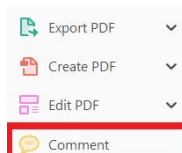


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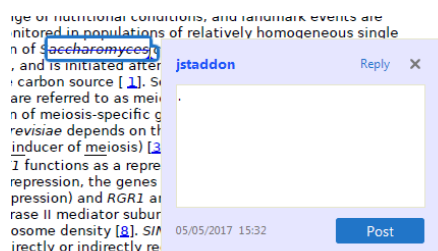
1. Replace (Ins) Tool – for replacing text.



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How to use it:

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- Click on .
- Type the replacement text into the blue box that appears.




2. Strikethrough (Del) Tool – for deleting text.



Strikes a red line through text that is to be deleted.

How to use it:

- Highlight a word or sentence.
- Click on .
- The text will be struck out in red.

experimental data if available. For ORFs to be had to meet all of the following criteria:



1. Small size (35-250 amino acids).
2. Absence of similarity to known proteins.
3. Absence of functional data which could not be the real overlapping gene.
4. Greater than 25% overlap at the N-terminus terminus with another coding feature; over both ends; or ORF containing a tRNA.

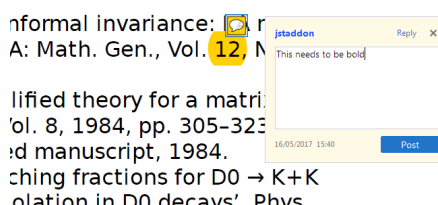
3. Commenting Tool – for highlighting a section to be changed to bold or italic or for general comments.



Use these 2 tools to highlight the text where a comment is then made.

How to use it:

- Click on .
- Click and drag over the text you need to highlight for the comment you will add.
- Click on .
- Click close to the text you just highlighted.
- Type any instructions regarding the text to be altered into the box that appears.




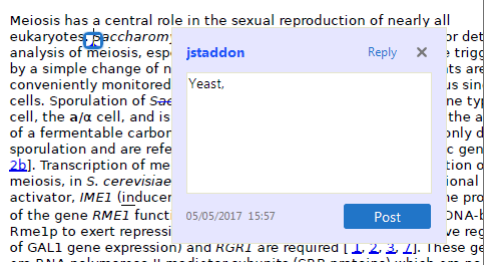
4. Insert Tool – for inserting missing text at specific points in the text.



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How to use it:

- Click on .
- Click at the point in the proof where the comment should be inserted.
- Type the comment into the box that appears.




5. Attach File Tool – for inserting large amounts of text or replacement figures.



Inserts an icon linking to the attached file in the appropriate place in the text.

How to use it:

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- Click on the proof to where you'd like the attached file to be linked.
- Select the file to be attached from your computer or network.
- Select the colour and type of icon that will appear in the proof. Click OK.

The attachment appears in the right-hand panel.


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6. Add stamp Tool – for approving a proof if no corrections are required.



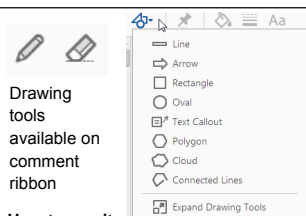
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How to use it:

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- Fill in any details and then click on the proof where you'd like the stamp to appear. (Where a proof is to be approved as it is, this would normally be on the first page).

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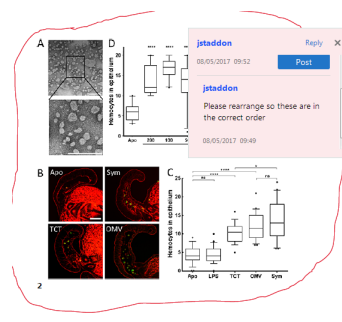


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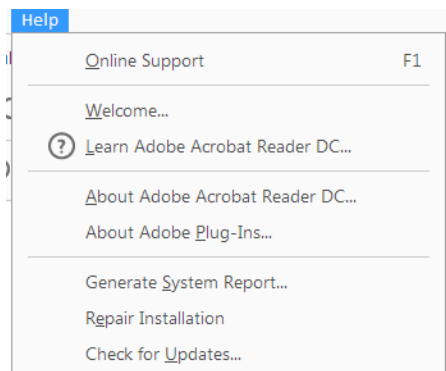
- Click on one of the shapes in the [Drawing Markups](#) section.
- Click on the proof at the relevant point and draw the selected shape with the cursor.
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7. Drawing Markups Tools – for drawing shapes, lines, and freeform annotations on proofs and commenting on these marks.

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The partitioning of gross primary production for young *Eucalyptus tereticornis* trees under experimental warming and altered water availability

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Received: 20 September 2018

Accepted: 20 November 2018

New Phytologist (2018)

doi: 10.1111/nph.15629

Key words: allocation, climate change, drought, partitioning, respiration, warming.

Summary



- The allocation of carbon (C) is an important component of tree physiology that influences growth and ecosystem C storage. Allocation is challenging to measure, and its sensitivity to environmental changes such as warming and altered water availability is uncertain.
- We exposed young *Eucalyptus tereticornis* trees to +3°C warming and elimination of summer precipitation in the field using whole-tree chambers. We calculated C allocation terms using detailed measurements of growth and continuous whole-crown CO₂ and water exchange measurements.
- Trees grew from small saplings to nearly 9 m height during this 15-month experiment. Warming accelerated growth and leaf area development, and it increased the partitioning of gross primary production (GPP) to aboveground respiration and growth while decreasing partitioning below ground. Eliminating summer precipitation reduced C gain and growth, but it did not impact GPP partitioning. Trees utilized deep soil water and avoided strongly negative water potentials.
- Warming increased growth respiration, but maintenance respiration acclimated homeostatically. The increasing growth in the warmed treatment resulted in higher rates of respiration, even with complete acclimation of maintenance respiration. Warming-induced stimulations of tree growth likely involve increased C allocation above ground, particularly to leaf area development, whereas reduced water availability may not stimulate allocation to roots.

Introduction

The carbon (C) economy of trees and forests depends not only on the amount of C fixed via photosynthesis, but how that fixed C is used. Ecosystem C storage is affected by the allocation of C to long-lived C pools, such as wood, relative to C allocation to pools with higher turnover rates, such as fine roots (DeLucia *et al.*, 2005). C allocation also affects the acquisition of light, nutrients and water (H₂O), which influences ecosystem C cycling and tree survival (Litton *et al.*, 2007; Epron *et al.*, 2012; Franklin *et al.*, 2012; De Kauwe *et al.*, 2014; Trugman *et al.*, 2018). The allocation of C below ground affects soil C and nutrient cycling (Högberg *et al.*, 2001; Epron *et al.*, 2012) in part because below-ground C allocation can affect soil organic matter decomposition and the acquisition of limiting nutrients by trees (Drake *et al.*, 2011; Finzi *et al.*, 2015). The importance of C allocation and the relative difficulty of its study contribute to its role as an

important unknown for modeling the biogeochemistry of ecosystems (Roux *et al.*, 2001; Franklin *et al.*, 2012; De Kauwe *et al.*, 2014; Dietze *et al.*, 2014).

Terminology regarding allocation has been a source of some confusion. Here, we follow Litton *et al.* (2007) and use 'allocation' as a term of broad definition encompassing three specific aspects of study: (1) *ratios* of biomass pool sizes, (2) *fluxes* of C to a given component and (3) *partitioning*, the C flux to a given component as a fraction of gross primary production (GPP). These areas of study are similar, but not equivalent. For example, old and large trees have a large wood mass fraction relative to small, young trees (Poorter *et al.*, 2015), but this reflects the low turnover of woody tissues relative to leaves, not a higher partitioning of GPP to wood in old trees (Duursma & Falster, 2016). Thus, it is often inappropriate to infer C partitioning from biomass ratios (Reich, 2002; Litton *et al.*, 2007). Surprisingly, partitioning of photosynthate is relatively poorly understood

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despite its direct relevance to ecosystem models (Epron *et al.*, 2012; Franklin *et al.*, 2012; De Kauwe *et al.*, 2014).

Several schemes have been used to conceptualize and model C allocation. The simplest approach is to assume that trees partition a constant fraction of fixed C to each use (e.g. growth, respiration). This is supported by linear relationships between production terms in some systems (Gower *et al.*, 2001). However, fixed allocation schemes cannot capture ontogenetic effects (Poorter *et al.*, 2015; Duursma & Falster, 2016) or dynamic temporal responses (De Kauwe *et al.*, 2014; Doughty *et al.*, 2014). Another approach is to assume a functional balance between tree organs via allometric relationships, Huber values, or root to leaf mass fractions (Landsberg & Waring, 1997; Mäkelä *et al.*, 2008; Feng *et al.*, 2012). Finally, there is the concept that trees increase C partitioning towards the acquisition of the primary limiting resource (McMurtrie & Dewar, 2013). This approach appears sensible and has been implemented in several models (e.g. Running & Gower, 1991; Friedlingstein *et al.*, 1999), but direct evidence supporting this concept is scarce, given the challenges involved in measuring allocation (Poorter & Sack, 2012; Poorter *et al.*, 2015). However, optimization approaches have been used to constrain dynamic allocation schemes with some success (Franklin *et al.*, 2012; McMurtrie & Dewar, 2013).

Temperature is a fundamental aspect of climate that affects many aspects of tree physiology, including photosynthesis, respiration, and growth (Way & Oren, 2010; Lu *et al.*, 2013). Several lines of evidence suggest that warming may increase tree C allocation above-ground at the expense of C allocation below-ground. Experimental warming of forest soils increased aboveground biomass while reducing or not affecting belowground biomass (Strömgren & Linder, 2002; Melillo *et al.*, 2011), which has been attributed to a warming-induced increase in soil nitrogen (N) availability (Melillo *et al.*, 2002). A recent ^{13}C -CO₂ labeling study indicated that warming increased allocation above-ground and reduced C allocation below-ground in beech saplings via a direct effect on tree physiology, without an altered soil N cycle (Blessing *et al.*, 2015). Also, environmental gradients in mean annual temperature are strongly correlated with the distribution of biomass; forests have a lower root mass fraction (i.e. the proportion of live forest biomass contained in roots) in warm climates than in cold climates (Reich *et al.*, 2014). Meta-analyses of warming experiments generally find an increase in aboveground plant growth (Rustad *et al.*, 2001) that is slightly larger than the increase in belowground plant growth (Lu *et al.*, 2013), although such experiments have exclusively involved small-stature vegetation given the logistical challenges of warming tall forests. Thus, prior research suggests a shift in C allocation above-ground with experimental warming, although direct tests in the field with large trees have not yet been performed.

H₂O availability also impacts tree growth and physiology (Mencuccini, 2003; Nemani *et al.*, 2003; Farooq *et al.*, 2009; Martínez-Vilalta *et al.*, 2009; Yang *et al.*, 2018), and the effects of drought are of particular concern (Burke *et al.*, 2006; Sillmann *et al.*, 2013; Jump *et al.*, 2017). Although it appears sensible that trees would increase C allocation to roots in dry regions or during drought periods (Poorter *et al.*, 2012), there is limited support

for this idea. Reich *et al.* (2014) found no correlation between root mass fraction and aridity across a global dataset of > 6200 forests. Additionally, Amazonian forests responded to droughts in 2005 and 2010 with a shift away from fine-root growth and increased C partitioning to aboveground growth and respiration, particularly in the year following the drought (Doughty *et al.*, 2014, 2015). However, drought has been observed to increase root mass fractions for small plants grown in artificial conditions (Reich, 2002; Poorter *et al.*, 2012), and increased C allocation below-ground under drought is consistent with some ^{13}C -CO₂ labeling studies (Hommel *et al.*, 2016) but not with others (Hartmann *et al.*, 2015). Although the simple expectation of increased allocation to roots during drought is appealing, C allocation responses to drought are likely more complex and merit further study.

Interactions between temperature and drought effects may also be important for tree C allocation. Warmer temperatures may exacerbate tree H₂O loss during drought and increase mortality risk (Allen *et al.*, 2015). If warmer temperatures reduce C allocation below-ground, then the ability of trees to acquire soil H₂O may also be impaired. However, an open-top chamber experiment with young oak saplings found no interaction between experimental warming and drought on tree transpiration or biomass (Kuster *et al.*, 2013). Other studies have found limited evidence for interactive effects on plant growth rates (e.g. Edler *et al.*, 2015; Taeger *et al.*, 2015), although there are also exceptions (Munir *et al.*, 2015; Leon-Sanchez *et al.*, 2016). A 6-yr warming and precipitation redistribution experiment with two tree species found complex growth responses (Volder *et al.*, 2013), with a strongly interactive effect on the relative growth rate of *Quercus stellata* monocultures. Thus, it is challenging to generalize how the interactive effects of drought and warming affect tree physiology and growth.

To address these knowledge gaps concerning, temperature, H₂O availability, and C allocation, we studied C allocation in young eucalypt trees as they grew from roughly 1–9 m in height in an experiment that manipulated both temperature and H₂O availability. We used whole-tree chambers (WTCs) in the field in southeastern Australia to grow *Eucalyptus tereticornis* trees under experimental warming of +3°C for > 1 yr, crossed with a summer drought for 3 months. We continuously measured whole-crown CO₂ and H₂O exchange and measured aboveground biomass production every 2 wk. From these intensive measurements, we derived GPP, aboveground net primary production (NPP), aboveground autotrophic respiration (R_a), and the residual C that must have been partitioned below-ground for each fortnightly interval. We use these data to test the predictions that warming decreases C partitioning below ground and drought increases C partitioning below ground.

Materials and Methods

Site and experiment

We implemented a warming and drought experiment using 12 WTCs in Richmond, New South Wales (Australia; 33°36'40"S,

150°44'26.5"E). The WTCs were large cylindrical structures topped with a cone (3.25 m in diameter, 9 m in height, volume of $c. 53 \text{ m}^3$) that enclosed a single tree rooted in soil. The WTCs controlled atmospheric CO_2 concentration, air temperature T_{air} , relative humidity RH , and irrigation while continuously measuring the net exchange of CO_2 and H_2O between entire tree crowns and the atmosphere (Barton *et al.*, 2010, 2012; Duursma *et al.*, 2011, 2014; Aspinwall *et al.*, 2016; Drake *et al.*, 2016b).

The roots of each tree were compartmentalized with a barrier extending vertically below-ground to 100 cm depth. A cemented layer of manganese nodules and clay was present at 90–100 cm depth, providing a natural horizontal barrier at the bottom of the rooting volume. Thus, the rooting volume of each tree was isolated from surrounding trees. However, some trees extended roots through this layer and acquired deep soil H_2O in a previous experiment (Duursma *et al.*, 2011). Soil was collected from an adjacent paddock and placed in the chambers in two layers (0–25 cm and from 25 cm to the hard layer) on 10 July 2012. Soils at the site were an alluvial formation of low-fertility sandy loam (Clarendon sand).

Nursery seedlings of a local provenance of *E. tereticornis* Sm. were established in 251 pots inside the WTCs using the same soil. *E. tereticornis* was chosen because it is a widespread and abundant tree across eastern Australia (Drake *et al.*, 2015). Six potted trees were placed in each chamber on 5 December 2012; a single tree was selected based on size similarity within each treatment and planted in the chamber center on 12 March 2013. Trees assigned to the ambient and warmed temperature treatments had equivalent height and basal diameter when potted seedlings were placed into the WTCs in December 2012 (heights of 41.5 ± 0.8 (SE) and 40.2 ± 1.8 cm; diameters of 2.4 ± 0.1 and 2.5 ± 0.1 mm in ambient and warmed treatment, respectively). Thus, the experiment began with small seedlings, but subsequent growth over the 15-month study period was rapid, and trees quickly grew into larger size classes.

Six chambers tracked ambient T_{air} and six chambers tracked ambient $T_{\text{air}} + 3^\circ\text{C}$ warming ($n=6$; 'ambient' and 'warmed', respectively); treatments started on 12 December 2012 (Aspinwall *et al.*, 2016; Drake *et al.*, 2016b). The average warming achieved was $+2.9^\circ\text{C}$ (SD ± 0.3 across 466 d) for T_{air} , $+2.2^\circ\text{C}$ (SD ± 0.4) for soil temperature at 5 cm depth, $+2.0^\circ\text{C}$ (SD ± 0.4) for soil temperature at 20 cm depth, and $+1.4^\circ\text{C}$ (SD ± 0.2) for soil temperature at 50 cm depth. Trees were irrigated equally every 15 d with half the mean monthly rainfall, which we consider the control treatment. An H_2O exclusion treatment was applied to half of the trees on 12 February 2014, resulting in a 2×2 factorial design between the experimental treatments of warming and drought ($n=3$; abbreviated A-Con, A-Dry, W-Con, and W-Dry hereafter). Trees assigned to the drought treatment received no irrigation from 12 February 2014 through 5 May 2014, representing a summer drought of nearly 3 months. A rainfall record from 1881 to 2014 from this area (Australian Bureau of Meteorology station 67021; station is < 2 km from site) indicates that only three years ($c. 2\%$ of the record) had < 20 mm precipitation during the months of the drought treatment, and 80% of the years had > 100 mm

precipitation. This suggests that our drought was relatively strong in terms of the surface H_2O addition during these months.

Plant H_2O status and soil H_2O content

Predawn leaf H_2O potentials $\Psi_{\text{L-PD}}$ were measured monthly before the drought and every 1–2 wk during the drought treatment. Three leaves were measured per tree on each date using a Scholander-type pressure chamber (1505D-EXP; PMS Instrument Company, Albany, OR, USA). Leaves were placed in sealed and humidified plastic bags, placed in a dark cool box, and measured within 1 h of collection in a nearby laboratory.

Soil volumetric H_2O content was measured by three sensors in each chamber (CS650 time-domain reflectometers; Campbell Scientific, Logan, UT, USA). Sensors were installed horizontally at three depths: in the surface soil (10 cm depth), at 30 cm depth, and just above the hard layer of cemented manganese ($c. 100$ cm depth). Soil temperature was measured with thermocouples at 5, 20, and 50 cm within the center of each chamber.

A single neutron probe tube per chamber (503DR, Hydroprobe; InstroTek, Research Triangle Park, NC, USA) was used to measure soil H_2O content to a depth of 425 cm (at 25 or 50 cm steps) approximately every 2 wk (Duursma *et al.*, 2011). Note that high neutron probe counts in deep soil (150–400 cm depth) partially reflect a change in soil texture towards a higher clay content.

Whole-tree crown flux measurements

An automated system measured the net exchange of CO_2 and H_2O between each crown and its chamber airspace (Barton *et al.*, 2010). Measurements began on 13 September 2013 when suspended plastic floors were sealed around the stem of each tree at $c. 45$ cm height, when the trees were $c. 3$ m tall. Flux measurements finished on 26 May 2014, when the trees were nearly 9 m tall. We report $> 70\,000$ hourly flux observations aggregated into > 3000 daily sums across 12 trees.

We partitioned the net CO_2 fluxes into the components of GPP and R_a using an analytical technique common to eddy-covariance research (Reichstein *et al.*, 2005). We used direct measurements of whole-crown R_a and its temperature dependence at night to predict R_a for each hourly measurement as a function of T_{air} . For daylight hours, we then calculated GPP as the sum of the measured net CO_2 flux and the predicted R_a given the measured T_{air} . We assumed GPP was zero when photosynthetic photon flux density was zero; in such conditions, the measured net C flux was used as the measure of R_a . Note that the chamber airspaces were continuously well mixed and R_a fluxes were directly measured at night, avoiding some of the issues inherent in eddy-covariance partitioning. The underlying flux data and the partitioning approach were published previously (Drake *et al.*, 2016a,b).

Final harvest

The dry mass of all trees was measured destructively at the end of the experiment (26 May 2014), 15 months after seedlings were planted. At this time, the trees had heights of 8.8 ± 0.14 m and

diameters of 6.6 ± 0.2 cm. Total tree dry mass was measured as the sum of five components: leaves, branches, stem, coarse roots, and fine roots.

The crown of each tree was divided into three equal heights. All branches were cut flush to the stem and all leaves were separated from branches. A random subsample of 100 leaves per layer was measured for total leaf area (LI-3100C leaf area meter; LiCor, Lincoln, NE, USA), dry mass, and specific leaf area (SLA). The stem was cut into three segments, and a single 1 cm thick cross-section (cookie) was sampled for bark depth, wood density, and bark density at the stem base, between the first and second layers, and between the second and third crown layers. Bark and wood densities were measured on cookie subsamples (Thomas *et al.*, 2007). Wood and bark densities were similar (0.44 g cm^{-3} and 0.37 g cm^{-3} , respectively). Bark depth increased with stem diameter ($\log_{10}(\text{bark depth, mm}) = -1.48 + 1.23 \times \log_{10}(\text{diameter, cm})$, $P < 0.001$, $r^2 = 0.92$), whereas wood and bark density (g cm^{-3}) decreased with stem diameter (cm) (wood density = $0.50 - 0.001 \times \text{diameter}$, $P = 0.007$, $r^2 = 0.17$; bark density = $0.45 - 0.001 \times \text{diameter}$, $P < 0.001$, $r^2 = 0.48$). The reduction in density with increasing stem diameter reflected increasing densities in the upper layers of the tree crowns. Warming and drought treatments did not alter these relationships (analysis of covariance (ANCOVA), $P > 0.05$). Total stem, branch, and leaf mass were measured directly after drying at 70°C ; some samples required > 2 wk of drying to reach a stable dry mass.

Fine roots were measured using soil cores. The soil surface area was divided into four equal quadrants and two 50 mm diameter cores were taken within each quadrat on 29 May 2014, just after the crown harvest. Cores were separated into two depths: (1) 0–25 cm and (2) from 25 cm to the hard layer, which varied from 70 to 100 cm depth. Samples within each quadrat and depth category were composited (eight samples per chamber). Fine roots were isolated by washing samples through 2 mm and then 1 mm brass sieves; fine roots were defined as all roots < 2 mm diameter. Fine root dry mass was measured after drying at 70°C . Total fine root dry mass was calculated as the product of fine root density in the soil (g m^{-3}) and soil volume (m^3) in each layer.

Coarse roots were destructively harvested by fully excavating the soil volume of each chamber. Soil was shoveled out of the chamber onto a conveyor belt that transported the soil to a series of 5 mm steel sieves. Roots were collected by hand, washed, sorted into two size categories (2–10 mm, > 10 mm diameter), and weighed after drying at 70°C . During the excavation, several small roots (≈ 1 cm diameter) were observed to have penetrated through the hard layer and into deeper soil (J. E. Drake, personal observations).

Growth measurements

Aboveground biomass was estimated every 2 wk for each tree as the sum of leaf, branch, wood, and bark mass. The following paragraphs describe the methodologies for each component in detail. Aboveground net primary production NPP_a was estimated as the fortnightly difference in aboveground biomass plus fortnightly litterfall, assuming a constant biomass C fraction of 0.5

of dry mass. Allocation to reproduction was not explicitly measured, but it can be considered zero as these trees did not produce any reproductive structures.

Tree height and stem volume were measured fortnightly; diameter was measured at 30 cm intervals along each tree stem from a basal height of 15 cm (before floor installation) or 65 cm (after floor installation) to the tree apex. The volume of stem wood and bark was estimated for each stem segment as the frustum of a cone, corrected for bark depth (see earlier). Wood and bark mass were calculated as the product of volume and density. This approach assumes that the wood and bark density did not change through time in this experiment.

An allometric relationship was developed to predict branch wood mass from branch diameter. Four branches from each experimental tree were destructively sampled near the end of the study (13 May 2014 and 22 May 2014); branch mass was strongly correlated with branch diameter ($\log_{10}(\text{branch mass, g}) = -1.299 + 2.722 \times \log_{10}(\text{branch diameter, mm})$, $P < 0.001$, $r^2 = 0.91$, $n = 48$ branches). This allometry did not differ between treatments (ANCOVA, $P > 0.1$) and was used to predict total branch mass on three dates when the diameters of all branches were measured (24 October 2013, 15 January 2014, and 22 May 2014). Total branch mass and stem volume were strongly correlated in a chamber-specific manner ($\log\text{--}\log$ ANCOVA, $P < 0.001$, $r^2 = 0.95$), which was used to estimate branch mass as a function of stem volume.

Standing leaf area and leaf mass production were estimated as previously at this site (Barton *et al.*, 2012; Drake *et al.*, 2016b). Standing leaf area was measured for each tree by counting all the leaves and multiplying by a tree-specific mean leaf size measured across the crown of each tree with a handheld leaf area meter (LI-3000; $n = 86\text{--}102$ leaves per tree). These measurements were performed before chamber floor installation (9 September 2013) and at the beginning of the drought treatment (10 February 2014). A third direct measurement of standing leaf area was calculated from the final harvest data (26 May 2014) by multiplying total crown leaf dry mass by SLA weighted by the leaf dry mass in each layer. Litterfall was collected, dried, and weighed fortnightly for each tree, although relatively few leaves fell as litter ($\approx 5\%$ of the total leaf mass). Total tree leaf mass was estimated for each set of fortnightly size measurements by dividing leaf area by the crown-weighted SLA measured at harvest.

Calculating C partitioning

A major goal of this study was to calculate the partitioning of photosynthetically fixed C into components for each fortnightly interval. We quantified GPP, NPP_a , and R_a separately, as described earlier. We calculated the residual between GPP and the sum of NPP_a and R_a :

$$\text{GPP} = \text{NPP}_a + R_a + \text{residual} \quad \text{Eqn 1}$$

The residual term is a mass-balance calculation of all C put below ground to root production, respiration, and exudation, but this term is also affected by measurement error in GPP, NPP_a ,

and R_a . We calculated the partitioning of GPP directly for each fortnightly interval as NPP_a/GPP , R_a/GPP , and residual/GPP.

Growth and maintenance R_a

Given the evidence for thermal acclimation of tissue-specific respiration rates to experimental warming in this experiment (Aspinwall *et al.*, 2016; Drake *et al.*, 2016b), we investigated growth and maintenance respiration as drivers of R_a (McCree, 1970; Tjoelker *et al.*, 1999; Amthor, 2000; Adu-Bredu & Hagi-hara, 2003). Separating growth and maintenance components of R_a is not possible in all C-cycle studies, but we were able to couple direct measures of tree growth with continuous measurements of integrated whole-tree aboveground respiration to enable quantification of growth and maintenance components. We evaluated the relationship between R_a per unit tree C and relative growth rate RGR; the slope reflects the growth component of R_a , whereas the y -intercept reflects the maintenance component of R_a . If R_a does not acclimate to warming, we expect the warmed treatment to have a higher y -intercept than the ambient treatment. If R_a acclimates homeostatically, we expect the ambient and warmed treatments to have equivalent intercepts. We also directly estimated coefficients associated with growth and maintenance components of R_a (Amthor, 2000);

$$R_a = R_g + R_m = g_r G + m_r W$$

Eqn 2

where R_g (grams of C per day, $g_C d^{-1}$) is the growth respiration rate, R_m ($g_C d^{-1}$) is the maintenance respiration rate, G ($g_C d^{-1}$) is biomass growth, W (gram of C, g_C) is the standing biomass weight, g_r (grams of C respired per gram of C growth, $g_C g_C^{-1}$) is the growth respiration coefficient, and m_r (grams of C respired per gram of C standing biomass per day, $g_C g_C^{-1} d^{-1}$) is the maintenance respiration coefficient.

Data analysis

Data were analyzed following a completely randomized design with the single treatment of warming ($n=6$ for 6 months, then $n=3$ for the drought period). Longitudinal analyses were performed using the 'lme' function within the NLME R package with a random tree effect and fixed effects of date, temperature treatment, and H₂O treatment. Treatment means were estimated after adjustment for other terms in the model (i.e. least-square means) with the LSMEANS package in R v.3.2.2 (R Development Core Team, 2012; Pinheiro *et al.*, 2013). Analyses were evaluated to test assumptions of residual normality and homoscedasticity; transformations were often necessary. Datasets that were not longitudinal were analyzed as a 2×2 ANOVA using the 'lm'

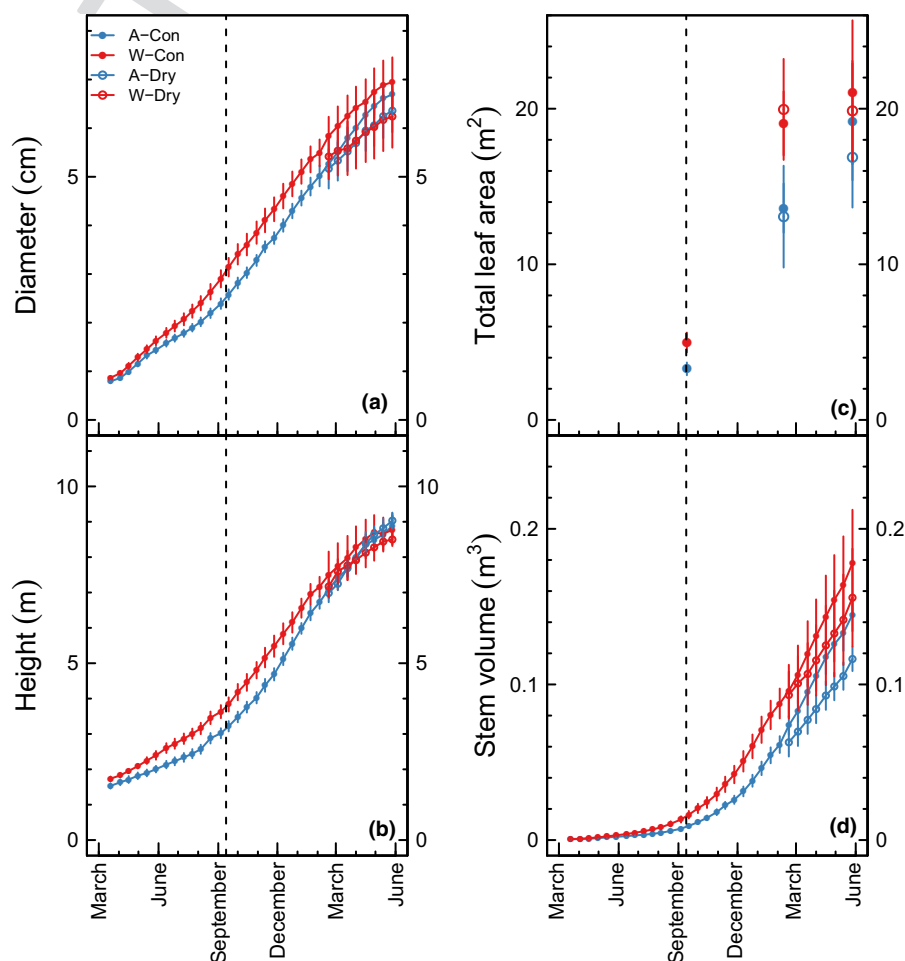


Fig. 1 Growth of *Eucalyptus tereticornis* trees exposed to warming and drought. Trees were either exposed to ambient T_{air} ('A', blue) or warming of +3°C ('W', red), and either a well-watered control ('Con', solid points) or drought conditions ('Dry', open points). Stem diameter (a) was measured at 65 cm height, and height reflects total stem length (b). Total leaf area was directly measured on three dates (c), and stem volume was calculated from diameter measurements along the stem of each tree (d). The vertical dashed line denotes when CO₂ and water flux measurements began. Points reflect the mean, error bars denote one SEM ($n=6$ until February 2014, when the drought treatment began and $n=3$).

function in R. Eqn 2 was fit using the NLME R package with a random tree effect.

Results

Growth

Experimental warming increased the rates of diameter and height growth (Fig. 1a,b), particularly during the austral winter and spring. Trees in the warmed treatment were larger than trees in the ambient treatment when the CO₂ and H₂O flux measurements began (13 September 2013; vertical dashed line in Fig. 1). On that date, warming had increased diameter by 21% ($P < 0.01$; Fig. 1a), height by 19% ($P < 0.01$; Fig. 1b), total leaf area by 53% ($P < 0.01$; Fig. 1c), and stem volume by 79% ($P < 0.01$; Fig. 1d). During the warm summer, the diameter and height of the ambient and warmed treatment trees converged (Fig. 1a,b), but total stem volume continued to be larger in the warmed

treatment (Fig. 1d). This effect was driven by a difference in stem taper – the warmed trees had wider stem diameters throughout the crown than the ambient trees (not shown). The drought treatment reduced tree diameter but not height growth (Fig. 1a,b), modestly reduced total leaf area (Fig. 1c), and reduced stem volume increment (Fig. 1d). Notably, there was no interactive effect of warming and drought on growth (e.g. $P > 0.4$ for volume increment).

CO₂ and H₂O fluxes

Experimental warming increased photosynthetic C uptake and H₂O loss via transpiration early in the experiment (Fig. 2a,c). This was expected, given the strong increase in tree growth and total leaf area with experimental warming during this period (Fig. 1). However, the rates of C uptake and H₂O loss converged between the ambient and warmed treatments during the summer (January; Fig. 2a,c), despite the fact that the warmed trees were

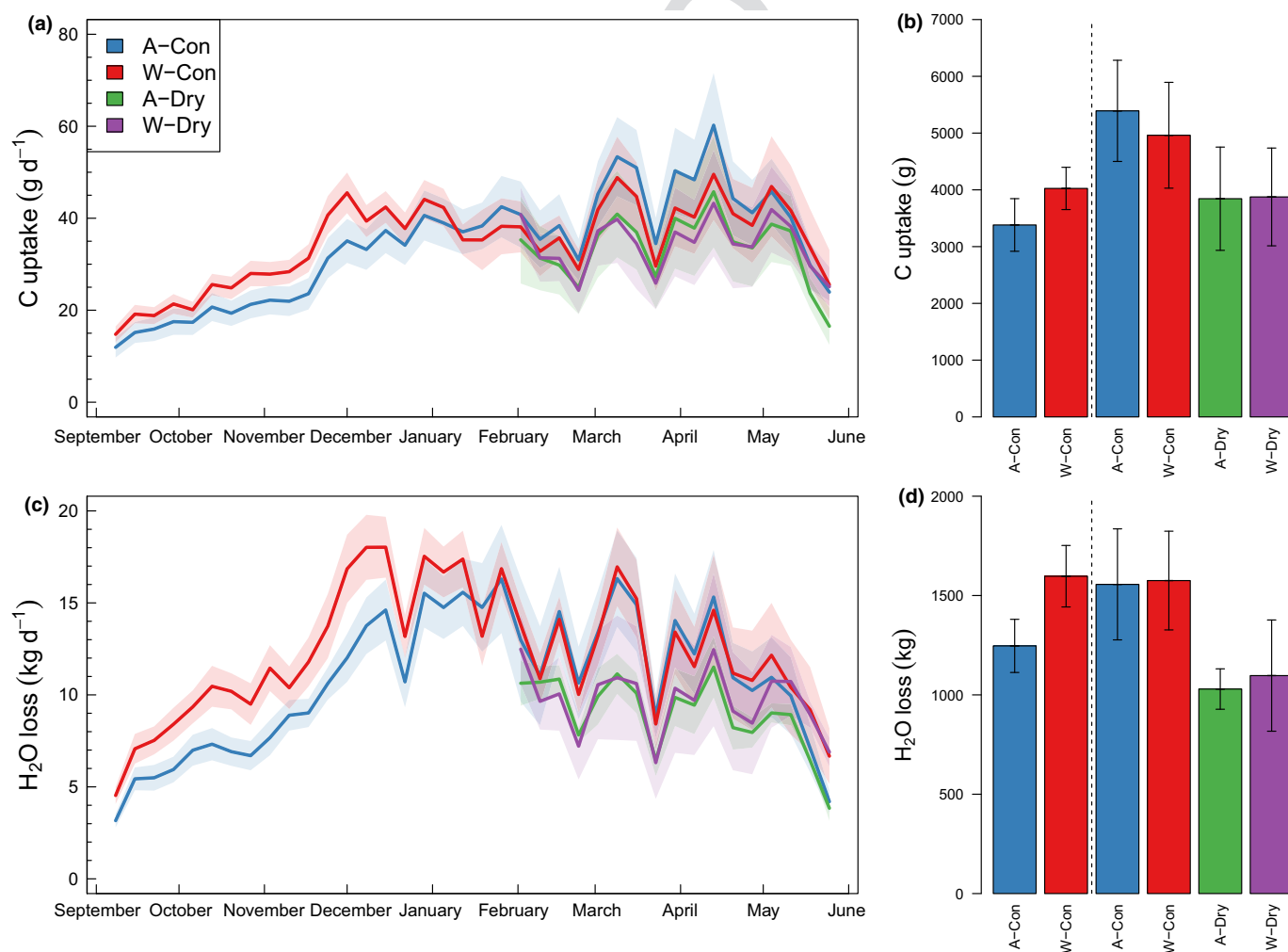


Fig. 2 Summary of measured CO₂ and water (H₂O) fluxes for 12 *Eucalyptus tereticornis* trees exposed to ambient (A) or warmed (W) air temperatures in 2013 and 2014. All trees were maintained in control well-watered conditions (Con) until mid-February, when half of the trees were subjected to a soil drydown (Dry). We show weekly averages of the measured daily net carbon (C) uptake (a) and the sum of net C uptake for the two measurement periods (pre-drought, drought; b). We also show weekly averages of the measured daily net H₂O loss to transpiration (c) and the H₂O loss to transpiration summed across the two measurement periods (d). In (a) and (c), lines reflect the mean and shaded areas reflect the SE. The dotted vertical lines in (b) and (d) separate the pre-drought (left) and drought periods (right). These plots reflect > 580 000 individual flux measurements at 15 min resolution.

larger and had more leaf area. This may have been influenced by warming-induced reductions in photosynthetic rates per unit leaf area (Drake *et al.*, 2016b).

We imposed an experimental drought in which all surface irrigation was withheld from trees in the dry treatments for nearly 3 months. Total C uptake during the drought period was reduced 25%, while total H₂O loss was reduced 32% (Fig. 2b,c; main effects of drought, $P < 0.01$; no interaction with warming, $P > 0.5$). Thus, the drought strongly and significantly reduced whole-crown fluxes of C uptake and H₂O loss. On the other hand, these fluxes were maintained at moderate values during the drought, despite the complete lack of H₂O addition.

Final harvest

The final biomass did not significantly differ between the warming or drought treatments (Fig. 3a). The lack of difference in final mass between the ambient and warmed treatments may have arisen from the convergence of tree diameter and height across treatments (Fig. 1a,b). The difference in stem volume between ambient and warmed trees (Fig. 1d) was apparent in the harvest biomass (Fig. 3a) but was not statistically significant at this level of replication ($n = 3$; $P > 0.05$). The only biomass component that was affected by the experimental treatments at harvest was fine root biomass, for which there was a significant interaction between warming and drought ($P < 0.05$). The A-Dry trees had higher fine root biomass than the A-Con trees, whereas the W-Dry trees had slightly lower fine root biomass than the W-Con trees (Fig. 3a). This interaction was also present in the tree root mass ratios; experimental drought increased the root mass ratio,

but only in the ambient temperature treatment ($P < 0.05$; Fig. 3b).

Plant and soil H₂O status

The drought reduced soil volumetric H₂O content from the 10–100 cm depth to values approaching 0.05 m³ m^{−3} (Fig. 4a–c). Predawn leaf H₂O potentials Ψ_{L-PD} were reduced in the dry treatments relative to the control treatments (Fig. 4d; $P < 0.01$). However, this effect was modest; Ψ_{L-PD} was -0.29 ± 0.02 in the control and -0.48 ± 0.05 in the dry treatments. Thus, the drought trees had moderate Ψ_{L-PD} (Fig. 4d) and moderate rates of transpiration (Fig. 2c) despite extremely dry surface soils.

Trees likely utilized deep soil H₂O during the drought treatment. We observed a few roots of *c.* 1 cm diameter penetrating through the cemented manganese layer at *c.* 100 cm depth during the complete soil excavation (J. E. Drake, personal observations). Neutron probe measurements down to 400 cm depth indicated that soil H₂O was removed from the profile in the dry treatment chambers during the drought, particularly from the 50–200 cm depth (Supporting Information Fig. S1). Thus, trees in the dry treatments likely transpired deep soil H₂O during the summer drought, consistent with a previous drought study of *Eucalyptus saligna* at this site (Duursma *et al.*, 2011).

Fluxes of GPP, NPP_a, R_a and allocation below ground

We derived GPP and its partitioning to NPP_a, R_a , and the residual, which we attribute to C allocation below ground and to measurement error.

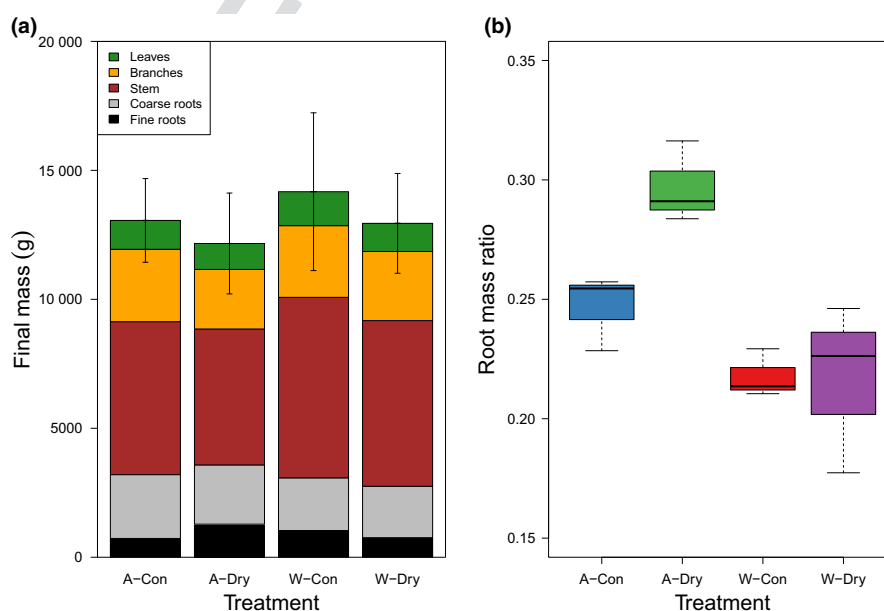


Fig. 3 Biomass components at final harvest for 12 *Eucalyptus tereticornis* trees exposed to ambient (A) or warmed (W) air temperatures and either control well-watered conditions (Con) or a soil drydown treatment (Dry). Note that these data reflect grams of dry mass. Each of the measured biomass components (a) reflects the mean of three trees per treatment, the error bars reflect the SE of the total measured mass. The root mass ratio (b) reflects the sum of coarse and fine roots relative to total tree mass (boxplot of three observations per category; the thick line reflects the median value and the whiskers reflect the other two values; the box shows the 25th and 75th percentiles). Warming reduced the root mass ratio, whereas the drought treatment increased root mass ratio in the ambient temperature treatment only. The root mass ratio interaction primarily follows the response of fine roots, although stem wood and coarse roots also contributed.

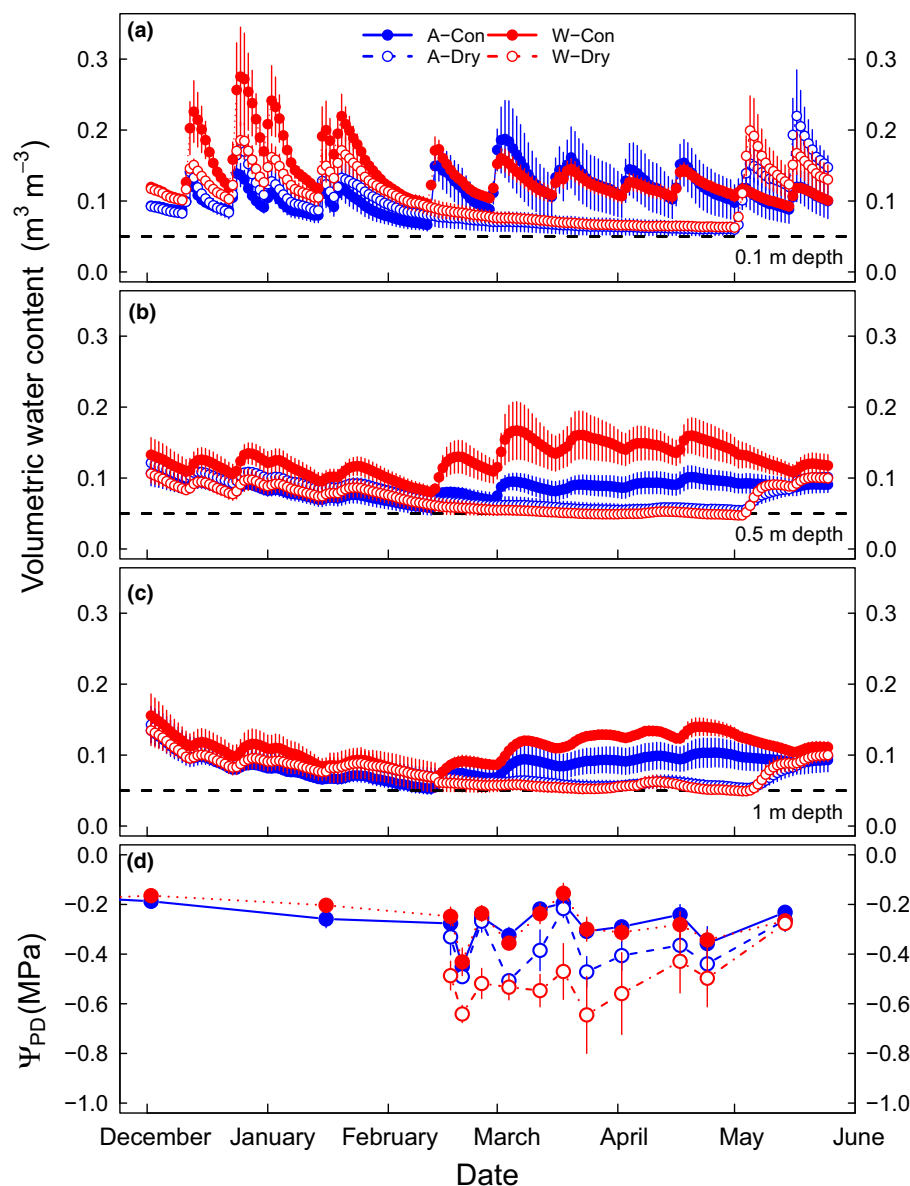


Fig. 4 Soil volumetric water content and predawn leaf water potential Ψ_{PD} for 12 *Eucalyptus tereticornis* trees exposed to ambient (A) or warmed (W) air temperatures. All trees were maintained in control well-watered conditions (Con) until mid-Feb, when half of the trees were subjected to a soil drydown (Dry). We show daily averages of the measured volumetric water content in surface soils (c. 0.1 m depth; a), an intermediate depth (c. 0.5 m depth; b), and in deep soils just above the hard layer of partially cemented manganese nodules (c. 1 m depth; c). The horizontal dashed line reflects the volumetric water content at which soil matric potential drops to -1.5 MPa. We also show leaf Ψ_{PD} measured throughout the drydown (d). Points reflect the mean and error bars reflect the SE ($n = 6$ or 3). Note that Ψ_{PD} was moderate in all treatments.

GPP was increased by experimental warming early in the experiment (+22%, $P < 0.01$), but GPP between ambient and warmed treatments converged beginning in mid-summer (late January; Fig. 5a). The drought treatment reduced GPP in both temperature treatments (-15% , $P < 0.01$). These results follow the net C flux measurements (Fig. 2a,c). The response of NPP_a (Fig. 5b) closely followed the results for GPP, with a warming effect early in the experiment (+36%, $P < 0.01$) and a reduction with drought in both temperature treatments (-25% , $P < 0.01$). The response of R_a (Fig. 5c) also followed GPP, with a stimulation by warming early in the experiment (+39%, $P < 0.01$) and a modest reduction with drought that was equivalent across temperature treatments (-13% , $P < 0.05$). The allocation of C below ground, as measured by the residual, was decreased by experimental warming throughout the experiment (-11% ,

$P < 0.05$) and was unchanged by the drought treatment (+3%, $P > 0.1$; Fig. 5d).

GPP partitioning

Given these flux measurements, we derive the partitioning of GPP into three components: NPP_a/GPP , R_a/GPP , and residual/GPP (Figs 6, 7). Warming increased NPP_a/GPP in a way that was stronger early in the experiment (+11%, $P = 0.01$) relative to the entire experiment (+3%; $P > 0.1$; Fig. 6a,b). Similarly, warming increased R_a/GPP (+12%; $P < 0.1$) but decreased residual/GPP (-15% ; $P < 0.05$) before the drought (Figs 6, 7). Thus, experimental warming increased the partitioning of GPP to aboveground components (Fig. 7a,b) and decreased partitioning below-ground (Fig. 7c). The experimental drought had weak

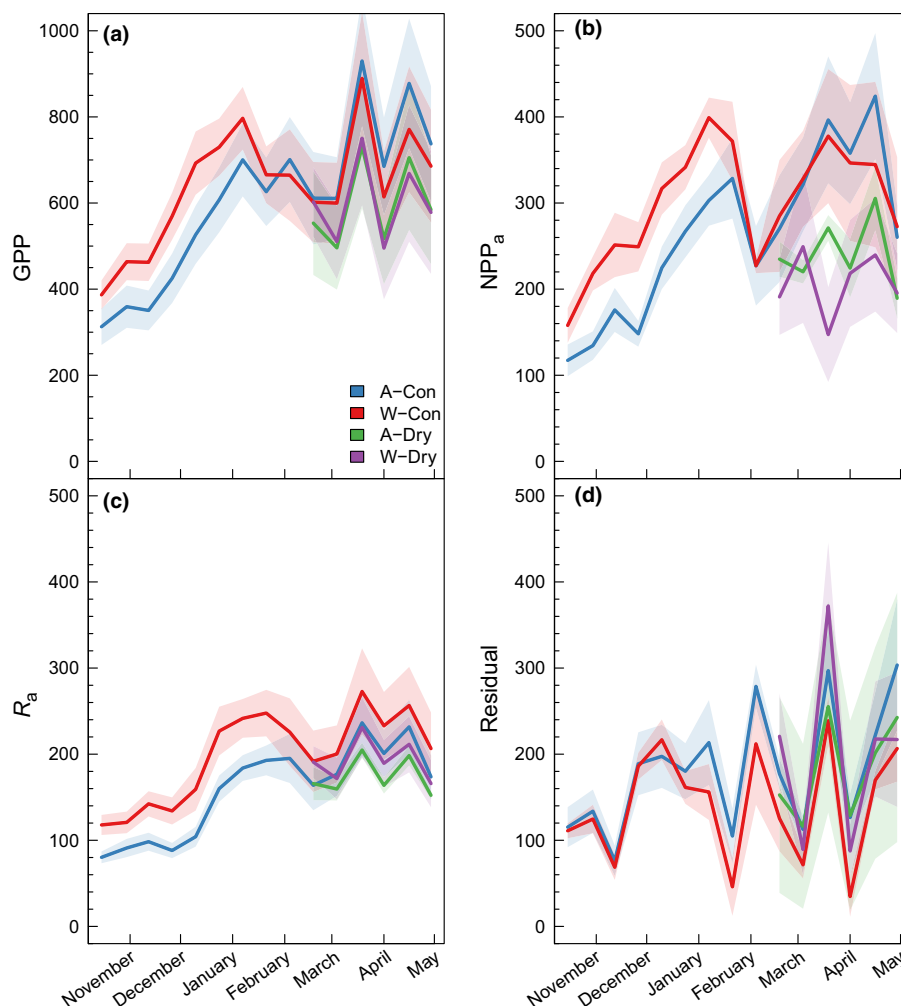


Fig. 5 Fortnightly carbon (C) fluxes for 12 *Eucalyptus tereticornis* trees exposed to ambient (A) or warmed (W) air temperatures. All fluxes are presented in units of grams of C per tree per fortnight. All trees were maintained in control well-watered conditions until mid-February (Con), when half of the trees were subjected to a soil drydown (Dry). Solid lines reflect the mean of fortnightly data (i.e. 2-wk periods) and shaded areas reflect one SEM. Measurements include gross primary production GPP (a), aboveground net primary production NPP_a (b), aboveground autotrophic respiration R_a (c), and the residual (d). The residual reflects belowground C flux and measurement error. Note that the y-axis scale is twice as large for GPP relative to the other fluxes.

effects on partitioning, none of which were statistically significant ($P > 0.1$).

Growth and maintenance respiration

We combined growth and respiratory measurements to infer changes in respiratory C efflux attributable to growth vs maintenance respiration (Amthor, 2000). There was a strong and linear relationship between R_a per unit aboveground tree C and relative growth rate (RGR; Fig. 8). Neither the slope nor the intercept of this relationship was affected by experimental treatments (all $P > 0.1$). Thus, we present a common relationship across all measurements. The y-intercept of this relationship was positive (mean of 0.0213, 95% confidence interval of 0.0157–0.0268), indicating significant R_a in the absence of aboveground growth, reflecting maintenance respiration. The lack of a warming effect on this y-intercept is consistent with respiratory temperature acclimation; trees in the ambient and warmed treatments expended similar amounts of C on maintenance respiration, despite the increased temperature in the warmed treatment. The slope of the relationship (Fig. 8) was strongly positive (mean of 0.0059, 95% confidence interval of 0.0053–0.0065), indicating that much of the R_a observed at the crown scale was attributable to

construction respiration. Observations during the drought period followed the general relationship, with lower values on both axes (Fig. 8). Thus, the experimental drought reduced R_a primarily via a reduction in growth respiration.

We also directly estimated coefficients for growth and maintenance respiration by fitting Eqn 2 to the fortnightly dataset of standing biomass, growth rate, and respiration. We estimate the growth respiration to consume $\approx 0.3 \text{ g}_C$ per gram of biomass C produced, and maintenance respiration to consume $\approx 0.015 \text{ g}_C$ per gram of standing biomass C per day (Table 1). These coefficients did not differ across the ambient and warmed treatment ($P > 0.3$).

Discussion

Summary

We studied the experimental effects of warming and drought on the C allocation of *E. tereticornis* trees using a combination of growth and whole-crown flux measurements. This was the first study to directly measure allocation changes of relatively large trees (as large as 9 m tall) in an ecologically relevant field setting with detailed and continuous measurements.

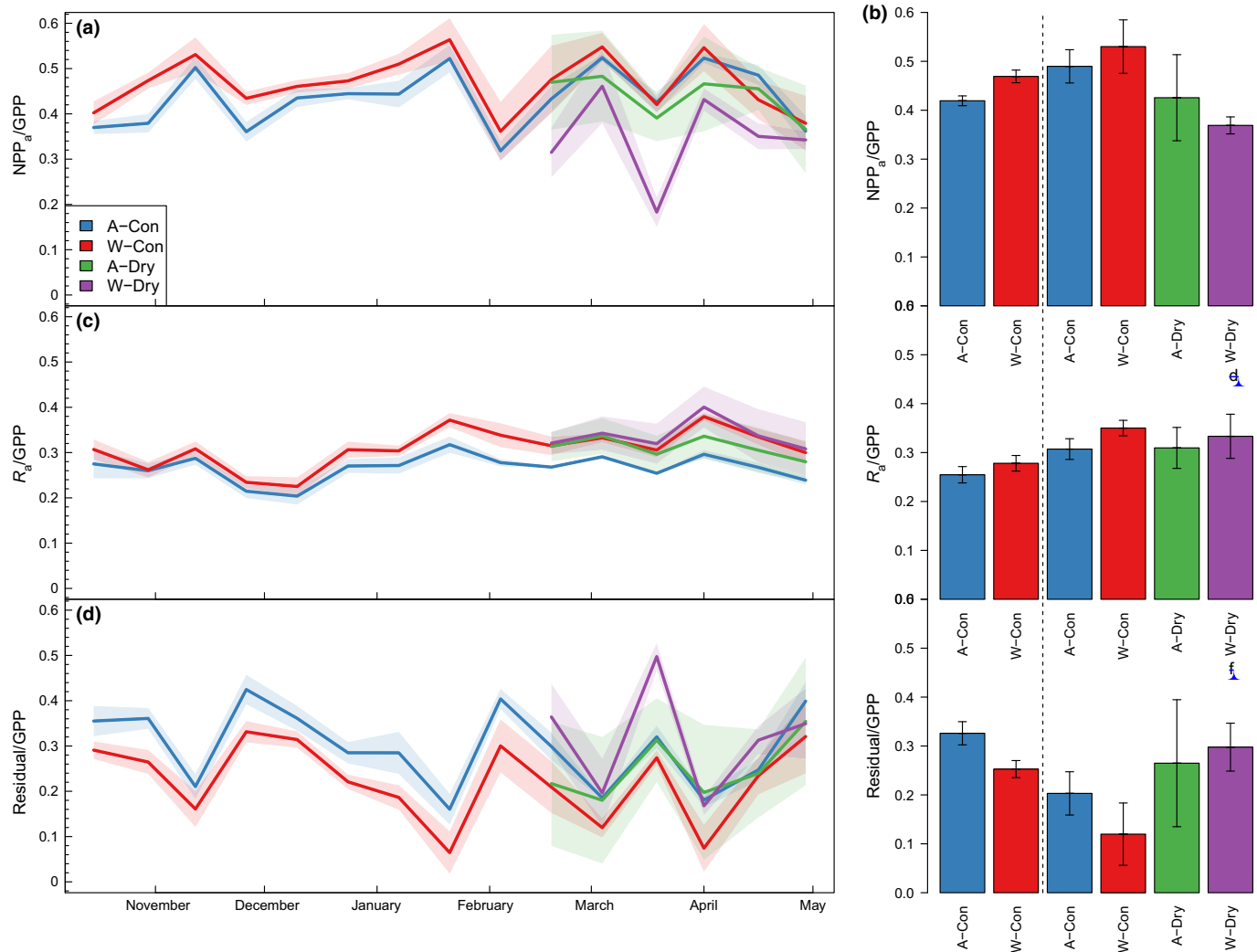


Fig. 6 The fractional partitioning of gross primary production GPP for 12 *Eucalyptus tereticornis* trees. GPP was partitioned into aboveground net primary production NPP_a (a, b), aboveground autotrophic respiration R_a (c, d), and the residual carbon (C), which includes belowground C allocation and measurement error (e, f). The shaded areas in a–c reflect ± 1 SEM. All trees were maintained in control well-watered conditions (Con) until mid-January, when half of the trees were subjected to a soil drydown (Dry). Bar charts of flux partitioning terms (b, d, f) represent the mean (± 1 SEM), and the dotted vertical lines separate the pre-drought (left) and drought periods (right). A, ambient air temperatures; W, warmed air temperatures.

Experimental warming increased the proportion of GPP that was allocated to aboveground uses and decreased the proportion of GPP that was allocated below-ground. This was consistent with a reduced root mass fraction in the warmed treatments at the final harvest. The experimental drought reduced CO_2 and H_2O fluxes but did not affect the allocation of C, perhaps because tree access to deep soil H_2O prevented them from experiencing strongly negative H_2O potentials. There were no interactions between warming and drought on C partitioning terms, so we discuss the impacts of warming and drought separately.

Effects of experimental warming on C allocation

Experimental warming strongly affected several aspects of tree C allocation. Warming increased the fractional partitioning of GPP to aboveground uses, including growth and respiration,

at the expense of C partitioning below-ground. This observation is consistent with some soil warming experiments (e.g. Melillo *et al.*, 2002, 2011) that attributed this effect to a warming-induced stimulation of soil nutrient availability. However, it is also possible that experimental warming directly stimulated the activity of meristems above ground, such that a smaller remainder of fixed C was available for transport and use below-ground. Such a mechanism would imply an aboveground priority in tree C allocation, consistent with previous work on forest C budgets and elevated atmospheric CO_2 treatments (Palmroth *et al.*, 2006). This mechanism also makes sense given the structural arrangement of tree phloem, as aboveground tissues have the opportunity to remove sucrose from the phloem before belowground tissues do (Lemoine *et al.*, 2013; Furze *et al.*, 2018). The mechanisms regarding soil nutrient availability and aboveground metabolic activity are not mutually exclusive. For example, enhanced N

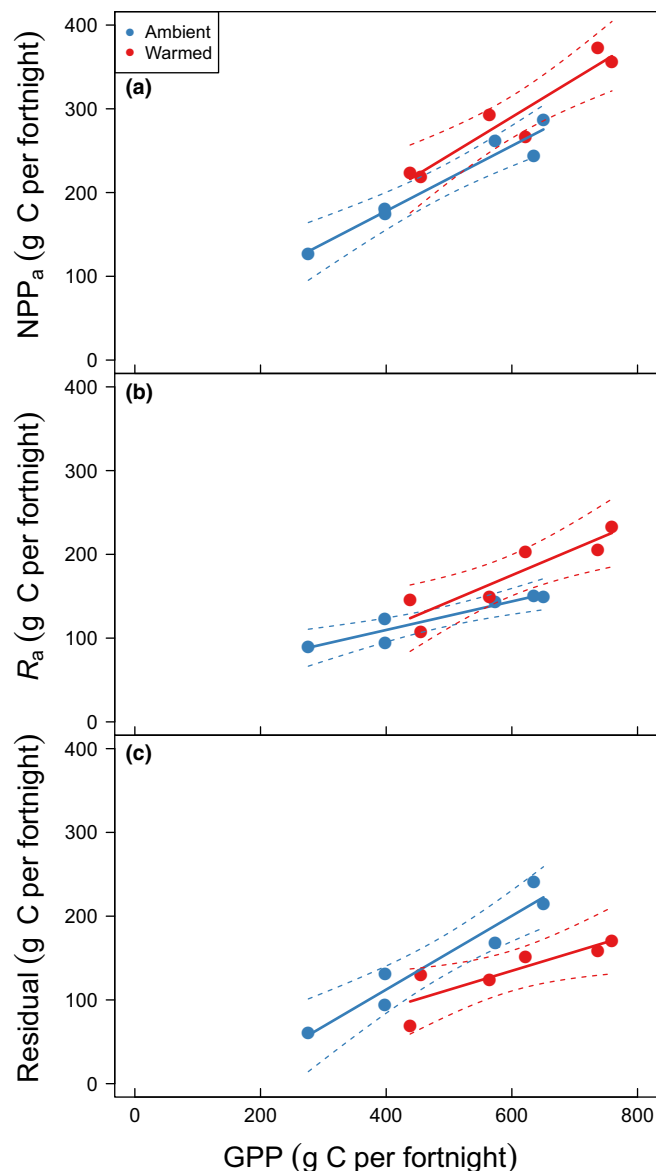


Fig. 7 The fractional partitioning of gross primary production GPP for 12 *Eucalyptus tereticornis* trees grown under ambient (A) and elevated (W) temperature. Data for each tree were averaged across the pre-drought period; each point reflects an individual tree ($n = 6$). GPP was partitioned into aboveground net primary production NPP_a (a), aboveground autotrophic respiration R_a (b), and the residual carbon (C), which includes belowground C allocation (c). Solid lines reflect linear models fit to each treatment; dashed lines reflect the 95% confidence interval. Con, control well-watered; Dry, soil drydown.

supply from soil N mineralization may have enabled the increased aboveground metabolism in the warmed treatment, which may have resulted in the larger consumption of GPP above-ground in the warmed relative to the ambient treatment.

We also acknowledge that warming may have influenced allocation indirectly via ontogenetic drift. A meta-analysis documented a decline in root-to-shoot ratios as trees grew larger, possibly reflecting an ontogenetic effect on belowground allocation (Mokany *et al.*, 2006), although we hesitate to infer

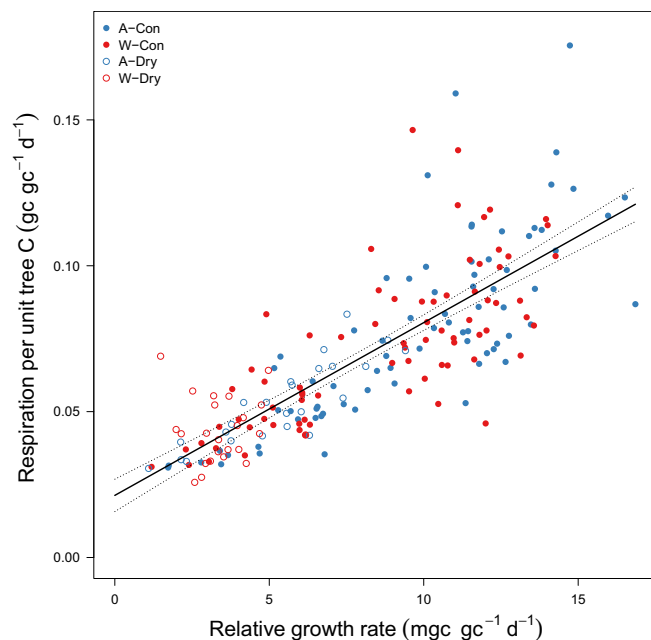


Fig. 8 Partitioning of aboveground respiration into maintenance and growth components for *Eucalyptus tereticornis* trees in a warming and drought experiment. Each point reflects a tree during a fortnightly growth interval. Note that the y-intercept reflects the maintenance respiration component and the slope reflects the growth respiration component. Neither the slope nor the intercept was affected by experiment treatments (mixed effects model with random intercepts for each chamber, $P > 0.5$). The solid lines reflect models fit to the ambient temperature (A) and warmed temperature (W) data, and dashed lines reflect the 95% confidence interval. All of the data were well described by a single linear function ($Y = 0.021 + 0.0059x$, $r^2 = 0.64$, $P < 0.001$). Con, control well-watered; Dry, soil drydown.

allocation directly from root-to-shoot ratios (Reich, 2002; Litton *et al.*, 2007). In this experiment, the allocation terms (e.g. NPP_a/GPP , R_a/GPP , and residual/GPP) were not significantly correlated with any metric of tree size, suggesting that the warming effect is unlikely to reflect ontogeny.

We previously demonstrated that aboveground autotrophic respiration acclimated nearly homeostatically to experimental warming in this experiment, both at the leaf scale (Aspinwall *et al.*, 2016) and at the whole-crown scale (Drake *et al.*, 2016b). As such, the demonstration that warming increased R_a (Fig. 5) may appear contradictory. We emphasize that our previous presentations of autotrophic respiration were expressed per unit leaf area (Aspinwall *et al.*, 2016; Drake *et al.*, 2016b), whereas the current study shows the total fluxes per tree (Fig. 5c). Experimental warming increased R_a primarily by increasing growth and tree size early in the experiment. Furthermore, the common relationship between relative growth rate and R_a per unit tree mass for the ambient and warmed treatment is indicative of homeostatic acclimation of maintenance respiration in this experiment (Fig. 8). The stimulation of whole-crown R_a by warming was primarily attributable to an increase in respiration to support growth. Thus, we suggest that this study is in agreement with previous published work from this experiment (Aspinwall *et al.*, 2016; Drake *et al.*, 2016b), where homeostatic acclimation of

Table 1 Estimate of aboveground growth and maintenance respiration coefficients for *Eucalyptus tereticornis* trees derived from Eqn 2, with SEs and 95% confidence intervals (CIs); parameters were statistically equivalent across ambient and warmed treatments (all $P > 0.3$).

Term	Units	Ambient		Warmed	
		Mean (SE)	95% CI	Mean (SE)	95% CI
g_r (growth respiration rate)	g C respired per g C growth	0.32 (0.02)	0.27–0.37	0.28 (0.03)	0.21–0.36
m_r (maintenance respiration rate)	g C respired per g C standing aboveground biomass per day	0.015 (0.001)	0.012–0.019	0.017 (0.002)	0.013–0.021

respiration to experimental warming prevented a warming-induced increase in maintenance respiration, whereas a warming effect on growth stimulated growth respiration and increased whole-crown R_a .

The increased allocation of C above-ground in the warmed treatment, combined with homeostatic acclimation of maintenance respiration, likely contributed to the observed warming-induced stimulation in growth during the first half of this experiment (Fig. 1). Experimental warming had neutral or negative effects on leaf-level photosynthetic rates in this study (Aspinwall *et al.*, 2016; Drake *et al.*, 2016b), so a warming-induced stimulation of growth was somewhat surprising. We suggest that an increase in C partitioning above-ground (Fig. 6a) was associated with accelerated leaf development early in the experiment in these young and rapidly growing trees (Fig. 1c), such that trees exposed to the warmed treatment had higher rates of crown-scale photosynthesis (Fig. 5a) primarily through a warming effect on total crown leaf area. That is, the warmed treatment got a head start in leaf area production, which compounded over time. This is consistent with nutrient fertilization studies, in which increases in leaf area, rather than changes in leaf N or leaf function, often dominates the growth responses of rapidly growing plants (Sinclair & Horie, 1989; Gastal & Lemaire, 2002; Lovelock *et al.*, 2004; Wang *et al.*, 2012), although there are exceptions (Santiago *et al.*, 2012). This is also consistent with other experimental manipulations, in which a stimulation of leaf area development early in an experiment can strongly affect exponential growth trajectories (Tjoelker *et al.*, 1998; Kirschbaum, 2011; Drake *et al.*, 2017).

Effects of drought on C allocation

We did not detect any significant effects of drought on C partitioning. The drought appeared to reduce all C fluxes proportionally, such that the ratios of C fluxes to GPP were unchanged. We recognize that our ability to resolve C partitioning below-ground was limited by the nature of the measurements based on the residual, and our lack of root biomass measurements through time. The biomass in roots relative to the total at the harvest (root mass ratio; Fig. 3b) did indicate an interactive effect of drought and warming, possibly via small differences in the partitioning of GPP to roots that accumulated over time. Perhaps the ambient temperature trees had sufficient carbohydrate reserves to fuel additional root growth in the drought treatment, whereas warmed temperature trees were consuming more carbohydrates above-ground and were thus unable to increase root growth in

the drought. This speculative process may explain the observed interaction between warming and drought on root mass ratio (Fig. 3b).

Trees acquired H_2O from deep in the soil profile during the drought. Leaf predawn H_2O potential declined to only $c. -0.5$ MPa, which is a moderate value that is not indicative of pronounced H_2O stress. Thus, although we successfully implemented a drought that dried the soils from the surface to 1 m depth, the trees successfully avoided acute physiological drought stress by reducing growth and transpiration rates while also acquiring deeper soil H_2O . Previous studies have shown that groundwater use enables vegetation to mitigate production declines under conditions of surface moisture limitation (Baldocchi *et al.*, 2010; Barbeta *et al.*, 2015), and several eucalypt species are well-known users of groundwater (Mensforth *et al.*, 1994; Pfautsch *et al.*, 2011, 2015; Eamus *et al.*, 2015; Zolfaghar *et al.*, 2017). Furthermore, Koirala *et al.* (2017) demonstrated correlations between GPP and groundwater table depth that were present over $c. 70\%$ of the vegetated surface of the Earth, suggesting that vegetation–groundwater interactions are common and globally relevant. Our study demonstrates that some trees may utilize access to soil H_2O at depth to maintain moderate rates of photosynthetic C uptake and growth during extended droughts that lead to dry surface soils.

Implications for mathematical models

Many ecosystem and Earth system models begin their simulation of ecosystem C cycling by predicting GPP as a function of leaf area and environmental drivers. GPP is then partitioned into component terms, including R_a and the production of leaf, wood, and root mass. Our observations suggest that tree C allocation of GPP to these terms can be influenced by environmental drivers such as temperature. That is, the observations presented here are not consistent with static partitioning schemes with fixed and constant partitioning of GPP into component fluxes. However, when aggregated over longer time periods, our average partitioning coefficients ($c. 30\%$ of GPP to aboveground respiration, 43% of GPP to aboveground growth, and 27% to belowground fluxes) are comparable to values used in some ecosystem models (De Kauwe *et al.*, 2014). The observations presented here are also not consistent with a dynamic C partitioning scheme based on Sprenkel and Leibig's law of the minimum (van der Ploeg *et al.*, 1999), where C would be preferentially allocated to increase the acquisition of the factor most limiting primary production. Furthermore, Aspinwall *et al.* (2016) recently documented strong

seasonal variation in carbohydrate storage in these evergreen trees, characterized by the buildup of high starch concentrations during the winter and a drawdown of these reserves during the summer. It appears that these trees partially utilize a nonstructural carbohydrate storage reserve to fuel growth and metabolism during the hot summer. Based on these observations, we suggest that a dynamic allocation scheme incorporating a dynamic carbohydrate reserve may be appropriate for future investigation (e.g. Fatichi *et al.*, 2014; Pugh *et al.*, 2016).

Conclusions

We used a novel combination of growth and whole-crown flux measurements to study the effects of warming and drought on the C allocation of young *E. tereticornis* trees. These trees did not substantially alter C allocation in response to the drought treatment, as reduced growth, reduced transpiration, and the facultative use of deep soil H₂O allowed the trees to avoid physiological drought stress. Experimental warming increased the proportion of GPP that was allocated to aboveground uses and decreased the proportion of GPP that was allocated below ground. Such a change in tree C allocation may have important implications for tree growth, forest C storage, and soil nutrient cycling in a warmer world. In particular, increased allocation above-ground in a warmer world may stimulate leaf area development and aboveground growth during conditions of sufficient soil resource supply.

Acknowledgements

We thank Burhan Amiji (Western Sydney University) for maintaining the site, for collecting much of the growth and harvest data, and for his excellent research support. This experiment was made possible through a collaboration with Sune Linder and the Swedish University of Agricultural Sciences, who designed, built, and generously provided the WTCs. We also gratefully acknowledge Courtney Company (Colgate University) for his measurements of fine root biomass, as well as Renee Smith and Carrie Drake (Western Sydney University) for their help with the whole-tree harvests. This research was supported by the Australian Research Council (Discovery, DP140103415), a New South Wales government Climate Action Grant (NSW T07/CAG/016), the Hawkesbury Institute for the Environment, and Western Sydney University.

Author contributions

JED co-led the experimental design, contributed to data collection for the chamber flux, and led the data analysis, modeling, and writing. MGT was the senior scientific lead; he co-led the experimental design and made large contributions to analysis, interpretation, and writing. MJA contributed to the experimental design, data collection and interpretation, and writing. PBR contributed to the experimental design, interpretation, and writing. SP contributed to experimental design, data collection and interpretation, and writing. CVMB contributed to the measurements

of chamber flux, and contributed to experimental design, data analysis, and writing.

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References

- Adu-Bredu S, Hagihara A. 2003. Long-term carbon budget of the above-ground parts of a young hinoki cypress (*Chamaecyparis obtusa*) stand. *Ecological Research* 18: 165–175.
- Allen CD, Breshears DD, McDowell NG. 2015. On underestimation of global vulnerability to tree mortality and forest die-off from hotter drought in the Anthropocene. *Ecosphere* 6: 129.
- Amthor JS. 2000. The McCree–de Wit–Penning de Vries–Thornley respiration paradigms: 30 years later. *Annals of Botany* 86: 1–20.
- Aspinwall MJ, Drake JE, Company C, Varhammar A, Ghannoum O, Tissue DT, Reich PB, Tjoelker MG. 2016. Convergent acclimation of leaf photosynthesis and respiration to prevailing ambient temperatures under current and warmer climates in *Eucalyptus tereticornis*. *New Phytologist* 212: 354–367.
- Baldocchi DD, Ma S, Rambal S, Misson L, Ourcival J-M, Limousin J-M, Pereira J, Papale D. 2010. On the differential advantages of evergreenness and deciduousness in Mediterranean oak woodlands: a flux perspective. *Ecological Applications* 20: 1583–1597.
- Barbeta A, Mejia-Chang M, Ogaya R, Voltas J, Dawson TE, Peñuelas J. 2015. The combined effects of a long-term experimental drought and an extreme drought on the use of plant-water sources in a Mediterranean forest. *Global Change Biology* 21: 1213–1225.
- Barton CVM, Duursma RA, Medlyn BE, Ellsworth DS, Eamus D, Tissue DT, Adams MA, Conroy J, Crous KY, Liberloo M *et al.* 2012. Effects of elevated atmospheric [CO₂] on instantaneous transpiration efficiency at leaf and canopy scales in *Eucalyptus saligna*. *Global Change Biology* 18: 585–595.
- Barton CVM, Ellsworth DS, Medlyn BE, Duursma RA, Tissue DT, Adams MA, Eamus D, Conroy JP, McMurtrie RE, Parsby J *et al.* 2010. Whole-tree chambers for elevated atmospheric CO₂ experimentation and tree scale flux measurements in south-eastern Australia: the Hawkesbury Forest Experiment. *Agricultural and Forest Meteorology* 150: 941–951.
- Blessing CH, Werner RA, Siegwolf R, Buchmann N. 2015. Allocation dynamics of recently fixed carbon in beech saplings in response to increased temperatures and drought. *Tree Physiology* 35: 585–598.
- Burke EJ, Brown SJ, Christidis N. 2006. Modeling the recent evolution of global drought and projections for the twenty-first century with the Hadley Centre climate model. *Journal of Hydrometeorology* 7: 1113–1125.
- De Kauwe MG, Medlyn BE, Zaehle S, Walker AP, Dietze MC, Wang Y-P, Luo Y, Jain AK, El-Masri B, Hickler T *et al.* 2014. Where does the carbon go? A model–data intercomparison of vegetation carbon allocation and turnover processes at two temperate forest free-air CO₂ enrichment sites. *New Phytologist* 203: 883–899.
- DeLucia EH, Moore DJ, Norby RJ. 2005. Contrasting responses of forest ecosystems to rising atmospheric CO₂: implications for the global C cycle. *Global Biogeochemical Cycles* 19: GB3006.
- Dietze MC, Sala A, Carbone MS, Czimczik CI, Mantooth JA, Richardson AD, Vargas R. 2014. Nonstructural carbon in woody plants. *Annual Review of Plant Biology* 65: 667–687.
- Doughty CE, Malhi Y, Araujo-Murakami A, Metcalfe DB, Silva-Espejo JE, Arroyo L, Heredia JP, Pardo-Toledo E, Mendizabal LM, Rojas-Landivar VD

- et al.* 2014. Allocation trade-offs dominate the response of tropical forest growth to seasonal and interannual drought. *Ecology* 95: 2192–2201.
- Doughty CE, Metcalfe D, Girardin CA, Farfan Amezcua F, Galiano Cabrera D, Huaraca Huasco W, Silva-Espejo JE, Araujo-Murakami A, da Costa MC, Rocha W *et al.* 2015. Drought impact on forest carbon dynamics and fluxes in Amazonia. *Nature* 519: 78–140.
- Drake JE, Aspinwall MJ, Pfautsch S, Rymer PD, Reich PB, Smith RA, Crous KY, Tissue DT, Ghannoum O, Tjoelker MG. 2015. The capacity to cope with climate warming declines from temperate to tropical latitudes in two widely distributed *Eucalyptus* species. *Global Change Biology* 21: 459–472.
- Drake JE, Gallet-Budynek A, Hofmockel KS, Bernhardt ES, Billings SA, Jackson RB, Johnson KS, Lichter J, McCarthy HR, McCormack ML *et al.* 2011. Increases in the flux of carbon belowground stimulate nitrogen uptake and sustain the long-term enhancement of forest productivity under elevated CO₂. *Ecology Letters* 14: 349–357.
- Drake J, Tjoelker M, Aspinwall MJ. 2016a. https://figshare.com/articles/Drake_NewPhyt_2016_WTC3_RtoGPP_forfigshare_zip/3122104
- Drake_NewPhyt_2016_WTC3_RtoGPP_forfigshare.zip.doi: 10.6084/m9.figshare.3122104.v1
- Drake JE, Tjoelker MG, Aspinwall MJ, Reich PB, Barton CVM, Medlyn BE, Duursma RA. 2016b. Does physiological acclimation to climate warming stabilize the ratio of canopy respiration to photosynthesis? *New Phytologist* 211: 850–863.
- Drake JE, Vårhammar A, Kumarathunge D, Medlyn BE, Pfautsch S, Reich PB, Tissue DT, Ghannoum O, Tjoelker MG. 2017. A common thermal niche among geographically diverse populations of the widely distributed tree species *Eucalyptus tereticornis*: no evidence for adaptation to climate-of-origin. *Global Change Biology* 23: 5069–5082.
- Duursma RA, Barton CVM, Eamus D, Medlyn BE, Ellsworth DS, Forster MA, Tissue DT, Linder S, McMurtrie RE. 2011. Rooting depth explains [CO₂] × drought interaction in *Eucalyptus saligna*. *Tree Physiology* 31: 922–931.
- Duursma RA, Barton CVM, Lin Y-S, Medlyn BE, Eamus D, Tissue DT, Ellsworth DS, McMurtrie RE. 2014. The peaked response of transpiration rate to vapour pressure deficit in field conditions can be explained by the temperature optimum of photosynthesis. *Agricultural and Forest Meteorology* 189: 2–10.
- Duursma RA, Falster DS. 2016. Leaf mass per area, not total leaf area, drives differences in above-ground biomass distribution among woody plant functional types. *New Phytologist* 212: 368–376.
- Eamus D, Zolfaghari S, Villalobos-Vega R, Cleverly J, Huete A. 2015. Groundwater-dependent ecosystems: recent insights from satellite and field-based studies. *Hydrology and Earth System Sciences* 19: 4229–4256.
- Elder B, Buerger J, Breitschneider L, Steinmann H-H, Iselstein J. 2015. Growth responses to elevated temperature and reduced soil moisture during early establishment of three annual weeds in four soil types. *Journal of Plant Diseases and Protection* 122: 39–48.
- Epron D, Bahn M, Derrien D, Lattanzi FA, Pumpanen J, Gessler A, Höglberg P, Maillard P, Dannoura M, Gérard D *et al.* 2012. Pulse-labelling trees to study carbon allocation dynamics: a review of methods, current knowledge and future prospects. *Tree Physiology* 32: 776–798.
- Farooq M, Wahid A, Kobayashi N, Fujita D, Basra SMA. 2009. Plant drought stress: effects, mechanisms and management. *Agronomy for Sustainable Development* 29: 185–212.
- Fatichi S, Leuzinger S, Koerner C. 2014. Moving beyond photosynthesis: from carbon source to sink-driven vegetation modeling. *New Phytologist* 201: 1086–1095.
- Feng L, de Reffye P, Dreyfus P, Auclair D. 2012. Connecting an architectural plant model to a forest stand dynamics model – application to Austrian black pine stand visualization. *Annals of Forest Science* 69: 245–255.
- Finzi AC, Abramoff RZ, Spiller KS, Brzostek ER, Darby BA, Kramer MA, Phillips RP. 2015. Rhizosphere processes are quantitatively important components of terrestrial carbon and nutrient cycles. *Global Change Biology* 21: 2082–2094.
- Franklin O, Johansson J, Dewar RC, Dieckmann U, McMurtrie RE, Brännström Å, Dybzinski R. 2012. Modeling carbon allocation in trees: a search for principles. *Tree Physiology* 32: 648–666.
- Friedlingstein P, Joel G, Field CB, Fung IY. 1999. Toward an allocation scheme for global terrestrial carbon models. *Global Change Biology* 5: 755–770.
- Furze ME, Trumbore S, Hartmann H. 2018. Detours on the phloem sugar highway: stem carbon storage and remobilization. *Current Opinion in Plant Biology* 43: 89–95.
- Gastal F, Lemaire G. 2002. N uptake and distribution in crops: an agronomical and ecophysiological perspective. *Journal of Experimental Botany* 53: 789–799.
- Gower ST, Krankina O, Olson RJ, Apps M, Linder S, Wang C. 2001. Net primary production and carbon allocation patterns of boreal ecosystems. *Ecological Applications* 11: 1395–1411.
- Hartmann H, McDowell NG, Trumbore S. 2015. Allocation to carbon storage pools in Norway spruce saplings under drought and low CO₂. *Tree Physiology* 35: 243–252.
- Höglberg P, Nordgren A, Buchmann N, Taylor AF, Ekblad A, Höglberg MN, Nyberg G, Ottosson-Löfvenius M, Read DJ. 2001. Large-scale forest girdling shows that current photosynthesis drives soil respiration. *Nature* 411: 789–792.
- Hommel R, Siegwolf R, Zavadlav S, Arend M, Schaub M, Galiano L, Haeni M, Kayler ZE, Gessler A. 2016. Impact of interspecific competition and drought on the allocation of new assimilates in trees. *Plant Biology* 18: 785–796.
- Jump AS, Ruiz-Benito P, Greenwood S, Allen CD, Kitzberger T, Fensham R, Martínez-Vilalta J, Lloret F. 2017. Structural overshoot of tree growth with climate variability and the global spectrum of drought-induced forest dieback. *Global Change Biology* 23: 3742–3757.
- Kirschbaum MUF. 2011. Does enhanced photosynthesis enhance growth? Lessons learned from CO₂ enrichment studies. *Plant Physiology* 155: 117–124.
- Koirala S, Jung M, Reichstein M, de Graaf IEM, Camps-Valls G, Ichii K, Papale D, Ráduly B, Schwalm CR, Tramontana G *et al.* 2017. Global distribution of groundwater–vegetation spatial covariation. *Geophysical Research Letters* 44: 2017GL072885.
- Kuster TM, Arend M, Bleuler P, Günthardt-Goerg MS, Schulm R. 2013. Water regime and growth of young oak stands subjected to air-warming and drought on two different forest soils in a model ecosystem experiment. *Plant Biology* 15: 138–147.
- Landsberg JJ, Waring RH. 1997. A generalised model of forest productivity using simplified concepts of radiation-use efficiency, carbon balance and partitioning. *Forest Ecology and Management* 95: 209–228.
- Lemoine R, La Camera S, Atanassova R, Dedaldechamp F, Allario T, Pourtau N, Bonnemain J-L, Laloi M, Coutos-Thevenot P, Maurousset L *et al.* 2013. Source-to-sink transport of sugar and regulation by environmental factors. *Frontiers in Plant Science* 4: e272.
- Leon-Sanchez L, Nicolas E, Nortes PA, Maestre FT, Querejeta JJ. 2016. Photosynthesis and growth reduction with warming are driven by nonstomatal limitations in a Mediterranean semi-arid shrub. *Ecology and Evolution* 6: 2725–2738.
- Litton CM, Raich JW, Ryan MG. 2007. Carbon allocation in forest ecosystems. *Global Change Biology* 13: 2089–2109.
- Lovelock CE, Feller IC, McKee KL, Engelbrecht BMJ, Ball MC. 2004. The effect of nutrient enrichment on growth, photosynthesis and hydraulic conductance of dwarf mangroves in Panama. *Functional Ecology* 18: 25–33.
- Lu M, Zhou X, Yang Q, Li H, Luo Y, Fang C, Chen J, Yang X, Li B. 2013. Responses of ecosystem carbon cycle to experimental warming: a meta-analysis. *Ecology* 94: 726–738.
- Mäkelä A, Valentine HT, Helmisaari H-S. 2008. Optimal co-allocation of carbon and nitrogen in a forest stand at steady state. *New Phytologist* 180: 114–123.
- Martínez-Vilalta J, Cochard H, Mencuccini M, Sterck F, Herrero A, Korhonen JFJ, Llorens P, Nikinmaa E, Nolé A, Poyatos R *et al.* 2009. Hydraulic adjustment of Scots pine across Europe. *New Phytologist* 184: 353–364.
- McCree KJ. 1970. An equation for the rate of respiration of white clover grown under controlled conditions. *Prediction and measurement of photosynthetic productivity. Proceedings of the IBP/PP Technical Meeting, Trebon*. PUDOC, Wageningen, the Netherlands, 221–229.
- McMurtrie RE, Dewar RC. 2013. New insights into carbon allocation by trees from the hypothesis that annual wood production is maximized. *New Phytologist* 199: 981–990.
- Melillo JM, Butler S, Johnson J, Mohan J, Steudler P, Lux H, Burrows E, Bowles F, Smith R, Scott L *et al.* 2011. Soil warming, carbon–nitrogen

- interactions, and forest carbon budgets. *Proceedings of the National Academy of Sciences, USA* 108: 9508–9512.
- Melillo JM, Steudler PA, Aber JD, Newkirk K, Lux H, Bowles FP, Catricala C, Magill A, Ahrens T, Morrisseau S. 2002. Soil warming and carbon-cycle feedbacks to the climate system. *Science* 298: 2173–2176.
- Mencuccini M. 2003. The ecological significance of long-distance water transport: short-term regulation, long-term acclimation and the hydraulic costs of stature across plant life forms. *Plant, Cell & Environment* 26: 163–182.
- Mensforth LJ, Thorburn PJ, Tyerman SD, Walker GR. 1994. Sources of water used by riparian *Eucalyptus camaldulensis* overlying highly saline groundwater. *Oecologia* 100: 21–28.
- Mokany K, Raison RJ, Prokushkin AS. 2006. Critical analysis of root : shoot ratios in terrestrial biomes. *Global Change Biology* 12: 84–96.
- Munir TM, Perkins M, Kaing E, Strack M. 2015. Carbon dioxide flux and net primary production of a boreal treed bog: responses to warming and water-table-lowering simulations of climate change. *Biogeosciences* 12: 1091–1111.
- Nemani RR, Keeling CD, Hashimoto H, Jolly WM, Piper SC, Tucker CJ, Myrneni RB, Running SW. 2003. Climate-driven increases in global terrestrial net primary production from 1982 to 1999. *Science* 300: 1560–1563.
- Palmroth S, Oren R, McCarthy HR, Johnsen KH, Finzi AC, Butnor JR, Ryan MG, Schlesinger WH. 2006. Aboveground sink strength in forests controls the allocation of carbon below ground and its [CO₂]-induced enhancement. *Proceedings of the National Academy of Sciences, USA* 103: 19362–19367.
- Pfautsch S, Dodson W, Madden S, Adams MA. 2015. Assessing the impact of large-scale water table modifications on riparian trees: a case study from Australia. *Ecohydrology* 8: 642–651.
- Pfautsch S, Keitel C, Turnbull TL, Braimbridge MJ, Wright TE, Simpson RR, O'Brien JA, Adams MA. 2011. Diurnal patterns of water use in *Eucalyptus vitrix* indicate pronounced desiccation–rehydration cycles despite unlimited water supply. *Tree Physiology* 31: 1041–1051.
- Pinheiro J, Bates D, DebRoy S, Sarkar D, R-Development Core Team. 2013. NLME: linear and nonlinear mixed effects models. R package v.3.1-108.
- van der Ploeg RR, Böhm W, Kirkham MB. 1999. On the origin of the theory of mineral nutrition of plants and the law of the minimum. *Soil Science Society of America Journal* 63: 1055–1062.
- Poorter H, Jagodzinski AM, Ruiz-Peinado R, Kuyah S, Luo Y, Oleksyn J, Usoltsev VA, Buckley TN, Reich PB, Sack L. 2015. How does biomass distribution change with size and differ among species? An analysis for 1200 plant species from five continents. *New Phytologist* 208: 736–749.
- Poorter H, Niklas KJ, Reich PB, Oleksyn J, Poot P, Mommer L. 2012. Biomass allocation to leaves, stems and roots: meta-analyses of interspecific variation and environmental control. *New Phytologist* 193: 30–50.
- Poorter H, Sack L. 2012. Pitfalls and possibilities in the analysis of biomass allocation patterns in plants. *Frontiers in Plant Science* 3: 259.
- Pugh TAM, Mueller C, Arneith A, Haverd V, Smith B. 2016. Key knowledge and data gaps in modelling the influence of CO₂ concentration on the terrestrial carbon sink. *Journal of Plant Physiology* 203: 3–15.
- R Core Team. 2012. *R: a language and environment for statistical computing*. Version R 3.2.2. Vienna, Austria: R Foundation for Statistical Computing.
- Reich PB. 2002. Root–shoot relations: optimality in acclimation and adaptation or the ‘Emperor’s New Clothes’. In: Waisel Y, Eshel A, Kafkafi U, eds. *Plant roots: the hidden half*. New York, NY, USA: Marcel Dekker, 205–220.
- Reich PB, Luo Y, Bradford JB, Poorter H, Perry CH, Oleksyn J. 2014. Temperature drives global patterns in forest biomass distribution in leaves, stems, and roots. *Proceedings of the National Academy of Sciences, USA* 111: 13721–13726.
- Reichstein M, Falge E, Baldocchi D, Papale D, Aubinet M, Berbigier P, Bernhofer C, Buchmann N, Gilmanov T, Granier A *et al.* 2005. On the separation of net ecosystem exchange into assimilation and ecosystem respiration: review and improved algorithm. *Global Change Biology* 11: 1424–1439.
- Roux XL, Lacomte A, Escobar-Gutiérrez A, Dizès SL. 2001. Carbon-based models of individual tree growth: a critical appraisal. *Annals of Forest Science* 58: 469–506.
- Running SW, Gower ST. 1991. FOREST-BGC, a general model of forest ecosystem processes for regional applications. II. Dynamic carbon allocation and nitrogen budgets. *Tree Physiology* 9: 147–160.
- Rustad L, Campbell J, Marion G, Norby R, Mitchell M, Hartley A, Cornelissen J, Gurevitch J, GCCTE-NEWS. 2001. A meta-analysis of the response of soil respiration, net nitrogen mineralization, and aboveground plant growth to experimental ecosystem warming. *Oecologia* 126: 543–562.
- Santiago LS, Wright SJ, Harms KE, Yavitt JB, Korine C, Garcia MN, Turner BL. 2012. Tropical tree seedling growth responses to nitrogen, phosphorus and potassium addition. *Journal of Ecology* 100: 309–316.
- Sillmann J, Kharin VV, Zwiers FW, Zhang X, Bronaugh D. 2013. Climate extremes indices in the CMIP5 multimodel ensemble: part 2. Future climate projections. *Journal of Geophysical Research: Atmospheres* 118: 2473–2493.
- Sinclair TR, Horie T. 1989. Leaf nitrogen, photosynthesis, and crop radiation use efficiency: a review. *Crop Science* 29: 90.
- Strömberg M, Linder S. 2002. Effects of nutrition and soil warming on stemwood production in a boreal Norway spruce stand. *Global Change Biology* 8: 1194–1204.
- Taeger S, Sparks TH, Menzel A. 2015. Effects of temperature and drought manipulations on seedlings of Scots pine provenances. *Plant Biology* 17: 361–372.
- Thomas DS, Montagu KD, Conroy JP. 2007. Temperature effects on wood anatomy, wood density, photosynthesis and biomass partitioning of *Eucalyptus grandis* seedlings. *Tree Physiology* 27: 251–260.
- Tjoelker MG, Oleksyn J, Reich PB. 1998. Temperature and ontogeny mediate growth response to elevated CO₂ in seedlings of five boreal tree species. *New Phytologist* 140: 197–210.
- Tjoelker MG, Oleksyn J, Reich PB. 1999. Acclimation of respiration to temperature and CO₂ in seedlings of boreal tree species in relation to plant size and relative growth rate. *Global Change Biology* 5: 679–691.
- Trugman AT, Detto M, Bartlett MK, Medvigy D, Anderegg WRL, Schwalm C, Schaffer B, Pacala SW. 2018. Tree carbon allocation explains forest drought-kill and recovery patterns. *Ecology Letters* 21: 1552–1560.
- Volder A, Briske DD, Tjoelker MG. 2013. Climate warming and precipitation redistribution modify tree–grass interactions and tree species establishment in a warm-temperate savanna. *Global Change Biology* 19: 843–857.
- Wang D, Maughan MW, Sun J, Feng X, Miguez F, Lee D, Dietze MC. 2012. Impact of nitrogen allocation on growth and photosynthesis of miscanthus (*Miscanthus × giganteus*). *Global Change Biology Bioenergy* 4: 688–697.
- Way DA, Oren R. 2010. Differential responses to changes in growth temperature between trees from different functional groups and biomes: a review and synthesis of data. *Tree Physiology* 30: 669–688.
- Yang J, Medlyn BE, Kauwe MGD, Duursma RA. 2018. Applying the concept of ecohydrological equilibrium to predict steady state leaf area index. *Journal of Advances in Modeling Earth Systems* 10: 1740–1758.
- Zolfaghar S, Villalobos-Vega R, Zeppel M, Cleverly J, Rumman R, Hingee M, Boulain N, Li Z, Eamus D. 2017. Transpiration of *Eucalyptus* woodlands across a natural gradient of depth-to-groundwater. *Tree Physiology* 37: 961–975.

Supporting Information

Additional Supporting Information may be found online in the Supporting Information section at the end of the article:

Fig. S1 The change in volumetric water content (VWC) throughout the soil profile.

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