Lye, as F. keniaeensis Kük.	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
F. consanguinea Kunth	$C_4$	Hofstra et al. 1972 (A. $\Gamma$ )
F. corynocarya F. Muell.	$C_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C)
F. cymosa R. Br., as F. cymosa subsp. spa-	$C_4$	Hofstra et al. 1972 (A. Γ); Prakash et al. 1976 (A) <sup>b</sup> ; Meinzer 1978 (A) <sup>b</sup> ;
thacea (Roth) T. Koyama <sup>a</sup> , as F. spa- thacea Roth <sup>b</sup>	-4	Hnatiuk 1980 (A); Hesla et al. 1982 (δ¹³C); Kuoh and Chiang 1984 (A); Takeda et al. 1985 (Af); Ueno and Takeda 1992 (A)³
F. densa S. T. Blake	$\mathbf{C}_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C)
F. denudata R. Br.	$C_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C); Bruhl et al. 1987 (Af. B)
F. depauperata R. Br.	$C_4$	Takeda et al. 1985 (Af)
F. dichotoma (L.) Vahl	$\overset{-4}{\mathrm{C}_4}$	Hattersley et al. 1977 (Af. B [IL]); Gilliland and Gordon-Gray 1978 (USf); Hesla et al. 1982 (δ¹³C); Kuoh and Chiang 1984 (A); Takeda et al. 1985 (Af); Ueno et al. 1986 (Af. B); Bruhl et al. 1987 (Af. B); Ueno e al. 1988b (USf); Ueno and Takeda 1992 (A. Γ); Ueno 1998a (US); LR (δ¹³C: −10.5, Bon 2186)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
F. diphylloides Makino	$C_4$	Ueno et al. 1986 (Af. B); Ueno and Takeda 1992 (A. Γ)
F. dipsacea (Rottb.) C. B. Clarke subsp.	$C_4$	Ueno et al. 1986 (Af. B); Ueno and Takeda 1992 (A)
verrucifera (Maxim.) T. Koyama		
F. ferruginea (L.) Vahl	$\mathbb{C}_4$	Hofstra et al. 1972 (A. $\Gamma$ ); Hnatiuk 1980 (A); Hesla et al. 1982 ( $\delta^{13}$ C); Take
	_	da et al. 1985 (Af. $\delta^{13}$ C); LR ( $\delta^{13}$ C: $-11.6$ , <i>Jacques-Félix 7218</i> )
F. fimbristyloides (F. Muell.) Druce	$C_4$	Ueno and Takeda 1992 (A)
F. furva R. Br.a, as F. capitata R. Br.b	$C_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C) <sup>a,b</sup>
F. hirsutifolia Govindarajalu	$C_4$	Govindarajalu 1990a ([A])
F. kadzusana Ohuai	$C_4$	Ueno and Takeda 1992 (A)
F. leptoclada Benth. F. leucocolea Benth.	$C_4$	Ueno and Takeda 1992 (A) Telepho et al. 1985 (Af. SIG)
F. littoralis Gaud.	$egin{array}{c} { m C}_4 \\ { m C}_4 \end{array}$	Takeda et al. 1985 (Af. δ <sup>13</sup> C) Akita et al. 1969 ([A]); Carolin et al. 1977 (USf); Takeda et al. 1980 (Af.
r. morans Gaud.	$C_4$	Γ); Takeda et al. 1985 (Af); Ueno et al. 1986 (Af. B)
F. microcarya F. Muell.	$\mathbb{C}_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C)
$F.\ miliacea\ (L.)\ Vahl,\ as\ F.\ quinquangular-$	$\mathbb{C}_4$	Hofstra et al. 1972 (A. Γ); Meinzer 1978 (A); Hesla et al. 1982 $(δ^{13}C)^a$ ;
is Kunth <sup>a</sup>		Kuoh and Chiang 1984 (A); Lin et al. 1993 (Af)
F. nelmesii Kern (Tylocarya cylindrostach- ya Nelmes)	$C_4$	JB (Af: <i>Kerr 21294</i> BM, L)
F. nuda Boeck.	$\mathbf{C}_4$	Takeda et al. 1985 (Af)
F. nutans (Retz.) Vahl	$\mathbf{C}_4$	Takeda et al. 1985 (Af); Ueno and Takeda 1992 (A)
F. obtusifolia (Lam.) Kunth	$\mathbf{C}_4$	LR (δ <sup>13</sup> C: -13.0, <i>Chevalier 20054</i> )
F. oligocephala W. V. Fitzg.	$C_4$	Takeda et al. 1985 (Af)
F. pallida S. T. Blake	$C_4$	Takeda et al. 1985 (Af)
F. pauciflora R. Br.	$\mathbf{C}_{\!\scriptscriptstyle{4}}$	Takeda et al. 1985 (Af); Ueno and Takeda 1992 (A)
F. phaeoleuca S. T. Blake	$\mathbf{C}_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C)
F. pierotii Miq.	$\mathbf{C}_4$	Ueno and Takeda 1992 (A. $\Gamma$ )
F. pilosa Vahl	$\mathbf{C}_4$	LR (δ <sup>13</sup> C: -12.1, <i>Pobéguin 419</i> )
F. polytrichoides (Retz.) R. Br.	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C); Kuoh and Chiang 1984 (A); Takeda et al. 1985 (Af. δ <sup>13</sup> C); Bruhl et al. 1987 (Af. B); JB (Af: <i>Bruhl 204, 443</i> CANB. δ <sup>13</sup> C: -11.1, -11.6, -11.7, <i>Bruhl 443</i> ); LR (δ <sup>13</sup> C: -15.3, <i>Sacleux 1652</i> )
F. pseudomicrocarya Govindarajalu	$\mathbf{C}_4$	Govindarajalu 1990a ([A])
F. pterygosperma R. Br.	$\mathbf{C}_4$	Takeda et al. 1985 (Af)
F. punctata R. Br.	$\mathbf{C}_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C)
F. rara R. Br.	$\mathbf{C}_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C)
F. recta F. M. Bailey	$\mathbf{C}_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C)
F. scabrida Schum. & Thonn.	$C_4$	LR (δ <sup>13</sup> C: -13.2, Chevalier 23501)
F. schoenoides (Retz.) Vahl	$C_4$	Takeda et al. 1985 (Af. $\delta^{13}$ C); Ehleringer et al. 1987 ( $\delta^{13}$ C)
F. schultzii Boeck.	$C_4$	Takeda et al. 1985 (Af)
F. sericea R. Br.	$C_4$	Ueno and Takeda 1992 (A. Γ); Ueno et al. 1986 (Af. B); JB (Af: s. coll. CANB 272779)
F. sieboldii Miq.	$C_4$	Kuoh and Chiang 1984 (A); Ueno et al. 1986 (Af. B); Ueno and Takeda 1992 (A. Γ)
F. solidifolia F. Muell.	$\mathbf{C}_4$	Takeda et al. 1985 (Af)
F. sp. (Carolin 8690)	$\mathbf{C}_4$	Carolin et al. 1977 (USf)
F. sphaerocephala Benth.	$C_4$	Takeda et al. 1985 (Af)
F. splendida C. B. Clarke	$\mathbf{C}_4$	LR (δ <sup>13</sup> C: -11.6, Sita 495)
F. squarrosa Vahl, as F. aestivalis (Retz.) Vahl subsp. squarrosa (Vahl) T. Koya- ma <sup>a</sup>	$C_4$	Takeda et al. 1985 (Af); Ueno et al. 1986 (Af. B) <sup>a</sup> ; Ueno et al. 1988 <i>b</i> (USf) <sup>a</sup> ; Ueno and Takeda 1992 (A) <sup>a</sup>
F. stauntonii Debeaux & Franch. subsp. to- nensis (Makino) T. Koyama	$C_4$	Ueno and Takeda 1992 (A)
F. subbispicata Nees & Meyen ex Nees, as F. tristachya subsp. subbispicata (Nees & Meyen) T. Koyama <sup>a</sup>	$\mathrm{C}_4$	Kuoh and Chiang 1984 (A); Ueno et al. 1986 (Af. B) <sup>a</sup> ; Ueno et al. 1988 $b$ (USf) <sup>a</sup> ; Ueno and Takeda 1992 (A. $\Gamma$ ) <sup>a</sup>
F. tenera Schult.	$\mathbf{C}_4$	Sabnis 1921 ([A])
F. tetragona R. Br.	$C_4$	Carolin et al. 1977 (USf); Takeda et al. 1985 (Af); Bruhl et al. 1987 (Af. B); Bruhl and Perry 1995 (USf)
F. trachycarya F. Muell.	$C_4$	Takeda et al. 1985 (Af)
F. trigastrocarya F. Muell.	$C_4$	Takeda et al. 1985 (Af)
F. tristachya R. Br., as F. marianna Gaud. <sup>a</sup>	$C_4$	Takeda et al. 1985 (Af); JB (Af: <i>Brown</i> MEL 1543831); LR ( $\delta^{13}$ C: $-12.1$ , <i>Poilane</i> 20844) <sup>a</sup>

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
F. uliginosa Hochst. ex Steud.	$\mathrm{C}_{\scriptscriptstyle{4}}$	Raghavendra and Das 1976 (A. Γ)
F. umbellaris (Lam.) Vahl, as F. globulosa (Retz.) Kunth <sup>a</sup>	$\mathrm{C}_4$	Kuoh and Chiang 1984 (A); Ueno and Takeda 1992 (A) <sup>a</sup>
F. variegata Gordon-Gray (Abildgaardia variegata (Gordon-Gray) K. Lye)	$C_3$	JB (A: Ward 1108 BRI)
F. velata R. Br.	$C_4$	Carolin et al. 1977 (USf); Bruhl et al. 1987 (Af. B)
F. xyridis R. Br.	$\mathbf{C}_4$	Takeda et al. 1985 (Af)
Fuirena ciliaris (L.) Roxb., as F. ciliaris var. ciliarisª	$C_3$	Govindarajalu 1969 <i>a</i> ([A]); Hesla et al. 1982 (δ <sup>13</sup> C); Takeda et al. 1985 (A); JB (A: <i>Hartley 13767</i> CANB); SCV (δ <sup>13</sup> C: -26.63, <i>Rogers 13216</i> ) <sup>a</sup>
F. glomerata Lam.	$\mathbb{C}_3$	Takeda et al. 1985 (A. $\delta^{13}$ C)
F. incrassata S. T. Blake	$\mathbf{C}_3$	Takeda et al. 1985 (A); JB (A: <i>Bruhl 445</i> CANB)
F. leptostachya Oliv.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
var. nudiflora C. B. Clarke	$C_3$	SCV (δ <sup>13</sup> C: -26.81, Swynnerton 16027)
F. obcordata P. L. Forbes F. ochreata Kunth, as F. calolepis K.	$ \begin{array}{c} C_3 \\ C_3 \end{array} $	SCV ( $\delta^{13}$ C: $-26.03$ , Maputaland Expedition 14313) Hesla et al. 1982 ( $\delta^{13}$ C)
Schum. $F.\ pachyrrhiza$ Ridl., as $F.\ pachyrrhiza$ var.	$C_3$	Hesla et al. 1982 (δ <sup>13</sup> C); SCV (δ <sup>13</sup> C: -25.04, <i>Pegler 309</i> ) <sup>a</sup>
pachyrrhiza <sup>a</sup>	C	Harland of 1002 (\$13C), LD (\$13C), 20 0 Trans. 1000
F. pubescens (Poir.) Kunth var. pergamentacea Fisch.	$C_3$	Hesla et al. 1982 (δ <sup>13</sup> C); LR (δ <sup>13</sup> C: -26.8, <i>Taton 1068</i> ) Govindarajalu 1969 <i>a</i> ([A])
Val. pergamentacea Fisch.  F. simplex Vahl	$C_3$ $C_3$	Covindarajatu 1909 $a$ ([A]) LR ( $\delta^{13}$ C: $-28.9$ , Stanley 20533, $-34.3$ , Serre Orsay cult., 1972)
F. squarrosa Michx.	$C_3$	JB (A: <i>Radford 15859</i> MEL)
F. stricta Steud.	$C_3$	Hesla et al. 1982 (δ <sup>13</sup> C); Ellery et al. 1992 (δ <sup>13</sup> C); SCV (δ <sup>13</sup> C: -25.86, <i>Eyles</i> 3850)
F. umbellata Rottb.	$C_3$	Govindarajalu 1969 <i>a</i> ([A]); Hesla et al. 1982 (δ <sup>13</sup> C); Takeda et al. 1985 (A); Bruhl et al. 1987 (A. B); JB (A: <i>Bruhl 214</i> CANB; <i>Dharmawardhana</i> 15 CANB. Γ: 43, <i>Bruhl 214</i> ); SCV (δ <sup>13</sup> C: -25.07, <i>Rogers 13277</i> )
F. uncinata Kunth	$C_3$	Govindarajalu 1969a ([A])
F. wallichiana Kunth	$C_3$	Govindarajalu 1969a ([A])
F. zambesiaca K. Lye	$\mathbf{C}_3$	Hesla et al. 1982 (δ <sup>13</sup> C)
Gahnia aspera (R. Br.) Spreng.	$\mathbf{C}_3$	Takeda et al. 1985 (A)
G. baniensis Benl	$\mathbb{C}_3$	LR (δ <sup>13</sup> C: -28.3, <i>Poilane 29003</i> )
G. clarkei Benl	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); KW (A: Wilson 7652)
G. deusta (R. Br.) Benth., as "G. densta" [sic]	$C_3$	Takeda et al. 1985 (A)
G. erythrocarpa R. Br.	$\mathbf{C}_3$	KW (A: Wilson 7653)
G. howeana R. O. Gardner	$C_3$	KW (A: Brown 2003/46)
G. javanica Zoll. & Mor. ex Mor.	$C_3$	Hofstra et al. 1972 (A. $\Gamma$ )
G. lacera (A. Rich.) Steud.	$C_3$	Troughton et al. 1974 ( $\delta^{13}$ C)
G. lanigera (R. Br.) Benth.	$C_3$	Takeda et al. 1985 (A)
G. melanocarpa R. Br.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C) Betts 1920 ([A])
G. procera J. R. Forst. & G. Forst. G. radula (R. Br.) Benth.	$C_3$	Takeda et al. 1985 (A)
G. sieberiana Kunth	$C_3$ $C_3$	Takeda et al. 1985 (A); KW (A: Wilson 7651)
G. subaequiglumis S. T. Blake	C <sub>3</sub>	Takeda et al. 1985 (A); JB (A: <i>Prober 161</i> CANB)
G. trifida Labill.	C <sub>3</sub>	Takeda et al. 1985 (A. $\delta^{13}$ C)
Gymnoschoenus anceps (R. Br.) Nees	$C_3$	KW (A: Jackson NSW 22510)
G. sphaerocephalus (R. Br.) Hook. f.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: <i>Bruhl 635</i> CANB); KW (A: <i>Coveny 6302</i> , <i>6322</i> )
Hellmuthia membranacea (Thunb.) Haines & K. Lye	$C_3$	JB (A: Arnold 705 PRE; van Jaarsveld 4491 PRE)
Hypolytrum bullatum C. B. Clarke	$C_3$	Alves et al. 2002 ([A])
H. compactum Nees & Mey. ex Kunth	$\mathbf{C}_{3}^{J}$	JB (A: van Royen 3212 CANB)
H. glaziovii Boeck.	$C_3$	Alves et al. 2002 ([A])
H. heteromorphum Nelmes	$C_3$	LR (δ <sup>13</sup> C: -29.5, Serre Orsay cult., 1972, -36.3, <i>Lorougnon 1272</i> )
H. jenmanii C. B. Clarke	$C_3$	Koyama 1967 ([A])
H. lancifolium C. B. Clarke	$\mathbf{C}_3$	LR ( $\delta^{13}$ C: $-36.0$ , Le Testu 9334)
H. longifolium (Rich.) Nees	$C_3$	Alves et al. 2002 ([A])
subsp. <i>rubescens</i> (C. B. Clarke) T. Koyama, as <i>H. sylvaticum</i> Poepp. & Kunth subsp. <i>rubescens</i> (C. B. Clarke) T. Koyama	$C_3$	Koyama 1967 ([A])

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
H. nemorum (Vahl) Spreng.	$C_3$	Hofstra et al. 1972 (A. $\Gamma$ ); Takeda et al. 1985 (A. $\delta^{13}$ C); Ehleringer et al. 1987 ( $\delta^{13}$ C); JB (A: <i>Bruhl 478</i> CANB); KW (A: <i>Farina s. n.</i> , Queensland NE)
H. nudum C. B. Clarke	$C_3$	Koyama 1967 ([A])
H. pulchrum (Rudge) H. Pfeiff.	$C_3$	Koyama 1967 ([A]); Alves et al. 2002 ([A]); LR (δ <sup>13</sup> C: -31.7, Sagot 889)
H. rigens Nees	$C_3$	Alves et al. 2002 ([A])
H. schraderianum Nees	$C_3$	Alves et al. 2002 ([A])
H. sp. nov. (Alves et al. 1915)	$\mathbf{C}_3$	Alves et al. 2002 ([A])
H. sphaerostachyum Boeck.	$C_3$	Alves et al. 2002 ([A])
H. stemonifolium T. Koyama	$\mathbf{C}_3$	Alves et al. 2002 ([A])
H. strictum Poepp. ex Kunth	$C_3$	Koyama 1967 ([A])
H. verticillatum T. Koyama	$C_3$	Alves et al. 2002 ([A])
Isolepis cernua (Vahl) Roem. & Schult., as Scirpus cernuus Vahl <sup>a</sup>	$C_3$	Troughton et al. 1974 ( $\delta^{13}$ C); Takeda et al. 1985 (A. $\delta^{13}$ C) <sup>a</sup> ; LR ( $\delta^{13}$ C: $-27.3$ , <i>Dieterlen 705</i> )
I. costata Hochst. ex A. Rich., as Scirpus constatus <sup>a</sup> [sic]	$C_3$	Hesla et al. 1982 $(\delta^{13}C)^a$ ; LR $(\delta^{13}C: -29.6, Peter 41441)$
I. crassiuscula Hook. f. (Eleogiton crassiusculus (Hook. f.) Benth.)	$C_3$	JB (A: van Royen 10857 CANB)
I. fluitans (L.) R. Br., as Eleogiton fluitans (L.) Link <sup>a</sup> , as Scirpus fluitans L. <sup>b</sup>	C <sub>3</sub>	Govindarajalu 1976 ([A]) <sup>b</sup> ; Hesla et al. 1982 (A. δ <sup>13</sup> C) <sup>b</sup> ; Takeda et al. 1985 (A) <sup>b</sup> ; JB (A: Evans 2778 CANB; Telford 10203 CANB; van Royen 11001 CANB); LR (δ <sup>13</sup> C: -29.7, Pappi 749) <sup>a</sup> ; SCV (δ <sup>13</sup> C: -25.71, Dummer 1620)
I. graminoides (R. Haines & K. Lye) K. Lye, as Scirpus graminoides R. Haines & K. Lye	$C_3$	Hesla et al. 1982 (δ <sup>13</sup> C)
I. habra (Edgar) Soják	$C_3$	JB (A: Bruhl ex-147 CANB)
I. hystrix (Thunb.) Nees	$C_3$	LR (8 <sup>13</sup> C: -24.1, <i>Drège 1601b</i> )
I. inundata R. Br., as Scirpus inundatus (R. Br.) Poir.	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C)
I. marginata (Thunb.) A. Dietr., as Scirpus antarcticus L.	$C_3$	Takeda et al. 1985 (A)
I. platycarpa (S. T. Blake) Soják, as Scir- pus platycarpus S. T. Blake	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C)
I. prolifera (Rottb.) R. Br., as Holoschoen- us prolifer (Rottb.) ined. <sup>a</sup>	$C_3$	JB (A: <i>Bruhl 126</i> CANB. Γ: 46, <i>Bruhl 126</i> ); LR (δ <sup>13</sup> C: -26.8, <i>Ecklon s. n.</i> , -34.8, Serre Orsay cult., 1972) <sup>a</sup>
I. sepulcralis Steud., as Scirpus chloros- tachys Levyns <sup>a</sup> , as Isolepis sp. (Rich- ards 17045) <sup>b</sup>	$C_3$	Hesla et al. 1982 $(\delta^{13}C)^a$ ; LR $(\delta^{13}C: -27.1, Richards 17045)^b$
I. setacea (L.) R. Br., as Scirpus setaceus L.ª	$C_3$	Hesla et al. 1982 $(\delta^{13}C)^a$ ; LR $(\delta^{13}C: -28.4, Koernicke FGGE 1774)$
I. tenuissima (Nees) Kunth	$C_3$	SCV (δ <sup>13</sup> C: -24.03, Esterhuysen 28973)
Karinia mexicana (Britton) Reznicek & McVaugh	$C_3$	JB (A: Kral 27601 MO); KW (A: Arsène 8431 P)
Khaosokia caricoides Simpson et al.	$C_3$	Simpson et al. 2005 (δ <sup>13</sup> C); JB (A: Simpson, De Wilde et al. 1886 TCD)
Kobresia laxa Nees	$C_3$	Sharma and Mehra 1970 ([A])
K. myosuroides (Vill.) Fiori, as K. bellardii (Allioni) Degland ex Loiseleur <sup>a</sup>	$C_3$	JB (A: <i>Clokey</i> MEL 1543850; <i>Clokey</i> MEL 1543852; <i>Asplund</i> , 8 Aug 1946 MEL); LR (δ¹³C: -25.9, <i>Humbert s. n.</i> , 1911, Col Arsine) <sup>a</sup>
K. nitens C. B. Clarke	$C_3$	Sharma and Mehra 1970 ([A])
Koyamaea neblinensis W. W. Thomas & G. Davidse	$C_3$	JB (A: Stein 1668 MO)
Kyllinga alata Nees (Cyperus alatus (Nees) F. Muell.)	$C_4$	Getliffe Norris 1983 (A)
K. alba Nees (Cyperus cristatus (Kunth) Mattf. & Kük.)	$\mathbf{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C); Getliffe Norris 1983 (A); KW (Ac: O'Connor 4)
K. auroealata (K. Lye) ined., as Kyllinga alata [auct. non Nees]	$\mathbf{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
K. brevifolia Rottb., as Cyperus brevifolius (Rottb.) Hassk. <sup>a</sup> , as C. breviformis [sic] <sup>b</sup> , as K. colorata (L.) Druce <sup>c</sup>	$C_4$	Akita et al. 1969 ([A]); Govindarajalu 1969b ([A]) <sup>a</sup> ; Carolin et al. 1977 (USc) <sup>a</sup> ; Gilliland and Gordon-Gray 1978 (USc) <sup>c</sup> ; Getliffe Norris 1983 (A) <sup>c</sup> ; Takeda et al. 1985 (Ac) <sup>a</sup> ; Li 1993 (A) <sup>b</sup> ; Bruhl et al. 1987 (Ac. B) Lin et al. 1993 (Ac); JB (Ac: <i>Bruhl 162, Bruhl 163</i> CANB. δ <sup>13</sup> C: –10.4, <i>Bruhl 163</i> , –10.8, <i>Bruhl 162</i> ); KW (Ac: <i>Wilson 678, 3302</i> )

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
subsp. <i>leiolepis</i> (Franch. & Sav.) T. Koya-	$\mathbf{C}_4$	Ueno and Takeda 1992 (A. Γ)
K. bulbosa P. Beauv. (Cyperus richardii Steud.), as K. macrocephala A. Rich.	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
K. cartilaginea K. Schum. (Cyperus cartilagineus (K. Schum.) Mattf. & Kük.)	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
K. chrysantha K. Schum. (Cyperus aureostramineus Mattf. & Kük.)	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
K. crassipes Boeck. (Cyperus bulbipes Mattf. & Kük.)	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
K. elata Steud.	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
K. elatior Kunth (Cyperus pinguis (C. B. Clarke) Mattf. & Kük.)	$\mathrm{C}_4$	Gilliland and Gordon-Gray 1978 (USc); Hesla et al. 1982 (A. δ <sup>13</sup> C); Getliffe Norris 1983 (A); SCV (δ <sup>13</sup> C: -10.31, <i>Medley Wood 3993</i> )
K. erecta C. B. Clarke (Cyperus erectus (K. Schum.) Mattf. & Kük.), as K. colorata [auct.] <sup>a</sup>	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; Getliffe Norris 1983 (A); JB (Ac: <i>Gibbs Russell</i> 2869 BRI)
K. intermedia R. Br. (Cyperus sphaero- ideus L. Johnson & O. Evans)	$\mathrm{C}_4$	KW (Ac: Wilson 1453)
K. intricata Cherm. (Cyperus brevifolius subsp. intricatus (Cherm.) K. Lye), as K. aurata [sensu Napper non Nees]	$\mathrm{C}_4$	Hesla et al. 1982 (A. $\delta^{13}$ C)
K. comosipes (Mattf. & Kük.) Napper (Cyperus comosipes Mattf. & Kük.)	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
<ul> <li>K. melanosperma Nees subsp. bifolius</li> <li>(Miq.) ined., as Cyperus melanospermus (Nees) Valck. Sur. subsp. bifolius</li> <li>(Miq.) Kern</li> </ul>	$C_4$	Govindarajalu 1969b ([A])
subsp. melanosperma, as Cyperus melanospermus (Nees) Valck. Sur.	$\mathrm{C}_4$	Govindarajalu 1969 $b$ ([A]); Hesla et al. 1982 ( $\delta^{13}$ C); Getliffe Norris 1983 (A)
K. microstyla C. B. Clarke (Cyperus microstylis (C. B. Clarke) Mattf. & Kük.)	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
K. nemoralis (Forst. & Forst. f.) Dandy ex Hutch. & Dalziel, as Cyperus kyllin- gia Endl. <sup>a</sup>	$\mathrm{C}_4$	Govindarajalu 1969 <i>b</i> ([A]) <sup>a</sup> ; Hofstra et al. 1972 (A. Γ); Getliffe Norris 1983 (A); LR (δ <sup>13</sup> C: -14.4, <i>Squires 790</i> , -14.6, Serre Orsay cult., 1972); SCV (δ <sup>13</sup> C: -9.53, <i>Fourcade 1966</i> )
<ul><li>K. nervosa Steud. (Cyperus oblongus (C.</li><li>B. Clarke) Kük. subsp. nervosus (Steud.) K. Lye)</li></ul>	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C); McNaughton et al. 1983 (A)
K. odorata Vahl, as Cyperus sesquiflorus (Torr.) Mattf. & Kük. <sup>a</sup> , as K. cylindri- ca Nees <sup>b</sup> , as K. sesquiflora Torr. subsp. cylindrica (Nees) T. Koyama <sup>c</sup>	$C_4$	Govindarajalu 1969 <i>b</i> ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); Hesla et al. 1982 (δ <sup>13</sup> C) <sup>b</sup> εtliffe Norris 1983 (A); Takeda et al. 1985 (Ac) <sup>a</sup> ; Ueno and Takeda 1992 (A) <sup>c</sup> ; KW (Ac: <i>Wilson 1557</i> ); LR (δ <sup>13</sup> C: -13.6, <i>Adam 5802</i> )
K. pauciflora Ridl. (Cyperus ridleyi Mattf. & Kük.)	$\mathrm{C}_4$	Getliffe Norris 1983 (A)
K. polyphylla Willd. ex Kunth, as Cyperus aromaticus (Ridl.) Mattf. & Kük.ª	$\mathrm{C}_4$	Prakash et al. 1976 (A) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); Getliffe Norris 1983 (A); Bruhl et al. 1987 (Ac. B); JB (Ac: <i>Bruhl 512</i> CANB; <i>Dharmawar-dhana 25</i> CANB. δ <sup>13</sup> C: −11.1, −11.2, <i>Bruhl 512</i> ); KW (Ac: <i>Parham 9611</i> )
K. pulchella Kunth (Cyperus teneristolon Mattf. & Kük.)	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C); Getliffe Norris 1983 (A); KW (Ac: <i>Schlechter</i> 4030); SCV ( $\delta^{13}$ C: $-9.99$ , <i>Sister Stephany</i> 27728)
K. pumila Michx., as Cyperus tenuifolius (Steud.) Dandy <sup>a</sup>	$\mathbb{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C); Li et al. 1999 (A) <sup>a</sup>
K. squamulata Vahl, as Cyperus metzii (Hochst. ex Steud.) Mattf. & Kük.	$\mathrm{C}_4$	Rikli 1895 ([A]); Govindarajalu 1969 <i>b</i> ([A]) <sup>a</sup>
K. tenuifolia Steud., as Cyperus triceps (Rottb.) Endl. <sup>a</sup> , as K. triceps Rottb. <sup>b</sup>	$\mathrm{C}_4$	Govindarajalu 1969 $b$ ([A]) <sup>a</sup> ; Raghavendra and Das 1976 (A. $\Gamma$ ) <sup>b</sup>
K. welwitschii Ridl. (Cyperus welwitschii (Ridl.) K. Lye)	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
Kyllingiella microcephala (Steud.) Haines & K. Lye, as Scirpus microcephalus (Steud.) Dandy <sup>a</sup> , as Isolepis microce- phala (Steud.) K. Lye <sup>b</sup>	$C_3$	Druyts-Voets 1970 ([A]) <sup>a</sup> ; Hesla et al. 1982 $(\delta^{13}C)^a$ ; JB (A: Wanntorp 405 PRE); LR $(\delta^{13}C: -29.3, Adam\ 12362)^b$

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
K. polyphylla (A. Rich.) K. Lye, as Isole- pis polyphylla A. Rich. <sup>a</sup>	$C_3$	JB (A: Gillett 12991 EA); LR (δ <sup>13</sup> C: -27.3, Pappi 3859) <sup>a</sup>
Lagenocarpus guianensis Nees	$C_3$	JB (A: Broadway 759 K)
L. rigidus (Kunth) Nees	$C_3$	LR ( $\delta^{13}$ C: $-27.7$ , Irwin et al. 13523)
L. verticillatus (Spreng.) T. Koyama & Maguire	$C_3$	JB (A: Steyermark 89702 BRI)
Lepidosperma aphyllum R. Br.	$C_3$	KW (A: Whaite 4323A; Wilson 2853)
L. avium K. L. Wilson	$\mathbf{C}_3$	KW (A: Forde 905)
L. brunonianum Nees	$\mathbf{C}_3$	KW (A: Newbey 4676; Tindale 150; Wilson 2796)
L. canescens Boeck.	$C_3$	Takeda et al. 1985 (A); KW (A: Beauglehole 37932, Beauglehole 39126)
L. carphoides F. Muell. ex Benth.	$C_3$	Takeda et al. 1985 (A); KW (A: Johnson 7924; Streimann 3312; Whibley 3605; Wilson 3060)
L. concavum R. Br.	$C_3$	KW (A: Beauglehole 25305; Blake 22763; Blakely NSW 463671; Canning 2543C; Durrington 1222; McKay NSW 150413; Phillips NSW 464140; Wilson 464, 2204, 2241, 2391)
L. congestum R. Br.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); KW (A: Symon 6395; Wilson 1079)
L. costale Nees	$C_3$	KW (A: Coveny 7967)
L. curtisiae K. L. Wilson & D. I. Morris	$C_3$	KW (A: Boorman NSW 517775; Wilson 1708)
L. drummondii Benth.	$C_3$	KW (A: Fitzgerald NSW 19781)
L. effusum Benth.	$\mathbb{C}_3$	JB (A: Crisp 5231 CBG); KW (A: Crisp 5231 NSW)
L. elatius Labill.	$C_3$	KW (A: Wilson 10203)
L. ensiforme (Rodway) D. I. Morris	$C_3$	KW (A: Wilson 10200)
L. evansianum K. L. Wilson	$C_3$	KW (A: Wilson 8626)
L. filiforme Labill.	$C_3$	KW (A: Armstrong 866; Beauglehole 25024; Boyd 1871; Constable 7340, 7352, NSW 53920; Coveny 4901; Evans NSW 136815; Henshall SYD 367116; McGillivray 148)
L. forsythii A. A. Hamilton	$C_3$	KW (A: Beauglehole 30224; Coveny 6286, 7374, 10051)
L. gladiatum Labill.	$C_3$	Takeda et al. 1985 (A); KW (A: Beard 7735; Cheel NSW 150436; Jones NSW 150435; Pickard 1123; Telford 1859)
L. gracile R. Br.	$C_3$	KW (A: Blake 18027; Tindale 3913)
L. gunnii Boeck.	$C_3$	KW (A: Adams 1841; Constable 5027; Johnson 7053, 8536; McBarron 12361)
L. inops Rodway ex F. Muell.	$C_3$	JB (A: Bruhl 630 CANB); KW (A: Ratkowsky 545)
L. latens K. L. Wilson	$\mathbb{C}_3$	KW (A: Coveny 598; Evans 2610; Moore 1932; Wilson 8631)
L. laterale R. Br.ª, as L. lineare R. Br.b	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C) <sup>a,b</sup> ; KW (A: Barry 27; Blake 5312; Blaxell 814; Briggs NSW 466982; Campbell & Pickard 1221; Constable 5503; Evans NSW 128084; Johnson 2132, NSW 20431, 156454; McKee 11566; Milthorpe & Cunningham 5523; Salasoo 3663; Wilson 822, 2319, 3992, 3696, 4419) <sup>a</sup>
L. leptostachyum Benth. var. asperatum Kük.	$C_3$	KW (A: Newbey 4678)
L. limicola N. A. Wakefield	$C_3$	KW (A: Constable 4356; Coveny 6132, 6310; Gregson NSW 464158; Johnson 622; Wilson 3206)
L. longitudinale Labill.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); KW (A: <i>Briggs 4278</i> ; <i>Betche</i> NSW 295546; <i>Johnson</i> NSW 79147; <i>Kenneally 7178</i> ; <i>Lucas</i> NSW 150437; <i>Williamson</i> NSW 295762; <i>Wilson 1190, 1601, 2168, 2404</i> )
L. neesii Kunth	$C_3$	KW (A: Coveny 4903; McBarron 10516; Opie & van Rees 128; Wilson 4029)
L. obtusum Kük.	$C_3$	KW (A: Wilson 8882)
L. oldfieldii Hook. f.	$C_3$	KW (A: Rodway NSW 150438)
L. perteres C. B. Clarke	$C_3$	LR (8 <sup>13</sup> C: -25.7, Raynal & Jaffré 16520)
L. pruinosum Kük. var. rigidulum Kük.	$\mathbf{C}_3$	KW (A: Newbey 4699; Wilson 2579)
L. pubisquameum Steud.	$\mathbf{C}_3$	KW (A: Salasoo 4117; Wilson 2834)
L. quadrangulatum A. A. Hamilton	$C_3$	KW (A: Coveny 4893; McGillivray 2298)
L. scabrum Nees	$C_3$	KW (A: Wilson 2695)
L. semiteres F. Muell. ex Boeck.	$C_3$	Takeda et al. 1985 (A); KW (A: Beauglehole 43906; Blake 16830)
L. sp. A1 (Wilson 2578)	$C_3$	KW (A)
L. sp. aff. elatius (Fallding NSW 150145)	$C_3$	KW (A: Boorman NSW 519847; Fallding NSW 150145; Williams K4)
L. sp. B2 (Crisp 4833)	$C_3$	KW (A)
L. sp. E3 (Whaite 4105) L. sp. E4 (Wilson 2703)	$ \begin{array}{c} C_3 \\ C_3 \end{array} $	KW (A) KW (A)
*		
L. sp. F ( <i>Pulley 1481</i> )	$C_3$	KW (A: Pulley 1481; Koch 1208)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
L. sp. F1 (Fitzgerald NSW 19769)	$C_3$	KW (A)
L. sp. I (Wilson 3015)	$\mathbf{C}_3$	KW (A)
L. sp. P (Tindale 166A)	$\mathbf{C}_3$	KW (A)
L. sp. Q (Wilson 2717)	$\mathbb{C}_3$	KW (A)
L. sp. S2 (Coveny 7871)	$\mathbb{C}_3$	KW (A)
L. sp. T2 (Wilson 2982)	$\mathbb{C}_3$	KW (A)
L. sp. U1 ( <i>Tindale 3846</i> )	$\mathbb{C}_3$	KW (A)
L. sp. U3 (Blake 18076)	$\mathbb{C}_3$	KW (A)
L. sp. Z (Wilson 9102)	$\mathbb{C}_3$	KW (A)
L. squamatum Labill.	$C_3$	KW (A: Whaite 4318; Wilson 2954)
L. striatum R. Br.	$C_3$	KW (A: Wilson 2946)
L. tenue Benth.	$C_3$	KW (A: Wilson 2764)
L. tetraquetrum Nees	$C_3$	KW (A: Wilson 3022)
L. tortuosum F. Muell.	$C_3$	KW (A: Fuhrer & Beauglehole 39756; Johnson 7062; Ratkowsky 1606; Tin dale NSW 83944)
L. tuberculatum Nees	$C_3$	Takeda et al. 1985 (A); KW (A: Fitzgerald NSW 19796, NSW 19798)
L. urophorum N. A. Wakefield	$C_3$	Takeda et al. 1985 (A); KW (A: Beauglehole 32798; Coveny 959; Wilson 2277, 3119)
L. ustulatum Steud.	$C_3$	KW (A: Wilson 2949)
L. viscidum R. Br.	$C_3$	Takeda et al. 1985 (A); KW (A: Beauglehole 37332; Coveny 10067; Johnson NSW 365502; Mulham W832; Wilson 1091, 8628)
Lepironia articulata (Retz.) Domin <sup>a</sup> , as L. mucronata Rich. <sup>b</sup>	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C) <sup>a,b</sup> ; JB (A: <i>Bruhl 526</i> CANB; <i>Lazarides 8120</i> CANB); KW (A: <i>Wilson 10195</i> ) <sup>a</sup> ; LR (δ <sup>13</sup> C: −28.1, <i>Poilane 23083</i> ) <sup>a</sup>
Lipocarpha albiceps Ridl.	$\mathbf{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C); LR ( $\delta^{13}$ C: $-11.3$ , Audru 774)
L. chinensis (Osb.) Kern	$\mathrm{C}_4$	Govindarajalu 1974 ([A]); Hesla et al. 1982 (δ¹³C); Ueno and Takeda 1992 (A); SCV (δ¹³C: -10.30, <i>Jeague 520</i> )
L. hemisphaerica (Roth) Goetgh., as Hemi- carpha isolepis Nees <sup>a</sup>	$\mathrm{C}_4$	LR (δ¹³C: -10.4, <i>Leprieur s. n.</i> , St Louis, Senegal) <sup>a</sup> ; SCV (δ¹³C: -9.45, <i>Rogers 6024</i> )
L. kernii (Raym.) Goetgh., as Rikliella ker- nii (Raym.) J. Raynal	$\mathrm{C}_4$	Raynal 1973 (Ac); LR ( $\delta^{13}$ C: $-12.7$ , Schweinfurth 2572) <sup>a</sup>
L. micrantha (Vahl) G. C. Tucker, as Hemicarpha subsquarrosa (Muhl.) Nees	$\mathrm{C}_4$	LR ( $\delta^{13}$ C: $-11.1$ , <i>Hall s. n.</i> , 1866)
L. microcephala (R. Br.) Kunth	$\mathrm{C}_4$	Takeda et al. 1985 (Ac); Ueno et al. 1986 (Ac. B); Bruhl et al. 1987 (Ac. B); Ehleringer et al. 1987 (δ <sup>13</sup> C); Ueno and Takeda 1992 (A); JB (Ac. <i>Bruhl 181, 287</i> CANB)
L. nana (A. Rich.) Cherm.	$\mathbf{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C); SCV ( $\delta^{13}$ C: $-10.34$ , Bolus 6025)
L. occidentalis (A. Gray) G. C. Tucker	$C_4$	JB (Ac: s. coll. MEL 1543861)
(Hemicarpha occidentalis A. Gray)	C	ID (\$13C) 11.0 Extina 1996)
L. prieuriana Steud.	$C_4$	LR ( $\delta^{13}$ C: -11.9, Fotius 1886)
L. raynaliana Govindarajalu	$C_4$	Govindarajalu 1981 ([A])  Povnel 1073 (Ao) <sup>3</sup> : Heele et al. 1082 (\$13C) <sup>3</sup> : IP (Ao: <i>Tenlor</i> : 10652 K): I.P.
L. rehmannii (Ridl.) Goetgh., as Rikliella rehmannii (Ridl.) J. Raynal <sup>a</sup>	$C_4$	Raynal 1973 (Ac) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C) <sup>a</sup> ; JB (Ac: <i>Taylor 10652</i> K); LR (δ <sup>13</sup> C: -12.3, <i>Dinter 7560</i> ) <sup>a</sup>
renmannii (Kidi.) J. Kayllai	$C_3+$	SCV (δ <sup>13</sup> C: -24.57, Bolus 4529)
L. squarrosa (L.) Goetgh., as Rikliella		Sharma 1972 ([A]); Raynal 1973 (Ac); Govindarajalu and Raynal 1976
squarrosa (L.) J. Raynal <sup>a</sup>	$C_4$	([A]); LR ( $\delta^{13}$ C: $-13.5$ , Couderc s. n., 1920, Cambodia)
* · · · · · · · · · · · · · · · · · · ·	C	****
Machaerina anceps (Poir.) Boj. M. falcata (Nees) T. Koyama	$C_3$	LR (8 <sup>13</sup> C: -26.4, Bosser 147)  JB (A: Sleumer BW14012 CANB)
M. insularis (Benth.) T. Koyama	$C_3$ $C_3$	JB (A: Hoogland 8807 CANB); KW (A: Brown 2003/35)
Mapania baldwinii Nelmes	$C_3$ $C_3$	LR ( $\delta^{13}$ C: $-33.5$ , Serre Orsay cult., 1972)
M. bancana (Miq.) Benth. & Hook. f. ex	$C_3$	Koyama 1967 ([A]) <sup>a</sup> ; JB (A: <i>Jacobs 5647</i> CANB); LR (δ <sup>13</sup> C: -28.0, <i>Beccal</i>
B. D. Jacks., as <i>Thoracostachyum</i> bancanum (Miq.) Kurz <sup>a</sup>	$C_3$	3332) <sup>a</sup>
M. coriandrum Nelmes	$C_3$	LR (\delta^{13}C: -31.3, Serre Orsay cult., 1972, -37.4, Lorougnon 1260)
M. cuspidata (Miq.) Uittien, as M. humilis auct. non (Steud.) P. Villar	$C_3$	Koyama 1967 ([A])
M. cuspidata (Miq.) Uittien var. petiolata (C. B. Clarke) Uittien	$C_3$	JB (A: s. coll. MEL 1543834)
M. effusa (C. B. Clarke) T. Koyama, as  Mapaniopsis effusa C. B. Clarke	$C_3$	Koyama 1967 ([A]); Metcalfe 1971 ([A])
M. macrantha (Boeck.) H. Pfeiff.	$C_3$	LR (δ <sup>13</sup> C: -33.4, Serre Orsay cult., 1972)
M. macrocephala (Gaud.) K. Schum.	C <sub>3</sub>	Takeda et al. 1985 (A. $\delta^{13}$ C)
M. macrophylla (Boeck.) H. Pfeiff.	C <sub>3</sub>	Koyama 1967 ([A])

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
M. mannii C. B. Clarke	$C_3$	LR (δ <sup>13</sup> C: -32. ?[illegible], Serre Orsay cult., -36.2, Farron 4099)
M. soyauxii (Boeck.) H. Pfeiff.	$\mathbf{C}_3$	LR (δ <sup>13</sup> C: -35.6, <i>Hallé &amp; Villiers 5429</i> , -35.8, Serre Orsay cult., 1972)
M. sumatrana (Miq.) Benth., as Thoracos-	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C) <sup>a</sup> ; JB (A: <i>Bruhl 308</i> CANB)
tachyum sumatranum (Miq.) Kurza		
M. sylvatica Aubl.	$C_3$	Koyama 1967 ([A]); LR (δ <sup>13</sup> C: -35.4, <i>Mangenot 3</i> )
Mesomelaena graciliceps (C. B. Clarke) K. L. Wilson	$C_3$	KW (A: Newbey 4976; Wilson 2888, 2924, 2957)
M. preissii Nees, as M. stygia <sup>a</sup> [voucher	$C_3$	Takeda et al. 1985 (A)a; KW (A: Canning WA/68 2493; Koch 1729; Wilson
Coveny 8296 re-determined by KLW in NSW]	- 3	2606, 2771, 2782)
M. pseudostygia (Kük.) K. L. Wilson	$C_3$	KW (A: Blake 18149; Coveny 3119; Wilson 2632)
M. stygia (R. Br.) Nees subsp. deflexa (Kük.) K. L. Wilson	$C_3$	KW (A: Hnatiuk 8000012; Mueller s. n. B)
M. stygia subsp. stygia	$C_3$	KW (A: Tindale 3849; Weston 8209; Wilson 2897, 2907, 2919, 2993)
M. tetragona (R. Br.) Benth.	$C_3$	Takeda et al. 1985 (A); JB (A: <i>Bailey</i> CANB 63655); KW (A: <i>Tindale 284</i> ; <i>Wilson 2918</i> ); LR ( $\delta^{13}$ C: $-25.0$ , <i>Home s. n.</i> , Australia)
Microdracoides squamosus Hua	$\mathbf{C}_3$	Chermezon 1933 ([A]); JB (A: <i>Morton K685</i> K. δ <sup>13</sup> C: -26.6, -27.1, <i>Morton K685</i> ); LR (δ <sup>13</sup> C: -28.4, <i>Leeuwenberg 5451</i> , -33.7, Serre Orsay cult., 1972)
Morelotia affinis (Brongn.) S. T. Blake	$\mathbb{C}_3$	JB (A: Bagnall 56270 NSW)
M. gahniiformis Gaud.	$C_3$	JB (A: Henrickson 3490; Ordoney, 14 Jul 1940 CANB)
Neesenbeckia punctoria (Vahl) Levyns	$C_3$	JB (A: Orchard 36 K; Taylor 3266 PRE); LR (δ <sup>13</sup> C: -29.0, McOwan 1688): SCV (δ <sup>13</sup> C: -26.01, Levyns 8328)
Nelmesia melanostachya Van der Veken, as Nelmesia <sup>a</sup>	$C_4$	Lerman and Raynal 1972 (A) <sup>a</sup> ; LR (δ <sup>13</sup> C: -12.8, <i>Gérard 57</i> )
Nemum spadiceum (Lam.) Desv. ex Hamilton, as N. angolensis (C. B. Clarke) J. Raynal ined.	$C_4$	LR (δ <sup>13</sup> C: -11.6, <i>Le Testu 3384</i> )
N. equitans (Kük.) J. Raynal	$C_4$	Raynal 1973 (Af); LR (8 <sup>13</sup> C: -13.2, Robinson 3912)
Oreobolopsis inversa Dhooge & Goetgh.	$C_3$	Dhooge and Goetghebeur 2002 ([A])
O. tepalifera T. Koyama & Guaglianone	$C_3$	Koyama and Guaglianone 1987 ([A])
Oreobolus acutifolius S. T. Blake	$C_3$	JB (A: Bruhl 626 CANB)
O. ambiguus Kük. & Steenis	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C:); JB (A: <i>Hope ANU 16067</i> CANB)
O. distichus F. Muell.	$C_3$	Takeda et al. 1985 (A); JB (A: <i>Gray 4834</i> CANB)
O. kuekenthalii Steenis	$C_3$	JB (A: Nooteboom 2023 CANB)
O. obtusangulus Gaud.	$C_3$	LR ( $\delta^{13}$ C: $-24.1$ , Holm & Iltis 571)
O. oligocephalus W. M. Curtis (Schoeno- ides oligocephalus (W. M. Curtis) O. Seberg)	$\mathbf{C}_3$	JB (A: Bruhl 626 CANB)
O. oxycarpus S. T. Blake	$C_3$	Takeda et al. 1985 (A)
O. pumilio R. Br. subsp. pumilio, as O. pumilio <sup>a</sup>	$C_3$	Takeda et al. 1985 (A) <sup>a</sup> ; JB (A: <i>Telford 3686</i> CBG)
Oxycaryum cubense (Poepp. & Kunth) K. Lye	$C_3$	JB (A: <i>Smith 611</i> PRE; <i>Ward 8044</i> PRE); KW (A: <i>Krapovickas 24620</i> P); LR (δ <sup>13</sup> C: -27.6, <i>Trochain 2135</i> )
Paramapania parvibractea (C. B. Clarke) Uittien	$C_3$	Takeda et al. 1985 (A)
P. radians (C. B. Clarke) Uittien	$C_3$	JB (A: <i>Jacobs 5597</i> CANB; <i>Ramos</i> , Aug 1915 BRI 002215. δ <sup>13</sup> C: -29.2, <i>Ramos</i> , Aug 1915); LR (δ <sup>13</sup> C: -30.4, <i>Ramos 23642</i> )
P. simplex (Ridl.) Uittien	$C_3$	JB (A: Brass 13481 BRI. δ <sup>13</sup> C: -31.7, Brass 13481)
Phylloscirpus acaulis (Philippi) Goetgh. & D. A. Simpson, as Scirpus acaulis Philippi <sup>a</sup>	$C_3$	Ponessa et al. 1997 ([A]); BDW (A: <i>Laegaard S-54783</i> AAU)
P. boliviensis (Barros) Dhooge & Goetgh.	$C_3$	BDW (A: Beck 22360 GENT)
P. deserticola (Philippi) Dhooge & Goetgh.	$C_3$	BDW (A: Laegaard S-54816 AAU)
Pleurostachys gaudichaudii Brongn.	$C_3$	LR (δ <sup>13</sup> C: -30.2, Riedel s. n., 1823)
Principina grandis Uittien	$C_3$	JB (A: Exell 703 BM)
Pseudoschoenus inanus (Thunb.) Oteng- Yeboah	$C_3$	JB (A: <i>Muller 619</i> K)
Ptilothrix deusta (R. Br.) K. L. Wilson, as Ptilanthelium deustum (R. Br.) Kük. <sup>a</sup>	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C) <sup>a</sup> ; JB (A: <i>Bruhl 71</i> CANB; <i>Prober 356</i> CANB); KW (A: <i>Coveny 6687</i> )
Pycreus aethiops (Ridl.) C. B. Clarke (Cyperus aethiops Ridl.)	$C_4$	Hesla et al. 1982 ( $\delta^{13}$ C)

Appendix 1. Continued.

2 .	Photosynthetic	
Species	pathway	References (method: value [as appropriate], voucher [if new record])
P. atroglumosus (Govindarajalu) P. Singh & V. Singh, as Cyperus atroglumosus Govindarajalu	$\mathbf{C}_4$	Govindarajalu 1978 ([A])
P. bipartitus (Torr.) C. B. Clarke, as Cyperus bipartitus Torr.	$C_4$	Li et al. 1999 (A. δ <sup>13</sup> C)
P. compressiformis Cherm. (Cyperus compressiformis (Cherm.) Kük.)	$C_4$	KW (Ac: Leandri 1029 P)
P. diandrus (Torr.) C. B. Clarke, as Cyperus diandrus Torr.	$C_4$	Li et al. 1999 (A. δ <sup>13</sup> C)
P. divulsus (Ridl.) C. B. Clarke (C. divulsus Ridl.)	$C_4$	KW (Ac: Bosser 15364 PRE)
P. fibrillosus (Kük.) Cherm. (P. scaettae Cherm., Cyperus fibrillosus Kük.)	$C_4$	LR (δ <sup>13</sup> C: -12.2, <i>Le Testu 7452</i> )
P. filicinus (Vahl) T. Koyama, as Cyperus filicinus Vahl	$C_4$	Li et al. 1999 (A. $\delta^{13}$ C)
P. flavescens (L.) Beauv. ex Rchb., as Cyperus flavescens L.ª	$\mathrm{C}_4$	Meinzer 1978 (A) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); Kalapos et al. 1997 (δ <sup>13</sup> C); Li et al. 1999 (A. δ <sup>13</sup> C) <sup>a</sup> ; LR (δ <sup>13</sup> C: -11.9, <i>Chevalier 908</i> ); SCV (δ <sup>13</sup> C: -10.93, <i>Pegler 1089</i> )
P. flavescens (L.) Rchb., as Cyperus flavescens L. <sup>a</sup>	$C_3$ +	Li 1993 (A. δ <sup>13</sup> C)
P. flavicomus (Michx.) C. D. Adams, as Cyperus albomarginatus (Mart. & Schrad. ex Nees) Steud. <sup>a</sup> , as C. flavi- comus Michx. <sup>b</sup>	$\mathrm{C}_4$	Downton and Tregunna 1968 ( $\Gamma$ ) <sup>a</sup> ; Tregunna et al. 1970 (Ac. B. $\Gamma$ . $\delta$ <sup>13</sup> C) <sup>a</sup> ; Meinzer 1978 (A) <sup>a</sup> ; Li et al. 1999 (A) <sup>b</sup>
P. flavidus (Retz.) T. Koyama, as Cyperus flavidus Retz. <sup>a</sup> , as P. globosus (All.) Rchb. <sup>b</sup>	$\mathrm{C}_4$	Hofstra et al. 1972 (A. Γ) <sup>b</sup> ; Govindarajalu 1978 ([A]) <sup>a</sup> ; Saxena and Ramakrishnan 1984 (A) <sup>b</sup> ; Takeda et al. 1985 (A. δ <sup>13</sup> C); Ueno et al. 1986 (Ac. B) <sup>b</sup> ; Ueno et al. 1988 <i>b</i> (USc) <sup>b</sup> ; Ueno and Takeda 1992 (A. Γ); Li 1993 (δ <sup>13</sup> C) <sup>a</sup> ; JB (Ac: <i>Lazarides 7350</i> CANB)
P. flavidus (Retz.) T. Koyama, as Cyperus flavidus Retz. <sup>a</sup> , as P. globosus (All.) Rchb. <sup>b</sup>	C <sub>3</sub> +	Saxena and Ramakrishnan 1984 (A)
P. govindarajalui V. S. Raju., as Cyperus decumbens Govindarajalu (non P. de- cumbens T. Koyama)	$C_4$	Govindarajalu 1978 ([A])
P. hildebrandtii C. B. Clarke (Cyperus pseudohildebrandtii Kük.)	$C_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
P. intactus (Vahl) J. Raynal (Cyperus intactus Vahl), as P. ferrugineus (Poir.) C. B. Clarke	$\mathrm{C}_4$	Gilliland and Gordon-Gray 1978 (USc)
P. intermedius (Steud.) C. B. Clarke (Cyperus subintermedius Kük.)	$C_4$	KW (Ac: Audru 6062 P)
P. lanceolatus (Poir.) C. B. Clarke, as Cyperus lanceolatus Poir.	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
P. latespicatus (Boeck.) C. B. Clarke, as Cyperus latespicatus Boeck.	$C_4$	Govindarajalu 1978 ([A])
P. latevaginatus (Govindarajalu) P. & V. Singh, as Cyperus latevaginatus Govindarajalu	$\mathrm{C}_4$	Govindarajalu 1978 ([A])
P. longistolon (A. Peter & Kük.) Napper (Cyperus longistolon A. Peter & Kük.)	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C)
P. luridus (Govindarajalu) P. Singh & V. Singh, as Cyperus luridus Govindarajalu	$\mathrm{C}_4$	Govindarajalu 1978 ([A])
P. macranthus (Boeck.) C. B. Clarke (Cyperus macranthus Boeck.)	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C); KW (Ac: <i>Rudatis 702</i> )
P. macrostachyos (Lam.) J. Raynal, as Cyperus albomarginatus (Mart. & Schrad. ex Nees) Steud. <sup>a</sup> , as C. macrostachyos Lam. <sup>b</sup>	$\mathrm{C}_4$	Carolin et al. 1977 (USc) <sup>b</sup> ; Govindarajalu 1978 ([A]) <sup>b</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); Takeda et al. 1985 (Ac. δ <sup>13</sup> C) <sup>a</sup> ; KW (Ac: <i>Decary 16455</i> P) <sup>b</sup>
P. mundii Nees, as Cyperus mundtii [sic] (Nees) Kunth <sup>a</sup>	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C); Li 1993 (A. $\Gamma$ ) <sup>a</sup> ; LR ( $\delta^{13}$ C: $-12.0$ , Gaston 688)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
P. nervulosus (Kük.) ined., as Cyperus nervulosus (Kük.) S. T. Blake	$C_4$	Carolin et al. 1977 (USc)
P. niger (Ruiz & Pav.) Cufod., as Chloro- cyperus cimicinus (J. Presl & C. Presl) Rikli	$\mathbf{C}_4$	Rikli 1895 ([A])
P. niger (Ruiz & Pav.) Cufod. subsp. ele- gantulus (Steud.) K. Lye (Cyperus ni- ger Ruiz & Pav. subsp. elegantulus (Steud.) K. Lye), as P. elegantulus (Steud.) C. B. Clarke <sup>a</sup>	$C_4$	Lerman and Raynal 1972 (A) <sup>a</sup> ; Hesla et al. 1982 ( $\delta^{13}$ C) <sup>a</sup> ; LR ( $\delta^{13}$ C: $-12.2$ , Schimper 118) <sup>a</sup>
P. nigricans (Steud.) C. B. Clarke (Cyperus nigricans Steud.)	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C); KW (Ac: <i>Lye</i> 5288 P)
P. nitidus (Lam.) J. Raynal (Cyperus nitidus Lam.)	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C); Ellery et al. 1992 ( $\delta^{13}$ C); SCV ( $\delta^{13}$ C: $-10.02$ , $Ty$ - son 1681)
P. pelophilus (Ridl.) C. B. Clarke (Cyperus pelophilus Ridl.)	$\mathrm{C}_4$	Hesla et al. 1982 (δ <sup>13</sup> C); SCV (δ <sup>13</sup> C: -10.93, Russell 2076)
P. permutatus (Boeck.) Napper (Cyperus permutatus Boeck.)	$\mathrm{C}_4$	Hesla et al. 1982 ( $\delta^{13}$ C)
P. pervillei (Boeck.) C. B. Clarke (Cyperus pervillei Boeck.)	$\mathrm{C}_4$	KW (Ac: Humbert 4045 P)
P. plumbeonuceus (Govindarajalu) P. Singh & V. Singh, as Cyperus plumbeonuceus Govindarajalu	$\mathrm{C}_4$	Govindarajalu 1978 ([A])
P. plurinodosus (Govindarajalu) P. Singh & V. Singh, as Cyperus plurinodosus Govindarajalu	$\mathrm{C}_4$	Govindarajalu 1978 ([A])
P. polystachyos (Rottb.) Beauv., as Cyperus polystachyos Rottb. <sup>a</sup>	$\mathrm{C}_4$	Carolin et al. 1977 (USc) <sup>a</sup> ; Govindarajalu 1978 ([A]) <sup>a</sup> ; Takeda et al. 1985 (Ac) <sup>a</sup> ; Bruhl et al. 1987 (Ac. B); Hesla et al. 1982 (δ¹³C); Ueno et al. 1986 (Ac); Ueno and Takeda 1992 (A. Γ); Li 1993 (A. δ¹³C) <sup>a</sup> ; Lin et al. 1993 (Ac); Bruhl and Perry 1995 (USc); Li et al. 1999 (A. δ¹³C) <sup>a</sup> ; Soros and Dengler 2001 (Ac); JB (Ac: <i>Bruhl 190</i> CANB); KW (Ac: <i>Wilson 1447</i> )
var. laxiflorus Benth.	$C_4$	SCV (δ <sup>13</sup> C: -10.43, <i>McOwan 1326</i> )
P. pumilus (L.) Nees, as Cyperus pumilus L. <sup>a</sup>	$C_4$	Govindarajalu 1978 ([A]) <sup>a</sup> ; Hnatiuk 1980 (A); Hesla et al. 1982 (δ <sup>13</sup> C); LR (δ <sup>13</sup> C: -11.3, <i>Chevalier 9857</i> )
P. puncticulatus (Vahl) Nees, as Cyperus punticulatus Vahl	$C_4$	Govindarajalu 1978 ([A])
P. sanguinolentus (Vahl) Nees, as Cyperus sanguinolentus Vahl <sup>a</sup> , as C. sanguissolentus [sic] <sup>b</sup> , as Cyperus sanguinolentus subsp. sanguinolentus <sup>c</sup>	$C_4$	Hattersley et al. 1977 (Ac. B [IL]) <sup>a</sup> ; Govindarajalu 1978 ([A]) <sup>c</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); Ueno et al. 1986 (Ac); Ueno et al. 198 <i>b</i> (USc); Ueno and Takeda 1992 (A. Γ); Li and Jones 1994 (A) <sup>b</sup> ; KW (Ac: <i>Wilson 1444</i> )
subsp. cyrtostachys (Miq.) S. Karthikey- an, as Cyperus sanguinolentus subsp. cyrtostachys (Miq.) Kern	$\mathrm{C}_4$	Govindarajalu 1978 ([A])
var. micronux (C. B. Clarke) S. Karthi- keyan, as Cyperus sanguinolentus var. micronux (C. B. Clarke) Kük.	$\mathrm{C}_4$	Govindarajalu 1978 ([A])
P. stramineus (Nees) C. B. Clarke, as Cyperus substramineus Kük.	$\mathrm{C}_4$	Govindarajalu 1978 ([A])
P. stricticulmis (Govindarajalu) P. Singh & V. Singh, as Cyperus stricticulmis Govindarajalu	$\mathrm{C}_4$	Govindarajalu 1978 ([A])
P. sulcinux (C. B. Clarke) C. B. Clarke, as Cyperus sulcinux C. B. Clarke	$\mathrm{C}_4$	Govindarajalu 1978 ([A])
P. unioloides (R. Br.) Urb., as Cyperus unioloides R. Br. <sup>a</sup>	$\mathrm{C}_4$	Akita et al. 1969 ([A]); Govindarajalu 1978 ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C); JB (Ac: <i>Bruhl</i> , 16 Apr 1986 CANB)
Queenslandiella hyalina (Vahl) Ballard, as Cyperus hyalinus Vahl <sup>a</sup> , as Queenslan- diella <sup>b</sup>	$C_4$	Lerman and Raynal 1972 (A) <sup>b</sup> ; Govindarajalu 1975 <i>b</i> ([A]) <sup>a</sup> ; JB (Ac: <i>Bogdan 5353</i> K; <i>Cooray 69121001R</i> PDA; <i>van Oostroom 13596</i> CANB); LR (δ <sup>13</sup> C: -13.7, <i>Boivin s. n.</i> , ca. 1850)
Reedia spathacea F. Muell.	$C_3$	Takeda et al. 1985 (A); JB (A: <i>Maslin 1682c</i> CANB)
Rhynchocladium steyermarkii (T. Koyama) T. Koyama	$C_3$	JB (A: <i>Davidse 27377</i> NY)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
Rhynchospora affinis W. Fitzg.	$\mathrm{C}_4$	Takeda et al. 1980 (Ar); Takeda et al. 1985 (Ar. $\delta^{13}$ C); Ueno and Koyama 1987 (Ar)
R. alba (L.) Vahl	$C_3$	Takeda et al. 1980 (A. Γ); Ueno and Koyama 1987 (A)
R. albescens (Miq.) Kük.	$C_3$	Ueno and Koyama 1987 (A)
R. albiceps Kunth	$C_3$	Ueno and Koyama 1987 (A)
R. albomarginata Kük.	$C_3$	Ueno and Koyama 1987 (A)
R. albotuberculata Kük.	$C_3$ $C_4$	Ueno and Koyama 1987 (Ar)
R. amazonica Poepp. & Kunth	$C_4$ $C_3$	Ueno and Koyama 1987 (A)
R. andina Kük.	C <sub>3</sub>	Ueno and Koyama 1987 (A)
R. angustifolia Palla		
R. arechavaletae Boeck.	$C_3$	Ueno and Koyama 1987 (A) Ueno and Koyama 1987 (A)
"R. argentina Standley" [?= R. argentea Standley]	$C_3$ $C_3$	Ueno and Koyama 1987 (A)
R. aripoensis Britton	$C_3$	Ueno and Koyama 1987 (A)
R. aristata Boeck.	C <sub>3</sub>	Ueno and Koyama 1987 (A)
	-	
R. armerioides J. Presl & C. Presl	$C_4+$	Ueno and Koyama 1987 (Ac); JB (Ac: McKee 10847 CANB. δ <sup>13</sup> C: -10.0, McKee 10847)
R. baldwinii A. Gray	$C_3$	Ueno and Koyama 1987 (A)
R. barbata (Vahl) Kunth	$C_4+$	Ueno and Koyama 1987 (Ac); JB (Ac: <i>King 721</i> CANB; <i>McKee 10573</i> CANB. δ <sup>13</sup> C: -10.4, -10.8, <i>King 721</i> )
R. berteroi (Spreng.) C. B. Clarke, as R. pusilla (Sw.) Griseb.	$C_3$	Ueno and Koyama 1987 (A)
R. biflora Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. brachychaeta C. Wright	$C_3$	Ueno and Koyama 1987 (A)
R. brevirostris Griseb.	$C_3$	Ueno and Koyama 1987 (A)
R. brownii Roem & Schult., as R. rugosa <sup>a</sup> , as R. rugosa subsp. brownii (Roem. & Schult.) Koyama <sup>b</sup>	C <sub>3</sub> +	Govindarajalu 1975 $a$ ([A]) $^a$ ; Gilliland and Gordon-Gray 1978 (US) $^a$ ; Takeda et al. 1980 (A. $\Gamma$ ) $^b$ ; Ueno and Koyama 1987 (A) $^a$
R. cacuminicola Gale	$C_3$	Ueno and Koyama 1987 (A)
R. caduca Elliott	$C_3$	Ueno and Koyama 1987 (A)
R. californica Gale	$C_3$	Ueno and Koyama 1987 (A)
R. candida (Nees) Boeck.	$C_3$	Ueno and Koyama 1987 (A); LR (δ <sup>13</sup> C: -27.0, <i>Humbert 18779</i> )
R. capillacea Torr.	$C_3$	Ueno and Koyama 1987 (A)
R. capitata (Kunth) Roem. & Schult.	$C_4$	Ueno and Koyama 1987 (Ar)
R. caracasana (Kunth) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. cariciformis Nees	C <sub>3</sub>	Ueno and Koyama 1987 (A)
R. cephalantha A. Gray (incl. R. cephalan-	C <sub>3</sub>	Ueno and Koyama 1987 (A); JB (A: <i>Smith</i> , 17 Aug 1939 BRI. $\delta^{13}$ C: $-26.5$
tha var. pleiocephala Fernald & Gale)  R. cephalotes (L.) Vahl	C <sub>3</sub>	Smith, 17 Aug 1939) Ueno and Koyama 1987 (A); JB (A: McKee 10709 CANB. δ <sup>13</sup> C: -26.9,
R. cernua Griseb.		-27.5, McKee 10709)
	$C_3$	Ueno and Koyama 1987 (A)
R. chalarocephala Fernald & Gale	$C_3$	Ueno and Koyama 1987 (A)
R. chapmanii M. A. Curtis	$C_3$	Ueno and Koyama 1987 (A)
R. chinensis Nees & Meyen ex Wight R. fauriei Franch., as R. chinensis subsp. fauriei (Franch.) Koyama	$C_3$ $C_3$	Takeda et al. 1980 (A. $\Gamma$ ) Takeda et al. 1980 (A)
R. ciliaris (Michx.) Vahl	$C_3$	Ueno and Koyama 1987 (A)
R. ciliata Vahl, as R. nervosa subsp. ciliata (Vahl) T. Koyama <sup>a</sup> , as Dichromena ciliata Vahl <sup>b</sup>	$C_3$	Thomas 1984 (A) <sup>a</sup> ; LR (δ <sup>13</sup> C: -30.4, <i>Husnot 31</i> ) <sup>b</sup>
R. ciliolata Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. colorata (L.) H. Pfeiff., as R. stellata (Lam.) Griseb. <sup>a</sup>	$C_3$	Thomas 1984 (A); Ueno and Koyama 1987 (A) <sup>a</sup> ; JB (A: <i>s. coll.</i> MEL 1543827)
R. comata (Link) Roem. & Schult.	$C_3$	Ueno and Koyama 1987 (A)
R. compressa J. Carey ex Chapm.	$C_3$	Ueno and Koyama 1987 (A)
R. confinis (Nees) C. B. Clarke	C <sub>3</sub>	Ueno and Koyama 1987 (A)
R. confusa F. Ballard (Syntrinema brasi-	$C_3$ $C_4$	Ueno and Koyama 1987 (Ar); JB (Ar, <i>Luetzelburg 1223</i> M. δ <sup>13</sup> C: -10.0,
liense Radk. & H. Pfeiff.)		Luetzelburg 1223)
R. consanguinea (Kunth) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. coriifolia Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. corniculata (Lam.) A. Gray	$C_3$	Ueno and Koyama 1987 (A)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
R. corymbosa (L.) Britton, as R. aurea Vahl <sup>a</sup>	C <sub>3</sub>	Hofstra et al. 1972 (A. Γ); Govindarajalu 1975 <i>a</i> ([A]); Gilliland and Gordon-Gray 1978 (US); Prakash et al. 1976 (A) <sup>a</sup> ; Takeda et al. 1980 (A); Bruhl et al. 1987 (A. B); Ueno and Koyama 1987 (A); JB (A: <i>Bruhl 196</i> CANB; <i>Pullen 8152</i> CANB. Γ: 46, <i>Bruhl 196</i> )
R. cubensis Griseb.	$C_3$	Ueno and Koyama 1987 (A)
R. curtissii Britton	$C_3$	Ueno and Koyama 1987 (A)
R. curvula Griseb.	$C_4$	Ueno and Koyama 1987 (Ar)
R. cyperoides (Sw.) Mart.	$C_3$	Ueno and Koyama 1987 (A); JB (A: <i>Eggers</i> Jul 1881 BRI. δ <sup>13</sup> C: -25.5, <i>Eggers</i> , Jul 1881)
R. decurrens Chapm.	$C_3$	Ueno and Koyama 1987 (A)
R. dentinux C. B. Clarke	$C_4$	Ueno and Koyama 1987 (Ar)
R. diamantina C. B. Clarke ex Kük.	$\mathbb{C}_4$	Ueno and Koyama 1987 (Ar)
R. dissitiflora Steud.	$C_3$	Ueno and Koyama 1987 (A)
R. divergens Chapm. ex M. A. Curtis	$C_3$	Ueno and Koyama 1987 (A)
R. dives Standley, as R. orizabensis C. B. Clarke	$C_3$	Ueno and Koyama 1987 (A)
R. duckei R. Gross	$C_3$	Ueno and Koyama 1987 (A)
R. ebracteata (Standley) H. Pfeiff.	$\mathbb{C}_3$	Ueno and Koyama 1987 (A)
R. elatior Kunth	$C_4$	Ueno and Koyama 1987 (Ar)
R. elliottii A. Dietrich, as R. schoenoides (Elliott) A. Wood	$C_3$	Ueno and Koyama 1987 (A)
R. elongata Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. emaciata (Nees) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. exaltata Kunth	$C_3$	Ueno and Koyama 1987 (A)
R. eximia (Nees) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. faberi C. B. Clarke	$C_3$	Akita et al. 1969 ([A]); Takeda et al. 1980 (A. Γ); Ueno and Koyama 1987 (A)
R. fascicularis (Michx.) Vahl	$C_3$	Ueno and Koyama 1987 (A)
R. filifolia A. Gray	$C_3$	Ueno and Koyama 1987 (A)
R. filiformis Vahl, as R. podosperma C. Wright	$C_3$	Ueno and Koyama 1987 (A)
R. flexuosa C. B. Clarke	$C_3$	Ueno and Koyama 1987 (A)
R. fusca (L.) Aiton f.	$C_3$	Ueno and Koyama 1987 (A)
R. gigantea Link	$C_3$	Ueno and Koyama 1987 (A)
R. glaziovii Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. globosa (Kunth) Roem. & Schult.	$\mathbf{C}_4$	Ueno and Koyama 1987 (Ar)
R. globularis (Chapm.) Small	$C_3$	Ueno and Koyama 1987 (A)
R. glomerata (L.) Vahl R. gollmeri Boeck.	$C_3$	Ueno and Koyama 1987 (A) Ueno and Koyama 1987 (A)
R. gracilenta A. Gray	$C_3$	
R. gracillima Thwaites	$C_3$ $C_3$	Ueno and Koyama 1987 (A) Govindarajalu 1975a ([A]); Ueno and Koyama 1987 (A)
R. graminea Uittien	$C_3$	Ueno and Koyama 1987 (A)
R. grayi Kunth	$C_3$	Ueno and Koyama 1987 (A)
R. grisebachii Boeck. ex Urb.	C <sub>3</sub>	Ueno and Koyama 1987 (A)
R. hassleri C. B. Clarke	$C_3$	Ueno and Koyama 1987 (A)
R. heterocaulis C. B. Clarke	$C_4$	Ueno and Koyama 1987 (Ac)
R. heterochaeta S. T. Blake <sup>a</sup> , as R. longise- tis R. Br. [voucher re-determined by KLW at NSW] <sup>b</sup> , as R. wightiana (Nees) Steud. [voucher Ramos 21743 re-determined by KLW at NSW] <sup>c</sup>	$C_4$	Takeda et al. 1980 (Ar) <sup>a</sup> ; Takeda et al. 1985 (Ar. δ <sup>13</sup> C) <sup>a</sup> ; Ueno and Koyama 1987 (Ar) <sup>a,b,c</sup> ; JB (Ar, <i>Bruhl 213</i> CANB) <sup>a</sup>
R. hieronymii Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. hirsuta Vahl	$C_3$	Ueno and Koyama 1987 (A)
R. hirta (Nees) Boeck.	$C_4$	Ueno and Koyama 1987 (Ac)
R. hispidula (Vahl) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. holoschoenoides (Rich.) Herter	$C_3$	Gilliland and Gordon-Gray 1978 (US); Takeda et al. 1980 (A)
R. inexpansa (Michx.) Vahl	$C_3$	Ueno and Koyama 1987 (A)
R. joveroensis Britton	C <sub>3</sub>	Ueno and Koyama 1987 (A)
R. junciformis (Kunth) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. knieskernii J. Carey	$C_3$	Ueno and Koyama 1987 (A)
R. kunthii Nees ex Kunth	$C_3$	Ueno and Koyama 1987 (A)
R. lapensis C. B. Clarke	$C_3$	Ueno and Koyama 1987 (A)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
R. leae C. B. Clarke	$\mathrm{C}_4$	Takeda et al. 1980 (Ar); Takeda et al. 1985 (Ar. $\delta^{13}$ C); Ueno and Koyama 1987 (Ar)
R. lechleri Steud.	$C_3$	Ueno and Koyama 1987 (A)
2. leptorrhyncha C. Wright	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
2. lindeniana Griseb.	$C_3$	Ueno and Koyama 1987 (A)
. longibracteata Rottb.	$C_3$	Ueno and Koyama 1987 (A); JB (A: <i>McKee 107572</i> CANB. δ <sup>13</sup> C: -24.0. <i>McKee 107572</i> )
. longiflora C. Presl	$C_3$	Ueno and Koyama 1987 (A)
. longisetis R. Br.	$\mathbf{C}_4$	Takeda et al. 1980 (Ar); Takeda et al. 1985 (Ar. $\delta^{13}$ C)
. luzuliformis Boeck.	$C_3$	Ueno and Koyama 1987 (A)
. macrochaeta Steud.	$C_3$	Ueno and Koyama 1987 (A)
. malasica C. B. Clarke	$C_3$	Takeda et al. 1980 (A); Ueno and Koyama 1987 (A)
. marisculus Nees	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
megalocarpa A. Gray	$C_3$	Ueno and Koyama 1987 (A)
. mexicana (Liebm.) Steud.	$C_4$	Ueno and Koyama 1987 (Ac)
. micrantha Vahl	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
. microcarpa Baldwin ex A. Gray	$C_3$	Ueno and Koyama 1987 (A)
. miliacea (Lam.) A. Gray	$C_3$	Ueno and Koyama 1987 (A)
. mixta Britton	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
nardifolia (Kunth) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
. nervosa (Vahl) Boeck., as Dichromena	$C_3$	Ueno and Koyama 1987 (A); JB (A: s. coll. MEL 153840); LR (δ <sup>13</sup> C:
nervosa Vahla	3	-34.1, Serre Orsay cult., 1972) <sup>a</sup>
. nipensis Britton	$C_3$	Ueno and Koyama 1987 (A)
. nitens (Vahl) A. Gray	$C_3$	Ueno and Koyama 1987 (A)
. nivea Boeck.	$\mathbf{C}_{3}^{J}$	Ueno and Koyama 1987 (A)
. odorata C. Wright ex Griseb.	$C_3$	Ueno and Koyama 1987 (A)
. oligantha A. Gray	$C_3$	Ueno and Koyama 1987 (A)
organensis C. B. Clarke, as R. rostrata Lindm.	$C_3$	Ueno and Koyama 1987 (A)
. patuligluma C. B. Clarke ex Lindm., as R. pallida (Nees) Steud.	$C_3$	Ueno and Koyama 1987 (A)
. paraensis Schrad. ex Kunth	$C_3$	Ueno and Koyama 1987 (A)
. perrieri Cherm.	$C_3$	Gilliland and Gordon-Gray 1978 (US); Takeda et al. 1980 (A); Hesla et a
		1982 (δ <sup>13</sup> C); Ueno and Koyama 1987(A)
. pilosa (Kunth) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
. plumosa Elliott	$C_3$	Ueno and Koyama 1987 (A)
. polyantha Steud.	$C_3$	Ueno and Koyama 1987 (A)
. polyphylla Vahl	$C_3$	Ueno and Koyama 1987 (A)
. praecincta Maury	$\mathbb{C}_3$	Ueno and Koyama 1987 (A)
. pruinosa Griseb.	$C_3$	Ueno and Koyama 1987 (A)
. pterochaeta F. Muell.	$C_4$	Takeda et al. 1980 (Ar); Takeda et al. 1985 (Ar. δ <sup>13</sup> C); Ueno and Koyama 1987 (Ar); KW (A: <i>Blake 13395</i> )
. pubera (Vahl) Boeck.	$\mathbb{C}_3$	Thomas 1984 (A); Ueno and Koyama 1987 (A)
. punctata Elliott	$C_3$	Ueno and Koyama 1987 (A)
. pusilla Chapm. ex M. A. Curtis <sup>a</sup> , as R. intermixta C. Wright <sup>b</sup>	$C_3$	Ueno and Koyama 1987 (A) <sup>a,b</sup>
. racemosa C. Wright	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
2. radicans (Schlecht. & Cham.) H. Pfeiff.	$\mathbb{C}_3$	Ueno and Koyama 1987 (A)
2. radicans subsp. microcephala (Bertero ex Spreng.) W. W. Thomas, as R. mi- crocephala (Bertero ex Spreng.) Kük. <sup>a</sup>	$C_3$	Thomas 1984 (A); Ueno and Koyama 1987 (A) <sup>a</sup>
2. rariflora (Michx.) Elliott	$C_3$	Ueno and Koyama 1987 (A)
. recurvata (Nees) Steud.	$C_3$	Ueno and Koyama 1987 (A)
. reptans (Rich.) Boeck., as Dichromena reptans (Rich.) Pers. <sup>a</sup>	$C_3$	Thomas 1984 (A); Ueno and Koyama 1987 (A); LR ( $\delta^{13}$ C: $-28.5$ , <i>Smith</i> $2113$ ) <sup>a</sup>
. ridleyi C. B. Clarke	$C_3$	Ueno and Koyama 1987 (A)
. robusta (Kunth) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
2. roraimae Kük.	$C_3$	Ueno and Koyama 1987 (A)
2. rubra (Lour.) Makino subsp. rubra, as	$C_4$	Govindarajalu 1975 $a$ ([A]); Takeda et al. 1980 (Ar. $\Gamma$ ); Takeda et al. 1980
R. parva (Nees) Steud. var. boninensis (Nakai ex Tuyama) T. Koyama <sup>a</sup>	<b>C</b> 4	(Ar) <sup>a</sup> ; Bruhl et al. 1987 (Ar. B); Gilliland and Gordon-Gray 1978 (U Ueno and Koyama 1987 (Ar); Ueno et al. 1986 (Ar. B); Ueno et al. 1988 (USr); Bruhl and Perry 1995 (USr); Soros and Dengler 2001 (Ar); JB (Ar, <i>Bruhl 573</i> CANB)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
subsp. africana J. Raynal	$C_4$	Takeda et al. 1980 (Ar); SCV ( $\delta^{13}$ C: $-10.21$ , Schlechter 12090)
R. rufa (Nees) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. rugosa (Vahl) Gale	$\mathbf{C}_3$	Takeda et al. 1980 (A)
R. schiedeana (Schlecht.) Kunth	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
R. schomburgkiana (Boeck.) Koyama (Micropapyrus viviparoides Suess.)	$C_3$	Ueno and Koyama 1987 (A); JB (A: Luetzelburg 22381 M)
R. scirpoides (Torr.) Griseb.	$C_3$	Ueno and Koyama 1987 (A)
R. sclerioides Hook. & Arn.	$C_3$	Ueno and Koyama 1987 (A)
R. scutellata Griseb. (R. pringlei Greenman)	$C_3$	Ueno and Koyama 1987 (A); JB (A: s. coll. MEL 1543836)
R. seslerioides Griseb.	$C_3$	Ueno and Koyama 1987 (A)
R. setigera (Kunth) Boeck.	$C_3$	Ueno and Koyama 1987 (A); JB (A: <i>Montes 1173</i> . δ <sup>13</sup> C: -27.6, -27.8, <i>Montes 1173</i> )
R. shaferi Britton	$C_3$	Ueno and Koyama 1987 (A)
R. siguaneana Britton	C <sub>3</sub>	Ueno and Koyama 1987 (A)
R. simplex (Kük.) Kük.	$C_3$	Ueno and Koyama 1987 (A)
R. solitaria R. M. Harper	$C_3$	Ueno and Koyama 1987 (A)
R. sp. (McKee 10493 CANB)	$C_3$	JB (A. $\delta^{13}$ C: $-25.0$ , $-25.1$ )
R. sp. A (Wilson 5171), as R. exserta C. B. Clarke <sup>a</sup>	$C_4$	Ueno and Koyama 1987 (Ar) <sup>a</sup> ; KW (A)
R. sp. B (Cowie 1123)	$C_3$	KW (A: Cowie 1123, Craven 6196)
R. sp. C (Dunlop 5330)	$C_4$	KW (A)
R. splendens Lindm.	$C_3$	Ueno and Koyama 1987 (A)
R. stenocarpa Kunth	$C_3$	Ueno and Koyama 1987 (A)
R. stenophylla Chapm.	C <sub>3</sub>	Ueno and Koyama 1987 (A)
R. subimberbis Griseb.	$C_4$	Ueno and Koyama 1987 (Ac)
R. subplumosa C. B. Clarke	$C_4$	Ueno and Koyama 1987 (Ac#)
R. subquadrata Cherm.	$C_3$	Ueno and Koyama 1987 (A)
R. subtenuifolia Kük., as R. submarginata Kük. [voucher re-determined by KLW at NSW] <sup>a</sup> , as R. tenuifolia Benth. non Griseb. <sup>b</sup>	$C_4$	Takeda et al. 1980 (Ar) <sup>b</sup> ; Bruhl et al. 1987 (Ar. B); Ueno and Koyama 1987 (Ar) <sup>a</sup> ; Takeda et al. 1985 (Ar. δ <sup>13</sup> C) <sup>b</sup> ; JB (Ar, <i>Bruhl 344</i> CANB)
R. subtilis Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. tenella (Nees) Boeck.	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
R. tenerrima Nees ex Spreng. subsp. tener- rima, as R. setacea (Berg) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. tenuifolia Griseb.	$C_3$	Ueno and Koyama 1987 (A)
R. tenuis Link	$C_3$	Ueno and Koyama 1987 (A)
R. terminalis (Nees) Steud.	$C_4$	Ueno and Koyama 1987 (Ar)
R. torreyana A. Gray	$C_3$	Ueno and Koyama 1987 (A)
R. trichochaeta C. B. Clarke	$\mathbf{C}_4$	Ueno and Koyama 1987 (Ac)
R. triflora Vahl	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
R. trispicata (Nees) Schrad. ex Steud.	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
R. tuerckheimii C. B. Clarke	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
R. umbraticola Poepp. & Kunth	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
R. uniflora Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. velutina (Kunth) Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. viridilutea C. B. Clarke	$\mathbf{C}_3$	Ueno and Koyama 1987 (A)
R. vulcani Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. warmingii Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. wightiana (Nees) Steud.	$\mathbf{C}_4$	Bruhl et al. 1987 (Ar. B); Ueno and Koyama 1987 (A); JB (Ar, <i>Bruhl 404</i> CANB. Γ: 1, <i>Bruhl 404</i> )
	$C_3+$	Govindarajalu 1975a ([A])
R. wrightiana Boeck.	$C_3$	Ueno and Koyama 1987 (A)
R. yasudana Makino	$C_3$	Ueno and Koyama 1987 (A)
subsp. leviseta T. Koyama	$C_3$	Takeda et al. 1980 (A. Γ)
Schoenoplectus americanus (Pers.) Volkart ex Schinz & R. Keller, as Scirpus ol- neyi A. Gray	$C_3$	Bender 1971 (δ <sup>13</sup> C)
S. articulatus (L.) Palla, as Scirpus articulatus L.ª	$C_3$	Govindarajalu 1976 ([A]) <sup>a</sup> ; Hesla et al. 1982 ( $\delta^{13}$ C) <sup>a</sup> ; Takeda et al. 1985 (A. $\delta^{13}$ C) <sup>a</sup> ; LR ( $\delta^{13}$ C: $-29.2$ , <i>Boivin s. n.</i> , ca. 1850)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
S. brachyceras (Hochst. ex A. Rich.) K. Lye, as Scirpus brachyceras Hochst. ex A. Rich. <sup>a</sup> , as Scirpus inclinatus (Del.) Aschers. & Schweinf. <sup>b</sup>	<b>C</b> <sub>3</sub>	Govindarajalu 1976 ([A]) <sup>a</sup> ; Hesla et al. 1982 ( $\delta^{13}$ C) <sup>b</sup> ; SCV ( $\delta^{13}$ C: $-23.71$ , Rogers 6431)
S. californicus (C. A. Mey.) Soják, as S. riparius (J. Presl & C. Presl) Palla	$C_3$	LR (δ <sup>13</sup> C: -25.4, St Hilaire C2 2302, -27.5, Infantes 6228)
S. confusus (N. E. Br.) K. Lye, as Scirpus confusus N. E. Br.	$C_3$	Hesla et al. 1982 (δ <sup>13</sup> C)
subsp. <i>natalitius</i> J. Browning	$C_3$	SCV (δ <sup>13</sup> C: -24.11, <i>Gibbs 106</i> )
S. corymbosus (Roem. & Schult.) J. Raynal	$C_3$	LR (δ <sup>13</sup> C: -27.5, Chevalier 8999)
S. dissachanthus (S. T. Blake) J. Raynal, as Scirpus dissachanthus S. T. Blake	$C_3$	Takeda et al. 1985 (A)
S. junceus (Willd.) J. Raynal, as Scirpus aureiglumis S. S. Hooper	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
S. juncoides (Roxb.) Palla, as Scirpus juncoides Roxb.	$C_3$	Govindarajalu 1976 ([A])
S. lacustris (L.) Palla	$C_3$	Troughton et al. 1974 (δ <sup>13</sup> C); JB (A: Holm-Nielsen, 23 July 1970 BRI)
S. laevis (S. T. Blake) J. Raynal, as Scirpus laevis S. T. Blake	$C_3$	Takeda et al. 1985 (A)
S. lateriflorus (J. F. Gmel.) K. Lye, as Scir- pus lateriflorus J. F. Gmel. <sup>a</sup>	$C_3$	Govindarajalu 1976 ([A]) <sup>a</sup> ; Takeda et al. 1985 (A) <sup>a</sup> ; JB (A: <i>Bruhl 454</i> CANB)
	$C_4+$	Hofstra et al. 1972 (A. $\Gamma$ )
S. lineolatus (Franch. & Sav.) T. Koyama	$C_3$	Lin et al. 1993 (A)
S. litoralis (Schrad.) Palla, as Scirpus litoralis Schrad.a, as Scirpus littoralis [sic]b	$C_3$	Govindarajalu 1976 ([A]) <sup>a</sup> ; Hesla et al. 1982 (δ <sup>13</sup> C) <sup>b</sup> ; Takeda et al. 1985 (A) <sup>a</sup> ; Bruhl et al. 1987 (A. B); JB (A: <i>Bruhl 432</i> CANB. Γ: 46, <i>Bruhl 538</i> CANB)
S. mucronatus (L.) Palla ex Kerner, as Scirpus mucronatus L.ª	$C_3$	Govindarajalu 1976 ([A]) <sup>a</sup> ; JB (A: <i>Bruhl 460, 538</i> CANB); LR (δ <sup>13</sup> C: -27.2 <i>Pobéguin 2191</i> )
S. muricinux (C. B. Clarke) J. Raynal	$C_3$	KW (A: Smook 6866)
S. paludicola (Kunth) Palla S. praelongatus (Poir.) J. Raynal	$C_3$	KW (A: Musil 105) SCV (δ <sup>13</sup> C: -23.99, Bolus 9476)
S. pulchellus (Kunth) J. Raynal	$ ext{C}_3 \\  ext{C}_4 +$	SCV (δ <sup>13</sup> C: -10.59, <i>Potts 1076</i> )
S. pungens (Vahl) Palla, as Scirpus americanus Pers.	$C_3$	Takeda et al. 1985 (A)
S. purshianus (Fernald) M. T. Strong, as Scirpus juncoides Roxb.	$C_3$	Lin et al. 1993 (A)
S. roylei (Nees) Ovczinn & Czukav., as Scirpus roylei (Nees) Parker <sup>a</sup> , as Scirpus quinquefarius BuchHam. ex Boeck <sup>b</sup>	C <sub>3</sub>	Sabnis 1921 ([A]) <sup>b</sup> ; Govindarajalu 1976 ([A]) <sup>a</sup> ; Hesla et al. 1982 ( $\delta^{13}$ C) <sup>a</sup>
S. senegalensis (Steud.) J. Raynal, as Schoenoplectus jacobii (C. E. Fisch.) K. Lye <sup>a</sup> , as Scirpus jacobii C. E. Fisch. <sup>b</sup> , as Scirpus jacobii <sup>c</sup>	C <sub>3</sub>	Govindarajalu 1976 ([A]) <sup>b</sup> ; Hesla et al. 1982 ( $\delta^{13}$ C) <sup>c</sup> ; LR ( $\delta^{13}$ C: $-27.5$ , Heudelot 319) <sup>a</sup>
S. subulatus (Vahl) K. Lye	$C_3$	LR (δ <sup>13</sup> C: -27.2, <i>Trochain 3191</i> )
S. supinus (L.) Palla	$C_3$	LR (δ¹³C: −27.9, Sacleux 2561)
S. validus (Vahl) A. Löve & D. Löve, as Scirpus validus Vahl <sup>a</sup>	$C_3$	Bender 1971 (δ <sup>13</sup> C) <sup>a</sup> ; Govindarajalu 1976 ([A]) <sup>a</sup> ; JB (A: <i>Bruhl s. n.</i> , Sullivans Creek CANB)
S. wallichii (Nees) T. Koyama	$C_3$	Lin et al. 1993 (A)
Schoenoxiphium lehmannii (Nees) Steud.	$C_3$	LR (δ <sup>13</sup> C: -30.6, Napper 1931)
S. sparteum (Wahlenb.) C. B. Clarke	C <sub>3</sub>	JB (A: Smook 995 BRI. δ <sup>13</sup> C: -25.4, Smook 995); SCV (δ <sup>13</sup> C: -27.71, Pegler 1196)
Schoenus acuminatus (R. Br.) Nees	$C_3$	KW (A: Wilson 2960)
S. andrewsii W. V. Fitzg. S. apogon Roem. & Schult.	$C_3$ $C_3$	KW (A: Fitzgerald NSW 74075)  Takeda et al. 1985 (A. δ <sup>13</sup> C); Ueno and Takeda 1992 (A); JB (A: Bruhl, Black Mtn. CANB); KW (A: Gardner 924; Ratkowsky 1576)
S. armeria Nees	$C_3$	KW (A: Blake 18102)
S. asperocarpus F. Muell.	$C_3$	KW (A: Wilson 3033)
S. benthamii F. Muell.	$\mathbf{C}_{3}$	KW (A: Fitzgerald NSW 74033)
S. bifidus (Nees) Boeck.	$C_3$	KW (A: Wilson 3040)
S. breviculmis Benth.	$\mathbf{C}_3$	Takeda et al. 1985 (A)
var. tepperi (F. Muell.) Kük.	$\mathbf{C}_3$	KW (A: Beauglehole 49577; Wilson 3157)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
S. brevifolius R. Br.	$C_3$	Takeda et al. 1985 (A); KW (A: <i>Balansa 700</i> ; <i>Johnson 7476</i> ; <i>Petrie</i> NSW 149802); LR (δ <sup>13</sup> C: -28.6, <i>Filhol 829</i> )
S. brevisetis (R. Br.) Roem. & Schult.	$C_3$	KW (A: Wilson 3058)
S. caespititius W. V. Fitzg.	$C_3$	KW (A: Wilson 2884, 2978)
S. calostachyus (R. Br.) Roem. & Schult.	$C_3$	Takeda et al. 1985 (A); Ueno and Takeda 1992 (A); KW (A: <i>Henty &amp; Foreman</i> NGF 49405; <i>Wilson 3693</i> ; s. coll. NSW 149804)
S. calyptratus Kük.	$C_3$	KW (A: Ratkowsky 1583)
S. carsei Cheeseman	$C_3$	KW (A: Beauglehole 33397; Sinclair NSW 149803)
S. curvifolius (R. Br.) Roem. & Schult.	$C_3$	Takeda et al. 1985 (A); KW (A: Coveny 8185)
S. curvulus F. Muell.	$C_3$	KW (A: Elmer 11379; Frodin NGF 26818)
S. deformis (R. Br.) Roem. & Schult.	$C_3$	Takeda et al. 1985 (A); KW (A: Beauglehole 38249)
S. discifer Tate	$C_3$	KW (A: Newbey 4861)
S. efoliatus F. Muell.	$C_3$	KW (A: Coveny 8120; Wilson 2958)
S. ericetorum R. Br.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: <i>Blake 10782</i> BRI); KW (A: <i>Blake 15918</i> ; <i>Hamilton</i> NSW 74127)
S. evansianus K. L. Wilson	$C_3$	KW (A: Wilson 1709)
S. falcatus R. Br.	$C_3$	Takeda et al. 1985 (A); Ueno and Takeda 1992 (A); KW (A: <i>Beauglehole</i> 11486; Ramos BS32717)
S. ferrugineus L.	$C_3$	KW (A: Charpin NSW 149911)
S. fluitans Hook. f.	$C_3$	Takeda et al. 1985 (A)
S. grammatophyllus F. Muell.	$C_3$	KW (A: Blake 17984)
S. grandiflorus Nees ex Lehm.	$C_3$	KW (A: Fitzgerald NSW 4337; Salasoo 4017)
S. hexandrus F. Muell. & Tate	$C_3$	KW (A: Whaite 4063)
S. imberbis (R. Br.) Poir.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: <i>Bruhl</i> , Grose Road CANB); KW (A: <i>Melville 2769</i> ; <i>Tindale</i> NSW 18250)
S. insolitus K. L. Wilson	$C_3$	KW (A: Wilson 2690)
S. juvenis C. B. Clarke	$C_3$	KW (A: Jaffré 554)
S. kennyi F. M. Bailey	$C_3$	Takeda et al. 1985 (A); KW (A: Coveny 10065; Wilson 3453)
S. laevinux (Kük.) Ohwi	$C_3$	KW (A: <i>Croft</i> LAE 68984)
S. lanatus Labill.	$C_3$	KW (A: Coveny 7986)
S. latelaminatus Kük.	$C_3$	KW (A: Beauglehole 29865)
S. latitans S. T. Blake	$C_3$	KW (A: Wilson 2625)
S. lepidosperma (F. Muell.) K. L. Wilson subsp. lepidosperma	$C_3$	KW (A: Archer NSW 74168; Corrick 6176)
S. lepidosperma subsp. pachylepis (S. T. Blake) K. L. Wilson	$C_3$	KW (A: Coveny 10477; Hamilton NSW 74162)
S. maschalinus Roem. & Schult.	$C_3$	JB (A: Bruhl, 7 Oct 1986 CANB); KW (A: Wilson 3085)
S. melanostachys R. Br.	$C_3$	Takeda et al. 1985 (A); KW (A: Constable 5440, 5744)
S. microcephalus Kern	$C_3$	KW (A: McKee 7990)
S. minutulus F. Muell.	$C_3$	KW (A: Crisp 5213)
S. moorei Benth.	$C_3$	KW (A: Coveny 2323, Hamilton NSW 149805)
S. multiglumis Benth.	$C_3$	KW (A: Wilson 3007)
S. neocaledonicus C. B. Clarke	$C_3$	KW (A: MacKee 21092)
S. nigricans L.	$C_3$	Mateu Andres 1991 ([A]); KW (A: <i>Curtiss 130</i> ; <i>Kneucker 44a</i> )
S. nitens (R. Br.) Roem. & Schult.	$C_3$	Takeda et al. 1985 (A); KW (A: <i>Lucas</i> NSW 74160)
S. obtusifolius (Nees ex Lehm.) Boeck.	$C_3$	KW (A: Wilson 2975)
S. ornithopodioides (Kük.) S. T. Blake	$C_3$	KW (A: Johnson NSW 55308)
S. paludosus (R. Br.) Roem. & Schult. (Tricostularia paludosa (R. Br.) Benth.)	$C_3$	Takeda et al. 1985 (A); JB (A: <i>Burbidge</i> , 4 Apr 1948 CANB); KW (A: <i>Blake 13131; Wilson 3116</i> )
S. pauciflorus (Hook. f.) Hook. f.	$C_3$	KW (A: Briggs NSW 90812)
S. pedicellatus (R. Br.) Roem. & Schult.	$C_3$	KW (A: Fitzgerald NSW 74348)
S. pleiostemoneus F. Muell.	$C_3$	KW (A: Coveny 7811; Wilson 2626)
S. punctatus R. Br.	$C_3$	KW (A: Latz 7397)
S. racemosus J. Black	$C_3$	Takeda et al. 1985 (A)
S. rigens S. T. Blake	$C_3$	KW (A: Blake 17985)
S. scabripes Benth.	$C_3$	KW (A: Coveny 4961)
S. sculptus (Nees) Boeck.	$C_3$	Takeda et al. 1985 (A)
S. sesquispiculus C. B. Clarke	$C_3$	KW (A: Newbey 4207)
S. sp. nov. A1 (Crisp 5589)	$C_3$	KW (A: Crisp 5589)
S. sp. nov. A2 (Crisp 5209)	$C_3$	KW (A: Crisp 5209)
S. sp. aff. brevifolius (Wilson 3001)	$C_3$	KW (A: Wilson 3001)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
S. sp. aff. brevisetis (Wilson 2870)	$C_3$	KW (A: Fitzgerald NSW 74087; Newbey 6493; Whittaker & Niering D68-6 Wilson 2870, 2926, 2935, 2977, 3014)
S. sp. aff. elegans S. T. Blake (Wilson 3041)	$C_3$	KW (A: Wilson 3041)
S. sp. aff. falcatus (Lazarides 7859)	$C_3$	KW (A: Lazarides 7859)
S. sp. aff. laevigatus (Crisp 4966)	$C_3$	KW (A: Crisp 4966)
. sp. aff. lanatus (Crisp 5472)	$\mathbf{C}_3$	KW (A: Crisp 5472)
. sp. aff. pleiostemoneus (Wilson 2903)	$\mathbf{C}_3$	KW (A: Canning WA/68 7274; Coveny 3281, 3293a; Wilson 2903)
. sp. aff. punctatus (Dunlop 4444)	$\mathbf{C}_3$	KW (A: Dunlop 4444)
. sp. aff. sparteus (Henderson 1155)	$C_3$	KW (A: Henderson 1155)
sp. aff. subbarbatus (Crisp 5284)	$C_3$	KW (A: Crisp 5284)
sp. aff. subfascicularis (Wilson 2792)	$C_3$	KW (A: Wilson 2792, 2877)
sp. aff. trachycarpus (Wilson 2904)	$C_3$	KW (A: Wilson 2904)
sp. nov. 'Grey Rhizome' (Wilson 2922)	$C_3$	KW (A: Wilson 2922)
sp. nov. 'Murchison' (Haegi 1952)	$C_3$	KW (A: Haegi 1952)
S. sparteus R. Br.	$C_3$	Hesla et al. 1982 (8 <sup>13</sup> C); Takeda et al. 1985 (A); KW (A: <i>Blake 23116</i> ; <i>Henty of Foreman</i> NGF 49415)
. subaphyllus Kük.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); KW (A: Cunningham 3309; Pickard 2495)
. subbarbatus Kük.	$C_3$	KW (A: Wilson 2976)
S. subbarbatus Kük vel sp. nov. aff.	$C_3$	KW (A: Wilson 2864)
. subbulbosus Benth.	$C_3$	KW (A: Wilson 2959)
S. subfascicularis Kük.	$C_3$	KW (A: Wilson 2700, 2724)
S. subflavus Kük. vel sp. nov. aff.	$C_3$	KW (A: Wilson 2603, 2776, 2923)
. sublaxus Kük.	$C_3$	KW (A: Wilson 2885)
S. submicrostachyus Kük.	$C_3$	KW (A: Wilson 2871)
. tendo (Hook. f.) Hook. f. var. triander Kük.	$C_3$	KW (A: Franc 2174)
5. tesquorum J. Black	$C_3$	KW (A: Melville 1935)
. trachycarpus F. Muell.	$C_3$	KW (A: Melville 4408)
5. turbinatus (R. Br.) Roem. & Schult.	$C_3$	KW (A: Blake 7490; Rodway NSW 74069)
. unispiculatus F. Muell. ex Benth.	C <sub>3</sub>	KW (A: Blake 18101)
S. vaginatus F. Muell. ex Benth.	$C_3$	KW (A: Sharpe 2409)
G. villosus R. Br.	$C_3$	Takeda et al. 1985 (A); KW (A: <i>Blakely</i> NSW 74308; <i>Boorman</i> NSW 122302)
Scirpodendron ghaeri (Gaertn.) Merr.	$C_3$	Koyama 1967 ([A]); Takeda et al. 1985 (A); JB (A: White BSIP 75 CANB; Stevens LAE 58624); KW (A: Wilson 10194); LR (δ <sup>13</sup> C: -25.1, -25.6 Buwalda 5861)
Scirpoides holoschoenus (L.) Soják, as Ho- loschoenus vulgaris Link <sup>a</sup> , as Scirpus holoschoenus L. <sup>b</sup>	$C_3$	Mateu Andres 1991 ([A]) <sup>b</sup> ; JB (A: <i>Caine</i> NSW 181479); LR (δ <sup>13</sup> C: -27.4, <i>Bourgeau 490</i> ) <sup>a</sup>
Scirpus macrolepis Philippi [?= Phylloscir- pus acaulis; S. Dhooge, pers. comm.]	$C_3$	Ponessa et al. 1997 ([A])
5. polystachyus F. Muell.	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: Austin 86 CANB; Bruhl 25 CANB)
. sylvaticus L.	$\mathbf{C}_3$	LR (δ <sup>13</sup> C: -27.5, <i>Maire s. n.</i> , La Ferté Alais, 1841)
Scleria abortiva Nees ex Kunth	$C_3$	Chermezon 1926 ([A])
5. angusta Nees ex Kunth	$C_3$	SCV (δ¹³C: −29.19, Wood 3863)
S. bancana Miq.	$C_3$	Prakash et al. 1976 (A)
S. brownii Kunth	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C)
. bulbifera A. Rich.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
S. ciliaris Nees	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: <i>Bruhl 295, 505</i> CANB; <i>Hyland 8380</i> CANB. Γ: 46, <i>Bruhl 295, 52, Bruhl 505</i> )
S. corymbosa Roxb.	$C_3$	Govindarajalu 1975 <i>a</i> ([A])
S. distans Poir., as S. nutans Willd. ex Kunth	$C_3$	Hesla et al. 1982 (δ <sup>13</sup> C)
S. foliosa A. Rich.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
. graeffeana Boeck.	$C_3$	JB (A: Christan 3 CANB)
S. greigiifolia (Ridl.) C. B. Clarke (Acriulus greigiifolius Ridl.)	$C_3$	JB (A: Haines 129 K); LR (δ <sup>13</sup> C: -26.3, Angus 2725); SCV (δ <sup>13</sup> C: -25.98, Stohr 427)
5. iostephana Nelmes	$C_3$	LR ( $\delta^{13}$ C: $-28.6$ , Liben 2191)
S. levis Retz.	$C_3$	Govindarajalu 1975 <i>a</i> ([A]); Takeda et al. 1985 (A); Bruhl et al. 1987 (A. B. Ehleringer et al. 1987 (δ <sup>13</sup> C); JB (A: <i>Bruhl 522</i> CANB; <i>Dunlop 5877</i> CANB. Γ: 45, <i>Bruhl 227</i> CANB)

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
S. lithosperma (L.) Sw., as S. lithosperma	$C_3$	Govindarajalu 1975a ([A]) <sup>a</sup> ; Prakash et al. 1976 (A); Hesla et al. 1982
var. <i>lithosperma</i> <sup>a</sup>		$(\delta^{13}C)$ ; Takeda et al. 1985 (A. $\delta^{13}C$ )
	$C_4+$	Hofstra et al. 1972 (A. $\Gamma$ ); Raghavendra and Das 1976 (A)
var. linearis Benth.	$C_3$	Govindarajalu 1975a ([A])
var. multispiculata Govindarajalu	$C_3$	Govindarajalu 1975a ([A])
var. muricata Govindarajalu	$C_3$	Govindarajalu 1975a ([A])
S. mackaviensis Boeck.	$C_3$	Takeda et al. 1985 (A)
S. melanomphala Kunth	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
S. mikawana Makino	$C_3$	LR (8 <sup>13</sup> C: -32.0, Robinson 3582)
S. novaehollandiae Boeck.	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C)
S. poaeoides Ridl.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
S. poiformis Retz.	$C_3$	Govindarajalu 1975a ([A])
S. racemosa Poir.	$C_3$	Hesla et al. 1982 ( $\delta^{13}$ C)
S. rehmannii C. B. Clarke	$C_3$	SCV (δ <sup>13</sup> C: -23.86, <i>Bolus 1893</i> )
S. rugosa R. Br.	$C_3$	Takeda et al. 1985 (A)
S. sphacelata F. Muell.	$C_3$	Takeda et al. 1985 (A); JB (A: <i>Bruhl 515</i> CANB; <i>Craven 5599</i> CANB)
S. sumatrensis Retz.	$C_3$	Koyama 1967 ([A]); Govindarajalu 1975a ([A])
S. terrestris (L.) Fass.	$C_3$	Govindarajalu 1975a ([A]); Ehleringer et al. 1987 (δ <sup>13</sup> C)
S. tessellata Willd.	$C_3$	Govindarajalu 1975a ([A])
S. transvaalensis E. F. Franklin	$C_3$	SCV $(\delta^{13}C: -25.11, Meyer 15)$
S. tricuspidata S. T. Blake	$C_3$	Takeda et al. 1985 (A)
Sphaerocyperus erinaceus (Ridl.) K. Lye, as Cyperus erinaceus (Ridl.) Kük. <sup>a</sup>	$\mathrm{C}_4$	Druyts-Voets 1970 ([Ac]) <sup>a</sup> ; Lerman and Raynal 1972 (A) <sup>a</sup> ; JB (Ac: <i>Richards</i> 15066 K); KW (Ac: <i>Robinson</i> 3553 P); LR (8 <sup>13</sup> C: -11.7, <i>Gossweiler</i> 4229)
Sumatroscirpus junghuhnii (Miq.) Oteng- Yeboah	$C_3$	BW (A: de Wilde 15236 L)
Tetraria capillaris (F. Muell.) J. M. Black	$C_3$	Takeda et al. 1985 (A. δ <sup>13</sup> C); JB (A: <i>Blake 15846</i> CANB); KW (A: <i>Coveny 6244</i> ; <i>McBarron 11442</i> ); LR (δ <sup>13</sup> C: -28.1, <i>X s. n.</i> , Port Jackson, 1900)
T. compacta Levyns	$C_3$	SCV (δ <sup>13</sup> C: -24.49, <i>Levyns</i> 8726)
T. cuspidata (Rottb.) C. B. Clarke	$C_3$	LR (δ <sup>13</sup> C: -28.2, Schlechter 7429)
T. exilis Levyns	$C_3$	JB (A: Schlechter 7341); SCV (δ <sup>13</sup> C: -25.94, Levyns 6229)
T. natalensis (C. B. Clarke) Koyama	$C_3$	SCV (δ <sup>13</sup> C: -25.80, Rogers 19183)
T. octandra (Nees) Kük. (Tetrariopsis octandra (Nees) C. B. Clarke)	$C_3$	JB (A: Blake 2240 CANB; Seabrook 130 CANB; P.G. Wilson 3965 CANB)
Trachystylis stradbrokensis S. T. Blake	$C_3$	JB (A: <i>Blake 22673</i> BRI; <i>Clarkson 5156</i> BRI; <i>Perry 439</i> CANB. δ <sup>13</sup> C: –29.8, <i>Clarkson 5156</i> ); LR (δ <sup>13</sup> C: –26.7, <i>Blake 13201</i> )
Trianoptiles capensis (Steud.) Harvey	$C_3$	JB (A: Parker 4132 K); LR (δ <sup>13</sup> C: -27.6, Schlechter 9137); SCV (δ <sup>13</sup> C: -28.02, Levyns 7762)
T. solitaria (C. B. Clarke) Levyns	$C_3$	SCV (δ <sup>13</sup> C: -25.82, Esterhuysen 34682)
T. stipitata Levyns	$C_3$	SCV (δ <sup>13</sup> C: -29.24, <i>Levyns</i> 7678)
Trichophorum alpinum (L.) Pers.	$\mathbf{C}_3$	LR (δ <sup>13</sup> C: -26.1, De la Pylaie 1643)
T. cespitosum (L.) Hartm., (Baeothryon caespitosum (L.) A. Dietrich), as Scirpus cespitosus L. <sup>a</sup>	$C_3$	Bender 1971 ( $\delta^{13}$ C) <sup>a</sup> ; JB (A: <i>Townsend 73/154</i> PDA); LR ( $\delta^{13}$ C: $-27.1$ , <i>Lerman s. n.</i> , Oetztal, 1971)
T. subcaptitatum (Thwaites) D. A. Simpson, as Scirpus subcapitatus Thwaites	$C_3$	Govindarajalu 1976 ([A])
Trichoschoenus bosseri J. Raynal	$C_3$	LR (δ¹³C: -24.9, <i>Humbert 28576</i> )
Tricostularia compressa Nees ex Lehm.	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C)
T. pauciflora (F. Muell.) Benth.	$C_3$	Takeda et al. 1985 (A); JB (A: Willis, 1 Oct 1959 CANB)
T. undulata (Thwaites) J. Kern	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C); JB (A: <i>Bruhl 325</i> CANB); LR ( $\delta^{13}$ C: $-26.5$ , <i>Evrard 2315</i> )
Trilepis lhotzkiana Nees	$C_3$	JB (A: Harley 19425 K; LR (δ <sup>13</sup> C: -28.7, Weddell 471)
Uncinia angustifolia Hamlin	$C_3$	Kukkonen 1967 ([A])
U. brevicaulis Thouars	$C_3$	Kukkonen 1967 ([A])
U. compacta R. Br.	$C_3$	JB (A: Bruhl 634 CANB; Smith 15531 CANB); KW (A: Thompson 4048)
U. dawsonii Hamlin	$C_3$	LR (δ <sup>13</sup> C: -32.8, <i>MacKee 9783</i> )
U. divaricata Boott	$C_3$	Kukkonen 1967 ([A]); KW (A: Seppelt 12453)
U. elegans (Kük.) Hamlin	$C_3$	KW (A: <i>Rodway</i> NSW 52591)
U. erinacea (Cav.) Pers.	$C_3$	Kukkonen 1967 ([A])
U. flaccida S. T. Blake	$C_3$	Takeda et al. 1985 (A. $\delta^{13}$ C); KW (A: <i>Thompson 3025</i> )
U. hamata (Schwartz) Urb.	$C_3$	Kukkonen 1967 ([A])

Appendix 1. Continued.

Species	Photosynthetic pathway	References (method: value [as appropriate], voucher [if new record])
U. hookeri Boott	$C_3$	KW (A: Seppelt 12101, 12681)
U. nemoralis K. L. Wilson	$\mathbf{C}_3$	KW (A: Coveny 5913)
U. nervosa Boott	$C_3$	KW (A: Druce CHR 131588)
U. riparia R. Br.	$C_3$	KW (A: Ratkowsky 1596)
U. sp. nov. aff. filiformis Colenso ex Boott	$C_3$	KW (A: Blake 18413)
(Blake 18413) U. sulcata K. L. Wilson	C	WW (A. Bishand & Courses 2740)
U. tenella R. Br.	$C_3$	KW (A: Pickard & Coveny 2749)  Kylklonen 1067 (IA)), Takada et al. 1085 (A), KW (A: Cross 5402)
***************************************	$C_3$	Kukkonen 1967 ([A]); Takeda et al. 1985 (A); KW (A: <i>Gray 5403</i> )
U. uncinata (L. f.) Kük.	$C_3$	Kukkonen 1967 ([A])
Volkiella disticha Merxm. & Czech., as Volkiella <sup>a</sup>	$C_3$	Lerman and Raynal 1972 (A) <sup>a</sup> ; JB (A: <i>Mueller 493</i> PRE); LR (δ <sup>13</sup> C: -13.6, <i>Volk 1815</i> )
Websteria confervoides (Poir.) S. S. Hooper, as Websteria <sup>a</sup>	$\mathbf{C}_3$	Lerman and Raynal 1972 (A) <sup>a</sup> ; JB (A: <i>Smith 1797</i> PRE); LR (δ <sup>13</sup> C: -23.3, <i>Hallé &amp; Guillaumin s. n.</i> , Moossou, Côte d'Ivoire, 1960)
Zameioscirpus muticus Dhooge & Goetgh.	$C_3$	Dhooge et al. 2003 (A)

APPENDIX 2: Assessment of Conflicting Data on Photosynthetic Pathway Status in Cyperaceae

Our survey of literature on photosynthetic pathways in Cyperaceae found various inconsistent records. We have seen some but not all of the vouchers for these records. However, in most, perhaps all, of these cases we concluded that they resulted from misidentification of the material used or misapplication of names, rather than the species concerned being variable in this regard. See, for example, the discussion below about records of *Cyperus eragrostis*, which we suggest involved confusion in usage of that name. These suspect records are tagged in Appendix 1 with "+".

Carex.—Smith and Epstein's (1971)  $C_4$   $\delta^{13}C$  value for an unnamed *Carex* species is at variance with all other available data for that genus, including one biochemically typed species,  $\delta^{13}C$  value determinations for 33 species, anatomical data for 48 species and  $\Gamma$  values for eight species.

Cyperus s.l.—Cyperus aggregatus was reported as  $C_3$  by Li et al. (1999). This species, previously known as C. flavus (Vahl) Nees or C. cayennensis (Lam.) Britton, is  $C_4$ , as are all the other species so far as known in Kükenthal's (1935, 1936) Cyperus sect. Mariscus apart from C. deciduus (see below).

Cyperus albostriatus was reported as  $C_4$  by Sonnenberg and Botha (1992) but our samples (Appendix 1) show this to be  $C_3$ , as are all the other species sampled in Kükenthal's (1935, 1936) Cyperus sect. Diffusi.

Cyperus deciduus was treated by Kükenthal (1935, 1936) as a member of Cyperus subgen. Mariscus, mainly on account of its spikelets falling as a unit as in other species in that subgenus as traditionally circumscribed. However, its characteristics, including non-Kranz anatomy, suggest that it is better placed with the "C<sub>3</sub>" species of Cyperus in subgen. Pycnostachys (Wilson 1991). Vorster (1990, 1996) reported unusual anatomy in this species, with a partial inner chlorenchymatous sheath as in Kranz anatomy. However, the "maximum cells distant" count is in accord with non-Kranz anatomy. Further study of this unusual species is warranted.

Conflicting data have been presented in the literature for  $C.\ eragrostis$  (Appendix 1). The  $C_4$  values were obtained from one laboratory. We sampled two Australian and two New Zealand accessions of  $C.\ eragrostis$ . All four proved to be  $C_3$ , with  $C_3$  anatomy (Appendix 1), very low or undetectable levels of  $C_4$  acid decarboxylating enzymes (Bruhl et al. 1987) and with  $\Gamma$  and  $\delta^{13}C$  values (Appendix 1) typical of a  $C_3$  species. It seems, therefore, that  $C.\ eragrostis$  is  $C_3$ . The  $C_4$  records may result from the misapplication of that name to  $C.\ sanguinolentus$  Vahl, which was commonly

known as *C. eragrostis* Vahl (non Lam.) in Europe over the last couple of centuries (Kukkonen 1995).

Cyperus glaber was listed by Li (1993) as  $C_3$ . However, other records show this to be a  $C_4$  species, as are all the other species in Kükenthal's (1935, 1936) Cyperus sect. Compressi.

Cyperus glomeratus was reported to be  $C_3$  by Li (1993), but other studies record this as  $C_4$ , as are all the other species (so far as known) in Kükenthal's Cyperus sect. Distantes.

For *C. papyrus*, both  $C_3$  and  $C_4$  determinations were obtained from several laboratories (Appendix 1). Our own anatomical observations on two accessions support the  $C_4$  status of this species, in agreement with Lerman and Raynal (1972; Appendix 1) and Jones and Milburn (1978). We wonder whether the  $C_3$  determinations were made on the morphologically somewhat similar *C. prolifer* or *C. involucratus*, both of which are commonly cultivated and are sometimes known as "papyrus" (or "dwarf papyrus" in the case of the former species).

The  $C_4$  record for *C. pulchellus* of Lerman and Raynal (1972; Appendix 1) is at odds with  $C_3$   $\delta^{13}C$  values obtained for this species by Hesla et al. (1982) and Takeda et al. (1985) and anatomical observations of several workers. Plants of this species have been misidentified as species of *Lipocarpha* ( $C_4$ ), *Ascolepis* ( $C_4$ ) and *Kyllingiella* ( $C_3$ ) (Haines and Lye 1983).

Cyperus textilis was reported as  $C_4$  by Li (1993). However, another study records this as  $C_3$ , which is in line with the other species in Kükenthal's (1935, 1936) Cyperus sect. Vaginati.

Li and Jones (1994) reported a form of C<sub>4</sub> anatomy that they called Kranzkette (literally "chain of garlands") from the arrangement of the vascular bundles so that they form a "ring" around airspaces as seen in cross-section. This anatomical type is indeed interesting, but the paper is unsatisfactory in several ways. Firstly, in discussing this unusual anatomy, they fail to mention Metcalfe's (1971: 382) designation of the extreme form of this as "Mariscus A-type anatomy" (characterized by adaxial epidermal cells being of similar size to those of the abaxial layer; and with vascular bundles surrounding air spaces). This type of anatomy was also distinguished by Bruhl et al. (1992) as "vascular bundles forming 'ring' or 'horseshoe' patterns". As discussed by Wilson (1991), it is present in species of Cyperus sections Pinnati, Glutinosi, Thunbergiani and Turgiduli p.p. (sectional names as in Kükenthal [1935, 1936]). The form described by Li and Jones (1994) is the same as described and illustrated by Metcalfe (1971: 316, Fig. 42F) for C. serotinus under the name Juncellus serotinus (Rottb.) C. B. Clarke. Secondly, it is not clear what species the authors were studying. They named it C. japonicus Makino, but that name is a synonym of C. microiria Steud. which has "ordinary" C4 anatomy with a single row of vascular bundles in the two specimens examined by the current authors. Indeed, all the other species examined in *Cyperus* sect. *Iriae* (as updated by the current authors from Kükenthal [1935, 1936]: *C. amuricus, C. alulatus* Kern, *C. iria*) have this type of C<sub>4</sub> anatomy, except for *C. orthostachys*, which has *Mariscus* A-type ("Kranzkette") anatomy in the outer quarter of its leaves but a single row of vascular bundles closer to the midrib, as seen in cross-section. *Cyperus orthostachys* is also an Asian species, so perhaps Li and Jones were using that species. However, a more likely alternative is that their material was of *C. serotinus*, mentioned above, which also grows in Asia and exhibits a well-developed example of this type of anatomy.

Eleocharis.—The evidence for the  $C_4$  status of *E. retroflexa* is compelling with 20 records including assessment of anatomy, ultrastructure, biochemistry and  $\delta^{13}C$  value determinations (Appendix 1). By contrast the semi-diagrammatic drawing by Govindarajalu (1975*a*: Fig. 1a) suggests  $C_3$  status due to a "maximum cells distant" count of greater than one. His tissue map and description of the vascular bundles for this species do not help resolve this conflict which most likely stems from the breakdown of the "one cell distant" criterion in *Eleocharis*, as discussed in the main text.

We present three anatomical records and four  $\delta^{13}C$  value determinations for *E. subcancellata* that clearly indicate this species is  $C_4$ . In contrast, Ueno et al. (1989: 430) presented one anatomical and one  $\delta^{13}C$  value assessment of this species and assigned the species as " $C_3$ ?". Their typical  $C_3$  value of -23.2 for *E. subcancellata* indicates that either the specimen was incorrectly identified or this species is another member of the genus that is variable for photosynthetic pathway (see discussion on *Eleocharis* in the text) and worthy of detailed study.

Lipocarpha.—This genus is generally  $C_4$ , with reports for 12 species cited in Appendix 1. The report of  $C_3$  for a species of *Lipocarpha* (Stock et al. 2004) from a single  $\delta^{13}C$  value for *L. rehmannii* (Appendix 1) is at odds with four other records for the species based on anatomical observations and  $\delta^{13}C$  values, and with the other species in the genus. The specimen in question appears to be a collection of multiple individuals but with all components matching other material of *L. rehmannii* at BOL (A. Verboom, pers. comm., Aug 2004). These specimens are in need of anatomical study and broader sampling for  $\delta^{13}C$  values.

Pycreus.—Pycreus flavescens (as Cyperus flavescens) was reported as being C<sub>3</sub> by Li (1993). However, other reports for this species record it as C<sub>4</sub>, as are all known members of the genus Pycreus (Cyperus subgen. Pycreus).

Saxena and Ramakrishnan (1984) reported *P. flavidus* (as *P. globosus*) as anatomically  $C_3$ . By contrast, all other evidence (including

ultrastructural, physiological, biochemical, and further anatomical characteristics) reported for this species and the genus in general (Appendix 1) indicate  $C_4$ . It is possible that Saxena and Ramakrishnan sampled a specimen of the  $C_3$  species *Cyperus tenuispica*, which has at times in the past been known (erroneously) as *Cyperus flavidus*.

Rhynchospora.—The  $\Gamma$  values presented by Takeda et al. (1980) for 17 species of *Rhynchospora* include values that are higher than classic  $C_4$  values (for species with rhynchosporoid anatomy, e.g., *R. rubra*: 10  $\mu$ L liter<sup>-1</sup>), and that are lower than typical  $C_3$  values (e.g., *R. brownii*: 32  $\mu$ L liter<sup>-1</sup>). Such values are usually indicative of  $C_3$ - $C_4$  intermediates (Hattersley et al. 1986) (Appendix 1: cf. *Eleocharis pusilla*). Indeed the two values fall outside the range of values Takeda et al. (1980: 57) obtained for control species: i.e., "less than 10  $\mu$ l l<sup>-1</sup> (for  $C_4$  species) and . . . more than 40  $\mu$ l l<sup>-1</sup> for  $C_3$  species", though they did not query these results. More recent anatomical and biochemical investigations of *R. rubra* have, however, corroborated its  $C_4$  status (Bruhl et al. 1987; Ueno and Koyama 1987).

The  $\delta^{13}$ C values and C<sub>4</sub> anatomy (Appendix 1) for *R. armerioides* and *R. barbata* confirm that *Rhynchospora* species with chlorocyperoid anatomy are also consistently C<sub>4</sub>, as Ueno and Koyama (1987) initially reported (see also Bruhl et al. 1987; Bruhl 1995).

Schoenoplectus.—The listing by Hofstra et al. (1972) of *S. lateriflorus* as  $C_4$ , based on  $C_4$  anatomy and a low  $\Gamma$  value, conflicts with other observations for this species (Appendix 1), and for the 26 other species surveyed. Our anatomical observations do not indicate even remotely  $C_4$ -like anatomy for *S. lateriflorus*.

The  $C_4$   $\delta^{13}C$  value for *S. pulchellus* is the first report for this species. The specimen in question is not mixed and appears to have been correctly identified by cyperologist Jane Browning (A. Verboom, pers. comm., Aug 2004). Hayasaka (2002) puts *S. pulchellus* in the "*S. corymbosus* complex" of species (viz. *S. brachyceras, S. confusus, S. corymbosus, S. decipiens* (Nees) J. Raynal, *S. muricinux, S. muriculatus* (Kük.) J. Browning, *S. paludicola*, and *S. pulchellus*) on morphological grounds. The five other species of the complex so far sampled are  $C_3$  (Appendix 1), and we suspect that *S. pulchellus* will also prove to be  $C_3$ . Nevertheless, the photosynthetic pathway of all species of *Schoenoplectus* clearly merits assessment.

Scleria.—Variation in photosynthetic pathway has also been reported for *S. lithosperma* (Appendix 1), but "these discrepancies . . . may have resulted from identificatory error of plant materials" (Takeda et al. 1985: 405). Another 20 species of *Scleria* appear in the literature as  $C_3$ , and our anatomical observations and  $\Gamma$  values (Appendix 1) for *Scleria* also support the contention that the genus is wholly C