

# The partitioning of gross primary production for Eucalyptus tereticornis trees exposed to experimental warming and drought

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- 1 The partitioning of gross primary production for *Eucalyptus tereticornis* trees
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Policy.

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# Summary

- The allocation of C is an important component of tree physiology that influences growth and ecosystem C storage. Allocation is challenging to measure and the influences of environmental changes such as warming and drought are uncertain.
- We exposed *Eucalyptus tereticornis* trees to a factorial combination of +3°C warming and an extreme summer drought in the field using whole-tree chambers. We calculated C allocation terms using detailed measurements of growth and continuous whole-crown CO<sub>2</sub> and H<sub>2</sub>O exchange measurements.
- Warming accelerated growth and leaf area development, and increased the partitioning of Gross Primary Production (GPP) to aboveground respiration and growth, while decreasing partitioning belowground. The summer drought reduced C gain and growth, but did not impact GPP partitioning. Trees utilized deep soil water and avoided strongly negative water potentials during the drought.
- Warming increased growth respiration, but maintenance respiration acclimated homeostatically. The increasing growth rates of trees in the warmed treatment resulted in higher rates of respiration, even with complete acclimation of maintenance respiration.
- Warming-induced stimulations of tree growth likely involve increased C allocation aboveground, particularly to leaf area development, while reduced water availability may not stimulate allocation to roots.

Policy.

## Introduction

The C economy of trees and forests depends not only on the amount of C fixed via photosynthesis, but how that fixed C is used. Ecosystem C storage is affected by the allocation of C to long-lived C pools such as wood relative to C allocation to pools with higher turnover rates such as fine roots (DeLucia *et al.*, 2005). C allocation also affects the acquisition of light, nutrients, and water, which influences ecosystem C cycling (Litton *et al.*, 2007; Epron *et al.*, 2012; Franklin *et al.*, 2012; De Kauwe *et al.*, 2014). The allocation of C belowground affects soil C and nutrient cycling (Högberg *et al.*, 2001; Epron *et al.*, 2012) in part because belowground C allocation can affect soil organic matter decomposition and the acquisition of limiting nutrients by trees (Drake *et al.*, 2011; Finzi *et al.*, 2015). The importance of C allocation and the relative difficulty of its study contribute to its role as an important unknown for modeling the biogeochemistry of ecosystems (Roux *et al.*, 2001; Franklin *et al.*, 2012; Dietze *et al.*, 2014; De Kauwe *et al.*, 2014).

Terminology regarding allocation has been a source of some confusion. Here, we follow Litton *et al.* (2007) and use 'allocation' as a term of broad definition encompassing three specific aspects of study: (1) *ratios* of biomass pool sizes, (2) *fluxes* of C to a given component, and (3) *partitioning*, the C flux to a given component as a fraction of GPP. These areas of study are similar, but not equivalent. For example, old and large trees have a large wood mass fraction relative to small young trees (Poorter *et al.*, 2015), but this reflects the low turnover of woody tissues relative to leaves, not a higher partitioning of GPP to wood in old trees (Duursma & Falster, 2016). Thus it is often inappropriate to infer C partitioning from biomass ratios (Reich, 2002; Litton *et al.*, 2007). Surprisingly, partitioning of photosynthate is relatively poorly understood despite its direct relevance to ecosystem models (Epron *et al.*, 2012; Franklin *et al.*, 2012; De Kauwe *et al.*, 2014).

Several schemes have been used to conceptualize and model C allocation. The simplest approach is to assume that trees partition a constant fraction of fixed C to each use (e.g., growth, respiration). This is supported by linear relationships between production terms in some systems (Gower *et al.*, 2001). However, fixed allocation schemes cannot capture ontogenetic effects (Poorter *et al.*, 2015; Duursma & Falster, 2016) or dynamic temporal responses (De Kauwe *et al.*, 2014; Doughty *et al.*, 2014). Another approach is to assume a functional balance between tree organs via allometric relationships, Huber values, or root to leaf mass fractions (Landsberg & Waring, 1997; Mäkelä *et al.*, 2008; Feng *et al.*, 2012). Finally, there is the concept that trees increase C partitioning towards the acquisition of the primary limiting resource (McMurtrie & Dewar, 2013). This approach appears sensible and has been implemented in several models (e.g., Running & Gower, 1991; Friedlingstein *et al.*, 1999), but direct evidence supporting

this concept is scarce, given the challenges involved in measuring allocation (Poorter & Sack, 2012; Poorter *et al.*, 2015). However, optimization approaches have been used to constrain dynamic allocation schemes with some success (Franklin *et al.*, 2012; McMurtrie & Dewar, 2013).

Temperature is a fundamental aspect of climate that affects many aspects of tree physiology (Way & Oren, 2010; Lu et al., 2013). Several lines of evidence suggest that warming may increase tree C allocation aboveground at the expense of C allocation belowground. Experimental warming of forest soil increased aboveground biomass while reducing or not affecting belowground biomass (Strömgren & Linder, 2002; Melillo et al., 2011), which has been attributed to a warming-induced increase in soil N availability (Melillo et al., 2002). A recent <sup>13</sup>C-CO<sub>2</sub> labeling study indicated that warming increased allocation aboveground and reduced C allocation belowground in beech saplings via a direct effect on tree physiology, without an altered soil N cycle (Blessing et al., 2015). Also, environmental gradients in mean annual temperature are strongly correlated with the distribution of biomass; forests have a lower root mass fraction (i.e., the proportion of live forest biomass contained in roots) in warm climates than in cold climates (Reich et al., 2014). Meta-analyses of warming experiments generally found an increase in aboveground plant growth (Rustad et al., 2001) that is slightly larger than the increase in belowground plant growth (Lu et al., 2013), although such experiments have exclusively involved small stature vegetation given the logistical challenges of warming tall forests. Thus, prior research suggests a shift in C allocation aboveground with experimental warming, although direct tests in the field with large trees have not yet been performed.

Water availability also impacts tree growth and physiology (e.g., Mencuccini, 2003; Nemani *et al.*, 2003; Farooq *et al.*, 2009) and the effects of drought are of particular concern (Burke *et al.*, 2006; Sillmann *et al.*, 2013). While it appears sensible that trees would increase C allocation to roots in dry regions or during drought periods (Poorter *et al.*, 2012), there is limited support for this idea. Reich *et al.* (2014) found no correlation between root mass fraction and aridity across a global dataset of >6,200 forests. Additionally, Amazonian forests responded to droughts in 2005 and 2010 with a shift away from fine-root growth and increased C partitioning to aboveground growth and respiration, particularly in the year following the drought (Doughty *et al.*, 2014, 2015). However, drought has been observed to increase root mass fractions for small plants grown in artificial conditions (Reich, 2002; Poorter *et al.*, 2012), and increased C allocation belowground under drought is consistent with some <sup>13</sup>C-CO<sub>2</sub> labeling studies (Hommel *et al.*, 2016), but not others (Hartmann *et al.*, 2015). While the simple expectation of increased allocation to roots during drought is appealing, C allocation responses to drought are likely more complex and merit further study.

Interactions between temperature and drought effects may also be important for tree C allocation. Warmer temperatures may exacerbate tree H<sub>2</sub>O loss during drought and increase mortality risk (Allen *et al.*, 2015). If warmer temperatures reduce C allocation belowground, then the ability of trees to acquire soil water may also be impaired. However, an open top chamber experiment with young oak saplings found no interaction between experimental warming and drought on tree transpiration or biomass (Kuster *et al.*, 2013). Other studies have found limited evidence for interactive effects on plant growth rates (e.g., Edler *et al.*, 2015; Taeger *et al.*, 2015), although there are also exceptions (Munir *et al.*, 2015; Leon-Sanchez *et al.*, 2016). A six-year warming and precipitation redistribution experiment with two tree species found complex growth responses (Volder *et al.*, 2013) with a strongly interactive effect on the relative growth rate of *Quercus stellata* monocultures. Thus, it is challenging to generalize how the interactive effects of drought and warming affect tree physiology and growth.

To address these knowledge gaps concerning, temperature, water availability, and C allocation, we studied C allocation in 8 to 9-meter tall eucalypt trees growing in an experiment that manipulated both temperature and moisture supply. We used whole-tree chambers in the field in southeastern Australia to grow *Eucalyptus tereticornis* trees under experimental warming of +3 °C for more than one year, crossed with a summer drought for three months. We continuously measured whole-crown CO<sub>2</sub> and H<sub>2</sub>O exchange and measured aboveground biomass production every 2 weeks. From these intensive measurements, we derived GPP, aboveground net primary production (NPP<sub>a</sub>), aboveground autotrophic respiration (R<sub>a</sub>) and the residual C that must have been partitioned belowground for each fortnightly interval. We use these data to test the predictions that warming decreases C partitioning belowground, while drought increases C partitioning belowground.

### **Materials and Methods**

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131 Site and experiment 132 We implemented a warming and drought experiment using 12 whole-tree chambers (WTCs) in 133 Richmond, New South Wales (Australia; 33°36′40″S, 150°44′26.5″E). The WTCs were large cylindrical 134 structures topped with a cone (3.25 m in diameter, 9 m in height, volume of ~53 m<sup>3</sup>) that enclosed a single 135 tree rooted in soil. The WTCs controlled atmospheric CO<sub>2</sub> concentration, air temperature (T<sub>air</sub>), relative 136 humidity (RH), and irrigation while continuously measuring the net exchange of CO<sub>2</sub> and H<sub>2</sub>O between 137 entire tree crowns and the atmosphere (Barton et al., 2010; Duursma et al., 2011; Barton et al., 2012; 138 Duursma et al., 2014; Drake et al., 2016b; Aspinwall et al., 2016). 139 The roots of each tree were compartmentalized with a barrier extending vertically belowground to 140 100-cm-depth. A cemented layer of manganese nodules and clay was present at 90–100 cm depth, 141 providing a natural horizontal barrier at the bottom of the rooting volume. Thus, the rooting volume of 142 each tree was isolated from surrounding trees. However, some trees extended roots through this layer and 143 acquired deep soil water in a previous experiment (Duursma et al., 2011). Soil was collected from an 144 adjacent paddock and placed in the chambers in two layers (0–25 cm and from 25 cm to the hard layer) on 145 10 July 2012. Soils at the site were an alluvial formation of low-fertility sandy loam (Clarendon sand). 146 Nursery seedlings of a local provenance of *Eucalyptus tereticornis* Sm. were established in 25 L 147 pots inside the WTCs using the same soil. Eucalyptus tereticornis was chosen because it is a widespread 148 and abundant tree across eastern Australia (Drake et al., 2015). Six potted trees were placed in each 149 chamber on 5 December 2012; a single tree was selected based on size similarity within each treatment 150 and planted in the chamber center on 12 March 2013. Trees assigned to the ambient and warmed 151 temperature treatments had equivalent height and basal diameter when potted seedlings were placed into 152 the WTCs in December 2012 (heights of  $41.5 \pm SE$  of 0.8 and  $40.2 \pm 1.8$  cm; diameters of  $2.4 \pm 0.1$  and 153  $2.5 \pm 0.1$  mm in ambient and warmed treatment, respectively). 154 Six chambers tracked ambient  $T_{air}$  and six chambers tracked ambient  $T_{air} + 3^{\circ}C$  warming (n = 6; 155 'ambient' and 'warmed'); treatments started on 12 December 2012 (Drake et al. 2016b; Aspinwall et al. 156 2016). The average warming achieved was +2.9 °C (± sd of 0.3 across 466 days) for T<sub>air</sub>, +2.2 °C (± sd of 157 0.4) for soil temperature at 5-cm-depth, +2.0 °C (± sd of 0.4) for soil temperature at 20-cm-depth, and 158 +1.4 °C (± sd of 0.2) for soil temperature at 50-cm-depth. Trees were irrigated equally every 15 d with 159 half the mean monthly rainfall. A water exclusion treatment was applied to half of the trees on 12 160 February 2014, resulting in a 2x2 factorial design between the experimental treatments of warming and drought (n = 3; abbreviated A-Wet, A-Dry, W-Wet, and W-Dry hereafter). Trees assigned to the drought 161

treatment received no irrigation from 12 February 2014 through 5 May 2014, representing an extreme
 summer drought of nearly three months.
 Plant water status and soil water content
 Predawn leaf water potentials (Ψ<sub>L-PD</sub>) were measured monthly prior to the drought and every one to two

weeks during the drought treatment. Three leaves were measured per tree on each date using a Scholander-type pressure chamber (1505D-EXP; PMS Instrument Company, OR, USA). Leaves were placed in sealed and humidified plastic bags, placed in a dark cool box, and measured within one hour of collection in a nearby laboratory.

Soil volumetric water content was measured by three sensors in each chamber (CS650 time-domain reflectometers; Campbell Scientific, Logan, UT, USA). Sensors were installed horizontally at three depths: in the surface soil (10-cm-depth), at 30-cm-depth, and just above the hard layer of cemented manganese (~100-cm-depth). Soil temperature was measured with thermocouples at 5, 20, and 50-cm within the center of each chamber.

A single neutron probe per chamber (503DR, Hydroprobe, Instrotek, NC, USA) was used to measure soil water content to a depth of 425 cm (at 25 or 50 cm steps) approximately every two weeks (Duursma *et al.*, 2011). Note that high neutron probe counts in deep soil (150-400 cm depth) partially reflect a change in soil texture towards a higher clay content.

Whole tree crown flux measurements

An automated system measured the net exchange of  $CO_2$  and  $H_2O$  between each crown and its chamber airspace (Barton *et al.*, 2010). Measurements began on 13 September 2013 when suspended plastic floors were sealed around the stem of each tree at ~45 cm height, when the trees were ~3 m tall. Flux measurements finished on 26 May 2014, when the trees were nearly 9 m tall. We report >70,000 hourly flux observations aggregated into >3000 daily sums across 12 trees.

We partitioned the net  $CO_2$  fluxes into the components of GPP and  $R_a$  using an analytical technique common to eddy-covariance research (Reichstein *et al.*, 2005); see Drake *et al.* (2016b) for a complete description. We used direct measurements of whole-crown  $R_a$  and its temperature dependence at night to predict  $R_a$  for each hourly measurement as a function of  $T_{air}$ . For daylight hours, we then calculated GPP as the sum of the measured net  $CO_2$  flux and the predicted  $R_a$  given the measured  $T_{air}$ . We assumed GPP was zero when PPFD = 0; in such conditions, the measured net C flux was used as the measure of  $R_a$ . Note that the chamber airspaces were continuously well-mixed and  $R_a$  fluxes were directly measured at night, avoiding some of the issues inherent in eddy covariance partitioning. The underlying flux data and the partitioning approach were published previously (Drake *et al.*, 2016ab).

Final harvest

The dry mass of all trees was measured destructively at the end of the experiment (26 May 2014). Total tree dry mass was measured as the sum of five components: leaves, branches, stem, coarse roots, and fine roots.

The crown of each tree was divided into three equal heights. All branches were cut flush to the stem and all leaves were separated from branches. A random subsample of 100 leaves per layer was measured for total leaf area (LI-3100C leaf area meter, LiCor, Lincoln, NE, USA), dry mass, and specific leaf area (SLA). The stem was cut into three segments and 1-cm-thick cross-sections (cookies) were sampled for bark depth, wood density, and bark density at the stem base, between the first and second layers, and between the second and third crown layers. Bark and wood density was measured on cookie subsamples (Thomas *et al.*, 2007). Wood and bark densities were similar (0.44 and 0.37 g cm<sup>3</sup> for wood and bark, respectively). Bark depth increased with stem diameter (log<sub>10</sub>(bark depth, mm) = -1.48 + 1.23 × log<sub>10</sub>(diameter, cm), P < 0.001,  $r^2 = 0.92$ ) while wood and bark density decreased with stem diameter (wood density = 0.50 – 0.001 × diameter, P = 0.007, P = 0.17; bark density = 0.45 – 0.001 × diameter, P = 0.001, P = 0.48; densities in g cm<sup>3</sup>, diameter in cm). Warming and drought treatments did not alter these relationships (ANCOVA, P > 0.05). Total stem, branch, and leaf mass were measured directly after drying at 70 °C; some samples required >2 weeks of drying to reach a stable dry mass.

Fine roots were measured using soil cores. The soil surface area was divided into four equal quadrants and two 50-mm-diameter cores were taken within each quadrat on 29 May 2014, just after the crown harvest. Cores were separated into two depths: (1) 0-25 cm and (2) from 25 cm to the hard layer, which varied from 70 to 100 cm depth. Samples within each quadrat and depth category were composited (eight samples per chamber). Fine roots were isolated by washing samples through 2-mm and then 1-mm brass sieves; fine roots were defined as all roots < 2-mm-diameter. Fine root dry mass was measured after drying at 70 °C. Total fine root dry mass was calculated as the product of fine root density (g m<sup>-3</sup>) and soil volume (m<sup>3</sup>) in each layer.

Coarse roots were destructively harvested by fully excavating the soil volume of each chamber. Soil was shoveled out of the chamber onto a conveyor belt that transported the soil to a series of 5-mm steel sieves. Roots were collected by hand, washed, sorted into two size categories (2-10 mm, > 10 mm diameter), and weighed after drying at 70 °C.

Growth measurements

Aboveground biomass was estimated every two weeks for each tree as the sum of leaf, branch, wood, and bark mass; aboveground net primary production (NPP<sub>a</sub>) was estimated as the fortnightly difference in aboveground biomass plus fortnightly litterfall, assuming a constant biomass C fraction of 0.5 of dry mass.

Tree height and stem volume were measured fortnightly; diameter was measured at 30-cm-intervals along each tree stem from a basal height of 15-cm (prior to floor installation) or 65-cm (after floor installation) to the tree apex. The volume of stem wood and bark was estimated for each stem segment as the frustum of a cone, corrected for bark depth (*see above*). Wood and bark mass were calculated as the product of volume and density.

An allometric relationship was developed to predict branch wood mass from branch diameter based on destructively sampled branches on 13 May 2014 and 22 May 2014 ( $\log_{10}(\text{branch mass}) = -1.299 + 2.722 \times \log_{10}(\text{branch diameter})$ , P < 0.001,  $r^2 = 0.91$ , branch mass in g, branch diameter in mm, n = 48 branches). This allometry did not differ between treatments (ANCOVA, P > 0.1) and was used to predict total branch mass on three dates when the diameter of all branches was measured (24 Oct 2013, 15 Jan 2014, and 22 May 2014). Total branch mass and stem volume were strongly correlated in a chamber specific manner (log-log ANCOVA, P < 0.001,  $r^2 = 0.95$ ), which was used to estimate branch mass as a function of stem volume.

Standing leaf area and leaf mass production were estimated as previously at this site (Barton *et al.*, 2012; Drake *et al.*, 2016b). Standing leaf area was measured for each tree by counting all the leaves and multiplying by a tree-specific mean leaf size measured across the crown of each tree with a handheld leaf area meter (LI-3000; n = 86 to 102 leaves per tree). These measurements were performed prior to chamber floor installation (9 Sept 2013) and at the beginning of the drought treatment (10 Feb 2014). A third direct measurement of standing leaf area was calculated from the final harvest data (26 May 2014) by multiplying total crown leaf dry mass by SLA weighted by the leaf dry mass in each layer. Litterfall was collected, dried, and weighed fortnightly for each tree, although relatively few leaves fell as litter (~5% of the total leaf mass). Total tree leaf mass was estimated for each set of fortnightly size measurements by dividing leaf area by the crown-weighted SLA measured at harvest.

*Calculating C partitioning* 

A major goal of this study was to calculate the partitioning of photosynthetically fixed C into components for each fortnightly interval. We quantified GPP, NPP<sub>a</sub>, and R<sub>a</sub> separately, as described above. We calculated the residual between GPP and the sum of NPP<sub>a</sub> and R<sub>a</sub>:

$$GPP = NPP_a + R_a + residual$$
 (eq. 1)

The residual term is a mass-balance calculation of all C put belowground to root production, respiration, and exudation, but this term is also affected by measurement error in GPP, NPP<sub>a</sub>, and R<sub>a</sub>. We calculated the partitioning of GPP directly for each fortnightly interval as NPP<sub>a</sub>/GPP, R<sub>a</sub>/GPP, and residual/GPP.

Growth and maintenance  $R_a$ 

Given the evidence for thermal acclimation of tissue-specific respiration rates to experimental warming in this experiment (Drake et~al., 2016b; Aspinwall et~al., 2016), we investigated growth and maintenance respiration as drivers of  $R_a$  (McCree, 1970; Tjoelker et~al., 1999; Amthor, 2000; Adu-Bredu & Hagihara, 2003). We evaluated the relationship between  $R_a$  per unit tree C and relative growth rate (RGR); the slope reflects the growth component of  $R_a$ , while the y-intercept reflects the maintenance component of  $R_a$ . If  $R_a$  does not acclimate to warming, we expect the warmed treatment to have a higher y-intercept than the ambient treatment. If  $R_a$  acclimates homeostatically, we expect the ambient and warmed treatments to have equivalent intercepts. We also directly estimated coefficients associated with growth and maintenance components of  $R_a$  (Amthor, 2000);

$$R_a = R_a + R_m = g_r G + m_r W (eq. 2)$$

'NLME' R package with a random tree effect.

- where  $R_g$  is the growth respiration rate (gC d<sup>-1</sup>),  $R_m$  is the maintenance respiration rate (gC d<sup>-1</sup>), G is biomass growth (gC d<sup>-1</sup>), W is the standing biomass weight (gC),  $g_r$  is the growth respiration coefficient (gC respired per gC growth), and  $m_r$  is the maintenance respiration coefficient (gC respired per gC standing biomass d<sup>-1</sup>).
- 274 Data analysis

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275 Data were analyzed following a completely randomized design with the single treatment of warming (n = 276 6 for 6 months, then n = 3 for the drought period). Longitudinal analyses were performed using the 'lme' 277 function within the 'NLME' R package with a random tree effect and fixed effects of date, temperature 278 treatment, and water treatment. Treatment means were estimated after adjustment for other terms in the 279 model (i.e. least square means, or LS means) with the 'LSMEANS' package in R v.3.2.2 (R Development 280 Core Team, 2012; Pinheiro et al., 2013). Analyses were evaluated to test assumptions of residual 281 normality and homoscedasticity; transformations were often necessary. Datasets that were not 282 longitudinal were analyzed as a 2x2 ANOVA using the 'lm' function in R. Equation 2 was fit using the

## Results

285 Growth

Experimental warming increased the rates of diameter and height growth (Fig. 1a,b), particularly during the Austral winter and spring. Trees in the warmed treatment were larger than trees in the ambient treatment when the  $CO_2$  and  $H_2O$  flux measurements began (13 Sept 2013; vertical dashed line in Fig. 1). On that date, warming had increased diameter by 21% (P < 0.01; Fig. 1a), height by 19% (P < 0.01; Fig. 1b), total leaf area by 53% (P < 0.01; Fig 1c), and stem volume by 79% (P < 0.01; Fig 1d). During the warm summer, the diameter and height of the ambient and warmed treatment trees converged (Fig. 1a,b), but total stem volume continued to be larger in the warmed treatment (Fig. 1d). This effect was driven by a difference in stem taper- the warmed trees had wider stem diameters throughout the crown than the ambient trees (*not shown*). The drought treatment reduced tree diameter but not height growth (Fig. 1a,b), modestly reduced total leaf area (Fig. 1c), and reduced stem volume increment (Fig. 1d). Notably, there was no interactive effect of warming and drought on growth (e.g., P > 0.4 for volume increment).

#### $CO_2$ and $H_2O$ fluxes

Experimental warming increased photosynthetic C uptake and H<sub>2</sub>O loss via transpiration early in the experiment (Fig. 2a,c). This was expected, given the strong increase in tree growth and total leaf area with experimental warming during this period (Fig. 1). However, the rates of C uptake and H<sub>2</sub>O loss converged between the ambient and warmed treatments during the summer (January; Fig. 2a,c), despite the fact that the warmed trees were larger and had more leaf area. This may have been influenced by warming-induced reductions in photosynthetic rates per unit leaf area (Drake *et al.*, 2016b).

We imposed an experimental drought in which all surface irrigation was withheld from trees in the dry treatments for nearly three months. Total C uptake during the drought period was reduced 25% while total  $H_2O$  loss was reduced 32% (Fig. 2b,c; main effects of drought, P < 0.01; no interaction with warming, P > 0.5). Thus, the drought strongly and significantly reduced whole-crown fluxes of C uptake and  $H_2O$  loss. On the other hand, these fluxes were maintained at moderate values during the drought, despite the complete lack of water addition.

#### 310 Final harvest

The final biomass did not differ between the warming or drought treatments (Fig. 3a). The lack of difference in final mass between the ambient and warmed treatments may have arisen from the convergence of tree diameter and height across treatments (Fig. 1a,b). The difference in stem volume between ambient and warmed trees (Fig. 1d) was apparent in the harvest biomass (Fig. 3a), but was not statistically significant at this level of replication (n = 3; P > 0.05). The only biomass component that was affected by the experimental treatments at harvest was fine root biomass, for which there was a significant

interaction between warming and drought (*P* < 0.05). The A-Dry trees had higher fine root biomass than
the A-Wet trees, while the W-Dry trees had slightly lower fine root biomass than the W-Wet trees (Fig.
319 3a). This interaction was also present in the tree root mass ratios; experimental drought increased the root
mass ratio, but only in the ambient temperature treatment (*P* < 0.05; Fig. 3b).

\*\*Plant and soil water status\*\*
The drought reduced soil volumetric water content from 10-100 cm depth to values approaching 0.05 m<sup>3</sup>\*

The drought reduced soil volumetric water content from 10-100 cm depth to values approaching 0.05 m<sup>3</sup> m<sup>-3</sup> (Fig. 4a-c). Pre-dawn leaf water potentials ( $\Psi_{\text{L-PD}}$ ) were reduced in the dry treatments relative to the wet treatments (Fig. 4d; P < 0.01). However, this effect was modest;  $\Psi_{\text{L-PD}}$  was -0.29  $\pm$  0.02 in the control and -0.48  $\pm$  0.05 in the dry treatments. Thus, the drought trees had moderate  $\Psi_{\text{L-PD}}$  (Fig. 4d) and moderate rates of transpiration (Fig. 2c) despite extremely dry surface soils.

Trees likely utilized deep soil water during the drought treatment. We observed a few roots of approximately 1-cm-diameter penetrating through the cemented manganese layer at ~100 cm depth during the complete soil excavation (JE Drake, *personal observation*). Neutron probe measurements down to 400-cm-depth indicated that soil water was removed from the profile in the dry treatment chambers during the drought, particularly from 50- to 200-cm-depth (Fig. S1). Thus, trees in the dry treatments likely transpired deep soil water during the summer drought, consistent with a previous drought study of *Eucalyptus saligna* at this site (Duursma *et al.* 2011).

Fluxes of GPP, NPP<sub>a</sub>,  $R_a$  and allocation belowground

We derived gross primary production (GPP) and its partitioning to above ground net primary production  $(NPP_a)$ , above ground autotrophic respiration  $(R_a)$ , and the residual, which we attribute to C allocation below ground as well as measurement error.

GPP was increased by experimental warming early in the experiment (+22%, P < 0.01), but GPP between ambient and warmed treatments converged beginning in mid-summer (late January; Fig. 5a). The drought treatment reduced GPP in both temperature treatments (-15%, P < 0.01). These results follow the net C flux measurements (Fig. 2a,c). The response of NPP<sub>a</sub> (Fig. 5b) closely followed the results for GPP, with a warming effect early in the experiment (+36%, P < 0.01) and a reduction with drought in both temperature treatments (-25%, P < 0.01). The response of R<sub>a</sub> (Fig. 5c) also followed GPP, with a stimulation by warming early in the experiment (+39%, P < 0.01) and a modest reduction with drought that was equivalent across temperature treatments (-13%, P < 0.05). The allocation of C belowground, as measured by the residual, was decreased by experimental warming throughout the experiment (-11%, P < 0.05) and was unchanged by the drought treatment (+3%, P > 0.1; Fig. 5d).

*GPP partitioning* 

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Given these flux measurements, we derive the partitioning of GPP into three components; NPP<sub>a</sub>/GPP, R<sub>a</sub>/GPP, and Residual/GPP (Figs. 6-7). Warming increased NPP<sub>a</sub>/GPP in a way that was stronger early in the experiment (+11%, P = 0.01) relative to the entire experiment (+3%; P > 0.1; Fig. 6ab). Similarly, warming increased  $R_a/GPP$  (+12%; P < 0.1) but decreased Residual/GPP (-15%; P < 0.1) 0.05) prior to the drought (Figs 6-7). Thus, experimental warming increased the partitioning of GPP to aboveground components (Fig. 7a-b) and decreased partitioning belowground (Fig. 7c). The experimental drought had weak effects on partitioning, none of which were statistically significant (P > 0.1). Growth and maintenance respiration We combined growth and respiratory measurements to infer changes in respiratory C efflux attributable to growth versus maintenance respiration (Amthor, 2000). There was a strong and linear relationship between R<sub>a</sub> per unit aboveground tree C and relative growth rate (RGR; Fig. 8). Neither the slope nor the intercept of this relationship were affected by experimental treatments (all P > 0.1). Thus, we present a common relationship across all measurements. The y-intercept of this relationship was positive (mean of 0.0213, 95% CI of 0.0157 to 0.0268), indicating significant R<sub>a</sub> in the absence of aboveground growth, reflecting maintenance respiration. The lack of a warming effect on this y-intercept is consistent with respiratory temperature acclimation; trees in the ambient and warmed treatments expended similar amounts of C on maintenance respiration, despite the increased temperature in the warmed treatment. The slope of the relationship (Fig. 8) was strongly positive (mean of 0.0059, 95% CI of 0.0053 to 0.0065), indicating that much of the R<sub>a</sub> observed at the crown scale was attributable to construction respiration. Observations during the drought period followed the general relationship, with lower values on both axes (Fig. 8). Thus, the experimental drought reduced R<sub>a</sub> primarily via a reduction in growth respiration. We also directly estimated coefficients for growth and maintenance respiration by fitting equation 2 to the fortnightly dataset of standing biomass, growth rate, and respiration. We estimate the growth respiration to consume approximately 0.3 gC per gram of biomass C produced, and maintenance respiration to consume approximately 0.015 gC per g of standing biomass C per day (Table 1). These

coefficients did not differ across the ambient and warmed treatment (P > 0.3).

## **Discussion**

Summary

We studied the experimental effects of warming and drought on the C allocation of *Eucalyptus* tereticornis trees using a combination of growth and whole-crown flux measurements. Experimental warming increased the proportion of GPP that was allocated to aboveground uses and decreased the proportion of GPP that was allocated belowground. This was consistent with a reduced root mass fraction in the warmed treatments at the final harvest. The experimental drought reduced CO<sub>2</sub> and H<sub>2</sub>O fluxes but did not affect the allocation of C, perhaps because tree access to deep soil water prevented them from experiencing strongly negative water potentials. There were no interactions between warming and drought on C partitioning terms, so we discuss the impacts of warming and drought separately.

Effects of experimental warming on C allocation

Experimental warming strongly affected several aspects of tree C allocation. Warming increased the fractional partitioning of GPP to aboveground uses, including growth and respiration, at the expense of C partitioning belowground. This observation is consistent with some soil warming experiments (e.g., Melillo *et al.*, 2002, 2011) that attributed this effect to a warming-induced stimulation of soil nutrient availability. However, it is also possible that experimental warming directly stimulated the activity of meristems aboveground, such that a smaller remainder of fixed C was available for transport and use belowground. Such a mechanism would imply an aboveground priority in tree C allocation, consistent with previous work on forest C budgets and elevated atmospheric CO<sub>2</sub> treatments (Palmroth *et al.*, 2006). This mechanism also makes sense given the structural arrangement of tree phloem, as aboveground tissues have the opportunity to remove sucrose from the phloem before belowground tissues (Lemoine *et al.*, 2013; Furze *et al.*, 2018). The mechanisms regarding soil nutrient availability and aboveground metabolic activity are not mutually exclusive. For example, enhanced N supply from soil N mineralization may have enabled the increased aboveground metabolism in the warmed treatment, which may have resulted in the larger consumption of GPP aboveground in the warmed relative to the ambient treatment.

We also acknowledge that warming may have influenced allocation indirectly via ontogenetic drift. A meta-analysis documented a decline in root to shoot ratios as trees grew larger, possibly reflecting an ontogenetic effect on belowground allocation (Mokany *et al.*, 2006), although we hesitate to infer allocation directly from root to shoot ratios (Reich, 2002; Litton *et al.*, 2007). In this experiment, the allocation terms (e.g., NPPa/GPP, Ra/GPP, and residual/GPP) were not significantly correlated with any metric of tree size, suggesting that the warming effect is unlikely to reflect ontogeny.

We previously demonstrated that aboveground autotrophic respiration acclimated nearly homeostatically to experimental warming in this experiment, both at the leaf-scale (Aspinwall *et al.*, 2016) and at the whole-crown scale (Drake *et al.*, 2016b). As such, the demonstration that warming increased R<sub>a</sub> (Fig. 5) may appear contradictory. We emphasize that our previous presentations of autotrophic respiration were expressed per unit leaf area (Drake *et al.*, 2016b; Aspinwall *et al.*, 2016), while the current study shows the total fluxes per tree (Fig. 5c). Experimental warming increased R<sub>a</sub> primarily by increasing growth and tree size early in the experiment. Furthermore, the common relationship between relative growth rate and R<sub>a</sub> per unit tree mass for the ambient and warmed treatment is indicative of homeostatic acclimation of maintenance respiration in this experiment (Fig. 8). The stimulation of whole-crown R<sub>a</sub> by warming was primarily attributable to an increase in respiration to support growth. Thus, we suggest that this study is in agreement with previous published work from this experiment (Drake *et al.*, 2016b; Aspinwall *et al.*, 2016), where homeostatic acclimation of respiration to experimental warming prevented a warming-induced increase in maintenance respiration, while a warming effect on growth stimulated growth respiration and increased whole-crown R<sub>a</sub>.

The increased allocation of C aboveground in the warmed treatment, combined with homeostatic acclimation of maintenance respiration, likely contributed to the observed warming-induced stimulation in growth during the first half of this experiment (Fig. 1). Experimental warming had neutral or negative effects on leaf-level photosynthetic rates in this study (Drake et al., 2016b; Aspinwall et al., 2016), so a warming-induced stimulation of growth was somewhat surprising. We suggest that an increase in C partitioning aboveground (Fig. 6a) was associated with accelerated leaf development early in the experiment in these young and rapidly growing trees (Fig. 1c), such that trees exposed to the warmed treatment had higher rates of crown-scale photosynthesis (Fig. 5a) primarily through a warming effect on total crown leaf area. That is, the warmed treatment got a head start in leaf area production, which compounded over time. This is consistent with nutrient fertilization studies, in which increases in leaf area rather than changes in leaf N or leaf function, often dominates the growth responses of rapidly growing plants (Sinclair & Horie, 1989; Gastal & Lemaire, 2002; Lovelock et al., 2004; Wang et al., 2012), although there are exceptions (Santiago et al., 2012). This is also consistent with other experimental manipulations, in which a stimulation of leaf area development early in an experiment can strongly affect exponential growth trajectories (Tjoelker et al., 1998; Kirschbaum, 2011; Drake et al., 2017).

#### Effects of drought on C allocation

We did not detect any significant effects of drought on C partitioning. The drought appeared to reduce all C fluxes proportionally, such that the ratios of C fluxes to GPP was unchanged. We recognize

that our ability to resolve C partitioning belowground was limited by the nature of the measurements based on the residual, and our lack of root biomass measurements through time. The biomass in roots relative to the total at the harvest (root mass ratio; Fig 3b) did indicate an interactive effect of drought and warming, possibly via small differences in the partitioning of GPP to roots that accumulated over time. Perhaps the ambient temperature trees had sufficient carbohydrate reserves to fuel additional root growth in the drought treatment, while warmed temperature trees were consuming more carbohydrates aboveground and were thus unable to increase root growth in the drought. This speculative process may explain the observed interaction between warming and drought on root mass ratio (Fig. 3b).

Trees acquired water from deep in the soil profile during the drought. Leaf predawn water potential declined to only approximately -0.5 MPa, which is a moderate value that is not indicative of pronounced water stress. Previous studies have shown that groundwater use enables vegetation to avoid production declines under conditions of surface moisture limitation (Baldocchi *et al.*, 2010; Barbeta *et al.*, 2015), and several eucalypt species are well-known users of groundwater (Mensforth *et al.*, 1994; Pfautsch *et al.*, 2011, 2015; Eamus *et al.*, 2015; Zolfaghar *et al.*, 2017). Furthermore, Koirala *et al.* (2017) demonstrated correlations between GPP and groundwater table depth that were present over approximately 70% of the vegetated surface of the earth, suggesting that vegetation-groundwater interactions are common and globally relevant. Our study demonstrates that some trees may utilize access to soil water at depth to maintain moderate rates of photosynthetic C uptake and growth during extended droughts that lead to dry surface soils.

Implications for mathematical models

Many ecosystem and earth system models begin their simulation of ecosystem C cycling by predicting GPP as a function of leaf area and environmental drivers. GPP is then partitioned into component terms including R<sub>a</sub> and the production of leaf, wood, and root mass. Our observations suggest that tree C allocation of GPP to these terms can be influenced by environmental drivers such as temperature. That is, the observations presented here are not consistent with static partitioning schemes with fixed and constant partitioning of GPP into component fluxes. However when aggregated over longer time periods, our average partitioning coefficients (approximately 30% of GPP to aboveground respiration, 43% of GPP to aboveground growth, and 27% to belowground fluxes) are comparable with values used in some ecosystem models (De Kauwe *et al.*, 2014). The observations presented here are also not consistent with a dynamic C partitioning scheme based on Sprengel and Leibig's law of the minimum (van der Ploeg *et al.*, 1999), where C would be preferentially allocated to increase the acquisition of the factor most limiting primary production. Furthermore, Aspinwall *et al.* (2016) recently documented strong seasonal variation in carbohydrate storage in these evergreen trees, characterized by the buildup of high starch

473 concentrations during the winter and a drawdown of these reserves during the summer. It appears that 474 these trees partially utilize a non-structural carbohydrate storage reserve to fuel growth and metabolism 475 during the hot summer. Based on these observations, we suggest that a dynamic allocation scheme 476 incorporating a dynamic carbohydrate reserve may be appropriate for future investigation (e.g., Fatichi et 477 al., 2014; Pugh et al., 2016). 478 **Conclusions** 479 We used detailed measurements of growth and whole-crown flux measurements to study the effects of 480 warming and drought on the C allocation of *Eucalyptus tereticornis* trees. Experimental warming 481 increased the proportion of GPP that was allocated to aboveground uses and decreased the proportion of 482 GPP that was allocated belowground. Such a change in tree C allocation may have important implications 483 for tree growth, forest C storage, and soil nutrient cycling in a warmer world. In particular, increased 484 allocation aboveground in a warmer world may stimulate leaf area development and aboveground growth 485 during conditions of sufficient soil resource supply. Acknowledgements 486 487 We thank Burhan Amiji (Western Sydney University) for maintaining the site, for collecting much of the 488 growth and harvest data, and for his excellent research support. This experiment was made possible 489 through a collaboration with Sune Linder and the Swedish University of Agricultural Sciences, who 490 designed, built, and generously provided the whole tree chambers. We also gratefully acknowledge 491 Courtney Campany (Colgate University) for his measurements of fine root biomass as well as Renee 492 Smith, Carrie Drake (Western Sydney University), and Richard Harwood (Sydney University) for their 493 help with the whole-tree harvests. This research was supported by the Australian Research Council 494 (Discovery, DP140103415), a New South Wales government Climate Action Grant (NSW 495 T07/CAG/016), the Hawkesbury Institute for the Environment, and Western Sydney University. **Authorship statement** 496 497 JED co-led the experimental design, contributed to data collection for the chamber flux, and led 498 the data analysis, modeling, and writing. MGT was the senior scientific lead; he co-led the 499 experimental design and made large contributions to analysis, interpretation, and writing. MJA 500 contributed to the experimental design, data collection and interpretation, and writing. PBR 501 contributed to the experimental design, interpretation, and writing. SP contributed to 502 experimental design, data collection and interpretation, and writing. CVMB contributed to the

503 measurements of chamber flux, and contributed to experimental design, data analysis, and writing.

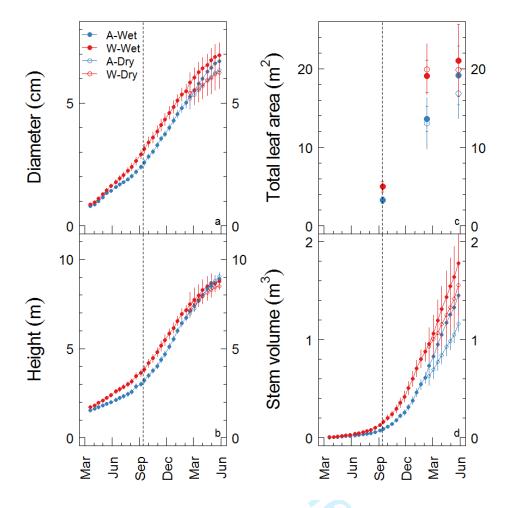
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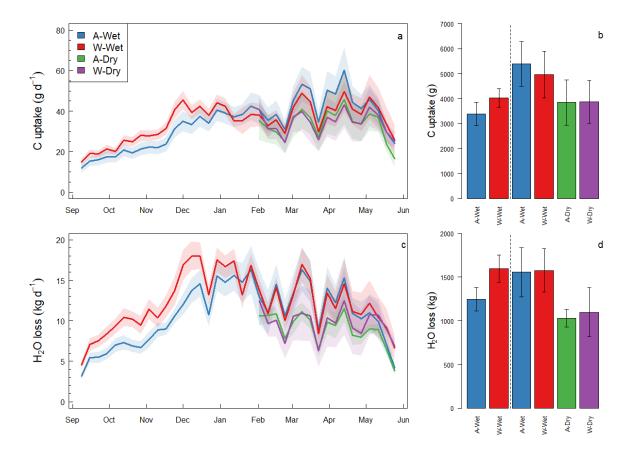
Table 1. Estimate of aboveground growth and maintenance respiration coefficients derived from equation 2, with standard errors (SE), and 95% confidence intervals; parameters were statistically equivalent across ambient and warmed treatments (all P > 0.3).

Term	Units	Ambient		Warmed	
		Mean (SE)	95% CI	Mean (SE)	95% CI
$g_{\rm r}$ (growth respiration rate)	g C respired per g C growth	0.32 (0.02)	0.27-0.37	0.28 (0.03)	0.21-0.36
$m_{\rm r}$ (maintenance respiration rate)	g C respired per g C standing aboveground biomass per day	0.015 (0.001)	0.012-0.019	0.017 (0.002)	0.013-0.021

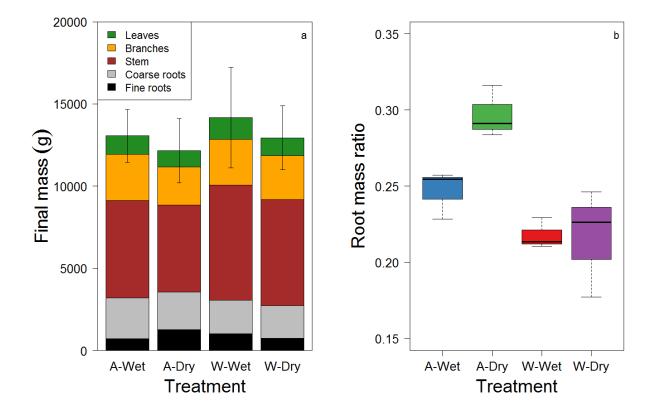




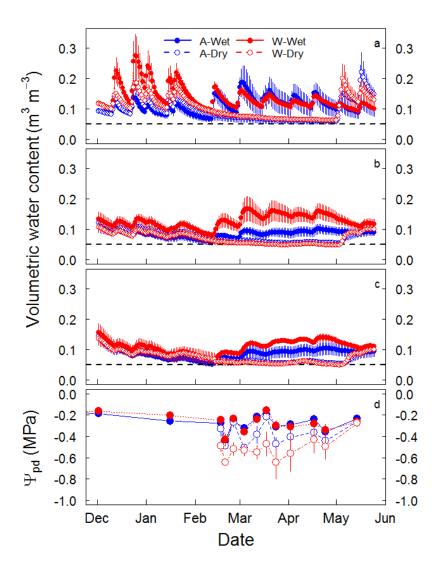
**Figure 1**. Growth of *Eucalyptus tereticornis* trees exposed to warming and drought. Trees were either exposed to ambient  $T_{air}$  ("A", blue) or warming of +3 °C ("W", red), and either well-watered ("Wet", solid points) or drought conditions ("Dry", open points). Stem diameter (a) was measured at 65-cm height, and height reflects total stem length (b). Total leaf area was directly measured on three dates (c) and stem volume was calculated from diameter measurements along the stem of each tree (d). The vertical dashed line denotes when  $CO_2$  and  $H_2O$  flux measurements began. Points reflect the mean, error bars denote 1SEM (n = 6 until Feb 2014, when the drought treatment began and n = 3).



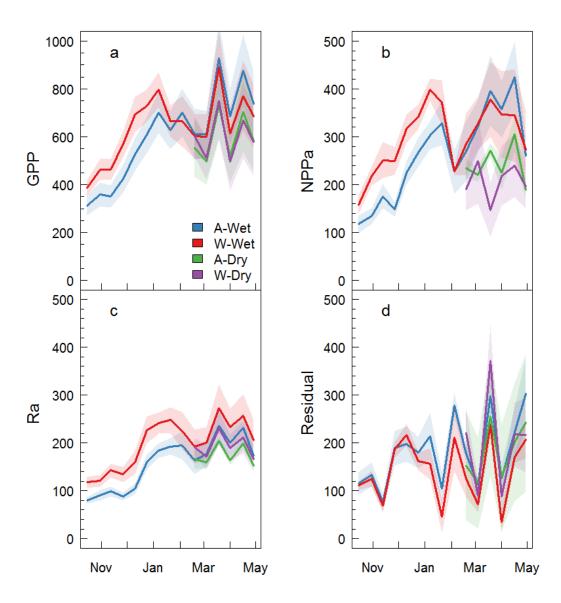
**Figure 2**. Summary of measured CO<sub>2</sub> and H<sub>2</sub>O fluxes for twelve *Eucalyptus tereticornis* trees exposed to ambient (A) or warmed (W) air temperatures in 2013 and 2014. All trees were maintained in well-water conditions (Wet) until mid-Feb, when half of the trees were subjected to a soil drydown (Dry). We show weekly averages of the measured daily net C uptake (a) and the sum of net C uptake for the two measurement periods (pre-drought, drought; b). We also show weekly averages of the measured daily net H<sub>2</sub>O loss to transpiration (c) and the H<sub>2</sub>O loss to transpiration summed across the two measurement periods (d). In (a) and (c), lines reflect the mean and shaded areas reflect the standard error. The dotted vertical lines in (b) and (d) separate the pre-drought (left) and drought periods (right). These plots reflect >580,000 individual flux measurements at 15-minute resolution.



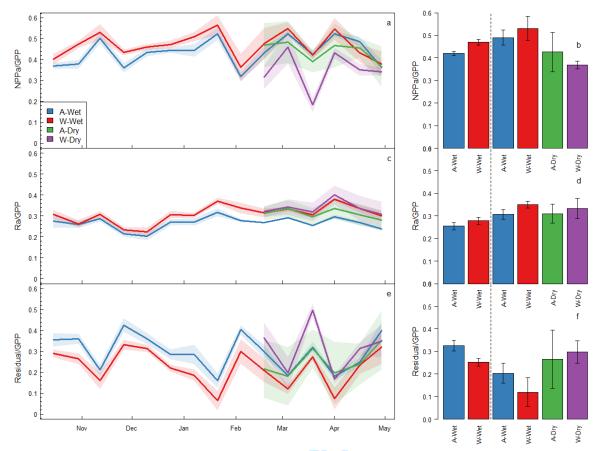
**Figure 3**. Biomass components at final harvest for twelve *Eucalyptus tereticornis* trees exposed to ambient (A) or warmed (W) air temperatures and either well-watered conditions (Wet) or a soil drydown treatment (Dry). Note that these data reflect grams of dry mass. Each of the measured biomass components (a) reflects the mean of three trees per treatment, the error bars reflect the standard error of the total measured mass. The root mass ratio (b) reflects the sum of coarse and fine roots relative to total tree mass. Warming reduced the root mass ratio, while the drought treatment increased root mass ratio in the ambient temperature treatment only. The root mass ratio interaction primarily follows the response of fine roots, although stem wood and coarse roots also contributed.



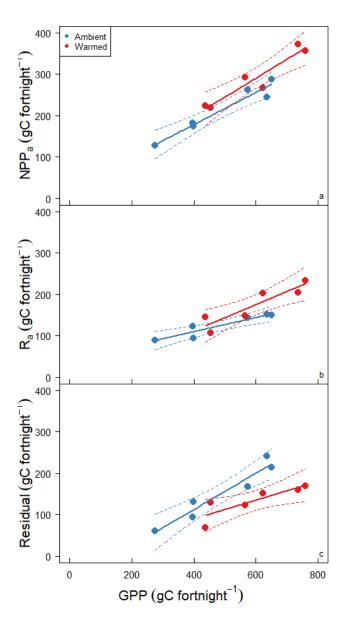
**Figure 4**. Soil volumetric water content and predawn leaf water potential ( $\Psi_{pd}$ ) for twelve *Eucalyptus tereticornis* trees exposed to ambient (A) or warmed (W) air temperatures. All trees were maintained in well-water conditions (Wet) until mid-Feb, when half of the trees were subjected to a soil drydown (Dry). We show daily averages of the measured volumetric water content in surface soils ( $\sim$ 0.1-m-depth; a), an intermediate depth ( $\sim$ 0.5-m-depth; b), and in deep soils just above the hard layer of partially cemented manganese nodules ( $\sim$ 1-m-depth; c). We also show leaf  $\Psi_{pd}$  measured throughout the drydown (d). Points reflect the mean and error bars reflect the standard error (n = 6 or 3). Note that  $\Psi_{pd}$  was moderate in all treatments.



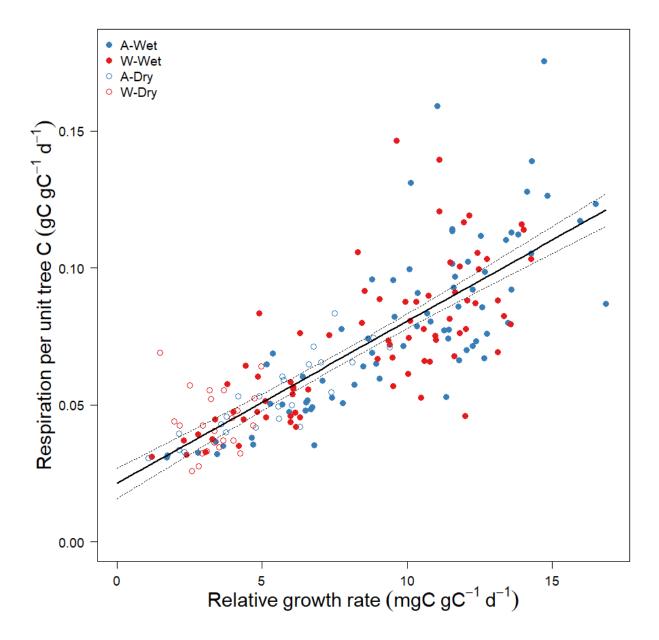
**Figure 5**. Fortnightly C fluxes for twelve *Eucalyptus tereticornis* trees exposed to ambient (A) or warmed (W) air temperatures. All fluxes are presented in units of g C tree<sup>-1</sup> fortnight<sup>-1</sup>. All trees were maintained in well-water conditions until mid-Feb (Wet), when half of the trees were subjected to a soil drydown (Dry). Solid lines reflect the mean of fortnightly data (i.e., two-weekly) and shaded areas reflect 1SEM. Measurements include gross primary production (GPP; a), aboveground net primary production (NPPa; b), aboveground autotrophic respiration (Ra; c), and the residual (d). The residual reflects belowground C flux and measurement error. Note that the y-axis scale is twice as large for GPP relative to the other fluxes.



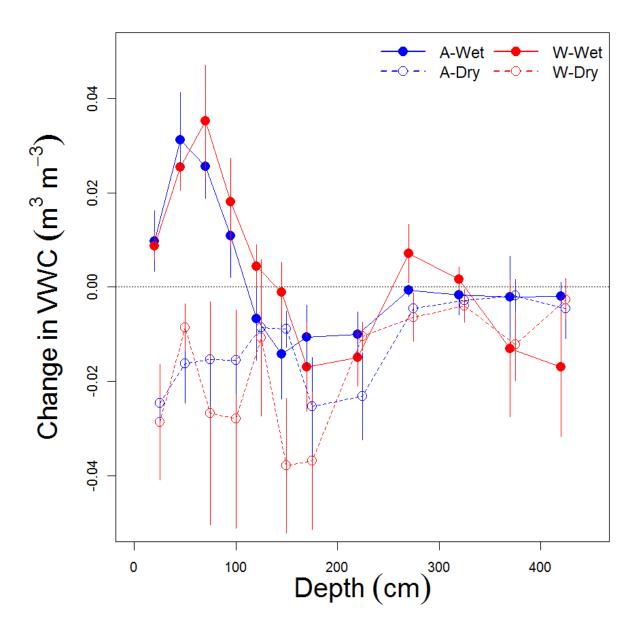
**Figure 6**. The fractional partitioning of gross primary production (GPP) for twelve *Eucalyptus tereticornis* trees. GPP was partitioned into aboveground net primary production (NPP $_a$ ; a-b), aboveground autotrophic respiration (R $_a$ ; c-d), and the residual C, which includes belowground C allocation and measurement error (e-f). All trees were maintained in well-water conditions (Wet) until mid-Jan, when half of the trees were subjected to a soil drydown (Dry). Bar charts of flux partitioning terms (b, d, f) represent the mean ( $\pm 1$ SEM), and the dotted vertical lines separate the pre-drought (left) and drought periods (right).



**Figure 7**. The fractional partitioning of gross primary production (GPP) for twelve *Eucalyptus tereticornis* trees grown under ambient and elevated temperature. Data for each tree were averaged across the pre-drought period; each point reflects an individual tree (n = 6). GPP was partitioned into aboveground net primary production (NPP<sub>a</sub>; a), aboveground autotrophic respiration ( $R_a$ ; b), and the residual C, which includes belowground C allocation (c). Solid lines reflect linear models fit to each treatment; dashed lines reflect the 95% confidence interval.



**Figure 8**. Partitioning of aboveground respiration into maintenance and growth components. Each point reflects a tree during a fortnightly growth interval. Note that the y-intercept reflects the maintenance respiration component and the slope reflects the growth respiration component. Neither the slope nor the intercept were affected by experiment treatments (mixed effects model with random intercepts for each chamber, P > 0.5). The solid lines reflects models fit to the ambient temperature (A) and warmed temperature (W) data and dashed lines reflect the 95% confidence interval. All of the data were well-described by a single linear function (Y = 0.021 + 0.0059x,  $r^2 = 0.64$ , P < 0.001; *not shown*).



**Figure S1**. The change in volumetric water content (VWC) throughout the soil profile, comparing the end of the drought to the beginning of the drought. VWC was measured with the neutron probe method; negative values reflect reductions in VWC from the beginning (17 Feb 2014) to the end of the drought (24 April 2014). Points reflect means and error bars reflect 1SEM (n = 3). Note that trees in the dry treatment likely acquired water from soils as deep as 200 cm during the drought. The positive values in the shallow soils of "Wet" trees reflects the addition of  $H_2O$  via surface irrigation.

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