tu13 DataWrangling

February 28, 2023

1 Data Wrangling

Data wrangling generally refers to the process of getting a data set ready for analysis. Why would we need to do that?

Real-world data can be messy. Data sets are recorded and assembled by humans, and humans make mistakes. A single data set might created and updated by multiple people who may decide to do things in slightly different ways. On a spreadsheet, one person might might decide to leave cells with missing data blank, another might enter "NaN", while a third may enter "missing". If the data has many many rows, one person might decide to repeat the column headers partway down so they don't have to scroll up to see them. Any of these things mean that the data set cannot be analyzed "as is" and wrangling will be required.

Even in a tightly controlled laboratory setting in which data are collected via computer and automatically written out to data files, some data wrangling might be required. There might be a separate data file for each subject or experimental session, meaning that these separate files will have to be combined into a single data set before analysis.

Our main wrangling tool is pandas, so we can go ahead and import it.

```
[1]: import pandas as pd
```

1.1 Loading

For our wrangling practice today, we'll look at a data set containing various measurements on breast cancer patients. The file is called breast_cancer_data.csv, and you should place it in the "data" folder you should already have in the same directory as this notebook.

Let's import it as a pandas dataframe.

```
[2]: bcd = pd.read_csv('./data/breast_cancer_data.csv')
bcd
```

[2]:	<pre>patient_id</pre>	clump_thickness	cell_size_uniformity	cell_shape_uniformity	\
0	1000025	5.0	1.0	1	
1	1002945	5.0	4.0	4	
2	1015425	3.0	1.0	1	
3	1016277	6.0	8.0	8	
4	1017023	4.0	1.0	1	
	•••	•••	***	***	

694 695 696 697 698	776715 841769 888820 897471	2 5 4	.0 .0 .0 .0	1.0 10.0 0 8.0			
0 1 2 3 4 694 695 696 697	 1 3 1	1 5 1 1 1 3 1 1 1 3 3	_ep_cell_si 	ze bare_nuc: 2 7 2 3 2 3 2 7 3 4	lei 1 10 2 4 1 2 1 3 4 5	bland_chromatin	\
0 1 2 3 4 694 695 696 697 698		mitoses	class benign benign benign benign benign benign benign malignant malignant malignant	doctor_name Dr. Doe Dr. Smith Dr. Lee Dr. Smith Dr. Wong Dr. Lee Dr. Smith Dr. Lee Dr. Smith Dr. Lee Dr. Lee Dr. Lee			

[699 rows x 12 columns]

Before we do any actual wrangling, let's get familiar with the data frame in its current form.

1.2 Exploring the Data Frame

We can explore the data frame by looking at it's attributes, such as its shape, column names, and data types:

```
[3]: bcd.columns
```

Use the cells below to get the shape and data types (dtypes) of our data frame.

[4]: bcd.shape

- [4]: (699, 12)
- [5]: bcd.dtypes
- [5]: patient_id int64 clump_thickness float64 cell_size_uniformity float64 cell_shape_uniformity int64 marginal_adhesion int64 single_ep_cell_size int64 bare_nuclei object bland_chromatin float64 normal_nucleoli float64 mitoses int64 object class doctor_name object dtype: object

In the cell below, use the describe() method to get a summary of the numerical columns.

[6]: bcd.describe()

[6]:		patient_id	clump_th	ickness	cell_size_	uniformity	\	
	count	6.990000e+02	698	.000000		698.000000		
	mean	1.071704e+06	4	.416905		3.137536		
	std	6.170957e+05	2	.817673		3.052575		
	min	6.163400e+04	1	.000000		1.000000		
	25%	8.706885e+05	2	.000000		1.000000		
	50%	1.171710e+06	4	.000000		1.000000		
	75%	1.238298e+06	6	.000000		5.000000		
	max	1.345435e+07	10	0.00000		10.000000		
		cell_shape_un	iformity	margina	l_adhesion	single_ep_	cell_size	\
	count	69	9.000000		699.000000	6	99.000000	
	mean		3.207439		2.793991		3.216023	
	std		2.971913		2.843163		2.214300	
	min		1.000000		1.000000		1.000000	
	25%		1.000000		1.000000		2.000000	
	50%		1.000000		1.000000		2.000000	
	75%		5.000000		3.500000		4.000000	
	max	1	0.000000		10.000000		10.000000	

	bland_chromatin	normal_nucleoli	mitoses
count	695.000000	698.000000	699.000000
mean	3.447482	2.868195	1.589413
std	2.441191	3.055647	1.715078
min	1.000000	1.000000	1.000000
25%	2.000000	1.000000	1.000000
50%	3.000000	1.000000	1.000000
75%	5.000000	4.000000	1.000000
max	10.000000	10.000000	10.000000

1.3 Modifying a text column

We'll often want to "tune up" columns that contain text. We might encounter, for example, a column containing full names that we need to break up into separate columns for the first and last names.

Let's look at the column for the doctors' names. Use the cell below to take a peek.

```
[7]: bcd['doctor_name']
[7]: 0
              Dr. Doe
     1
            Dr. Smith
     2
              Dr. Lee
     3
            Dr. Smith
     4
             Dr. Wong
     694
              Dr. Lee
     695
            Dr. Smith
     696
              Dr. Lee
     697
              Dr. Lee
     698
             Dr. Wong
     Name: doctor name, Length: 699, dtype: object
```

The doctors' name data are redundant; each one has a "Dr." in front of the actual name, but we already know these are doctors by the column name. Further, the entries have white space in them, which can cause us problems down the road. So let's modify this column so it only contains the surnames of the doctors.

One great thing about pandas is that it has versions of many of Python's string methods that operate *element-wise on an entire column of strings*. Here, we want to separate the "Dr." from the actual name, which is exactly what Python's **str.split()** function does. So chances are, pandas has a version of this function that operates element-wise on data frames.

String Splitting Review: Let's briefly remind ourselves of splitting up Python strings and extracting bits of them.

```
[8]: # Here's a string of the form: surname, first initial.
myStr = 'SirString, A.'
print(myStr)
```

SirString, A.

Let's say we wanted to get the surname. We could split this string into a Python list at the white space like this:

```
[9]: spltStr = myStr.split() # split() defaults to splitting at white space
print(spltStr)
```

```
['SirString,', 'A.']
```

We now have a list in which the items contain the text on either side of the split. This is close to what we want: the first entry in the list has the surname, but it also has an unwanted comma.

Let's split the string at the comma instead:

```
[10]: spltStr = myStr.split(',') # tell Python to split at commas
print(spltStr)
```

['SirString', ' A.']

Now we have isolated the last name, and we can fetch it by indexing:

```
[11]: surname = spltStr[0]
print(surname)
```

SirString

In the cell below, see if you can extract the surname from myStr in one line of code:

```
[12]: myStr
```

[12]: 'SirString, A.'

```
[13]: test_spr = myStr.split(',')[0]
test_spr
```

[13]: 'SirString'

Alright, time to replace the bcd['doctor_name'] column values with just the doctors' last names.

We could do this in one step, but let's break it out for clarity. First, let's copy the name column out into a new series.

```
[14]: dr_names = bcd['doctor_name']
      dr_names
[14]: 0
               Dr. Doe
      1
             Dr. Smith
      2
               Dr. Lee
      3
             Dr. Smith
      4
              Dr. Wong
      694
               Dr. Lee
             Dr. Smith
      695
               Dr. Lee
      696
      697
               Dr. Lee
      698
              Dr. Wong
      Name: doctor_name, Length: 699, dtype: object
```

Note: pandas objects behave like ordinary Python objects. So, strictly speaking, we have not created a new object (pandas Series), rather, we have created a new label that refers to the "doctor name" column of bcd.

In the cell below, use the id() function to compare the object IDs of dr_names and the corresponding column of bcd.

```
[15]: id(dr_names)
[15]: 140289344521120
[16]: id(bcd['doctor_name'])
[16]: 140289344521120
```

Now let's split all the names in the doctor_name column at the whitespace by using pandas DataFrame.str.split() function.

```
[17]: split_dr_names = dr_names.str.split()
split_dr_names
```

```
[17]: 0 [Dr., Doe]
1 [Dr., Smith]
2 [Dr., Lee]
3 [Dr., Smith]
4 [Dr., Wong]
...
694 [Dr., Lee]
695 [Dr., Smith]
```

```
696 [Dr., Lee]
697 [Dr., Lee]
698 [Dr., Wong]
Name: doctor_name, Length: 699, dtype: object
```

DataFrame.str.split(), however, does create a new object.

Use the cell below to confirm that the split() spawed a new object.

```
[20]: split_dr_names = dr_names.str.split('.')
split_dr_names
```

```
[20]: 0
                     Doe]
                [Dr,
      1
             [Dr,
                   Smith]
      2
                [Dr, Lee]
      3
             [Dr, Smith]
              [Dr, Wong]
      694
                [Dr, Lee]
      695
             [Dr, Smith]
      696
                [Dr,
                     Lee]
      697
                [Dr, Lee]
      698
              [Dr, Wong]
      Name: doctor_name, Length: 699, dtype: object
```

Now we have a column of lists, each with two elements. The first element of each list is the "Dr." bit, and the second consists of the surnames we want.

We can get these by using pandas string indexing, Series.str[index].

```
[23]: surnames = split_dr_names.str[1]
surnames
```

```
[23]: 0
                 Doe
      1
               Smith
      2
                 Lee
               Smith
      3
                Wong
      4
      694
                 Lee
      695
               Smith
      696
                 Lee
      697
                 Lee
      698
                Wong
      Name: doctor_name, Length: 699, dtype: object
```

Note that, like the splitting, the string indexing worked on the entire Series automatically. Now we can change the column in our main data frame, bcd.

```
[24]: bcd['doctor_name'] = surnames
[25]: bcd['doctor_name']
[25]: 0
                 Doe
      1
               Smith
      2
                 Lee
      3
               Smith
      4
                Wong
      694
                 Lee
      695
               Smith
      696
                 Lee
      697
                 Lee
      698
                Wong
      Name: doctor_name, Length: 699, dtype: object
[26]: bcd.head()
[26]:
         patient_id
                      clump_thickness
                                         cell_size_uniformity
                                                                 cell_shape_uniformity
             1000025
      0
                                    5.0
                                                            1.0
                                                                                        1
                                    5.0
                                                            4.0
                                                                                       4
      1
             1002945
      2
                                    3.0
             1015425
                                                            1.0
                                                                                       1
      3
             1016277
                                    6.0
                                                            8.0
                                                                                       8
      4
             1017023
                                    4.0
                                                            1.0
                                                                                       1
         marginal_adhesion
                              single_ep_cell_size bare_nuclei
                                                                  bland_chromatin
      0
                           1
                                                  2
                                                               1
                                                                                3.0
                           5
                                                  7
                                                              10
                                                                                3.0
      1
      2
                                                  2
                                                               2
                           1
                                                                                3.0
                                                  3
      3
                           1
                                                               4
                                                                                3.0
                           3
                                                  2
      4
                                                                                3.0
         normal_nucleoli mitoses
                                       class doctor_name
      0
                       1.0
                                   1
                                      benign
                                                      Doe
                       2.0
      1
                                   1
                                      benign
                                                    Smith
      2
                       1.0
                                   1
                                      benign
                                                      Lee
      3
                       7.0
                                      benign
                                                    Smith
      4
                       1.0
                                      benign
                                                     Wong
```

Success!

1.4 Converting a column type (and other aggravations)

Let's look at those data types again.

[27]: bcd.dtypes

[27]:	patient_id	int64
	clump_thickness	float64
	cell_size_uniformity	float64
	cell_shape_uniformity	int64
	marginal_adhesion	int64
	single_ep_cell_size	int64
	bare_nuclei	object
	bland_chromatin	float64
	normal_nucleoli	float64
	mitoses	int64
	class	object
	doctor_name	object
	dtype: object	

Notice that "class" and "doctor_name" are of dtype "object", which refers to a general purpose column type, and is how pandas imports text columns by default. Most of the others are numeric (integers or floats), except for "bare_nuclei".

In the cell below, take a quick glance at 'bcd' again, and see if the "bare_nuclei" column should be a different data type that, say "marginal_adhesion".

[37]: bcd.head()

[0/].	bc	u.neau()										
[37]:		patient_id	clump	_thickne	ss cel	l_size_	uniform	nity	cell_shape_unifor	mity	\	
	0	1000025		5	.0	1.0						
	1	1002945		5	.0	4.0			4			
	2	1015425		3	.0	1.0				1		
	3	1016277		6	.0	8.0			8			
	4	1017023		4	.0			1.0	1			
		marginal_adl		_	_ep_cel		bare_nu	_	bland_chromatin	\		
	0		1			2		1	3.0			
	1		5)		7		10	3.0			
	2		1			2		2	3.0			
	3		1	-		3		4	3.0			
	4		3	3		2		1	3.0			
		normal_nucle	eoli	mitoses	class	doctor	_name					
	0		1.0	1	benign		Doe					
	1		2.0	1	benign		Smith					
	2		1.0	1	benign		Lee					
	3		7.0	1	benign		Smith					
	4		1.0	1	benign		Wong					

It looks like "bare_nuclei" was intended to be a numeric column, so let's try and convert it using the DataFrame.astype() converter method.

```
[29]: bcd['bare_nuclei'] = bcd['bare_nuclei'].astype('int64')
```

```
Traceback (most recent call last)
Input In [29], in <cell line: 1>()
----> 1 bcd['bare_nuclei'] = bcd['bare_nuclei'].astype('int64')
File ~/opt/anaconda3/lib/python3.9/site-packages/pandas/core/generic.py:5912, i
 →NDFrame.astype(self, dtype, copy, errors)
           results = [
   5905
                self.iloc[:, i].astype(dtype, copy=copy)
   5906
   5907
                for i in range(len(self.columns))
   5908
   5910 else:
            # else, only a single dtype is given
   5911
            new_data = self._mgr.astype(dtype=dtype, copy=copy, errors=errors)
-> 5912
   5913
            return self._constructor(new_data).__finalize__(self,_

→method="astype")
   5915 # GH 33113: handle empty frame or series
File ~/opt/anaconda3/lib/python3.9/site-packages/pandas/core/internals/managers
 →py:419, in BaseBlockManager.astype(self, dtype, copy, errors)
    418 def astype(self: T, dtype, copy: bool = False, errors: str = "raise") -
 ⇔T:
--> 419
            return self.apply("astype", dtype=dtype, copy=copy, errors=errors)
File ~/opt/anaconda3/lib/python3.9/site-packages/pandas/core/internals/managers
 →py:304, in BaseBlockManager.apply(self, f, align_keys, ignore_failures,
 →**kwargs)
    302
                applied = b.apply(f, **kwargs)
    303
            else:
--> 304
                applied = getattr(b, f)(**kwargs)
    305 except (TypeError, NotImplementedError):
            if not ignore_failures:
    306
File ~/opt/anaconda3/lib/python3.9/site-packages/pandas/core/internals/blocks.p
 →580, in Block.astype(self, dtype, copy, errors)
    562 """
    563 Coerce to the new dtype.
    564
   (...)
    576 Block
   577 """
    578 values = self.values
--> 580 new_values = astype_array_safe(values, dtype, copy=copy, errors=errors)
```

```
582 new_values = maybe_coerce_values(new_values)
    583 newb = self.make_block(new_values)
File ~/opt/anaconda3/lib/python3.9/site-packages/pandas/core/dtypes/cast.py:
 ⇔1292, in astype array safe(values, dtype, copy, errors)
   1289
            dtype = dtype.numpy_dtype
   1291 try:
            new_values = astype_array(values, dtype, copy=copy)
-> 1292
   1293 except (ValueError, TypeError):
            # e.g. astype_nansafe can fail on object-dtype of strings
   1294
            # trying to convert to float
   1295
   1296
            if errors == "ignore":
File ~/opt/anaconda3/lib/python3.9/site-packages/pandas/core/dtypes/cast.py:
 →1237, in astype_array(values, dtype, copy)
            values = values.astype(dtype, copy=copy)
   1236 else:
            values = astype_nansafe(values, dtype, copy=copy)
-> 1237
   1239 # in pandas we don't store numpy str dtypes, so convert to object
   1240 if isinstance(dtype, np.dtype) and issubclass(values.dtype.type, str):
File ~/opt/anaconda3/lib/python3.9/site-packages/pandas/core/dtypes/cast.py:
 ⇔1154, in astype_nansafe(arr, dtype, copy, skipna)
   1150 elif is_object_dtype(arr.dtype):
   1151
            # work around NumPy brokenness, #1987
   1152
            if np.issubdtype(dtype.type, np.integer):
   1153
                return lib.astype_intsafe(arr, dtype)
-> 1154
            # if we have a datetime/timedelta array of objects
   1156
   1157
            # then coerce to a proper dtype and recall astype_nansafe
   1159
            elif is_datetime64_dtype(dtype):
File ~/opt/anaconda3/lib/python3.9/site-packages/pandas/_libs/lib.pyx:668, in_u
 →pandas._libs.lib.astype_intsafe()
ValueError: invalid literal for int() with base 10: '?'
```

And, argh, we get an error! If we look at the bottom of the error message, it seems that the error involves question marks ("?") in the data, which would also explain why this column imported as text rather than numbers in the first place.

Let's check.

In the cell below, use logical indexing to show the rows of bcd in which bcd[bare_nuclei] contains a question mark.

```
[41]: bcd['bare_nuclei'].str.find('?')
[41]: 0
            -1.0
      1
             -1.0
      2
            -1.0
      3
             -1.0
      4
            -1.0
      694
            -1.0
      695
            -1.0
      696
            -1.0
            -1.0
      697
      698
             -1.0
      Name: bare_nuclei, Length: 699, dtype: float64
```

Sure enough. Rather than leaving the cells of missing values empty, somebody has made the poor decision to enter question marks instead.

When you are dealing with other peoples' data, you'll find that this sort of the happens a LOT. It can be very aggravating, so we need to learn to treat these things as challenging puzzles instead of hassles!

Let's replace the question marks with nothing, so that this column becomes consistent with the rest. Fortunately, DataFrame (and Series) objects have a replace() function built in, so let's use that.

```
[34]: bcd['bare_nuclei'] = bcd['bare_nuclei'].replace('?', '')
```

In the cell below, confirm that we no longer have question marks in our "bare_nuclei" column.

```
[42]: bcd['bare_nuclei'].str.find('?')
[42]: 0
            -1.0
            -1.0
      1
      2
            -1.0
      3
            -1.0
            -1.0
      694
            -1.0
      695
            -1.0
      696
            -1.0
      697
            -1.0
      698
            -1.0
      Name: bare_nuclei, Length: 699, dtype: float64
```

Note: As mentioned above, extracting columns or other subsets of data from a pandas DataFrame or Series does not create a new object but rather a new label to the existing object.

So, for example, the_IDs = bcd['patient_id'] does not make a new object, but rather creates a second label referring to the original object (consistent with the behavior of base Python).

In general, however, pandas methods (functions) do create new objects. Thus, the step of assigning the output of .replace() back to the original data frame column is necessary.

In the cell below, confirm that the output of .replace() and bcd['bare_nuclei'] have different IDs.

```
[43]: id(bcd['bare_nuclei'] )
[43]: 140289448713760
[45]: id(bcd['bare_nuclei'].replace('?', ''))
[45]: 140288943200192

And now we can convert the column to numeric values.
[46]: bcd['bare_nuclei'] = pd.to_numeric(bcd['bare_nuclei'])
```

In the cell below, check the data types of columns in bcd.

```
[49]: bcd['bare_nuclei'].dtypes
[49]: dtype('float64')
```

Okay! We have now have gotten our data somewhat into shape, meaning:

- missing data are actually missing
- columns of numeric data are numeric in type
- the column of doctor names contains only last names

So now we can explore some ways to deal with missing values.

1.5 Dealing with missing data

1.5.1 Finding missing values

Even though this dataset isn't all that large:

```
[50]: bcd.shape
```

[50]: (699, 12)

 $699~{\rm rows}$ is lot to look through "by hand" in order to find missing values.

We can test for missing values using the DataFrame.isna() method.

[51]: bcd.isna()

[51]:		patient_id	clump_thickness		ss c	ell_size	_uniform	ity	cell_shape_uniformity	\
	0	False	False					lse	False	
	1	False	False			False			False	
	2	False	False				Fa	lse	False	
	3	False		Fal	se		Fa	lse	False	
	4	False	False				Fa	lse	False	
		•••	***			•••			•••	
	694	False		Fal		False			False	
	695	False		Fal	.se			lse	False	
	696	False		Fal	se		Fa	lse	False	
	697	False		Fal	.se		Fa	lse	False	
	698	False		Fal	se		Fa	lse	False	
		marginal adh	nesion	single	en ce	ell size	bare n	uclei	bland_chromatin \	
	0	8	False		_ · F _ ·	False		False	False	
	1		False			False		False	False	
	2		False			False		False	False	
	3		False			False		False	False	
	4		False			False		False	False	
			•••			•••			•••	
	694		False			False		False	False	
	695		False			False		False	False	
	696		False			False		False	False	
	697		False			False		False	False	
	698		False			False	•	False	False	
		normal_nucle	oli m	nitoses	clas	s docto	r name			
	0	-	alse	False	False		False			
	1		alse	False			False			
	2		alse	False			False			
	3		alse	False	False		False			
	4		alse	False	False		False			
			•••			•••				
	694	Fa	alse	False	False	е	False			
	695		alse	False			False			
	696	Fa	alse	False	False	е	False			
	697	Fa	alse	False	False	е	False			
	698	Fa	alse	False	False	е	False			

[699 rows x 12 columns]

By itself, that doesn't help us much. But if we combine it with summation (remember that True values count as 1 and False counts as zero):

```
[52]:
     bcd.isna().sum()
[52]: patient_id
                                  0
      clump thickness
                                  1
      cell_size_uniformity
                                  1
      cell shape uniformity
                                  0
      marginal_adhesion
                                  0
                                  0
      single_ep_cell_size
      bare_nuclei
                                 18
                                  4
      bland_chromatin
      normal_nucleoli
                                  1
                                  0
      mitoses
      class
                                  0
      doctor_name
```

dtype: int64

Now we have the counts by variable, and can easly see that there are missing values for a few of the variables.

The "bare_nuclei" variable we dealt with earlier has the most missing values, with "bland_chromatin" coming in a distant second.

Let's check some of the rows with missing values and make sure everything else looks normal in those rows. Notice above that the output of .isna() is Boolean, so we can use it to do logical indexing.

[53]: bcd[bcd['bland chromatin'].isna()] [53]: patient_id clump_thickness cell_size_uniformity cell shape uniformity 342 814265 2.0 1.0 1 343 814911 1.0 1.0 1 359 873549 10.0 3.0 5 365 2.0 1.0 1 897172 single_ep_cell_size bare_nuclei bland_chromatin marginal_adhesion 342 1 2 1.0 NaN 2 343 1 1.0 NaN 4 359 3 7.0 NaN 365 1 2 1.0 NaN normal_nucleoli mitoses class doctor_name 342 1.0 1 benign Lee 343 1.0 1 benign Doe malignant 359 5.0 3 Doe 365 1.0 1 benign Lee

In the cell below, check the rows that have missing values for either clump thickness or cell size uniformity. Do this in one go rather than separately (remember about the element-wise or operator, "|".

```
[55]: bcd[(bcd['clump_thickness'].isna()) | (bcd['cell_size_uniformity'].isna())]
[55]:
          patient_id
                       clump_thickness
                                        cell_size_uniformity cell_shape_uniformity
             1018099
      6
                                    1.0
                                                           NaN
                                                                                     1
      12
             1041801
                                   NaN
                                                           3.0
                                                                                     3
          marginal_adhesion
                              single_ep_cell_size
                                                    bare_nuclei
                                                                  bland_chromatin
      6
                                                            10.0
                           1
                           3
      12
                                                 2
                                                             3.0
                                                                               4.0
          normal_nucleoli
                            mitoses
                                          class doctor_name
      6
                       1.0
                                         benign
                                                         Doe
                                   1
      12
                       4.0
                                     malignant
                                                      Smith
```

So far so good. It looks like the rows that have missing values just have one missing value, and everything else seems fine. But let's do check that no rows have more than one missing value.

To do this, we can sum the number of missing values across the columns (i.e. within each row), and then see what the maximum number of missing values within a row is.

```
[56]: row_na_totals = bcd.isna().sum(axis = 1)
row_na_totals.max()
```

[56]: 1

So we see that no row has more than one missing value.

In the cell below, do the above calculation in one line.

```
[57]: bcd.isna().sum(axis = 1).max()
```

[57]: 1

1.5.2 Dealing with missing values

Now that we have determined that there are missing values, we have to determine how to deal with them.

Ignoring missing values elementwise One way to handle missing values is just to ignore them. Most of the standard math and statistical functions will do that by default.

So this:

```
[58]: bcd['clump_thickness'].mean()
```

[58]: 4.416905444126074

Computes the mean clump thickness ignoring the one missing value.

We can compute the mean (again ignoring missing values) for all the numeric columns like this:

```
[59]: patient_id
                                1.071704e+06
      clump_thickness
                                4.416905e+00
      cell_size_uniformity
                                3.137536e+00
      cell_shape_uniformity
                                3.207439e+00
      marginal_adhesion
                                2.793991e+00
      single_ep_cell_size
                                3.216023e+00
      bare_nuclei
                                3.538913e+00
      bland_chromatin
                                3.447482e+00
      normal_nucleoli
                                2.868195e+00
     mitoses
                                1.589413e+00
      dtype: float64
```

That worked, but the output is a little awkward because the patient ID is being treated as a numeric variable. We can fix that by converting the patient ID variable to a string variable.

```
[61]: bcd['patient_id'] = bcd['patient_id'].astype('string')
bcd['patient_id']
```

```
[61]: 0
              1000025
      1
              1002945
      2
              1015425
      3
              1016277
      4
              1017023
               776715
      694
      695
               841769
      696
               888820
      697
               897471
      698
               897471
      Name: patient_id, Length: 699, dtype: string
```

And now the means should look a little better because we won't have the mean for the ID column in the millions>

Recompute the mean for the numeric columns in the cell below.

```
[62]: bcd.mean(numeric_only = True) # the numeric_only refers to columns, not_
       ⇔missing values
[62]: clump_thickness
                               4.416905
      cell_size_uniformity
                               3.137536
      cell_shape_uniformity
                               3.207439
      marginal_adhesion
                               2.793991
      single_ep_cell_size
                               3.216023
      bare_nuclei
                               3.538913
      bland_chromatin
                               3.447482
     normal_nucleoli
                               2.868195
     mitoses
                               1.589413
      dtype: float64
```

Removing missing values We are about to start learning how to remove missing values from our data frame, *however...*

Before we start messing around too much with the values in our data frame, let's make sure we can easily "hit the reset button" and get back to a nice starting point. To do this, we'll want to

- reload the data
- modify the column of Dr. names
- set the patient ID to type str
- remove the question marks from the bare nuclei column
- set the bare nuclei column to numeric

This is a perfect job for a function!

In the cell below, finish writing the function to reset our data frame to the desired starting point.

```
bcd['bare_nuclei'] = pd.to_numeric(bcd['bare_nuclei'])  # Set the bare_
→nuclei column to numeric

return bcd

hit_reset()
```

[102]: [102]: patient_id clump_thickness cell_size_uniformity cell_shape_uniformity 0 1000025 5.0 1.0 1 1002945 5.0 4.0 4 2 1.0 1015425 3.0 1 3 8 1016277 6.0 8.0 4 1017023 4.0 1.0 1 694 776715 3.0 1.0 1 695 841769 2.0 1.0 1 696 888820 5.0 10.0 10 697 8.0 897471 4.0 6 698 897471 4.0 8.0 8 bare_nuclei bland_chromatin marginal_adhesion single_ep_cell_size 0 2 1.0 1 5 7 1 10.0 3.0 2 2 1 2.0 3.0 3 1 3 4.0 3.0 4 3 2 1.0 3.0 694 3 2.0 1.0 1 2 695 1.0 1.0 1 696 3 7 3.0 8.0 697 4 3 4.0 10.0 5.0 698 5 4 10.0 normal_nucleoli mitoses class doctor_name 0 1.0 Doe 1 benign 1 Smith 2.0 1 benign 2 1.0 1 Lee benign 7.0 3 1 benign Smith 4 1.0 1 benign Wong 694 1.0 1 benign Lee 695 1 benign Smith 1.0 696 2 10.0 malignant Lee 697 6.0 malignant Lee 698 4.0 malignant Wong

[699 rows x 12 columns]

[80]: bcd.dtypes

```
[80]: patient id
                                 string
      clump_thickness
                                float64
      cell_size_uniformity
                                float64
      cell_shape_uniformity
                                  int64
      marginal_adhesion
                                  int64
      single_ep_cell_size
                                  int64
      bare nuclei
                                float64
      bland_chromatin
                                float64
      normal_nucleoli
                                float64
                                  int64
      mitoses
      class
                                 object
      doctor_name
                                 object
      dtype: object
```

Removing rows with missing values Obviously, rows in which all values are missing won't do us any good, so we can drop them with:

```
[81]: bcd = bcd.dropna(how = 'all')
```

This drops rows in which *all* of the values are missing. This code ran without error, but we know it also didn't do anything in this case because we don't have any rows in which all the values are missing!

Sometimes a case can be made for throwing out all observations (rows) that are incomplete, that is, if they contain *any* missing values.

```
[82]: bcd = bcd.dropna(how = 'any')
```

In the cell below, check the (new) shape of bcd.

```
[84]: bcd.shape
```

[84]: (674, 12)

It should have fewer rows now.

And now is a perfect time to test our function! In the cell below, hit the reset button on bcd.

```
[85]: hit_reset()
```

```
[85]:
           patient_id clump_thickness
                                          cell_size_uniformity cell_shape_uniformity \
      0
              1000025
                                                               1.0
      1
                                      5.0
                                                              4.0
                                                                                          4
              1002945
      2
              1015425
                                      3.0
                                                              1.0
                                                                                           1
      3
                                                                                          8
                                      6.0
                                                              8.0
              1016277
      4
              1017023
                                      4.0
                                                               1.0
                                                                                           1
      . .
      694
               776715
                                      3.0
                                                              1.0
                                                                                           1
      695
               841769
                                      2.0
                                                              1.0
                                                                                          1
      696
                                                             10.0
                                                                                          10
               888820
                                      5.0
      697
               897471
                                      4.0
                                                              8.0
                                                                                          6
      698
               897471
                                      4.0
                                                              8.0
                                                                                          8
            marginal_adhesion
                                 single_ep_cell_size
                                                         bare_nuclei
                                                                       bland_chromatin
      0
                                                                  1.0
                                                                                     3.0
                              1
                              5
                                                      7
      1
                                                                 10.0
                                                                                     3.0
      2
                              1
                                                      2
                                                                  2.0
                                                                                     3.0
      3
                              1
                                                      3
                                                                  4.0
                                                                                     3.0
      4
                              3
                                                      2
                                                                  1.0
                                                                                     3.0
      694
                              1
                                                      3
                                                                  2.0
                                                                                     1.0
      695
                                                      2
                                                                  1.0
                                                                                     1.0
                              1
      696
                              3
                                                      7
                                                                  3.0
                                                                                     8.0
      697
                              4
                                                      3
                                                                  4.0
                                                                                    10.0
      698
                              5
                                                      4
                                                                  5.0
                                                                                    10.0
            normal_nucleoli mitoses
                                             class doctor_name
      0
                         1.0
                                            benign
                                                             Doe
                                      1
                         2.0
      1
                                                           Smith
                                      1
                                            benign
      2
                         1.0
                                      1
                                            benign
                                                             Lee
      3
                         7.0
                                      1
                                            benign
                                                           Smith
      4
                         1.0
                                      1
                                            benign
                                                            Wong
      694
                         1.0
                                      1
                                            benign
                                                             Lee
      695
                         1.0
                                      1
                                            benign
                                                           Smith
      696
                        10.0
                                      2
                                         malignant
                                                             Lee
                                         malignant
      697
                         6.0
                                      1
                                                             Lee
      698
                         4.0
                                         malignant
                                                            Wong
```

[699 rows x 12 columns]

Check the shape.

```
[86]: bcd.shape
```

[86]: (674, 12)

Check the data types of the columns.

```
[87]: bcd.dtypes
[87]: patient_id
                                 string
      clump_thickness
                                float64
      cell_size_uniformity
                                float64
      cell_shape_uniformity
                                   int64
      marginal_adhesion
                                   int64
      single_ep_cell_size
                                   int64
      bare_nuclei
                                 float64
      bland_chromatin
                                 float64
      normal nucleoli
                                float64
                                   int64
      mitoses
      class
                                 object
                                 object
      doctor_name
      dtype: object
     Check the doctor name column.
[88]: bcd['doctor_name']
[88]: 0
                Doe
              Smith
      1
      2
                Lee
      3
              Smith
      4
               Wong
      694
                Lee
      695
              Smith
      696
                Lee
      697
                Lee
      698
               Wong
      Name: doctor_name, Length: 674, dtype: object
```

Removing columns with missing values And we could do the same for columns if we wished, though this is less frequently done. We just need to change the axis (direction) over which DataFrame.dropna() works.

```
[89]: bcd = bcd.dropna(axis = 1, how = 'any') # drop columns rather than rows
```

This leaves us with only the complete columns.

```
[90]: bcd.shape
```

[90]: (674, 12)

Let's see which they are.

```
[91]: bcd.columns
```

Filling in missing values Occasionally, we may want to fill in missing values. This isn't very common, but might be useful if some other function you are using doesn't handle missing values gracefully.

Before filling in missing values, we need to restore our data frame so it actually has missing values. Good thing we wrote that function!

```
[92]: bcd = hit_reset()
```

We can fill in missing values with any single value we want, such as a zero.

```
[93]: bcd = bcd.fillna(0)
```

In the cell below, check to see that we no longer have missing values.

```
[104]: bcd.isna() bcd.isna().sum()
```

```
[104]: patient_id
                                  0
       clump_thickness
                                  0
       cell_size_uniformity
                                  0
       cell_shape_uniformity
                                  0
       marginal_adhesion
                                  0
       single_ep_cell_size
                                  0
       bare_nuclei
                                  0
       bland_chromatin
                                  0
       normal_nucleoli
                                  0
       mitoses
                                  0
       class
                                  0
                                  0
       doctor_name
       dtype: int64
```

In the cell below, reset the data and verify that the missing data are back.

```
[109]: bcd = hit_reset()
```

```
[108]: bcd.isna()
       bcd.isna().sum()
[108]: patient_id
                                   0
       clump_thickness
                                   1
       cell_size_uniformity
                                   1
       cell_shape_uniformity
                                   0
       marginal_adhesion
                                   0
                                   0
       single_ep_cell_size
       bare_nuclei
                                  18
       bland chromatin
                                   4
       normal nucleoli
                                   1
                                   0
       mitoses
                                   0
       class
       doctor_name
                                   0
       dtype: int64
      In the cell below, fill the missing values in each column with the column mean. (Hint: this is
      pandas, so this is actually easy!)
[111]:
      bcd.describe()
[111]:
                                                         cell_shape_uniformity \
               clump_thickness
                                 cell_size_uniformity
                    698.000000
                                            698.000000
                                                                     699.000000
       count
       mean
                      4.416905
                                              3.137536
                                                                       3.207439
       std
                      2.817673
                                              3.052575
                                                                       2.971913
       min
                      1.000000
                                              1.000000
                                                                       1.000000
       25%
                      2.000000
                                              1.000000
                                                                       1.000000
       50%
                      4.000000
                                              1.000000
                                                                       1.000000
       75%
                      6.000000
                                              5.000000
                                                                       5.000000
                     10.000000
                                             10.000000
                                                                      10.000000
       max
               marginal adhesion
                                   single_ep_cell_size
                                                          bare nuclei
                                                                        bland chromatin
                      699.000000
                                                                              695.000000
                                             699.000000
                                                           681.000000
       count
                        2.793991
                                               3.216023
                                                             3.538913
                                                                                3.447482
       mean
       std
                        2.843163
                                               2.214300
                                                             3.639493
                                                                                2.441191
                        1.000000
                                               1.000000
                                                             1.000000
                                                                                1.000000
       min
       25%
                                                                                2.000000
                        1.000000
                                               2.000000
                                                             1.000000
       50%
                        1.000000
                                               2.000000
                                                             1.000000
                                                                                3.000000
       75%
                        3.500000
                                               4.000000
                                                             6.000000
                                                                                5.000000
                                                            10.000000
                                                                               10.000000
       max
                       10.000000
                                              10.000000
               normal nucleoli
                                    mitoses
       count
                    698.000000
                                 699.000000
                      2.868195
                                   1.589413
       mean
```

1.715078

1.000000

3.055647 1.000000

std

min

```
25% 1.000000 1.000000

50% 1.000000 1.000000

75% 4.000000 1.000000

max 10.000000 10.000000
```

And now verify that there are no more missing values.

```
[116]: bcd.isna().sum()
[116]: patient_id
                                 0
       clump_thickness
                                 0
       cell_size_uniformity
                                 0
       cell_shape_uniformity
                                  0
       marginal_adhesion
                                  0
       single_ep_cell_size
                                  0
       bare_nuclei
                                  0
       bland_chromatin
                                  0
       normal_nucleoli
                                  0
                                  0
       mitoses
       class
                                  0
                                 0
       doctor_name
       dtype: int64
```

1.6 Summary

In this tutorial, we learned or remembered how to do some of the foundational data wrangling tasks. These are:

- importing data into pandas from a data file
- cleaning up the data in the columns
- $\bullet\,$ converting columns to the appropriate type
- removing or filling in missing values