

# JEAN CARLOS SERRANO

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## IN BRIEF

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Leveraging a strong foundation in biological transport phenomena and numerical simulations to engineer complex biological systems for pathophysiological research and preclinical studies, including organ-on-chip and organoid technologies. Integrated skills in microfabrication, quantitative imaging, and molecular biology enabling mechanistic modeling of disease processes and drug responses across multiple scales.

## EDUCATION

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**Massachusetts Institute of Technology** *September 2018 - June 2021*  
Ph.D. in Mechanical Engineering  
*Thesis: On-Chip Engineered Human Lymphatic Microvasculature for Physio-/Pathological Transport Phenomena Studies*

**Massachusetts Institute of Technology** *September 2016 - June 2018*  
M.S. in Mechanical Engineering  
*Thesis: Engineering 3D Lymphatic Vasculature On-Chip Through Biochemical and Mechanical Stimuli*

**University of Puerto Rico at Mayaguez** *August 2012 - June 2016*  
B.S. in Mechanical Engineering (Summa Cum Laude)  
*Honors Thesis: Adaptive Responses of Murine Osteoblasts Subjected to Coupled Mechanical Stimuli*

## WORK EXPERIENCE

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**Cellino Biotech** *May 2023 - Present*  
*Biomedical Fluidic Engineer*

### **Biophysical Characterization and Modeling of iPSCs Biomanufacturing**

- Derived analytical and numerical models on relevant heat and fluid transport phenomena, enabling the optimization of design principles for our laser-based cell editing platform.
- Developed and optimized biological assays with induced-pluripotent stem cells to evaluate the functional thresholds for cell editing utilizing our laser-based bioprocessing techniques.
- Designed and built instrumentation to measure the physicochemical properties of biocompatible ceramic thin-films, and an optical set-up to visualize flow dynamics in cell-culture chambers.
- Streamlined fluorescent data quantification from bioprocess outputs with custom Python-based image processing pipelines.

**Wyss Institute for Biologically Inspired Engineering** *July 2021 -May 2023*  
*Postdoctoral Research Fellow*

### **High-throughput, Micro-Patterned Organoid Systems**

- Developed microfluidic droplet-based techniques for single-cell encapsulation with patterned extracellular matrix droplets (patent pending), thus permitting high-throughput generation and screening of organoid systems. Commercialization of such platform is currently under development through a Wyss Institute Partnership.

**Harvard University**  
*Postdoctoral Research Fellow*

July 2021 - May 2023

***Microfluidic-based Bioassays for Next-Generation Diagnostics and Therapeutics***

- Built an integrated droplet digital PCR (ddPCR) with nanoplasmonic photothermal heating, thus achieving viral/bacterial DNA detection in less than 5 minutes. Furthermore, implemented low-cost, minimal equipment to facilitate its widespread use in clinics and at-home.
- Designed a novel microfluidic-based fluid mixer (patent pending) for high-throughput and uniform synthesis of lipid nanoparticles for mRNA vaccine delivery.

**Massachusetts Institute of Technology**  
*Graduate Student Researcher*

September 2016 - June 2021

***On-Chip Engineered, Physiologically-Functional Lymphatic Vasculature***

- Optimized the *in vitro*, angiogenic growth of lymphatic capillaries to mimic their *in vivo* morphology and function, in a versatile microfluidic platform implemented for disease models and drug screening.
- Developed analytical and computational models to study the relevant transport phenomena that drive protein drainage and inflammatory-immune signals by the engineered lymphatic vasculature.
- Collaborated with Amgen Inc. by utilizing our on-chip engineered lymphatics to screen and characterize vascular transport of their new monoclonal antibody candidates. Based on the measured transport parameters, developed a physiological-based pharmacokinetic (PBPK) framework predicting differences in bioavailability.
- Additional projects included the development of novel microfluidic systems to recapitulate biomechanical stimuli (microvascular flow and oxygen-tension gradients) and computational models characterizing chemotactic gradients during brain cancer metastasis.

**Harvard Medical School**  
*Undergraduate Student Researcher*

June 2015 - August 2015

***Engineered Flow-Activated Endothelial Cell Sensor for Atherosclerosis Studies***

- Characterized a transcriptionally-activated cellular sensor (KLF2-GFP promoter) capable of exhibiting a quantitative fluorescent response when endothelial cells are exposed to atherosclerosis-prone flow patterns, thus allowing real-time visualization of flow shear stress on cell physiology.
- Validated the versatility of the cell-based sensor as a fluorescent readout in drug screening studies for chemically inducing an atherosclerosis-protective endothelial phenotype despite the presence of atherosclerosis-prone flow patterns.

**Princeton University**  
*Undergraduate Student Researcher*

June 2014 - August 2014

***Characterizing Viscoelasticity of Bacterial Biofilms via Micro-Membrane Rheometry***

- Designed a microfluidic-based rheometer capable of measuring the elasticity of bacterial biofilms by the application of fixed air pressure to a micro-membrane in contact with the biofilm channel.
- Developed a COMSOL-based finite element analysis model to estimate the elasticity of the bacterial biofilm, based on the experimental measurements of the resultant deformations to the applied pressures.

**University of Puerto Rico at Mayaguez**  
*Undergraduate Student Researcher*

August 2013 - December 2015

***Adaptive Responses of Murine Osteoblasts Subjected to Coupled Mechanical Stimuli***

- Analyzed the orientational response of the actin cytoskeleton and expression of focal adhesion complexes in murine osteoblasts as a result of simultaneous mechanical cues (matrix stiffness and cyclic tensional strain) to induce preferential cellular alignment for functional bone tissue constructs.

## PEER-REVIEWED PUBLICATIONS

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Google Scholar Profile

1. **J.C. Serrano**, M. Gillrie, R. Li, R.D. Kamm, Microfluidic-Based Reconstitution of Functional Lymphatic Microvasculature: Elucidating the Role of Lymphatics in Health and Disease. *Advanced Science*. (2023)
2. G. Offeddu, **J.C. Serrano**, J. Z. Wan, et al, Microphysiological endothelial models to characterize subcutaneous drug absorption. *ALTEX-Alternatives to animal experimentation*. (2022)
3. C. Hajal, Y. Shin, L. Li, **J.C. Serrano**, T. Jacks, R.D. Kamm, The CCL2-CCR2 astrocyte-cancer cell axis in tumor extravasation at the brain. *Science Advances*. (2021)
4. G. Offeddu\*, **J.C. Serrano\***, S.W. Chen, S.E. Shelton, Y. Shin, R.D. Kamm, MicroHeart: A Microfluidic Pump for Functional Vascular Culture in Microphysiological Systems. *Journal of Biomechanics*. (2021) \*These authors contributed equally to this work.
5. **J.C. Serrano\***, S. Gupta\*, R.D. Kamm, M. Guo, In Pursuit of Designing Multicellular Engineered Living Systems: A Fluid Mechanical Perspective. *Annual Review of Fluid Mechanics*. (2021) \*These authors contributed equally to this work.
6. C. Hajal, L. Ibrahim, **J.C. Serrano**, G. Offeddu, R.D. Kamm, The effects of luminal and trans-endothelial fluid flows on the extravasation and tissue invasion of tumor cells in a 3D in vitro microvascular platform. *Biomaterials*. (2020)
7. R. Koens, Y. Tabata, **J.C. Serrano**, S. Aratake, D. Yoshino, R.D. Kamm, K. Funamoto, Microfluidic platform for three-dimensional cell culture under spatiotemporal heterogeneity of oxygen tension. *APL Bioengineering*. (2020)
8. R. Li, **J.C. Serrano**, H. Xing, T.A. Lee, H. Azizgolshani, M. Zaman, R.D. Kamm, Interstitial flow promotes macrophage polarization toward an M2 phenotype. *Molecular Biology of Cell*. (2018)
9. T. Osaki, **J.C. Serrano**, R.D. Kamm, Cooperative Effects of Vascular Angiogenesis and Lymphangiogenesis. *Regenerative Engineering and Translational Medicine*. (2018)
10. **J.C. Serrano**, J. Cora-Cruz, N. Diffoot, P. Sundaram, Adaptive Responses of Murine Osteoblasts Subjected to Coupled Mechanical Stimuli. *Journal of the Mechanical Behavior of Biomedical Materials*. (2018)

## INTELLECTUAL PROPERTY/PATENTS

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- *Single-cell derived organoids in extracellular matrix droplets*. (US patent pending)
- *Flexus Mixer: A microfluidic-based mixer for nanoparticle synthesis*. (US patent pending)
- *Microphysiological Model of the Brain*. (US patent pending)

## TECHNICAL STRENGTHS

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<b>Programming Languages:</b>	Python, MATLAB, R, LabVIEW, LaTeX, ImageJ1
<b>Simulation/Modeling:</b>	ODEs/PDEs, Compartmental (Phoenix NLME), Finite Element (COMSOL)
<b>Microscopy:</b>	Confocal, Epifluorescence, High-speed/Time-Lapse
<b>Molecular Biology:</b>	PCR, Immunofluorescence, Flow Cytometry, ELISA, Western Blot
<b>Microfabrication:</b>	AutoCAD, Photo/Soft-lithography, Micromachining, 3D Printing
<b>Culturing and Handling:</b>	Mammalian Cells & Tissue, Bacteria

## AWARDS AND HONORS

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**Invited Keynote Speaker: FluidicMEMS Consortium, Cambridge M.A.** *(2024)*

**El Mundo Boston's Latino 30 under 30** *(2022)*

**MIT University Center for Exemplary Mentoring (UCEM) Sloan Scholar** *(2018)*

**National Science Foundation (NSF) Graduate Research Fellowship** *(2017)*

**MIT Office of the Dean for Graduate Education (ODGE) Diversity Fellowship** *(2016)*

**NIH RISE 2 BEST Program** *(2013 - 2016)*

## LANGUAGES

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**English:** native, bilingual proficiency

**Spanish:** native, bilingual proficiency

**French:** elementary proficiency