

# Environmental and geographic variables are effective surrogates for genetic variation in conservation planning

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**Protected areas buffer species from anthropogenic threats and provide places for the processes that generate and maintain biodiversity to continue. However, genetic variation, the raw material for evolution, is difficult to capture in conservation planning, not least because genetic data require considerable resources to obtain and analyze. Here we show that freely available environmental and geographic distance variables are highly effective surrogates in conservation planning for representing adaptive and neutral intra-specific genetic variation. We obtained occurrence and genetic data from the IntraBioDiv project for 27 alpine plant species collected over the European Alps using a gridded sampling scheme. For each species, we identified loci that were potentially under selection using outlier loci methods, and mapped their main gradients of adaptive and neutral genetic variation across the grid cells. We then used the cells as planning units to prioritize protected area acquisitions. First, we verified that the spatial patterns of environmental and geographic variation were correlated, respectively, with adaptive and neutral genetic variation. Second, we showed that these surrogates can predict the proportion of genetic variation secured in randomly generated solutions. Finally, we discovered that solutions based only on surrogate information secured substantial amounts of adaptive and neutral genetic variation. Our work paves the way for widespread integration of surrogates for genetic variation into conservation planning.**

conservation | biodiversity | AFLP | genetic diversity

**P**rotected areas spearhead conservation efforts (1). They buffer species from anthropogenic impacts, providing places for them to persist. Since the resources available for conservation are limited, protected areas need to be sited in places that fulfill conservation objectives for minimal cost (2). To achieve this, conservation planning exercises often generate plans for entire networks of protected areas (prioritizations) to allocate resources cost-effectively and preserve broad-scale biodiversity processes (3). Intra-specific genetic variation is fundamental to long-term species persistence (4, 5), and as a consequence, there has been increasing interest in designing prioritizations that represent this aspect of biodiversity (6–10). Although the strength of natural selection is a continuous force, genetic variation is often classified as adaptive or neutral (reviewed in 11). Adaptive genetic variation is associated with loci that significantly affect fitness. Typically, such “adaptive” loci are detected due to anomalous patterns among individuals (eg. 12, 13), though phenotypic-genetic associations are more likely to identify functionally important genomic regions for individual species (eg. 14). By representing the full range of adaptive variation, protected areas can help ensure that populations with particularly beneficial adaptations are not lost (7, 15). In contrast, neutral genetic variation is associated

with loci that do not significantly affect fitness, but instead reflect the evolutionary history of different populations. By representing the full range of neutral variation, protected areas can safeguard against the adverse effects of low genetic diversity (6). Thus optimally sited protected area networks would aim to represent patterns of both adaptive and neutral genetic variation (6, 7, 15).

Recently, the use of surrogate data has been proposed to generate prioritizations that capture intra-specific genetic variation (16, 17)—without needing to utilize genetic data to achieve this. Since adaptation is ultimately driven by selection pressures, environmental variables have been proposed as surrogates for adaptive genetic variation (eg. 16, 18). Specifically, by conserving individuals in a broad range of environmental conditions, we might expect to capture individuals with a diverse set of local adaptations, and overall capture a large proportion of the adaptive genetic variation present in the species. However, the effectiveness of this approach remains unverified. Neutral genetic variation arises from a reduction in gene flow between populations. Over the last few years, the field of phylogeography has predominantly focused on describing neutral genetic diversity, and so the landscape factors that affect neutral variation are relatively well understood (reviewed in 19). Potential surrogates for neutral genetic variation have been based on variables that predict the level of connectivity between different areas. For instance, according to the isolation by distance model (IBD; 20), populations located further apart are predicted experience less gene flow, and in turn, share fewer neutral loci. Based on this idea, spreading conservation priorities evenly across the geographic distribu-

## Significance Statement

To protect biodiversity for the long term, nature reserves and other protected areas need to represent a broad range of different genetic types. However, genetic data are expensive and time consuming to obtain. Here we show that freely available environmental and geographic variables can be used as effective surrogates for genetic data in conservation planning. This means that conservation planners can, with some confidence, design protected area systems to represent intra-specific genetic diversity without investing in expensive programmes to obtain and analyze genetic data.

JOH performed the analyses. All authors developed the study and drafted the manuscript.

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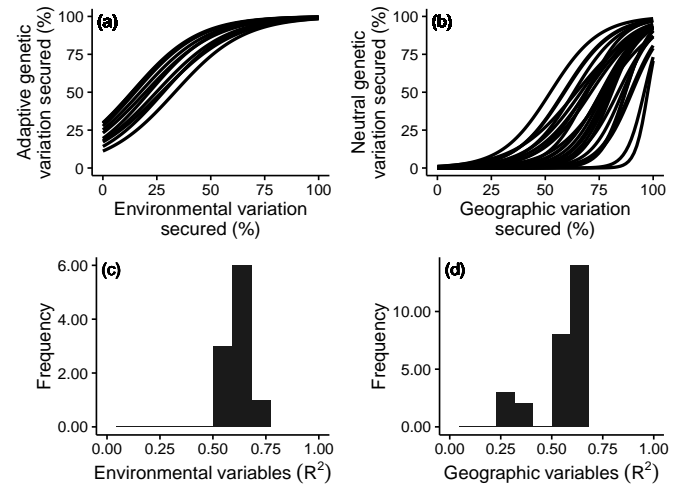
tion of an island network has been found to capture more neutral genetic variation (17). However this remains untested in spatially contiguous systems where connectivity is complicated by many additional factors (eg. land use; 21)—the typical situation in most conservation planning exercises.

Here we determine whether environmental and geographic surrogates capture adaptive and neutral genetic variation respectively in the context of conservation planning. We use distribution and genomic data for 27 alpine plant species in the European Alps that was obtained by the IntraBioDiv project (22). The data was collected following a gridded sampling scheme which we adopted as planning units for this study. We show that for most species the spatial patterns of adaptive and neutral genetic variation correlated with environmental and geographic variables. We also show that for most species the level of association is strong enough to be operational for conservation planning. Finally, we demonstrate that using these surrogates results in prioritizations that secure a substantial proportion of intra-specific genetic variation.

## Results

We detected putatively adaptive genetic variation in ten of the 27 plant species. We used two outlier detection methods—that solely used genetic data and did not use environmental data—to identify loci under selection. These methods returned reasonably consistent results (mean 87.92 % loci per species assigned the same classification  $\pm$  8.75 SD; Table S1). Of the species that were associated with adaptive genetic variation, only a small proportion of loci were classified as being adaptive (mean 3.01 % loci per species  $\pm$  1.95 SD). After identifying the loci showing strong signals of selection, we used non-metric multi-dimensional scaling (NMDS) analyses to identify the main gradients of adaptive (if detected) and neutral variation for each species. Generally, only a small number of continuous dimensions was needed to sufficiently describe their patterns of adaptive ( $K = 2$  for all species; Table S2) and neutral genetic variation ( $2 \leq K \leq 4$  for all species; Table S1). The resulting ordinations were used to construct an adaptive (if detected) and neutral genetic space for each species. The spatial distribution of these genetic spaces (Figures S1–S27) generally showed substantial spatial auto-correlation (eg. *Cerastium uniflorum*, Figure S34; *Dryas octopetala*, Figure S36), suggesting that the environmental and geographic variables have potential to be effective surrogates for genetic variation.

We verified that the spatial patterns in environmental variation correlated with the broad-scale patterns in adaptive genetic variation for each species. We also verified that there was a correlation between neutral genetic variation and variation in geographic position. For each species, we constructed dissimilarity matrices expressing differences between the planning units where individuals were detected based on the units' (i) environmental characteristics, (ii) geographic position, and also the (iii) adaptive (if adaptive loci detected), and (iv) neutral genetic characteristics of the individuals found inside them. The spatial patterns of environmental variation were significantly correlated with the pattern of adaptive genetic variation for eight of the ten species associated with adaptive variation ( $P < 0.05$ ; mean 0.09 marginal  $R^2 \pm 0.06$  SD for significant models; Table S3). Similarly, geographic distance between planning units was also correlated significantly with the spatial pattern of neutral genetic variation among planning units for

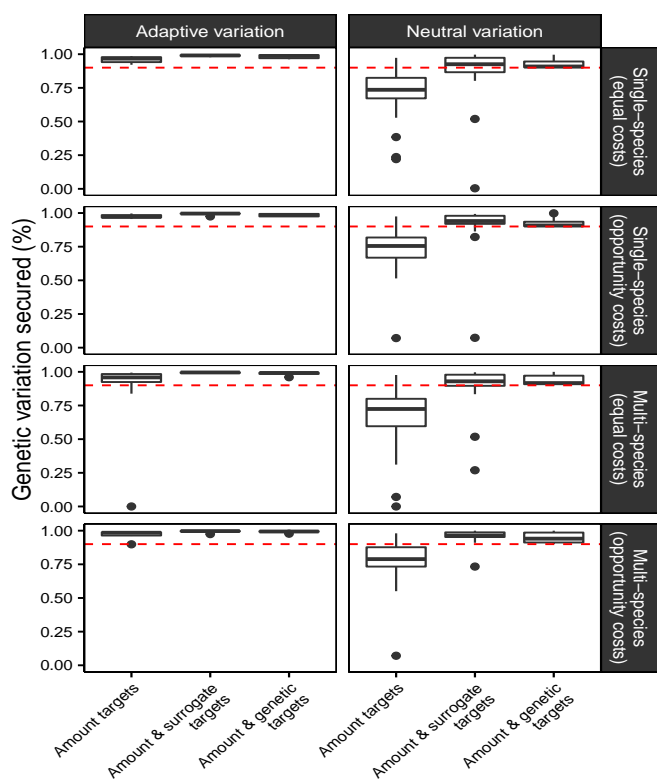


**Fig. 1.** The proportion of adaptive (a; 10 species) and neutral (b; 27 species) genetic variation represented in a suite of randomly generated prioritizations for different species as a function of the surrogate variation they also secured (lines). Panels (c) and (d) show the distribution of  $R^2$  values for the 37 models shown in (a) and (b).

26 of the 27 species ( $P < 0.05$ ; mean 0.2 marginal  $R^2 \pm 0.16$  SD for significant models; Table S3). Thus, for most species, planning units that contained different environmental conditions contained individuals with different adaptive genetic characteristics, and planning units that were further apart tended to contain individuals with different neutral genetic characteristics. After verifying these patterns were correlated, our next step was to determine whether these associations were strong enough to be effective for conservation planning.

For each species, we generated a suite of 10,000 random prioritizations, and calculated the proportion of environmental variation and adaptive (if present) genetic variation that each prioritization captured. We then repeated this process, and calculated the proportion of variation in geographic position and neutral genetic variation they sampled. The environmental and geographic variables were moderately effective predictors for the genetic variation represented by the randomly generated prioritizations (Figures 1, S28 and S29). The relationship between the proportion of genetic and surrogate variation secured in the prioritizations varied among the species (species  $\times$  proportion of environmental variation interaction term:  $\chi^2_9 = 6776.94$ ;  $P = 0.028$ ; species  $\times$  proportion of geographic variation interaction term:  $\chi^2_{26} = 4.670631 \times 10^4$ ;  $P < 0.001$ ). *Post-hoc* analyses showed that these relationships were positive for all species (environmental: minimum  $z = 40.34$ , maximum  $P < 0.001$ ; geographic: minimum  $z = 23.51$ , maximum  $P < 0.001$ ; Table S4). After establishing that the the proportion of surrogate variation secured in a prioritization also predicted the proportion of genetic variation it secured, our final step was to determine if prioritizations were more effective when generated using targets based on the surrogate variables.

To determine whether environmental and geographic targets could improve the effectiveness of prioritizations in representing genetic variation, we generated prioritizations using (i) “amount targets” reflecting the traditional approach of representing a certain proportion of each species’ geographic distribution, (ii) “amount and surrogate targets” where we targeted representation of the environmental and geographic



**Fig. 2.** The proportion of adaptive and neutral genetic variation secured for each species in solutions based on different targets and scenarios. Box plots show the median, and the 25th and 75th percentiles. Whiskers show the data at 1.5 times the inter-quartile range. Points show data outside the whiskers. Red lines show the target proportion of genetic variation.

surrogates as well as a certain proportion of each species' geographic distribution, and (iii) "amount and genetic targets" where we targeted representation of the directly measured genetic variation as well as a certain proportion of each species' geographic distribution. To determine if these patterns were robust, for each of the three combinations of the targets, we generated prioritizations under four scenarios: (i) single-species with equal costs, (ii) single-species with acquisition costs, (iii) multi-species with equal costs, and (iv) multi-species with acquisition costs (Figure S30).

The proportion of genetic variation secured in a prioritization depended on the targets used to generate the prioritization ( $\chi^2_2 = 67.81$ ;  $P < 0.001$ ). Most notably, prioritizations based on the surrogate variables represented significantly more genetic variation than amount-based prioritizations ( $93.24\% \pm 13.79$  SD overall genetic variation secured versus  $78.33\% \pm 23.62$  SD;  $Z_1 = 4.59$ ;  $P < 0.001$ ), and were not distinguishable from prioritizations based on measured genetic data ( $94.7\% \pm 3.8$  SD;  $Z_1 = 0.56$ ;  $P > 0.99$ ). These results indicate that environmental and geographic surrogates in this case far outperform traditional amount-based conservation planning, and perform almost as well as a conservation plan based directly on measured genetic data.

Overall, the prioritizations tended to secure a greater proportion of adaptive genetic variation than neutral genetic variation (adaptive: mean  $96.04\% \pm 7.61$  secured across all prioritizations, neutral: mean  $71.94 \pm 24.16$ ;  $\chi^2_1 = 1408.16$ ;  $P < 0.001$ ). Thus, regardless of the targets used to generate

a prioritization, or the planning context under which the prioritization was generated, prioritizations tend to secure more adaptive than neutral genetic variation.

The average proportion of genetic variation secured by prioritizations varied under different scenarios ( $\chi^2_3 = 34.45$ ;  $P < 0.001$ ). Specifically, prioritizations generated under single-species scenario (mean  $78.36\%$  genetic variation secured  $\pm 23.61$ ) secured less genetic variation than those under the single-species with opportunity costs (mean  $88.89\%$  genetic variation secured  $\pm 15.52$ ;  $Z_1 = 2.94$ ;  $P = 0.02$ ) or the multi-species with cost scenarios (mean  $91.89\%$  genetic variation secured  $\pm 12.31$ ;  $Z_1 = 3.68$ ;  $P = 0.001$ ). These results suggest the amount of genetic variation secured in a prioritization may be affected by the data used to represent acquisition cost.

## Discussion

We have shown that broad-scale environmental and geographic variables can be effective as surrogates for adaptive and neutral genetic variation in conservation planning. Our study is the first to show this using field measurements of genetic variation for a broad range of species. The spatial patterns of environmental and geographic surrogates were strongly correlated with the patterns of adaptive and neutral genetic variation for most species. Moreover, for most species, prioritizations generated using environmental and geographic targets secured a greater proportion of adaptive and neutral genetic variation than traditional conservation planning. These environmental and geographic surrogates were based on freely available data, and could be applied to any study region across the world.

Our results demonstrate that environmental and geographic variables are effective surrogates for most species considered here (Figure 2). Despite this, geographic distance was a surprisingly poor surrogate for neutral genetic variation for a few species (eg. *Gypsophila repens*:  $R^2 = 0.26$ ; *Luzula alpinopilosa*:  $R^2 = 0.28$ ; *Gentiana nivalis*  $R^2 = 0.29$ ). One explanation for this result is that geographic distance will be a poor surrogate for neutral genetic variation where spatial genetic structure is complicated by additional factors (21, 23). For example, some species can maintain relatively high connectivity between distant populations (eg. wind dispersing plants; 22), and in some places can be disrupted by landscape features (eg. anthropogenically modified land 21). Overall, such species were the exception in our analysis, and both surrogates performed moderately well for most species.

Our results show that environmental and geographic surrogates can be used to capture genetic variation in prioritizations, the next question is: what percentage of surrogate variation should planners target to preserve genetic variation? To secure at least 90 % of the species' neutral genetic variation, prioritizations needed to sample  $95.07\% \pm 6.1$  SD of the variation in geographic position among the planning units occupied by each species. Additionally, to secure at least 90 % of each species' adaptive genetic variation, prioritizations needed to sample  $56.43\% \pm 7.54$  SD of the environmental variation in planning units occupied each the species. For all species, however, it was possible to generate solutions that secured a large proportion of the surrogate variation ( $> 90\%$ ) and only a small proportion of genetic variation ( $< 20\%$ ; Figures S56–S57). Planners may avoid such outcomes by using both amount- and surrogate-based targets. Depending on the study area and the species of conservation interest, higher

373 targets may be needed to increase the likelihood that prior-  
 374 itizations will secure a large proportion of the intra-specific  
 375 genetic variation for all of the species in the planning exercise.

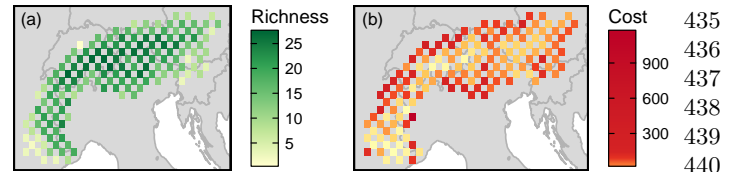
376 Our results suggest that conservation planners can secure  
 377 a reasonably representative sample of the intra-specific adap-  
 378 tive genetic variation using only amount-based targets (Figure  
 379 2). For most species, the spatial patterns of adaptive ge-  
 380 netic variation clustered into a few distinct groups (eg. *Carex*  
 381 *sempervirens* and *Cerastium uniflorum*; Figures S33 and S34  
 382 respectively). As a consequence, a prioritization would only  
 383 need to select a single planning unit in each cluster to secure  
 384 a representative sample. This mechanism explains why adap-  
 385 tive variation accumulated much more quickly than neutral  
 386 genetic variation in the randomly generated prioritizations.  
 387 Additionally, this mechanism also explains why the single-  
 388 species prioritizations generated using only amount targets  
 389 were often able to achieve high representation—they were  
 390 essentially a random selection and so frequently contained  
 391 planning units in different parts of the species' ranges (Fig-  
 392 ure 4a). Overall—and contrary to our original hypothesis—it  
 393 seems that conventional reserve selection methods might often  
 394 conserve species' adaptive genetic variation.

395 There are several limitations associated with our analysis.  
 396 Firstly, the size of the planning units we used (approx.  $23 \times 25$   
 397 km) is much larger than typically used in regional conservation  
 398 planning ( $1\text{--}10\text{ km}^2$ ). We used this resolution because the  
 399 genetic data were collected at this scale. Whilst we could have  
 400 interpolated the genetic data to a finer resolution and used  
 401 smaller planning units, this would have biased our analysis  
 402 by artificially introducing additional spatial auto-correlation.  
 403 Secondly, we used geographic distances as surrogates for neu-  
 404 tral genetic variation, yet distances that incorporate data on  
 405 topography may better describe connectivity in the study area.  
 406 However, such distances often require species-specific scaling  
 407 (eg. 21), and so cannot easily be utilized in multi-species plan-  
 408 ning exercises. Thirdly, we used amplified fragment length  
 409 polymorphism data (AFLP; 24) to describe genetic variation.  
 410 Whilst next-generation sequencing provides higher resolution  
 411 genetic information than AFLP data (11), we know of no  
 412 suitable multi-species genomic data set, and our methodology  
 413 would still have utilized only the main gradients of the genetic  
 414 variation to generate prioritizations in a feasible period of  
 415 time. Furthermore, even with modern population genomic  
 416 approaches, a survey can at best hope to identify markers that  
 417 are linked to functionally adaptive variants.

418 Broad-scale environmental and geographic variables are  
 419 generally effective surrogates for representing adaptive and  
 420 neutral genetic variation for most species in our study system.  
 421 Moreover, data to calculate the surrogate variables are freely  
 422 available for any location in the world. Careful use of such  
 423 surrogates in conservation planning could vastly improve the  
 424 chances of long-term biodiversity persistence for relatively  
 425 little additional cost.

## 427 Materials and Methods

429 **Study system.** We used data for 27 alpine plant species in  
 430 the European Alps collected by the IntraBioDiv project (22;  
 431 Figure 3a). All data, code, and results are stored in an online  
 432 repository to permit replication and validation of this study  
 433 ([www.github.com/jeffreyhanson/genetic-surrogates](https://github.com/jeffreyhanson/genetic-surrogates)). This data  
 434 set has been used to explore patterns of adaptive (eg. 25) and



**Fig. 3.** Map of the study area showing (a) the pattern of species richness, and (b) the distribution of acquisition cost, both plotted on a quantile-based color ramp. Squares denote planning units.

441 neutral genetic variation (eg. 26), and the potential for species  
 442 richness as a surrogate for genetic diversity (27). Data were  
 443 collected using a  $20'$  longitude by  $21'$  latitude grid (approx.  
 444  $22.3 \times 25\text{ km}$ ; Figure S31). Project surveyors visited every  
 445 second grid cell, and for each species, they collected samples  
 446 from three individuals if any individuals were found. They  
 447 genotyped samples using AFLPs, and constructed matrices  
 448 denoting the presence of polymorphisms at loci for each species  
 449 (mean  $130.7 \pm 54.9$  SD markers genotyped per species; for  
 450 more information see 26, Table 1). Thus the data set contains  
 451 information describing the genomic properties of individuals  
 452 for each species over a geographic grid. We used these data  
 453 because they comprised comparable genetic information for a  
 454 range of species with different evolutionary and life histories  
 455 collected using a standardized sampling scheme.

461 **Landscape data.** We adopted the sampling grid used to collect  
 462 data as planning units to develop prioritizations. To reduce  
 463 computational burden, we omitted cells that did not contain  
 464 samples (149 grid cells used in analysis). To examine the  
 465 effect of variation in cost, we calculated the total human  
 466 population density inside each planning unit ( $1\text{ km}^2$  resolution  
 467 from the Global Rural-Urban Mapping Project; 28) and used  
 468 this to represent acquisition cost (Figure 3b). All spatial and  
 469 statistical analyses were conducted in R (version 3.3.2; 29).

470 We created environmental and geographic surrogate vari-  
 471 ables for each species (Figure S32). To describe the geographic  
 472 location of each planning unit (Figure S33), we projected the  
 473 grid into an equi-distant coordinate system (Europe Equidistant  
 474 Conic; ESRI:102031), calculated the centroid of each  
 475 grid cell, and extracted their two-dimensional coordinates. To  
 476 describe the environmental characteristics of each planning  
 477 unit (Figure S34), we obtained 19 climatic layers ( $30''$  reso-  
 478 lution; 30), projected them and the planning units into an  
 479 equal-area coordinate system (Europe Lambert Conformal  
 480 Conic; ESRI:102014), and computed planning unit averages  
 481 for each climatic variable. To reduce dimensionality, for each  
 482 species, we subjected the climatic values associated with the  
 483 planning units they were found in to a principal components  
 484 analysis (PCA; Table S1). We used the first three principal  
 485 components to characterize climatic variation found across the  
 486 species' geographic distributions inside the study area (captur-  
 487 ing  $90.26\% \pm 0.99$  SD of the total climatic variation; Figure  
 488 S2). Thus we constructed a two-dimensional geographic space  
 489 as a potential surrogate for neutral genetic variation, and a  
 490 three-dimensional environmental space as a potential surrogate  
 491 for adaptive genetic variation for each species. In these spaces,  
 492 each planning unit was associated with a single point. Plan-  
 493 ning units with more comparable environmental conditions,  
 494 or higher spatial proximity in the real world, were associated  
 495 with points that were closer together in these environmental



497 or geographic spaces.

498

499 **Adaptive and neutral genetic data.** To investigate the effective-  
500 ness of our putative surrogates, we first needed to identify  
501 which of the sampled loci were adaptive (Figure S35). Follow-  
502 ing best practice, we used two outlier detection methods to  
503 achieve this (an individual- and a population-level method;  
504 31). The basic premise underpinning such methods is that  
505 neutral loci are expected to exhibit a certain level of variation,  
506 and loci that deviate from this expectation are likely to be  
507 under selection (11). The advantage of these methods—in  
508 contrast with environmental association analyses—is that they  
509 do not use environmental data, which would have introduced  
510 an element of circularity into our analysis and invalidated it.  
511 Loci identified by both outlier detection methods were treated  
512 as adaptive and the remainder were treated as neutral. To  
513 minimize false-positives, we omitted loci from both methods  
514 where the global frequency of the minor allele was less than  
515 10 % and treated them as neutral.

516 The first outlier detection method involved fitting  
517 multinomial-Dirichlet models implemented in **BayeScan** (ver-  
518 sion 2.1; 12). We adopted a similar methodology to Bothwell *et*  
519 *al.* (25) and applied it to each of the species separately. Follow-  
520 ing their methodology, we initially grouped conspecifics into  
521 genetic lineages to further minimize false-positives (Figures  
522 S36–S62), by fitting admixture and correlated alleles models  
523 implemented in **Structure** (version 2.3.4; 32; 20 replicates per  
524 species; 5,000 admixture burnin iterations; 300,000 burnin it-  
525 erations; 400,000 total iterations) using the number of lineages  
526 previously determined by Alvarez *et al.* (26) and combining  
527 replicate runs using **ClumPP** (version 1.1.2; 33; greedy algo-  
528 rithm based on the  $G'$  statistic; 1,000 iterations). We then  
529 ran **Bayescan** for each species using these lineages (1:1 prior  
530 odds; 4 replicates per species; 20 pilot runs; 100,000 burn-in  
531 iterations; 110,000 total iterations thinned by 10 iterations)  
532 using a suitable false discovery rate ( $q \leq 0.1$ ; 34). We omit-  
533 ted individuals if their population membership was uncertain  
534 (maximum membership probability  $< 0.75$ ).

535 The second outlier detection method involved fitting PCAs  
536 to identify outlier loci (implemented in the **pcadapt** R package;  
537 13). To enable comparisons between the two outlier detection  
538 methods, we used the same individuals in this analysis as  
539 in the **BayeScan** analysis. For each of the 27 species, we  
540 first imputed missing data by replacing missing values with  
541 the average frequency among conspecifics, then ran a PCA  
542 over the loci matrix, and extracted the minimum number of  
543 components needed to secure the over-arching population level  
544 variation among the loci (10 % of the variation in loci). We  
545 then computed  $q$ -values using Mahalanobis distances, and used  
546 the same false discovery rate used in the **BayeScan** analysis to  
547 identify loci under selection (using the **qvalue** R package; 35).

548 After classifying loci as adaptive or neutral, we mapped  
549 the main gradients of the adaptive (if detected) and neutral  
550 genetic variation for each species. We discarded the population  
551 groupings, and partitioned species' adaptive and neutral loci  
552 into separate matrices. We applied NMDS (implemented  
553 in the **vegan** R package; 36) using Gower distances (via the  
554 **cluster** R package; 37) to derive continuous variables that  
555 described the main gradients of adaptive and neutral genetic  
556 variation separately for each species (Table S2). To ensure that  
557 the ordinations described a sufficient amount of the genetic  
558 variation, we ran successive scaling analyses with increasing

dimensionality until a sufficient stress value was obtained  
(maximum stress value  $\leq 0.25$ ; 100 random starts for each  
analysis). Since each grid cell had multiple samples per species,  
we averaged the ordinated values for conspecifics in the same  
grid cell.

564 Thus we constructed an adaptive (if detected) and neu-  
565 tral genetic space for each species. For a given species, each  
566 planning unit in which it occurred was associated with a multi-  
567 dimensional point in the species' adaptive genetic space (if  
568 adaptive genetic variation was detected) and another multi-  
569 dimensional point in the species' neutral genetic space. Plan-  
570 ning units that were closer together in these genetic spaces  
571 were occupied by individuals with more comparable AFLP  
572 polymorphisms. By spreading out conservation effort across  
573 these genetic spaces, and in turn selecting planning units occu-  
574 pied by individuals with increasingly different polymorphisms,  
575 prioritizations can secure more genetic variation.

577 **Prioritization method.** We used the **raptr** R package to gen-  
578 erate solutions (38). This toolkit can identify the cheapest  
579 set of planning units required to preserve both a target pro-  
580 portion of the species' geographic range (using amount-based  
581 targets) and a broad sample of variation found across the  
582 species' range. We used the environmental and geographic  
583 surrogate spaces, and the adaptive (if detected) and neutral  
584 genetic spaces to generate and evaluate solutions. Solutions  
585 associated with negative values—because they secured very  
586 little genetic variation—were replaced with zeros to facilitate  
587 statistical analysis. We solved all reserve selection problems  
588 to within 10 % of optimality.

590 **Computational and statistical analyses.** Our first aim was to  
591 determine if the environmental and geographic variables corre-  
592 late with the spatial patterns of adaptive and neutral genetic  
593 variation. To achieve this, for each species, we created dissim-  
594 ilarity matrices using Euclidean distances and the data in the  
595 surrogate and genetic spaces. These matrices showed differ-  
596 ences between the planning units occupied by the species in  
597 terms of the units' (i) geographic position, (ii) environmental  
598 characteristics, and the (iii) adaptive (if detected) and (iv)  
599 neutral genetic characteristics of the individuals inside them.

600 We fitted maximum likelihood population-effects (MLPE)  
601 models using maximum likelihood to investigate correlations  
602 between the dissimilarity models (using the **lme4** R package;  
603 39). These models use random effects to accommodate the  
604 structure of dissimilarity matrices. For each species, we fitted  
605 a MLPE model to the species' dissimilarity matrices based on  
606 geographic position and neutral genetic variation. If adaptive  
607 loci were detected, we also fitted a MLPE model to the species'  
608 dissimilarity matrices describing environmental and adaptive  
609 genetic variation. All data variables were  $z$ -transformed to  
610 improve convergence. To test if the surrogates explained the  
611 genetic variation, we compared each model to its null model  
612 using a  $\chi^2$  test, and applied Bonferroni corrections.

613 Our second aim was to determine if the environmental  
614 and geographic variables were effective surrogates for adaptive  
615 and neutral genetic variation. For each species, we generated  
616 20,000 prioritizations by randomly selecting different combi-  
617 nations of planning units that the species occupied. For half  
618 of these random prioritizations, we calculated the proportion  
619 of geographic variation and neutral genetic variation they se-  
620 cured, and for the other half, we calculated the proportion of

environmental variation and adaptive genetic variation they secured. The proportion of surrogate and genetic variation captured in a prioritization was calculated using the **raptr** R package.

We fitted two full generalized linear models (GLM) with logit links. The first model was fit to the proportion of adaptive genetic variation secured in a prioritization using the proportion of environmental variation also secured in the prioritization. The second model was fit to the proportion of neutral genetic variation secured in a prioritization using the proportion of geographic variation also secured in the prioritization. Additionally, both models contained a variable indicating the species for which the prioritizations were generated, and an interaction term. They were subjected to backwards stepwise term deletion routines to assess term significance. To assess the performance of the surrogates, we refit these models separately for each species and computed the Cragg and Uhler's pseudo- $R^2$  value (using the **pscl** R package; 40).

Our third aim was to determine if surrogate-based targets improved representation of genetic variation in prioritizations. As previously mentioned, we generated prioritizations using different combinations of targets and conservation planning scenarios. We used 20 % amount-based targets for each species in all prioritizations to secure an adequate proportion of the species' distributions. Based on the results from our previous

analysis, we used 97.5 % surrogate-based targets and 90 % genetic-based targets. We generated a single solution for each target/scenario combination, except for the "single-species (equal costs) with amount-based targets" combination for which we generated 1,000 replicates because it had many optimal solutions. We computed the proportion of adaptive (if present) and neutral genetic variation secured for each species in the prioritizations.

We fitted generalized linear mixed-effects models (GLMMs) with logit links to evaluate the prioritizations (using the **lme4** R package). We fitted a full model to the proportion of genetic variation secured in a given prioritization. This model contained categorical variables indicating the targets (amount only, amount and surrogate targets, or amount and genetic targets), the planning scenario (single-species, multi-species, or multi-species with cost), the type of genetic variation measured (adaptive or neutral), and all interactions between them. Data for same species were accommodated using a random intercept term. The full model was subjected to a backwards stepwise term deletion routine to assess significance. A *post-hoc* analysis was conducted on the minimal adequate model using Tukey contrasts with a Bonferroni correction (using the **multcomp** R package; 41).

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