# UNIVERSIDADE FEDERAL DE JUIZ DE FORA

# Programa de Pós Graduação em Modelagem Computacional

# CGPGRN - User Guide 2.2

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## 1 Introduction

CGPGRN is a framework for inferring Gene Regulatory Networks (GRNs) using timeseries data which is the result of a Ph.D. Thesis. Initially developed for single-cell data, CGPGRN can be used for any data-type technology.

This user guide contains information about the usage of the CGPGRN framework for inferring Gene Regulatory Networks from time-series data.

### 1.1 CGPGRN pipeline

The general CGPGRN framework pipeline is presented in Figure 1.

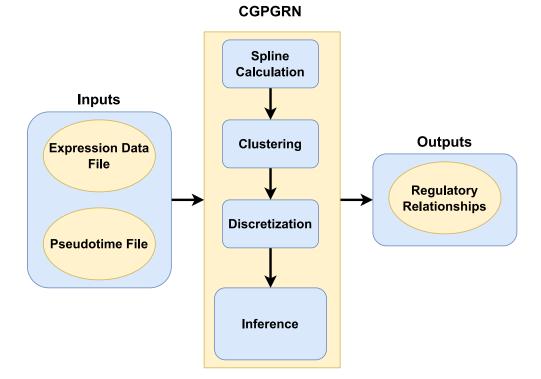


Figure 1: General CGPGRN framework pipeline.

Given expression data and pseudotime files, CGPGRN first calculates smoothing splines for each gene in order to obtain a single curve. This curve represents the expression level of the gene over time. Also, smoothing spline helps in removing or softening technical and biological variations. Then, it is possible to group the genes, according to a given similarity measure. Grouping genes helps in reducing the possible regulators for each gene. This step is called clustering. The user is able to choose between several grouping approaches and can also ignore the clustering step. After that, each gene expression is discretized in order to obtain transition state diagrams and, consequently, truth tables.

The truth tables are given to CGPGRN inference algorithm to evolve digital circuits. The evolved digital circuits represent the interactions (activation and inhibition) between genes. It is possible to choose between evolving one circuit for each gene or evolving the entire gene regulatory network depending on the selected clustering approach (KMeans, Agglomerative, and No Clustering). For the other clustering approaches, one circuit is evolved for each gene. Finally, the evolved circuits are analyzed to obtain the regulators for each target gene. The final results are presented in the form of a table with the regulator, target gene, and interaction strength of each regulatory relationship.

#### 1.2 What's New?

The complete Change Log can be found at https://github.com/jeduaardo/cgpgrn.

- New general requirements: G++ Compiler, Make and Bash.
- added support for multiple outputs and don't care recognizing;
- -kd default changed to False
- -run default changed to False
- added -gsl argument for automatically generating spline list files (default = False)
- added makeFile.py in includes for automatically generating Makefile
- Added CGPGRN inference algorithm parametrization:
  - -cgpn defines the number of nodes
  - cgpg defines the number of generations
- added timeStatistics for counting time in the entire framework
  - inference (mean and total)
  - complete framework (total)
- Minor fixes and more information on Log
- added mkdir function in utils
- all functions are now documented in utils

- noClustering now can be used with -fullTT
- fullBashScript is now generated in utils
  - automatically defines the number of nodes and generations of CGPGRN inference algorithm. However, the user still can define
- objects added for performing the CGPGRN framework main steps
  - spline
  - clustering
  - discretization

#### 1.3 CGPGRN structure

The folder and files structure of CGPGRN framework is presented in Figure 2a. Also, the source directory tree view is presented in Figure 2b.

The main path contains 3 files and 3 directories, as presented in Table 1. The description of the files in the folder *include* is presented in Table 2.

Name	Type	Description	
completePreProcessing.py	File	Main file of the CGPGRN framework	
${\bf ExpressionData\_mCAD.csv}$	File	Input dataset	
${\bf PseudoTime\_mCAD}$	File	Input pseudotime file	
include	Dir	folder with all scripts used with CGPGRN framework	
kernels	Dir	folder with OpenCL kernels for CGP inference procedure	
source	Dir	folder with CGPGRN inference algorithm source code	

Table 1: Files and folders description of the main path of CGPGRN.

Figure 2: CGPGRN structure.

```
CGPGRN.py
ExpressionData_mCAD.csv
PseudoTime_mCAD.csv
include/
    clusterData.py
    discretizeData.py
    generateOutputs.py
    makeFile.py
    objects.py
    postProcess.py
    preProcessing.py
    utils.py
    requirements/
        getRequirements.sh
        requirements.txt
kernels/
    evol.cl
    kernel.cl
    utils.cl
source/
```

(a) Main (root) directory structure.

```
cgp.cpp
cgp.h
constants.h
GPTime.cpp
GPTime.h
main.cpp
OCLConfig.cpp
OCLConfig.h
prog.exe
stack.cpp
stack.h
utils.cpp
utils.h
amd/
    include/
         CL/
              cl.h
              cl.hpp
cl_d3d10.h
              cl_d3d11.h
              cl_dx9_media_sharing.h
              cl_ext.h
              cl_gl.h
cl_gl_ext.h
              cl_platform.h
              opencl.h
    lib/
         x86/
              opencl.lib
         x86_64/
              opencl.lib
kernels/
     evol.cl
     kernel.cl
     utils.cl
```

(b) Source directory structure.

Name	Description		
clusterData.py	python script for clustering the data		
discretizeData.py	python script for discretizing the data		
generateOutputs.py	python script for generating the truth tables		
makeFile.py	python script for generating the make file		
objects.py	python script contaning the main objects of CGPGRN framework		
postProcess.py	python script for post processing the data		
utils.py	python script with utilities		
requirements/get Requirements. sh	bash script for get/update the CGPGRN's required python libraries		
requirements/requirements.txt	txt file with the name and version of python's required libraries		

Table 2: Files in the folder include.

## 2 CGPGRN Usage

In the following subsections, the CGPGRN framework usage is explained. Furthermore, Section 4 presents recommendations of the arguments and parameters to be used with CGPGRN framework.

### 2.1 Input Datasets

The expression data must be in *csv* format with genes as rows and cell IDs as columns. An example of a simple expression data file is presented in Figure 3. This file example is also present in the main path of CGPGRN framework as an example (Expression-Data mCAD.csv).

Figure 3: Expression Data file example. The file must contain genes as rows and cell IDs as columns.

	E950_416	E388_201	E984_309	 E1686_109	E571_69	E78_108
Coup	0.074369	0.109152	0.002359	0.159354	0.084578	0.051027
Emx2	0.000387	0.043068	0.014319	0.044069	0.013422	0.002859
Fgf8	1.226758	1.361096	2.009039	0.708046	0.017840	1.263805
Рахб	2.198534	1.503729	1.648089	1.463105	0.051140	1.765044
Sp8	2.306293	1.077755	2.273083	1.278887	2.510019	0.955908

For Pseudotime files, csv format is also needed and columns are pseudotimes and rows, cell IDs. Figure 4 shows examples of pseudotime files with one pseudotime (a) and two pseudotimes (b). The PseudoTime\_mCAD.csv present in the main path of CGPGRN framework is an example of pseudotime file with two pseudotimes.

Figure 4: Pseudotime file examples. Rows are cell IDs and columns are pseudotimes. NaN values are empty comma-separated fields.

	PseudoTime
H9_00hb4s_001	0.029945
H9_00hb4s_002	0.015237
H9_00hb4s_003	0.064590
H9_00hb4s_004	0.025702
H9_00hb4s_005	0.084345
H9_96h_187	0.243298
H9_96h_188	0.236680
H9_96h_189	0.233339
H9_96h_190	0.252850
H9_96h_192	0.244976

	PseudoTime1	<b>PseudoTime2</b>
E950_416	0.83333	NaN
E388_201	0.40161	NaN
E984_309	0.61847	NaN
E408_137	NaN	0.27309
E181_67	NaN	0.13253
E942_205	NaN	0.40964
E1770_131	0.26104	NaN
E1686_109	NaN	0.21687
E571_69	NaN	0.13655
E78_108	0.21486	NaN

(a) Pseudotime file example with one pseudo-(b) Pseudotime file example with two pseudotime.

## 2.2 Arguments

All possible arguments for CGPGRN framework are presented in Table 3. Further explanation about some arguments is presented in the following subsections.

For example, to perform CGPGRN framework on a dataset with expression data file ExpressionData.csv, pseudotime file PseudoTime.csv using KMeans as clustering method testing number of clusters at a maximum of 10. The problem name is mCAD and the suffix is 500cells. The discretization is made through BiKMeans and CGPGRN inference algorithm will perform 5 independent runs.

python CGPGRN.py -e ExpressionData.csv -p PseudoTime.csv -cm KMeans -nc 10 -pn mCAD -s 500cells -d BiKMeans -r 5 -run

Argument	Description
-cm	The cluster method used for grouping genes. (See Section 2.3)
-nc	The number of maximum clusters to be tested within -cm. (See Section 2.3)
-t	The minimum correlation coefficient to be used within -cm. (See Section 2.3)
-е	Expression data file.
-p	PseudoTime file.
-pn	Problem Name (string).
-S	Suffix: an identificator of the current problem. (e.g. 2000cells)
-d	Discretization approach used for discretizing spline data. (See Section 2.3.1)
-r	Number of independent runs of CGPGRN inference algorithm.
-run	Run or not the CGPGRN inference algorithm (default $=$ True).
-kd	Keep or not the source and spline data (default $=$ True).
-fd	To use or not full discretization. (See Section 2.3.1)
-sf	To use or not a user given spline data file. (See Section 2.3.2)
-sl	Identificator for multiple given spline data files. (See Section 2.3.2)
-gsl	Generate spline list file. (See Section 2.3.2)
-fullTT	To use full truth table (one GRN for all genes). (See Section 2.3).
-cgpn	Number of CGPGRN inference algorithm nodes. (See Section 2.4)
-cgpg	Number of maximum generations for CGPGRN inference algorithm. (See Section 2.4)

Table 3: All possible arguments for CGPGRN framework.

## 2.3 Clustering Methods

All implemented clustering methods are present in include/clusterData.py script. The available clustering approaches are presented in Table 4. Note that correlation methods require -t argument and clustering methods require -cm argument. Furthermore, when considering clustering methods, the maximum number of clusters to be tested (-nc) must be lower than the total number of genes present in the expression data file. If -nc is greater than the number of genes, CGPGRN framework will inform you.

It is possible to generate one GRN containing all genes or several partial GRNs for each gene and the final one is obtained by merging the partial GRNs. When using KMeans, Agglomerative, or None, one can generate one GRN, using -fullTT. If -fullTT is not considered, the partial GRNs are generated, and then, the final merged GRN. However

Method	Type	Description
Spearman	Correlation	Perform spearman correlation between the expression data of genes.
Pearson	Correlation	Perform pearson correlation between the expression data of genes.
KendallTau	Correlation	Perform kendall tau correlation between the expression data of genes.
KMeans	Clustering	Perform kmeans between the expression data of genes.
Agglomerative	Clustering	Perform agglomerative clustering between the expression data of genes.
None	-	Do not use grouping genes step. Default value.

Table 4: Possible values for -cm argument.

only partial GRNs a final one obtained by merging these partial GRNs is possible for all other clustering approaches.

#### 2.3.1 Discretization Approaches

All implemented discretization approaches are present in include/discretizeData.py script. The available discretization approaches are presented in Table 5. For further information about each discretization approach, we refer to [1].

Method	Discretization
Mean	The expression data are divided into two groups based on the mean of expression data.
Median	The expression data are divided into two groups based on the median of expression data.
EFD	The expression data is divided into two groups with the same number of data points.
TSD	Discretization based on the difference between successive time points.
BiKMeans	KMeans is performed on both rows and columns to determine the discrete value.

Table 5: Possible values for -d argument.

Note: The CGPGRN framework generates several files for each chosen clustering method. Some of them, such as correlation ones, generate one file per gene, containing the possible regulators for a given target gene. The discretization can be performed only on the spline files generated for each pseudotime or one can discretize every spline file per gene (computationally expensive). If you want to discretize every spline file you must use -fd argument.

#### 2.3.2 Spline Files

Spline files are generated through the pre-processing step of CGPGRN framework. However, you can give the framework your own spline files or store the spline files generated once to reuse them using different parameters.

If you have only one spline file, you must use -sf and pass the spline file name as an argument. Example:

python CGPGRN.py -cm Pearson -t 0.7 -e ExpressionData.csv -p
PseudoTime.csv -pn ProblemName -s Suffix -d TSD -r 5 -sf splineFile.csv
-run

However, if you want to use several spline files, for example when having multiple pseudotimes, you must pass a txt file containing one spline filename per line to -sf and use -sl to indicate that you are using a list of spline files. Example:

python CGPGRN.py -cm Pearson -t 0.7 -e ExpressionData.csv -p
PseudoTime.csv -pn ProblemName -s Suffix -d TSD -r 5 -sf
splineFileNames.txt -sl -run

Furthermore, this spline list file can be automatically generated if you use -gsl. Example:

python CGPGRN.py -cm KendallTau -t 0.7 -e ExpressionData.csv -p PseudoTime.csv -pn ProblemName -s Suffix -d BiKMeans -r 5 -run -gsl

Note: It is important to notice that spline files must be at the same format as expression data files. Furthermore, if you have multiple pseudotimes, each given spline file must have an indentificator of pseudotime, separated by an underscore (e.g. splineData pt0.csv, splineData pt1.csv).

#### 2.4 CGP

CGPGRN framework uses Cartesian Genetic Programming (CGP) as evolutionary search technique for evolving combinational logic circuits. In CGP, programs are represented by a directed acyclic graph, encoded by a matrix of processing nodes. Each node contains

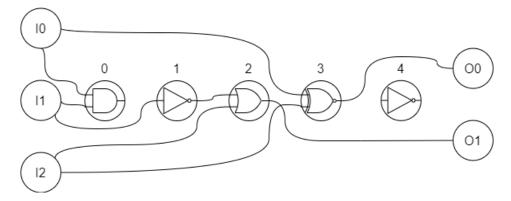


Figure 5: Representation of CGP individual with 3 inputs (I0, I1, and I2), 5 nodes (0, 1, 2, 3 and 4), and 2 outputs (O0 and O1).

inputs and a function that is performed on its inputs. Here, functions are boolean ones. A CGP's individual is presented in Figure 5.

Furthermore, a common stop criterion for evolutionary algorithms is a given maximum number of generations or the number of fitness function evaluations.

These parameters can be defined in CGPGRN framework by setting -cgpn for the number of nodes and -cgpg for the number of generations.

Note: If the user doesn't give these arguments values, CGPGRN framework will automatically define these parameters.

## 3 Post Processing

CGPGRN framework performs a post-processing step on the inferred GRNs. In post process, three main steps occur:

- Unify the different pseudotime and independent runs ranked edges,
- Regulatory relationships duplicate removal, and
- Generate output folder for the current problem containing all files generated during the CGPGRN framework operation and cleaning the main path.

Ranked edges are files containing the strength of each regulatory relationship (Gene2 is regulated by Gene1 with the strength of EdgeWeight). This standard structure is used on [2] for comparing different GRN inference algorithms.

All results are stored in the folder named finalResults\_problemName. If the folder already exists, a new folder with an index (e.g. finalResults\_problemName\_1) is created. The structure of directories in this folder is as follows:

- discretized Data: contains all discretized data files,
- executions\_parallel: contains all evolved circuits from CGPGRN inference procedure,
- kmeans, pearson, spearman, kendalltau: one of these names (-cm) is created an contains the spline files for each clustering approach,
- shellScripts: contains all the shell scripts used within CGPGRN framework,
- sourceData: contains the source expression data and pseudotime files,
- splineData: contains the general spline files generated from source expression data and pseudotime files,
- time\_counting: contains information about the spent time for evolving circuits in CGPGRN inference procedure,
- CGPGRN\_parameters.txt: log file containing information of the parameters used for inferring the GRN,
- CGPGRN\_times.txt: log file containing information of the time spent per step of the framework,
- LogFile\_PROBLEMNAME.txt: log file of the steps performed by the framework,
- rankedEdges\_problemName.csv: the final file containing the inferred regulatory relationships, and
- unified\_rankedEdges\_problemName.csv: a file containing the unified ranked edges inferred for each pseudotime and independent runs.

### 4 Recommendations

From all experiments performed using CGPGRN framework, we recommend the following parameters:

- -cm KMeans
- -nc 10
- -d BiKMeans
- -fullTT
- -r 1

Note that -nc 10 requires at least 11 genes. Using -fullTT, CGPGRN framework will generate one GRN containing all genes. Also, -fullTT automatically calculates the best parameters for CGP inference algorithm (number of nodes and generations). As standard, CGPGRN inference algorithm is run 5 times on each gene. For this reason, one can consider using -r 1. If you want to generate more results to use statistical tests, -r 5 is a good choice. Then, the full command line is given as:

python CGPGRN.py -cm KMeans -nc 10 -e ExpressionData.csv -p PseudoTime.csv -pn ProblemName -s Suffix -d BiKMeans -r 1 -fullTT -run

#### 5 Installation Guide

CGPGRN framework uses several python libraries, presented in Table 6. All these requirements can be found in <code>include/requirements/requirements.txt</code> and can be automatically installed using the getRequirements.sh bash script, present in the same folder. If you don't know how to run bash scripts on Windows, see Section 5.1.1. In Linux systems, usually, the third version of python is accessible by calling python3. For this reason, if your python is python3 in PATH, you must edit <code>getRequirements.sh</code>, replacing python with python3.

Furthermore, the CGPGRN inference method uses GPU OpenCL  $\geq 1.2$ . We recommend a GPU with 2GB+ of dedicated video memory.

Also, pandas==1.5.2 requires python-dateutils $\geq 2.8.1$ , which is automatically upgraded with the bash script.

The following sections explain in detail how to meet all the requirements for using CGPGRN framework.

Library	Version
csaps	1.1.0
matplotlib	3.6.2
numpy	1.23.5
pandas	1.5.2
psutil	5.9.4
$scikit_learn$	1.2.0
scipy	1.10.0

Table 6: CGPGRN python libraries requirements.

#### 5.1 Windows

This section gives information about how to meet the requirements for running CGPGRN on Windows.

#### 5.1.1 Bash

We recommend using Git Bash once you can use Git tools for cloning the CGPGRN repository. All Git Tools for Windows can be downloaded through the following link:

#### 5.1.2 G++ Compiler

For compiling the CGPGRN inference procedure you must have G++ compiler. For Windows, we recommend MinGW. You can use MSYS2, downloadable via:

#### 5.1.3 Make

For automatically compiling modifications of CGPGRN inference algorithm, CGPGRN framework uses make. For Windows, we recommend using Chocolatey. It can be downloaded through the link:

Once you have chocolatey installed, you must install make. For install make, open your command prompt and type:

#### choco install make

#### 5.1.4 OpenCL

On Windows, you must download and install OpenCL, available in:

https://github.com/GPUOpen-LibrariesAndSDKs/OCL-SDK/releases

#### 5.2 Linux

If you are using NVIDIA GPU, access https://developer.nvidia.com/cuda-downloads?target\_os=Linux and properly select your Linux configurations. After selecting everything, the website will return to you the prompt commands needed to install the CUDA Toolkit. An example of Linux with architecture x86\_64, using Ubuntu 22.04 to be installed with deb(network) is shown in Figure 6.

Figure 6: Example of the installation of CUDA Toolkit on Ubuntu 22.04 x86\_64 deb(network).

**Note:** When using -y you are accepting every prompt command question. It is highly recommended to not install drivers directly on the kernel because it can result in operating system initialization problems.

After that, just type the following command in your prompt to install nvidia open-cl. sudo apt-get install nvidia-opencl-dev For OpenCL on AMD processors and GPUs, type the following command in your prompt.

sudo apt-get install amd-opencl-dev

For more information, we refer to https://subscription.packtpub.com/book/app lication-development/9781849692342/1/ch01lvl1sec12/an-example-of-opencl-p rogram.

To verify if OpenCL was properly installed, type *clinfo* in your prompt command. It will list your OpenCL available devices. If you don't have *clinfo* install it by typing the following command in your command.

sudo apt install clinfo

**Note:** if your *clinfo* still returns 0 as available devices, try to restart your computer. If you further want to edit and compile the source code, you need to install the OpenCL headers by typing the following command in your prompt:

sudo apt-get install opencl-headers

## 6 Problem Solving

This section provides detailed information on problem-solving when running the CGP-GRN framework.

#### 6.1 ModuleNotFoundError

If your prompt return a value similar to that one presented in Figure 7, verify if your *include* folder is present in the main directory of CGPGRN.

Figure 7: An occurance of ModuleNotFoundError.

ModuleNotFoundError: No module named 'preProcessing'

This error commonly presents the name of the not found module. The *include* folder must contain all the scripts presented in Table 7.

File Name

clusterData.py
discretizeData.py
generateOutputs.py
makeFile.py
objects.py
postProcess.py
preProcessing.py
utils.py

Table 7: Files that must be in *include* folder.

#### 6.2 INVALID KERNEL

If your prompt return a value similar to that one presented in Figure 8, verify if your kernels folder is present in the main directory of CGPGRN.

Figure 8: An occurance of CL INVALID KERNEL error.

### CL\_INVALID\_KERNEL

This error refers to not finding the OpenCL kernel. The *kernels* folder must contain all the files presented in Table 8.

File Name
evol.cl
kernel.cl
utils.cl

Table 8: Files that must be in kernels folder.

## References

[1] C. A. Gallo, R. L. Cecchini, J. A. Carballido, S. Micheletto, and I. Ponzoni, "Discretization of gene expression data revised," *Briefings in bioinformatics*, vol. 17, no. 5, pp. 758–770, 2016.

[2] A. Pratapa, A. P. Jalihal, J. N. Law, A. Bharadwaj, and T. Murali, "Benchmarking algorithms for gene regulatory network inference from single-cell transcriptomic data," *Nature methods*, vol. 17, no. 2, pp. 147–154, 2020.