

**CHEM / BCMB 4190/6190/8189**

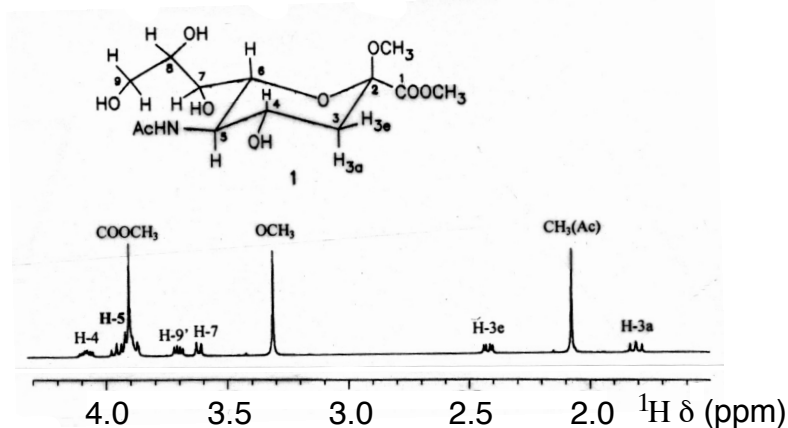
**Introductory NMR**

**Lecture 15**

## Introduction to 2D NMR

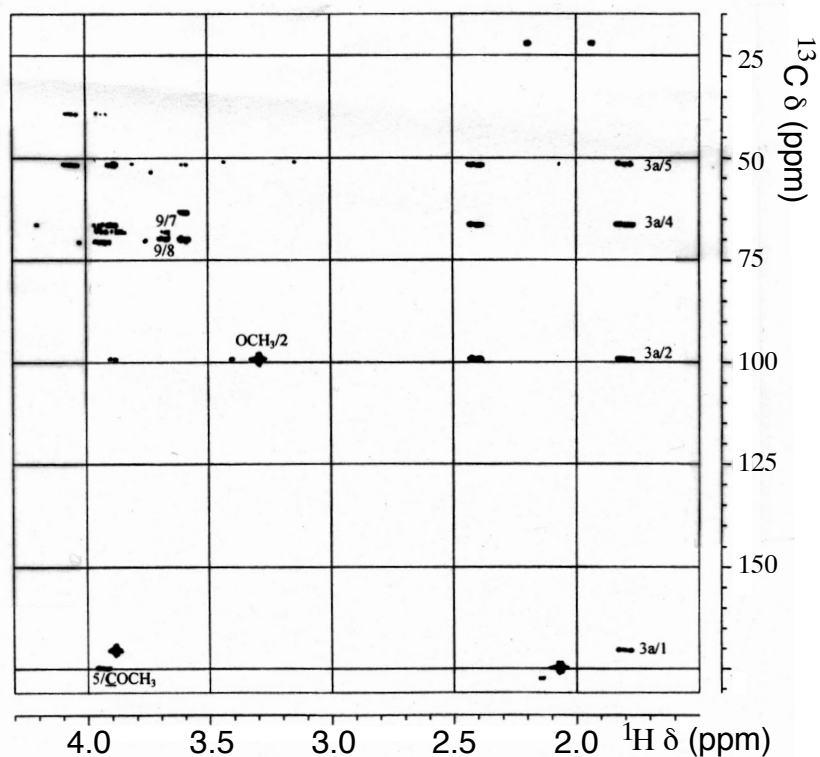
### 1D NMR spectrum:

- x axis is the directly detected frequency and y axis gives the intensity
- problem of overlap of the detected frequencies; defining connections between spins using selective excitation may be impossible



### 2D NMR spectrum:

- x and y are frequency axes and the intensity constitutes the z axis



### **2D J-resolved spectrum:**

- **x axis is the directly detected frequency**
- **y axis is the coupling constant**
- **resolves problems of overlap in 1D spectrum**

### **2D (shift) correlated NMR spectrum:**

- **x and y are frequency axes ( $^1\text{H} - ^1\text{H}$ ,  $^1\text{H} - ^{13}\text{C}$ ,  $^1\text{H} - ^{15}\text{N}$ , etc.)**
- **defines connections between nuclear spins**

### **3D NMR spectrum:**

- **2D NMR is good for M.W. up to 10-12 kD in studies of proteins: more dimensions increased the resolution for studies of larger systems.**
- **x, y, and z are frequency axes**
- **the intensity constitutes the fourth dimension**
- **for sensitivity reason,  $^1\text{H}$  is usually the detected frequency ( $\epsilon \propto \gamma^3$ )**

### **Connections between two dimensions:**

#### **1. Through-bond correlation:**

- **based of J coupling between nuclear dipoles**
- **gives information on the covalent structure (torsion angle constraint)**
- **Example: COSY, HETCOR, TOCSY, HSQC**

#### **2. Through-space correlation:**

- **based on dipolar interactions (NOE; nuclear Overhauser effect)**
- **gives information on the geometrical structure (distance constraint)**
- **Example: NOESY**

#### **3. Chemical exchange correlation:**

- **based on dynamic processes**
- **gives info on these dynamic processes**
- **Example: EXSY**

## General Experimental Scheme:

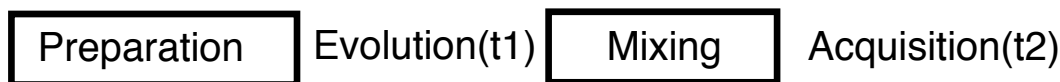
### In 1D NMR:



**PREPARATION period:** a single pulse in the most simple case and multiple pulses and delays in more complex pulse sequences (Example: J-modulated spin-echo experiment, INEPT, DEPT)

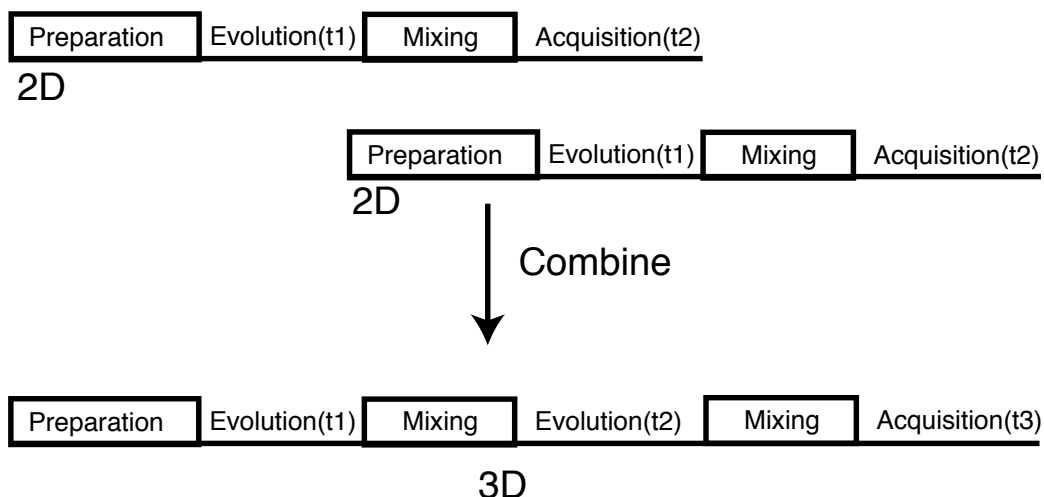
### In 2D NMR:

Two new elements, the **EVOLUTION** and **MIXING** periods are introduced between the preparation and the acquisition period. The **EVOLUTION** period introduces an indirectly-detected frequency dimension. During the **MIXING** period coherence is transferred from one spin to another.

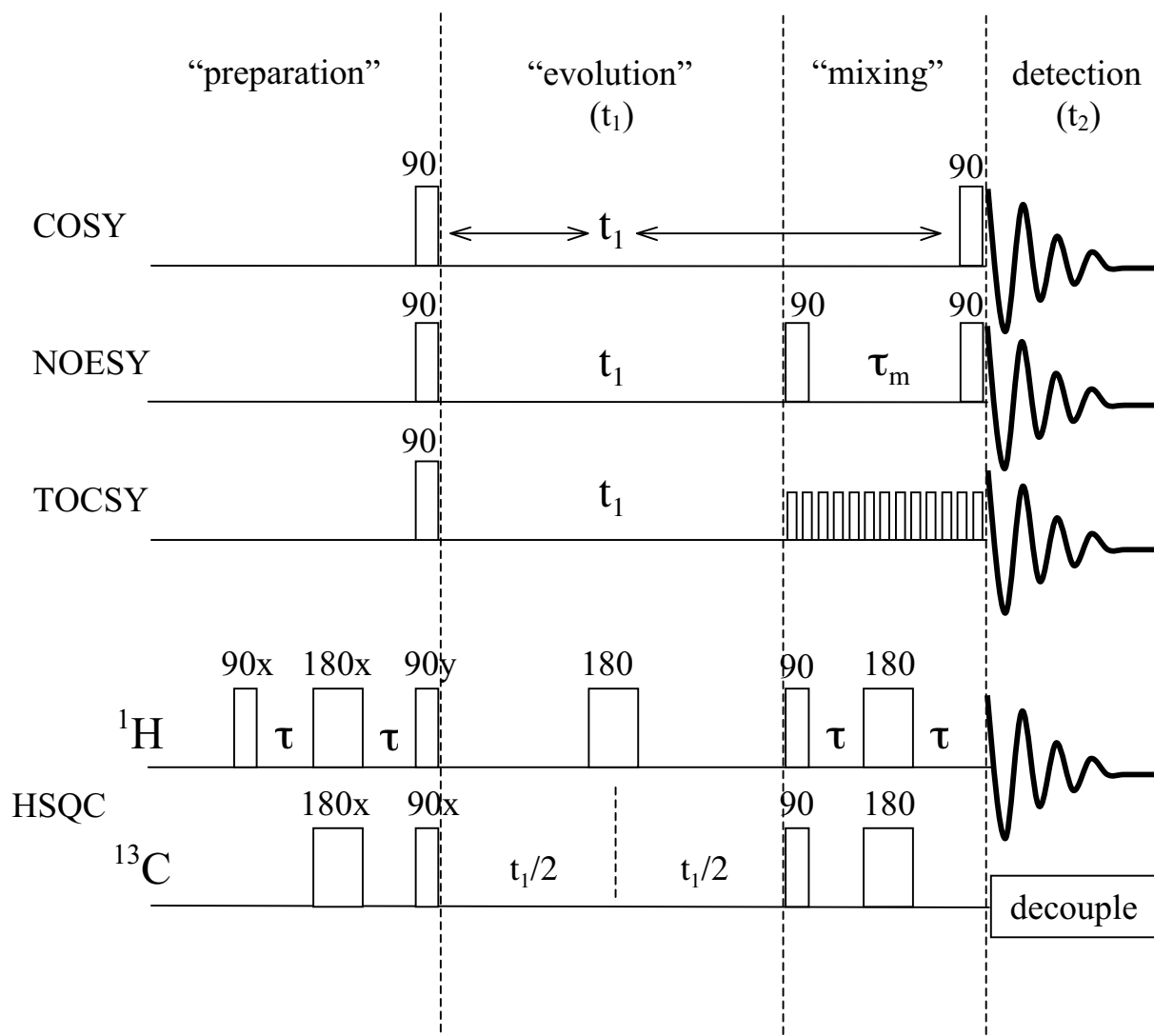


### In 3D NMR:

**Combination of 2D NMR experiments.**



**General Scheme for Two-Dimensional NMR:  
selected 2D experiments**



**COSY: Correlated Spectroscopy**

**NOESY: Nuclear Overhauser Effect Spectroscopy**

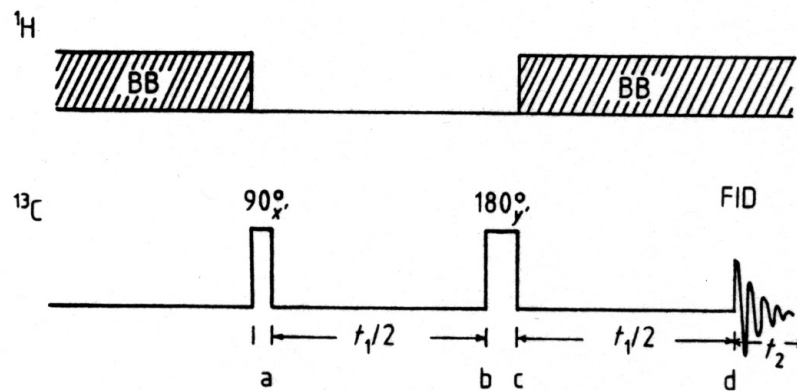
**TOCSY: Total Correlation Spectroscopy**

**HSQC: Heteronuclear Single Quantum Correlation**

## Heteronuclear Two-Dimensional J-Resolved NMR Spectroscopy::

### A) Pulse sequence

- Experiment similar to the J-modulated spin-echo pulse-sequence:  
The fixed delay  $\tau$  is now a variable delay  $t_1/2$



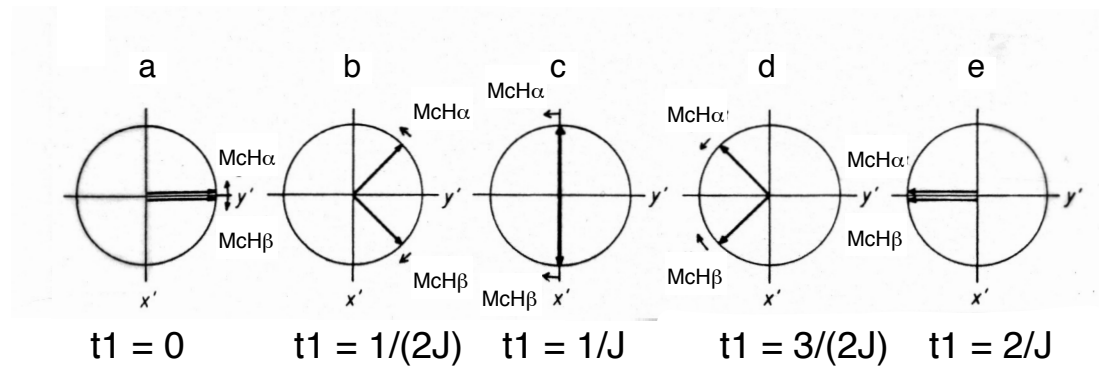
### B) Vector diagram

Lets consider a two-spin AX system with  $A = {}^1\text{H}$  and  $X = {}^{13}\text{C}$  as in  ${}^{13}\text{CHCl}_3$

$$\nu({}^{13}\text{CH}\alpha\text{Cl}_3) = \nu_c - 1/2 * J_{\text{CH}}$$

$$\nu({}^{13}\text{CH}\beta\text{Cl}_3) = \nu_c + 1/2 * J_{\text{CH}}$$

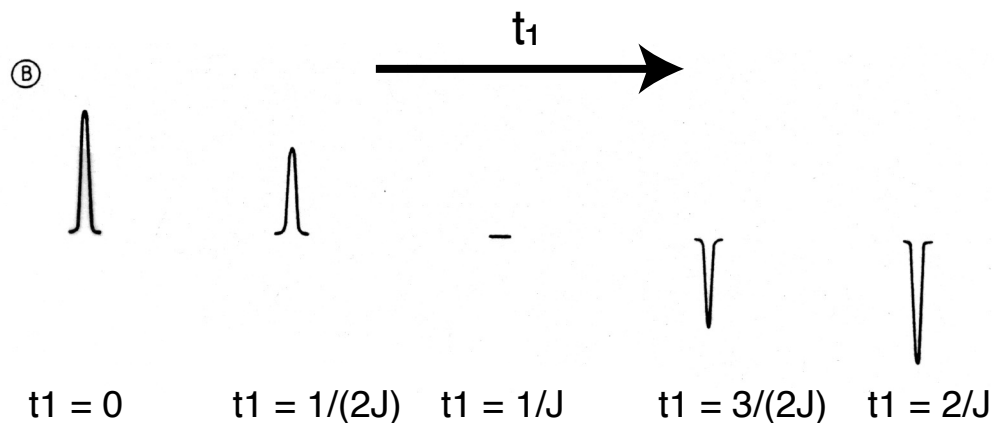
$$\Theta = 2\pi J_{\text{CH}} * (t_1/2)$$



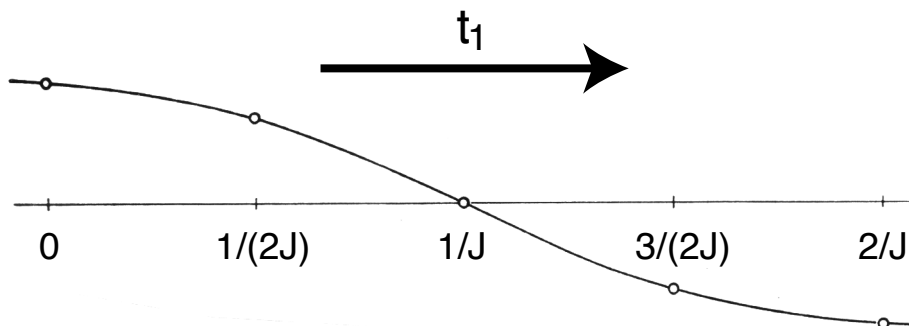
- J coupling evolves during the first  $t_1/2$  delay only
- Field inhomogeneity is refocused by the  $180^\circ y'$  pulse
- Chemical shift evolution in  $t_1$  is refocused by the  $180^\circ y'$  pulse
- For simplicity, ignore relaxation

### C) Fourier transform in $t_2$

We perform "n" experiments with various values of  $t_1$  and Fourier transform all "n" experiments

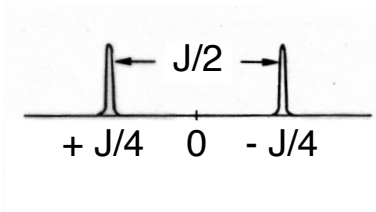


### D) Amplitude modulation in $t_1$



The amplitude is modulated in  $t_1$  at a frequency related to  $JCH$

### E) Second Fourier transform in $t_1$



A second Fourier transform with respect to  $t_1$  yields two frequencies with a separation of  $JCH/2$

## F) Summary

$$S(t_1, t_2) \xrightarrow{FT} S(t_1, F_2) \xrightarrow{FT} S(F_1, F_2)$$

**F2 dimension contains the  $^{13}\text{C}$  chemical shift**

**F1 dimension contains the coupling constant**

**For  $^{13}\text{CHCl}_3$ :**

- The signal induced in the receiver depends on the sum of the two vectors, which depends on JCH. After FT of the FID with respect to  $t_2$ , a single  $^{13}\text{C}$  signal is observed in the frequency spectrum F2 (no splitting).
- The  $^{13}\text{C}$  signal is modulated by the value of JCH and the value of  $t_1$ . After FT of the FID with respect to  $t_1$ , a doublet centered at zero frequency is observed in the frequency spectrum F1 with a splitting of  $J\text{CH} / 2$ .

**For more complex molecules:**

- In F2: Cq, CH, CH<sub>2</sub>, and CH<sub>3</sub> are all singlets
- In F1: Cq: singlet                      CH<sub>2</sub>: triplet  
CH: doublet                      CH<sub>3</sub>: quartet

## G) Choice of acquisition time in $t_1$

- Should be long enough to give satisfactory resolution in F1

$$\text{Digital resolution} = 1 / (\text{acquisition time in } t_1)$$

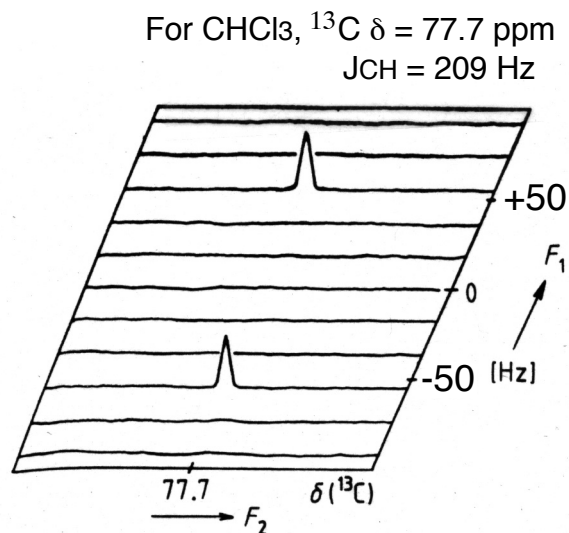
- Also limited by relaxation



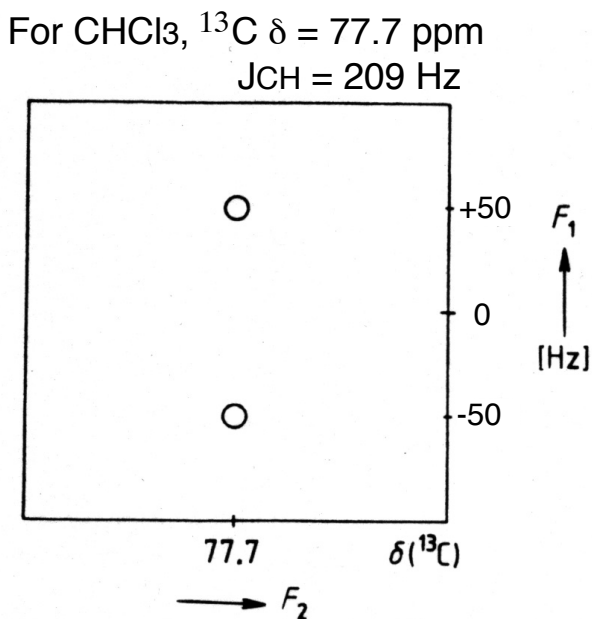
## Graphical Representation:

Two forms of display are generally used:

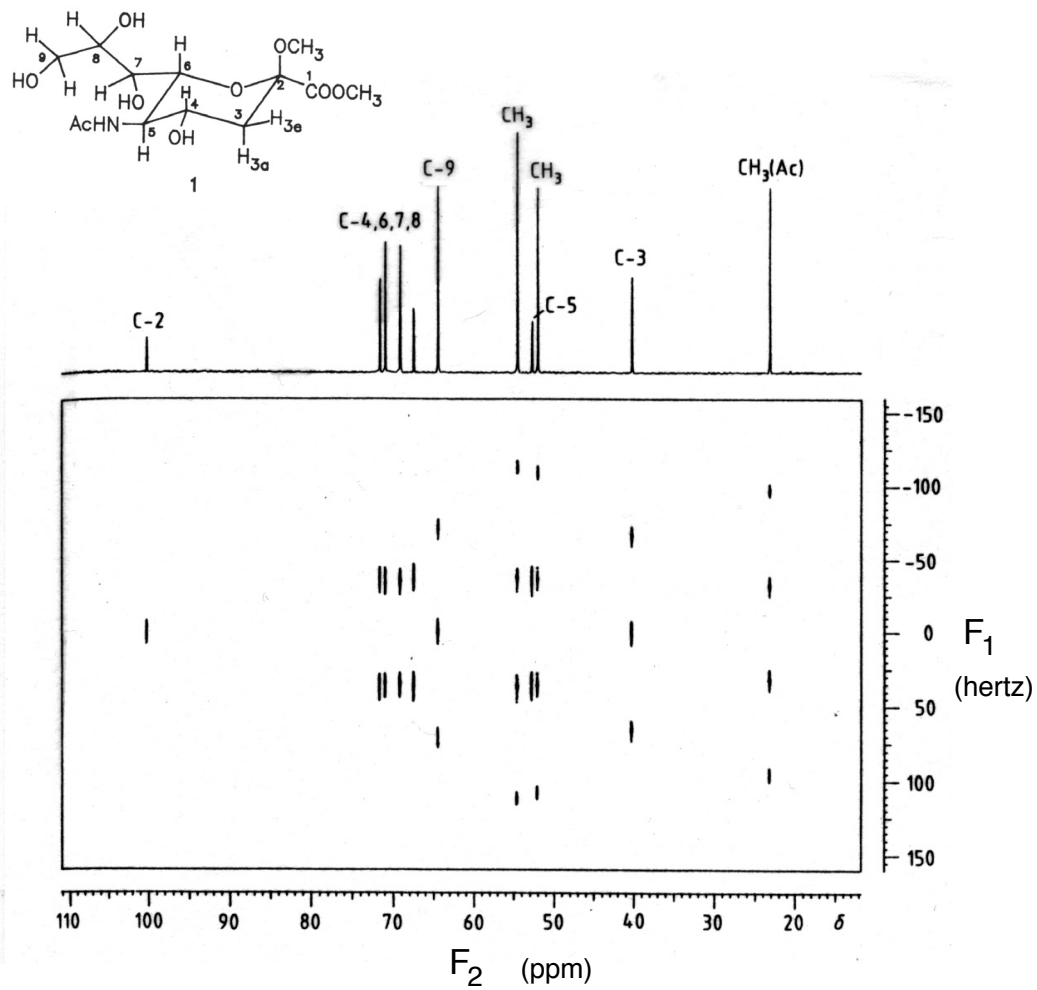
**Stacked plot:** a series of F2 spectra for different F1 values are plotted above one another. Each trace is shifted by a constant amount relative to the preceding one.



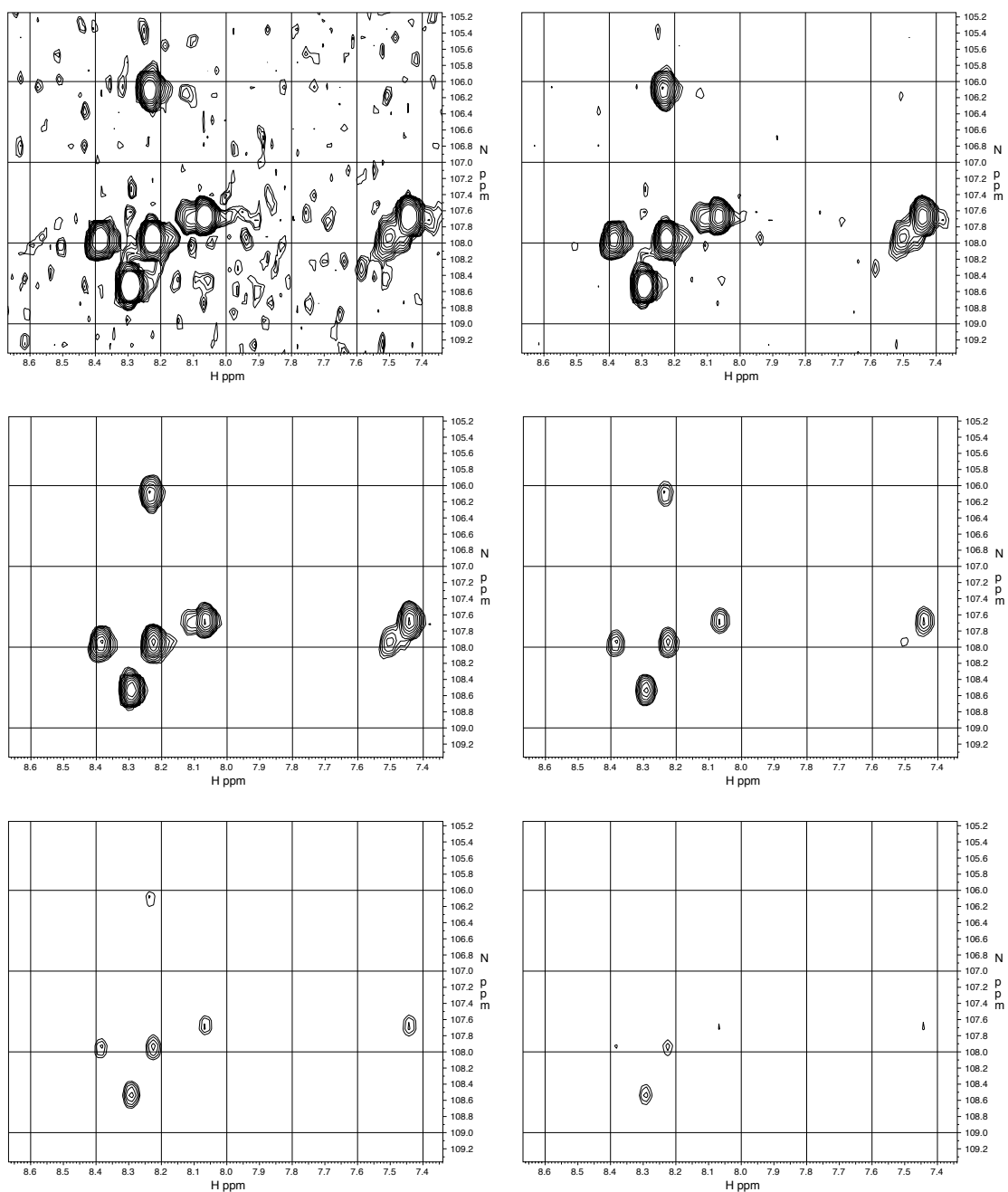
**Contour plot:** the peaks are seen from above as in a topographic map. The section is taken at certain height above the plane of the F1 and F2 axes, and the contours at that level are plotted



For more complex molecules



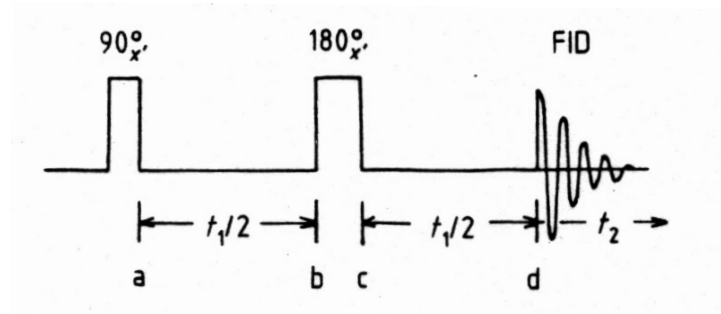
**Other example of contour plots: 2D  $^1\text{H}$ - $^{15}\text{N}$  correlation spectrum (HSQC) plotted at various levels**



## Homonuclear Two-Dimensional J-Resolved NMR Spectroscopy:

### A) Pulse sequence

- Experiment similar to the J-modulated spin-echo pulse sequence:  
The fixed delay  $\tau$  is now a variable delay  $t_1$
- In homonuclear case, not possible to use BB decoupling
- The  $180^\circ_{x'}$  pulse is applied to both spins (A and X)



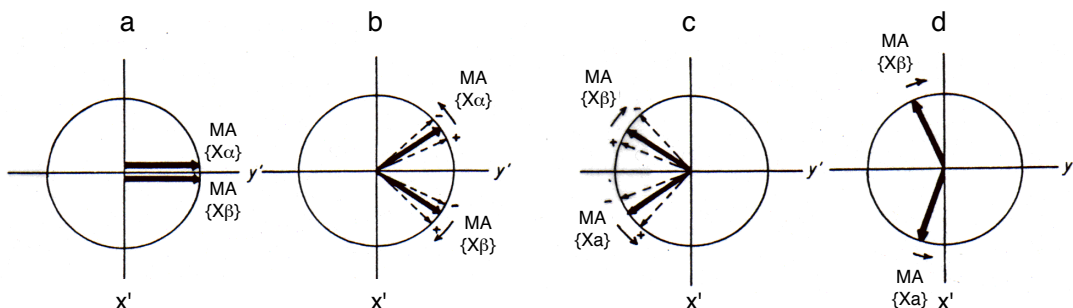
### B) Vector diagram

Lets consider a homonuclear two-spin AX system:

$$\nu_A (X\alpha) = \nu_A - 1/2 * J_{AX}$$

$$\nu_A (X\beta) = \nu_A + 1/2 * J_{AX}$$

$$\Theta = 2\pi J_{AX} * t_1$$



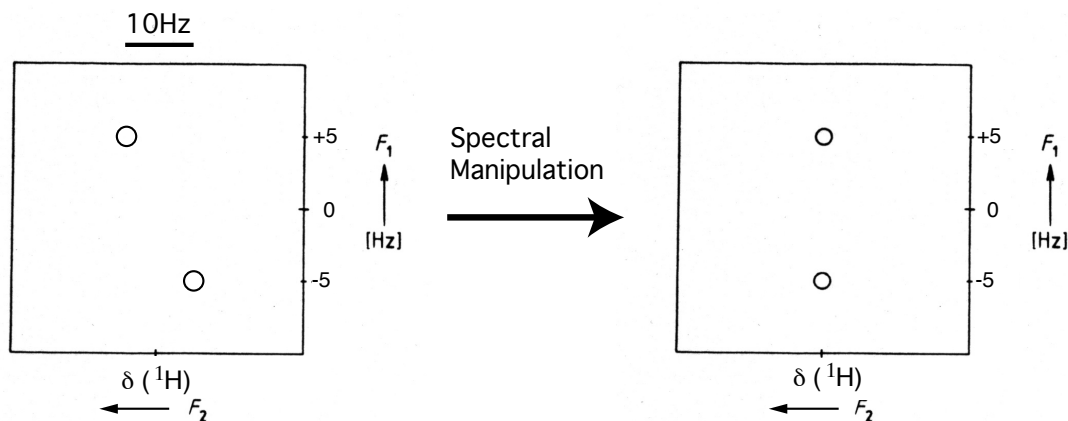
- J coupling evolves during both  $t_1/2$  delays
- Field inhomogeneity is refocused by the  $180^\circ_{x'}$  pulse
- Chemical shift evolution in  $t_1$  is refocused by the  $180^\circ_{x'}$  pulse
- For simplicity, ignore relaxation

## B) Contour plot

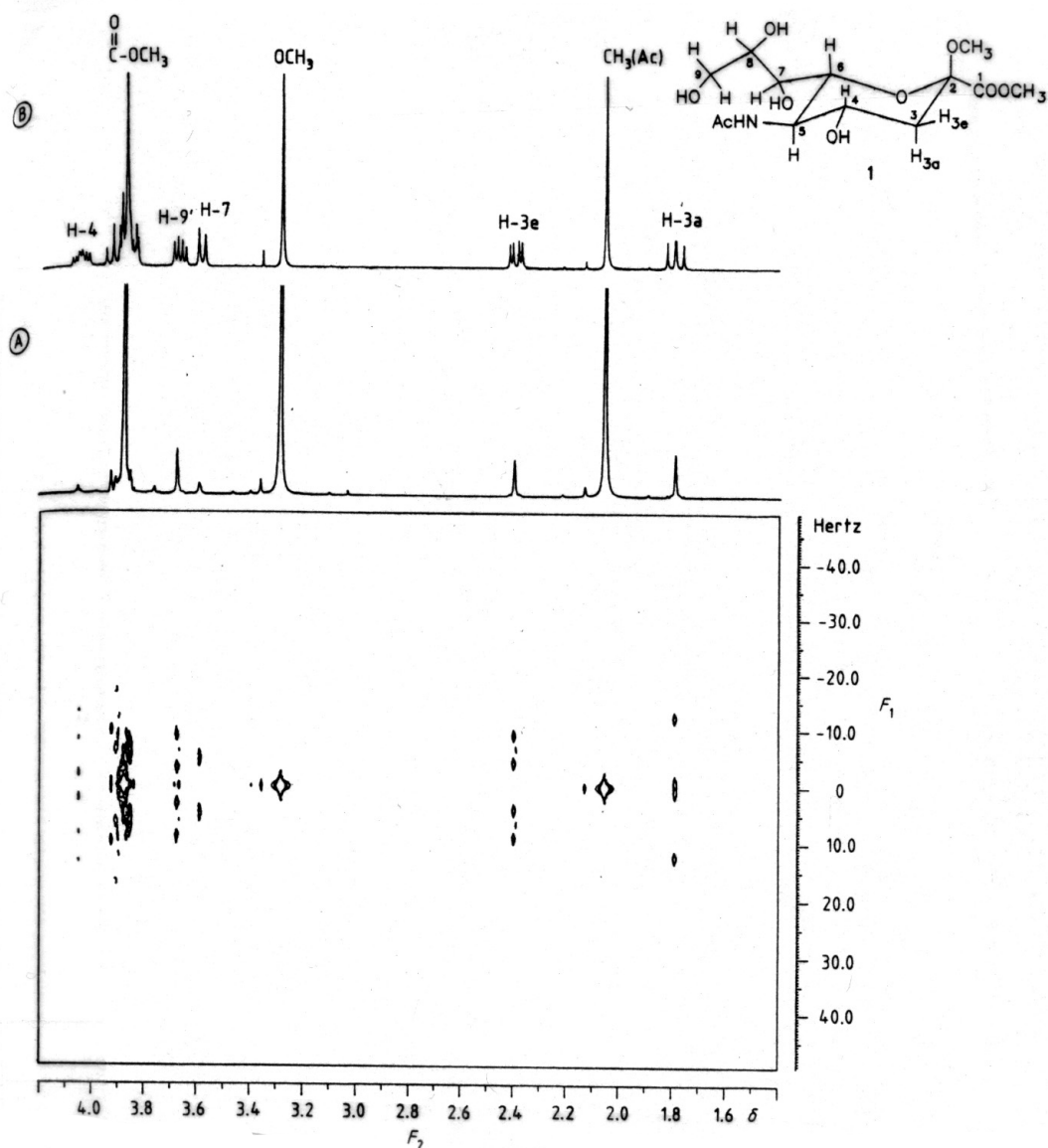
### For a $^1\text{H}$ AX system:

- The signal induced in the receiver depends on the sum of the two vectors, which depends on JAX. After FT of the FID with respect to  $t_2$ , two  $^1\text{H}$  peaks are observed. These two peaks are tilted relative to  $F_2$ , but the spectrum can be manipulated to bring all the component of the multiplet at the same  $F_2$  frequency (no splitting).
- The  $^1\text{H}$  signals are modulated by the value of JAX and the value of  $t_1$ . After FT of the FID with respect to  $t_1$ , a doublet centered at zero frequency is observed in the frequency spectrum  $F_1$  with a splitting of JAX.

$$JAX = 10 \text{ Hz}$$



**For more complex molecules:**



**Figure 9-8.**

Contour plot of the two-dimensional homonuclear *J*-resolved 400 MHz <sup>1</sup>H NMR spectrum of **1**.

A: Projection of the 2D spectrum onto the *F*<sub>2</sub>-axis. This is, in effect, a "decoupled" <sup>1</sup>H NMR spectrum.

B: Normal 400 MHz <sup>1</sup>H NMR spectrum of **1**.

(Experimental conditions:

20 mg of the compound in 0.5 ml D<sub>2</sub>O; 5 mm sample tube; 128 measurements with *t*<sub>1</sub> altered in 5.06 ms increments: each measurement with 48 FIDs and 4 K data points: total time 4.2 h.)