

Using readmers and hapmers in assessing phase switching after read error correction of Oxford Nanopore Sequences

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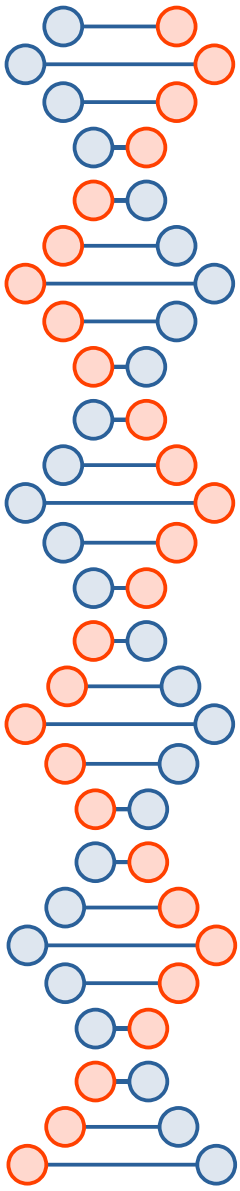
Institute of Medical
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David Horner

david.horner@meduniwien.ac.at

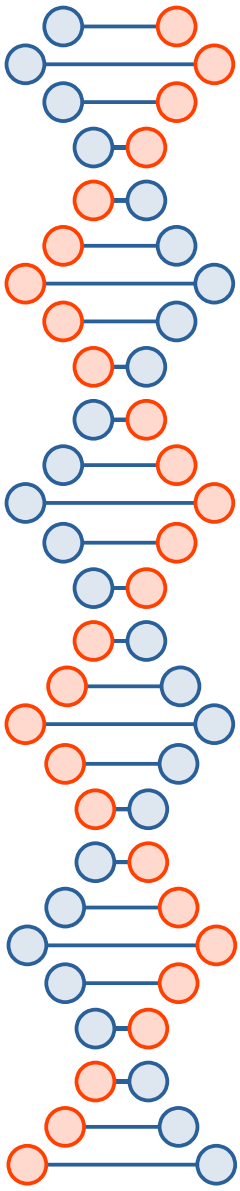
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Outline

- **Introduction**
- Methods
- Results
- Wrap up



Outline

- **Introduction**

k-mers

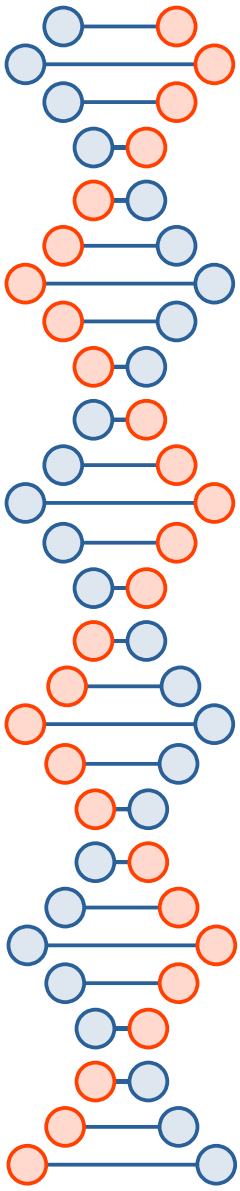
readmers

hapmers

phase switching

Error correction

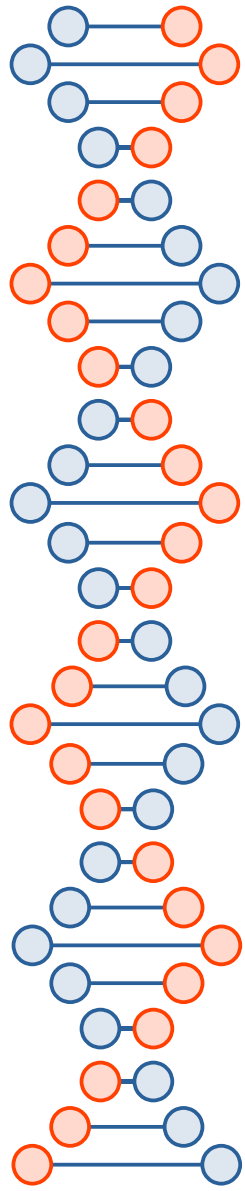
Why do we care? – Research Question



K-mers, Readmers, & Hapmers, oh my....

- DNA has no “words”
 - Can break DNA into pieces of “k-length” called k-mers
- k-mer length = 4

ACTGCA



K-mers, Readmers, & Hapmers, oh my....

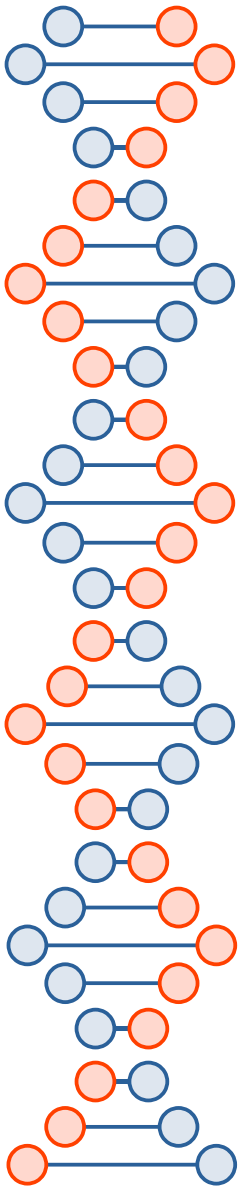
- DNA has no “words”
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ACTGCA

actg 4-mer #1

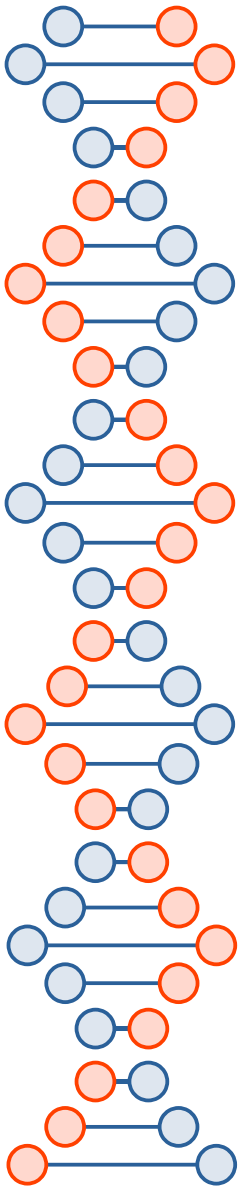
ctgc 4-mer #2

tgca 4-mer #3



K-mers, Readmers, & Hapmers, oh my....

- K-mers from sequencing reads ---
 - ▷ “Readmers”



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- K-mers from sequencing reads ---
 - ▷ “Readmers”

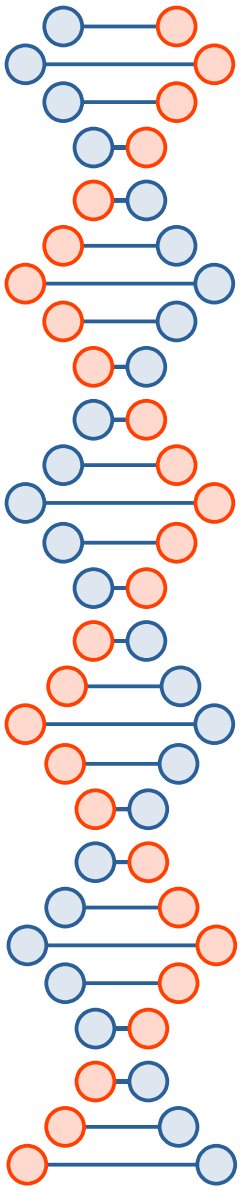
A FASTQ read:

@seq1

ACTGCATAGC

+

6655EEACDC

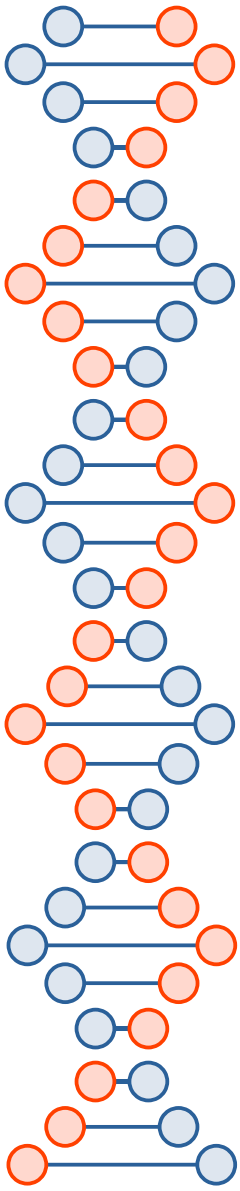


K-mers, Readmers, & Hapmers, oh my....

- K-mers unique to a haplotype ---

- ▷ “Hapmers”

using 8-mers on “+” strand



K-mers, Readmers, & Hapmers, oh my....

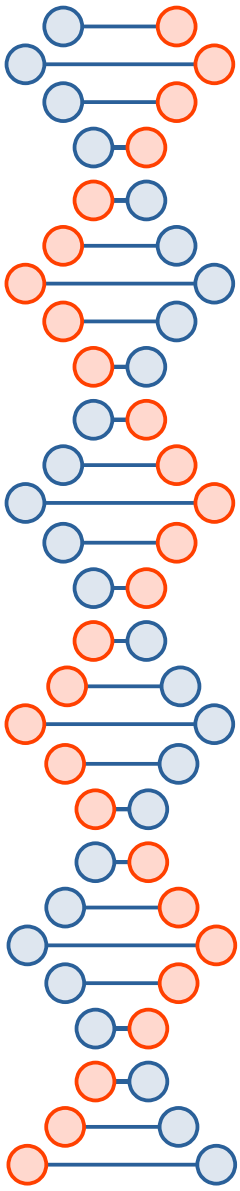
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using 8-mers on “+” strand

ATTGCATA + strand maternal haplotype

ACTGAATA + strand paternal haplotype



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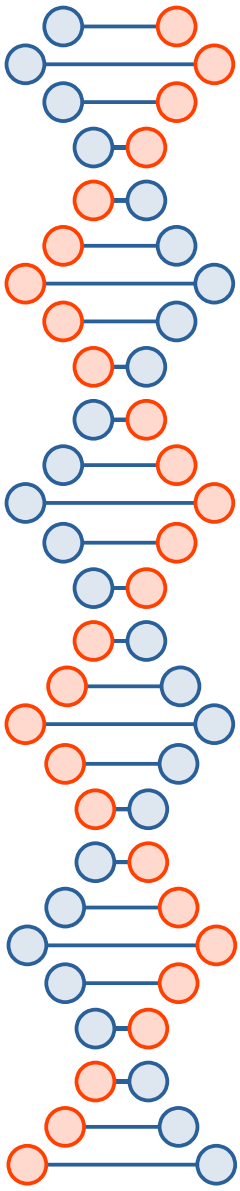
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attgcata

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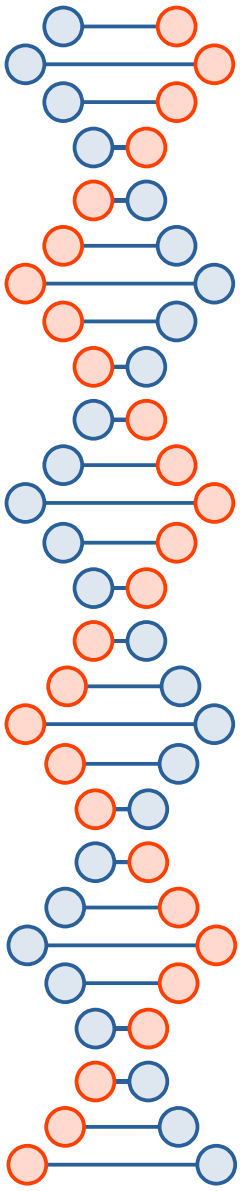
ATTGCATA “+” strand maternal haplotype

attgcata

ACTGAATA “+” strand paternal haplotype

actgaata

Hapmers

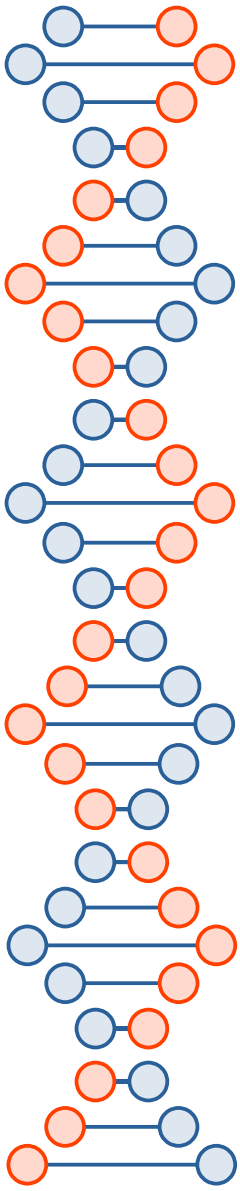


Phase Switching

- At the read level, read contains info of 1> haplotype

A**T**TGC**C**ATA “+” strand maternal haplotype

A**C**TGA**A**ATA “+” strand paternal haplotype



Phase Switching

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A**T**TGC**C**ATA “+” strand maternal haplotype

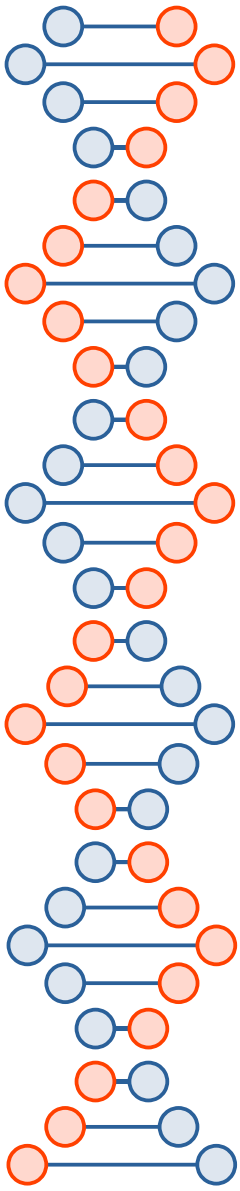
A**C**TG**A**AATA “+” strand paternal haplotype

@seq1

ACTGCATA

+

CCEEDDAD



Phase Switching

- At the read level, read contains info of 1> haplotype

A**T**TG**C**ATA “+” strand maternal haplotype

A**C**TG**A**ATA “+” strand paternal haplotype

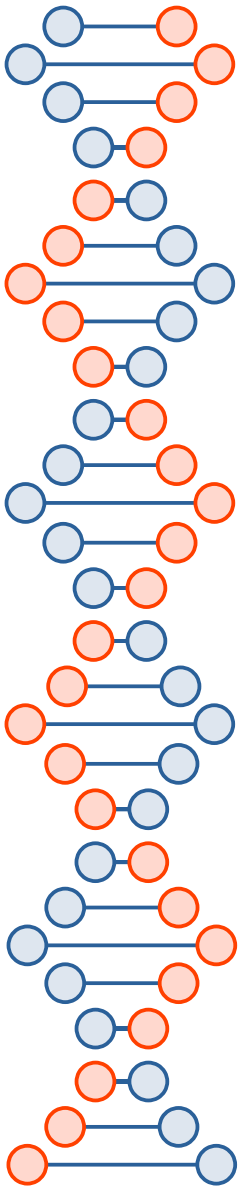
@seq1

A**C**TG**C**ATA

Phase Switching!

+

CCEEDDAD



Phase Switching

- At the read level, read contains info of 1> haplotype

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@seq1

A**C**TGC**C**ATA

+

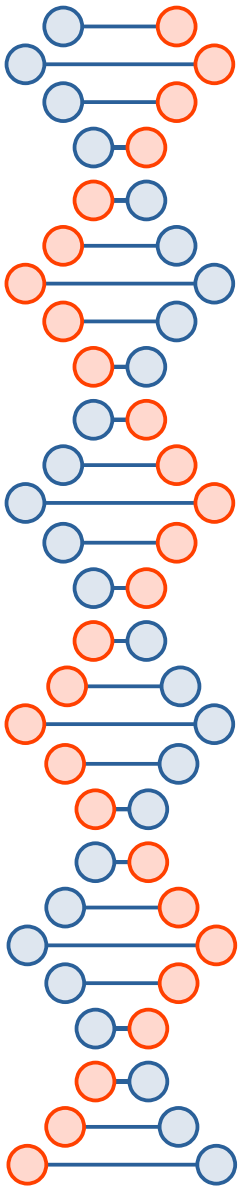
CCEEDDAD

In this example:



If counting 8-mers, then there will be

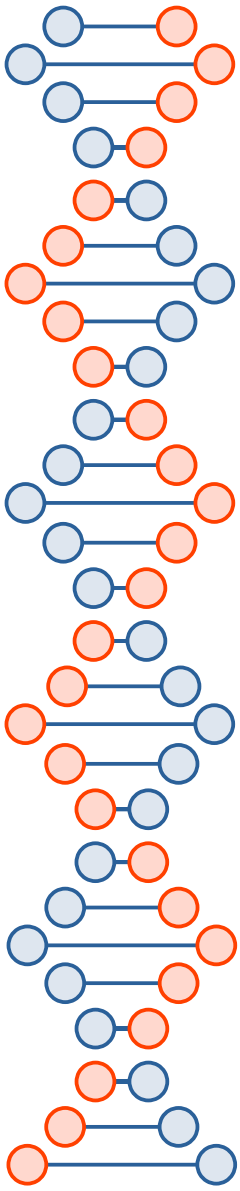
**No matching readmers to
hapmers**



Error correction

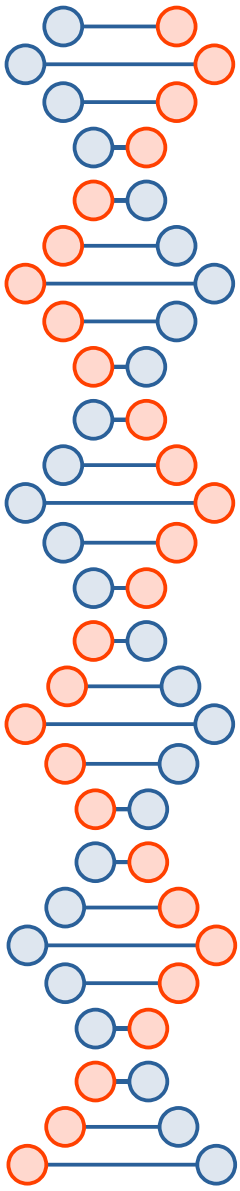
- Improve accuracy by “consensus”

1. Herro



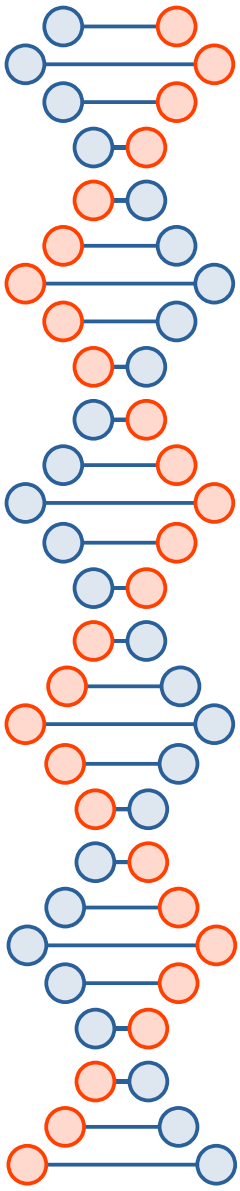
Error correction

- Improve accuracy by “consensus”
 1. Herro
 2. Brutal Rewrite



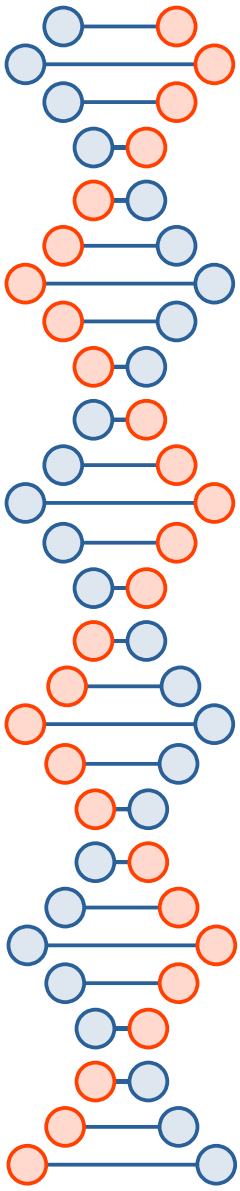
Error correction

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 1. Herro
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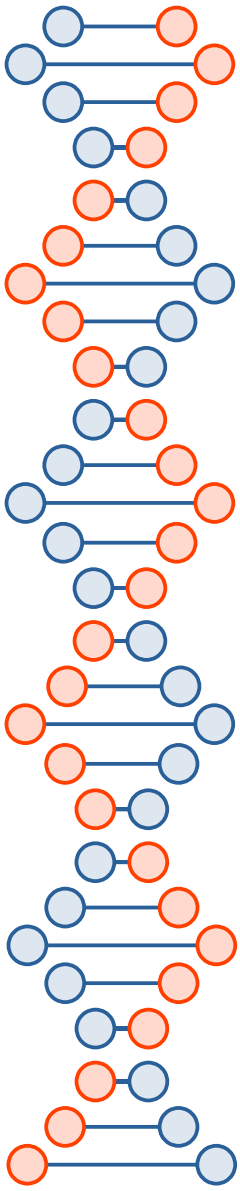
Error correction

- Improve accuracy by “consensus”
 1. Herro
 2. Brutal Rewrite
 3. Peregrine_2021
 4. DeChat



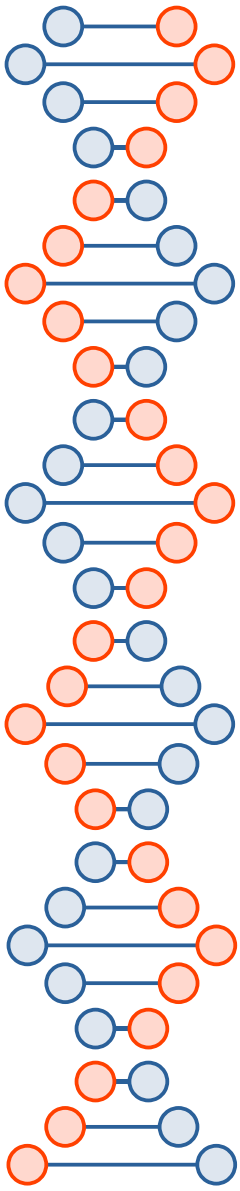
Question

- Since we do not know what sequences belong to which haplotype *a priori* , can error correcting reads cause phase switching?



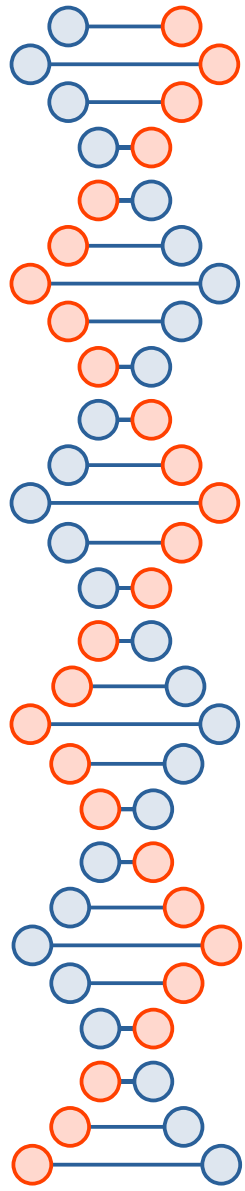
Question

- Since we do not know what sequences belong to which haplotype *a priori* , can error correcting reads cause phase switching?
- How can we estimate phase switching at the read level?



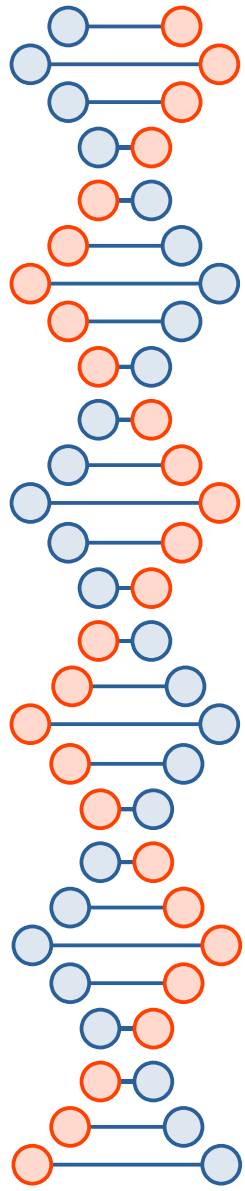
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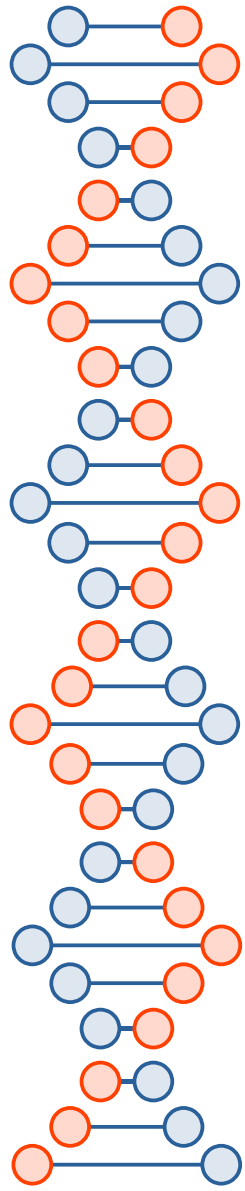
Outline

- **Methods**
 - error correction
 - read alignment
 - analyze alignments
 - k-mers



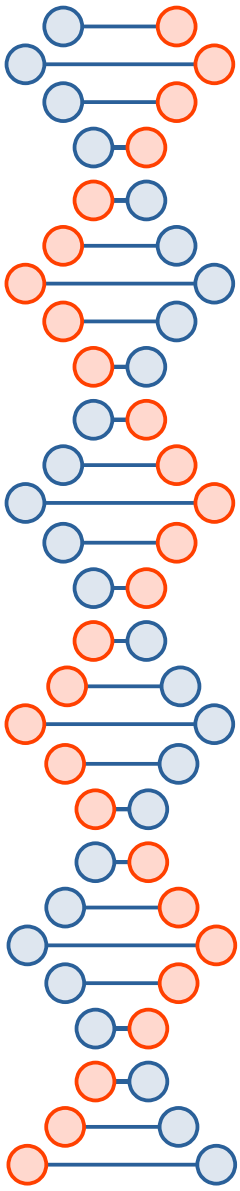
Methods

- Error correct reads



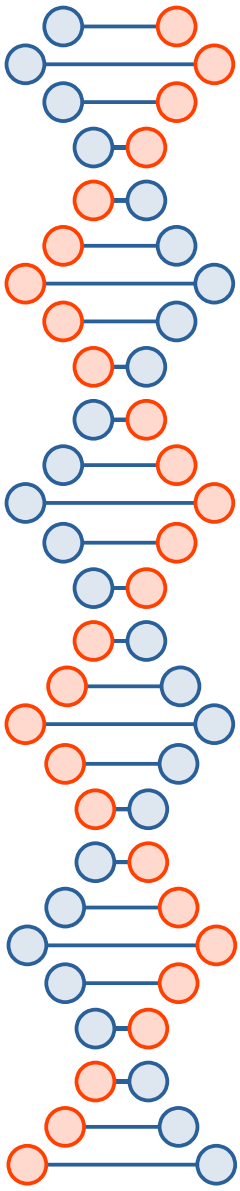
Methods

- Error correct reads
 1. Run Herro



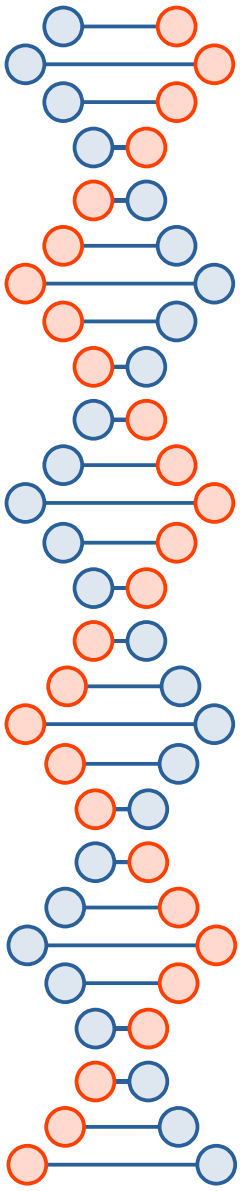
Methods

- Error correct reads
 1. Run Herro
 2. Run Brutal Rewrite on Herro-corrected reads



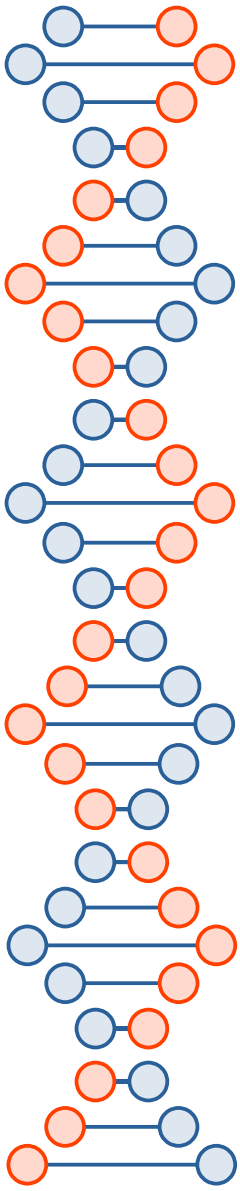
Methods

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 1. Run Herro
 2. Run Brutal Rewrite on Herro-corrected reads
 3. Run Peregrine_2021 on Brutal Rewrite-corrected reads



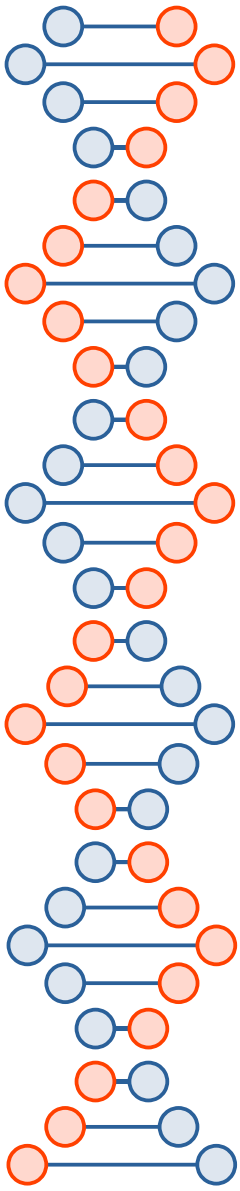
Methods

- Error correct reads
 1. Run Herro
 2. Run Brutal Rewrite on Herro-corrected reads
 3. Run Peregrine_2021 on Brutal Rewrite-corrected reads
 4. Run DeChat on Peregrine_2021-corrected reads



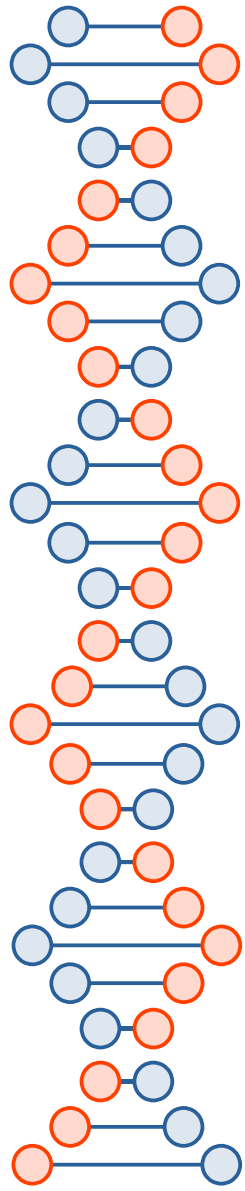
Methods

- Error correct reads
- Align raw & corrected reads to each HG002 haplotype separately



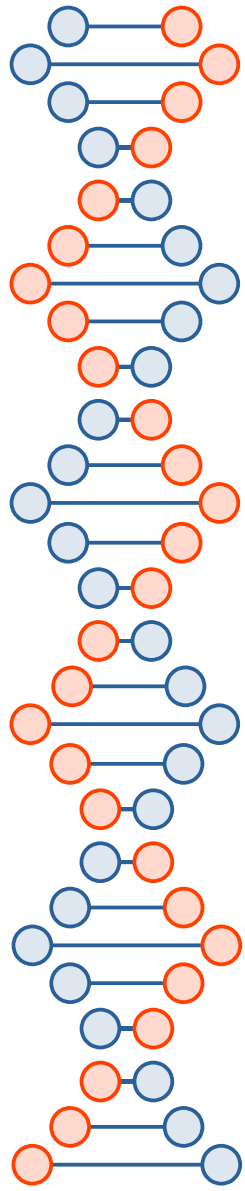
Methods

- Error correct reads
- Align raw & corrected reads to each HG002 haplotype separately
- Collect alignment info



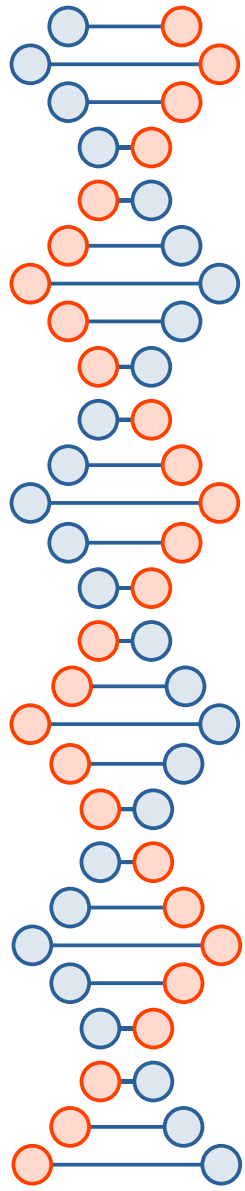
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- Collect alignment info
- Filter alignments (primary alignments, mapping quality Q60, longest alignment block per read)



Methods

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- Collect alignment info
- Filter alignments (primary alignments, mapping quality Q60, longest alignment block per read)
- Analyze k-mers for same reads (~ 1 million)

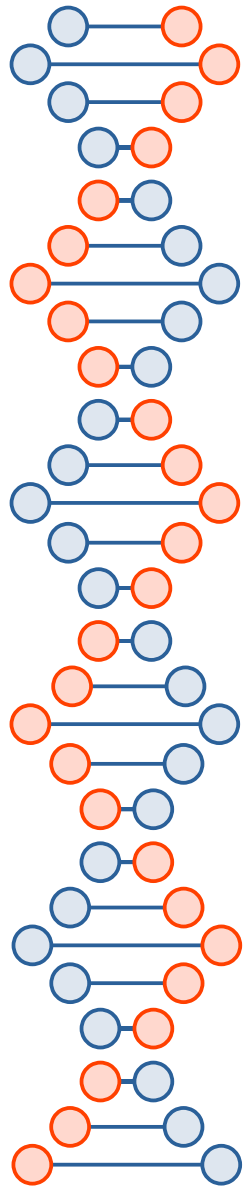


Methods

- Error correct reads
- Align raw & corrected reads to each HG002 haplotype separately
- Collect alignment info
- Filter alignments (primary alignments, mapping quality Q60, longest alignment block per read)
- Analyze k-mers for same reads (~ 1 million)
- Histograms (percent matching readers to hapmers)

K-mers and hapmers

- Keep alignments with same alignment block length on mat. and pat. chrom. (query_start/stop same)





K-mers and hapmers

- Keep alignments with same alignment block length on mat. and pat. chrom. (query_start/stop same)

Alignments to mat. chrom.

query_start query_stop

1 1000

Alignments to pat. chrom.

query_start query_stop

1 1000



K-mers and hapmers

- Count readmers from query_start to query_stop

Alignments to mat. chrom.

query_start query_stop

1 1000

Alignments to pat. chrom.

query_start query_stop

1 1000

K-mers and hapmers

- Count k-mers from each haplotype from each target_start to target_stop

Alignments to mat. chrom.

target_start

target_stop

10000

11000

Alignments to pat. chrom.

target_start

target_stop

20000

21001

K-mers and hapmers

- Count readmers from query_start to query_stop
- Count k-mers from each haplotype from each target_start to target_stop
- Determine hapmers

ATTGCATA “+” strand maternal haplotype

attgcata

ACTGAATA “+” strand paternal haplotype

actgaata

Hapmers



K-mers and hapmers

- Intersect readmers and hapmers

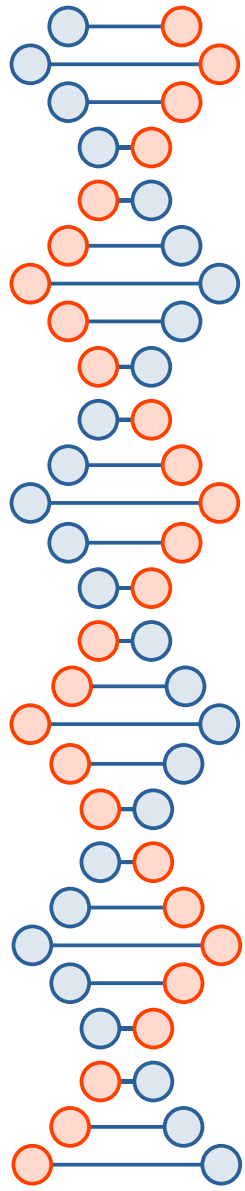
<u>read_id</u>	<u>%match_mat_hapmers</u>	<u>%match_pat_hapmers</u>
seq1	75	25
seq2	25	75
seq3	0	0
seq4	100	0



K-mers and hapmers

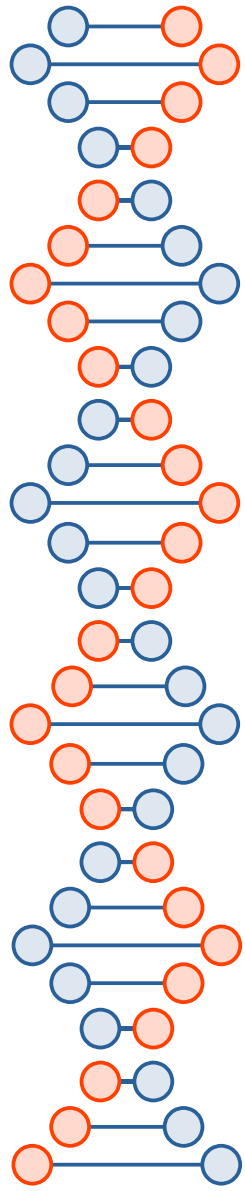
- Intersect readmers and hapmers

<u>read_id</u>	<u>%mat</u>	<u>%pat</u>	<u>calc Abs. Val of diff</u>
seq1	75	25	75-25 = 50%
seq2	25	75	25-75 = 50%
seq3	0	0	Discard
seq4	100	0	100-0 = 100%



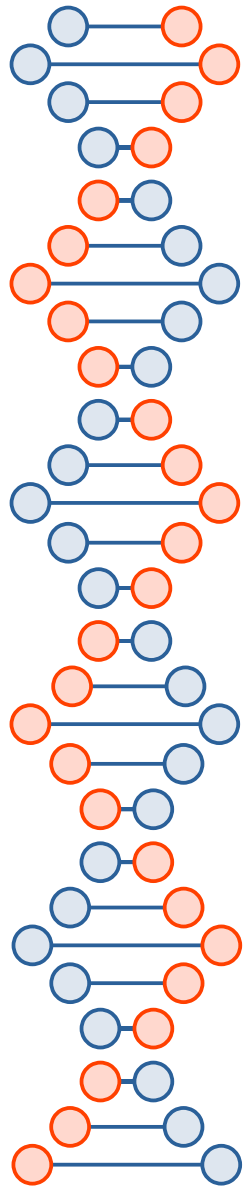
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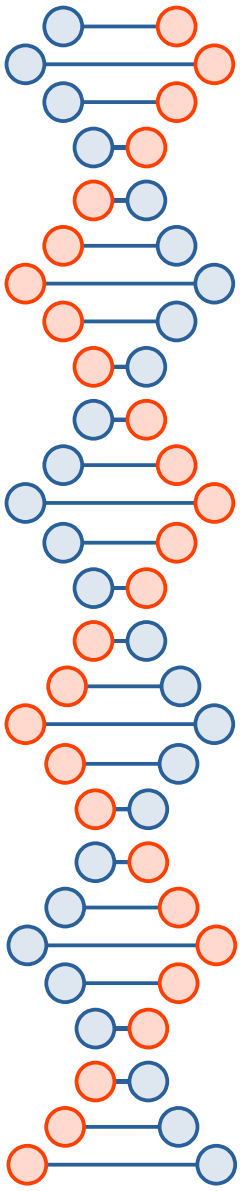
Outline

- **Results**
 - read alignments
 - matching hapmers
 - histograms
 - read error rates



Results

	Total Reads	Maternal Alignments	Paternal Alignments	N50	Coverage
Raw	15,048,314	4,029,514 (26.7772%)	3,962,042 (26.3288%)	21,268	40.3x
Herro	4,578,144	1,733,156 (37.8572%)	1,705,413 (37.2512%)	24,437	26.3x
Brutal Rewrite	4,578,144	1,733,152 (37.8571%)	1,705,385 (37.2506%)	24,437	26.3x
Peregrine_2021	4,490,689	1,724,488 (38.4014%)	1,697,130 (37.7922%)	24,152	25.7x
DeChat	4,490,689	1,724,727 (38.4067%)	1,697,341 (37.7969%)	24,153	25.7x

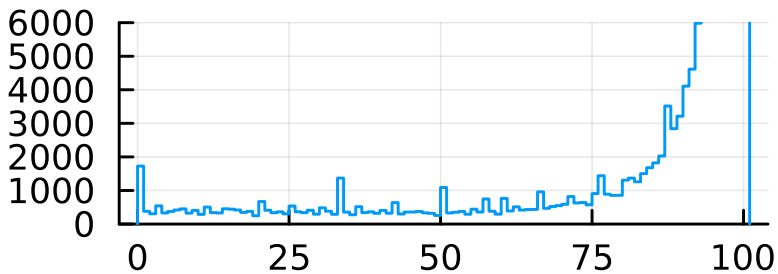


Results

	Read Alignments with Readmers Matching Hapmers	Read Alignments with Readmers Matching 0 Hapmers
Raw	1,009,418	136,325
Herro	1,009,411	136,332
Brutal Rewrite	1,009,411	136,332
Peregrine_2021	1,007,110	138,633
DeChat	1,007,150	138,593

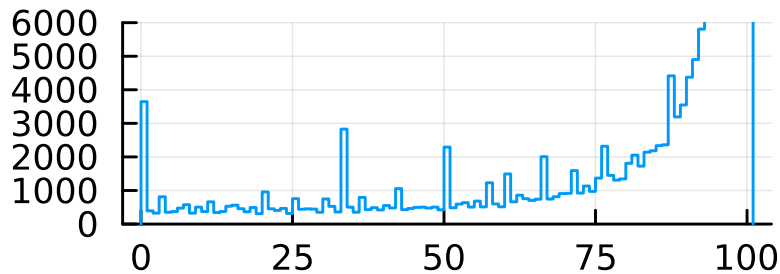
SUP

Num. Reads



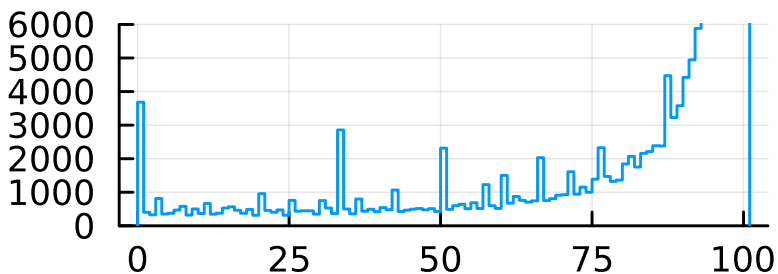
Herro

Num. Reads



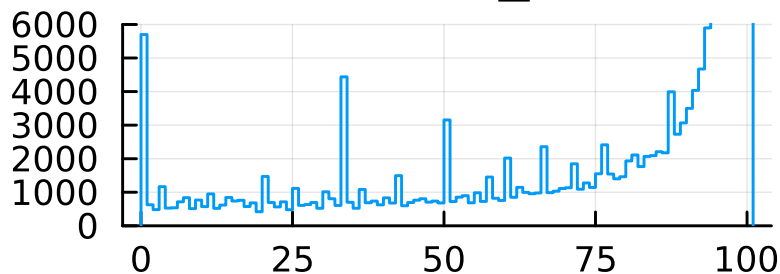
Brutal Rewrite

Num. Reads



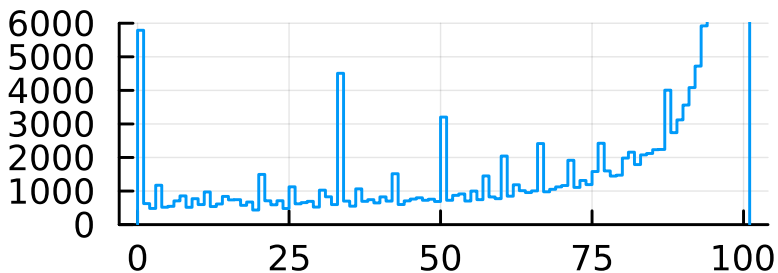
Peregrine_2021

Num. Reads

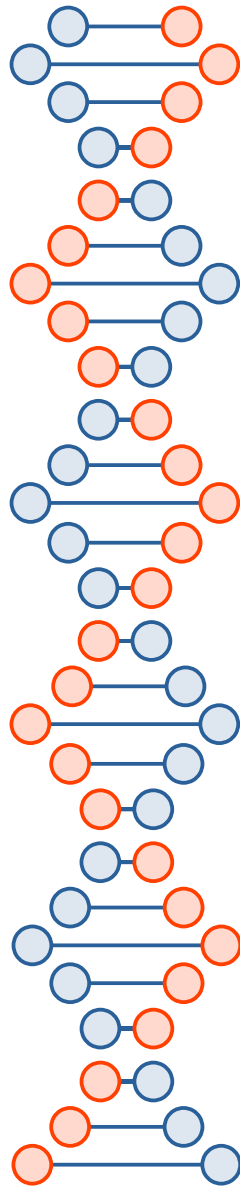


Dechat

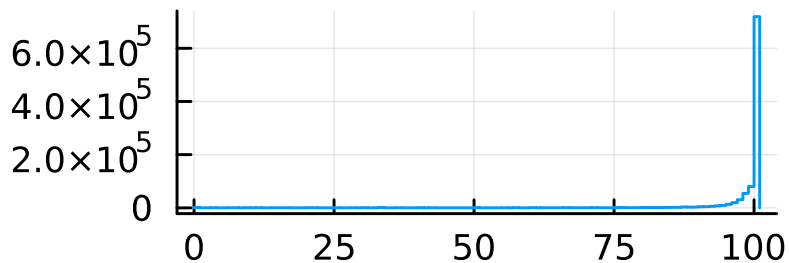
Num. Reads



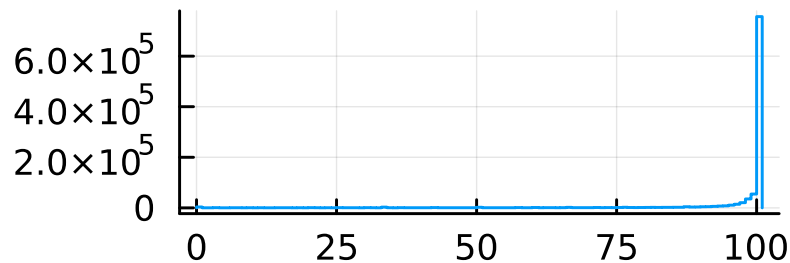
Absolute value of difference in percentage matching hapmers



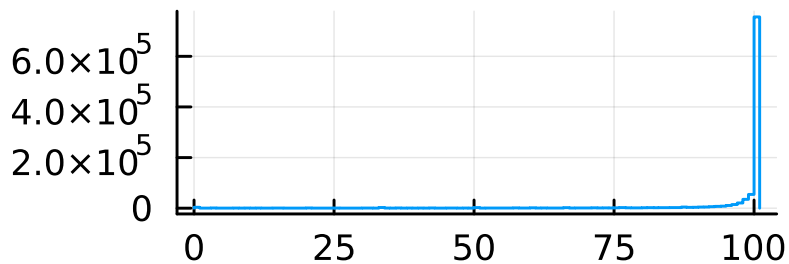
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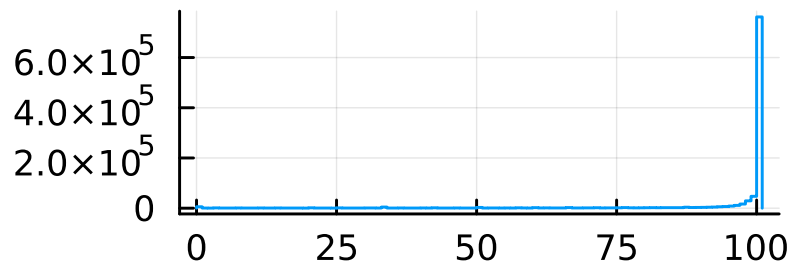
Num. Reads



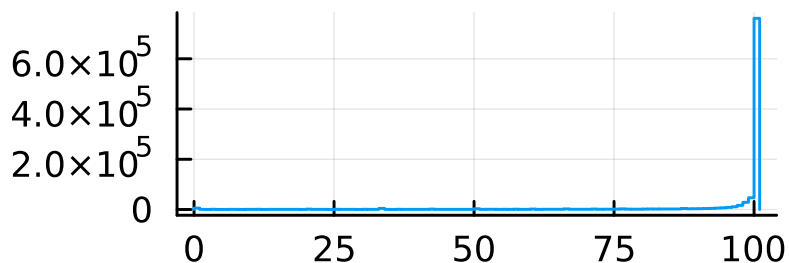
Num. Reads



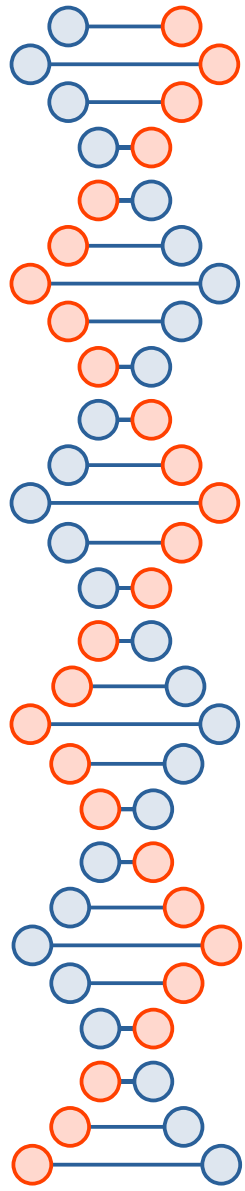
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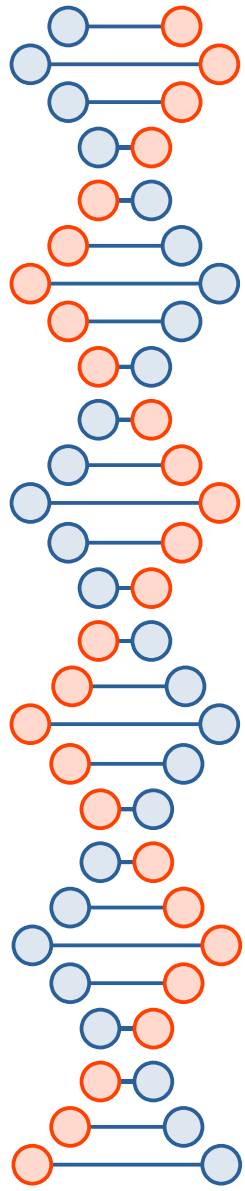
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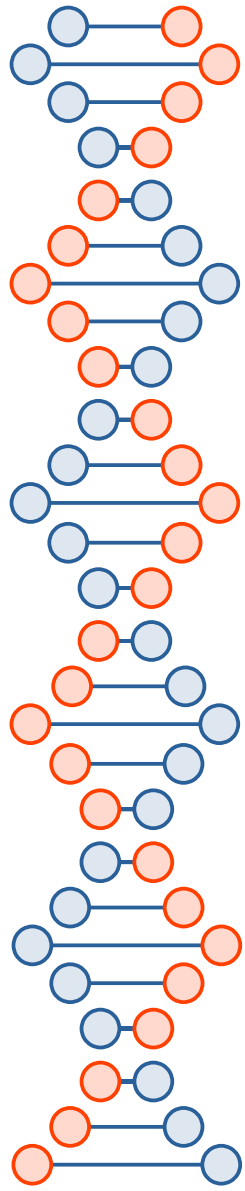
Absolute value of difference in percentage matching hapmers



	Identity	Identity Quality Value	Gap Compres Identity
Raw	0.980335	17.062956	0.984901
Herro	0.999637	34.396470	0.999770
Brutal Rewrite	0.999637	34.398948	0.999772
Peregrine_2021	0.999726	35.619356	0.999830
DeChat	0.999741	35.862487	0.999842
Illumina	0.994539	22.627025	0.994579



	Matches per Kbp	Mismatches per Kbp	Non-hp Inser per Kbp
Raw	985.021819	7.253553	2.826602
Herro	999.725438	0.052275	0.033748
Brutal Rewrite	999.725298	0.051433	0.033581
Peregrine_2021	999.790755	0.031596	0.024639
DeChat	999.803664	0.027125	0.023584
Illumina	994.562660	5.376677	0.01667



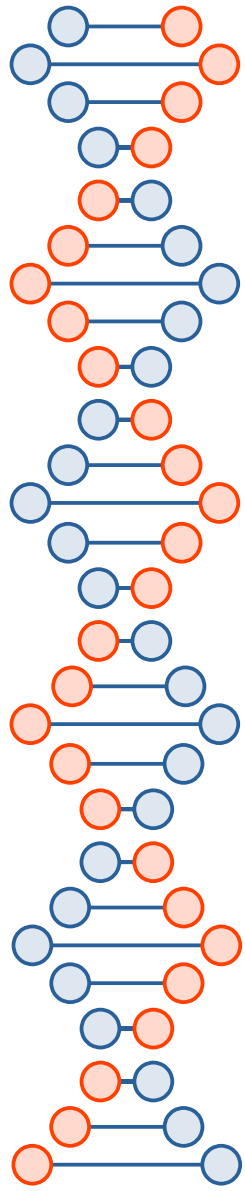
	Non-hp Del per Kbp	Hp Inser per Kbp	Hp Del per Kbp
Raw	3.413819	1.954716	4.310809
Herro	0.044347	0.055095	0.177940
Brutal Rewrite	0.044946	0.054916	0.178323
Peregrine_2021	0.026350	0.040332	0.151300
DeChat	0.022643	0.039365	0.146568

Illumina

0.029777

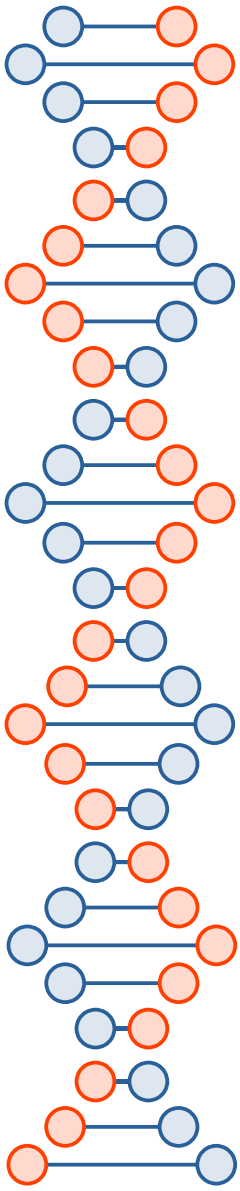
0.007443

0.030885



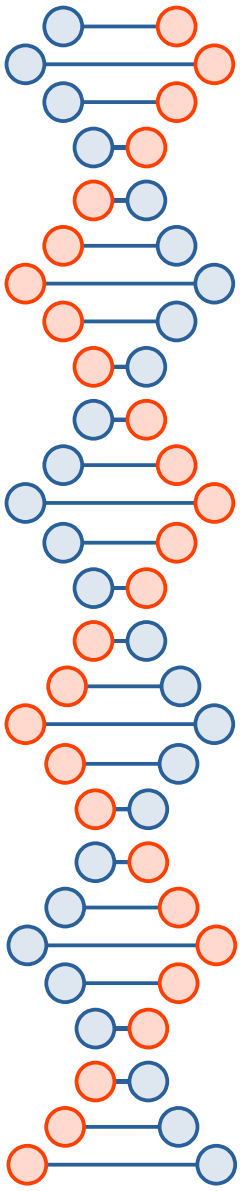
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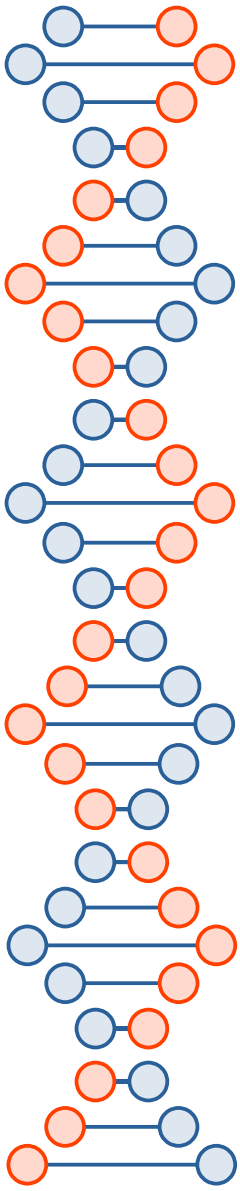
Wrap up

- Can assess phase switching at read level



Wrap up

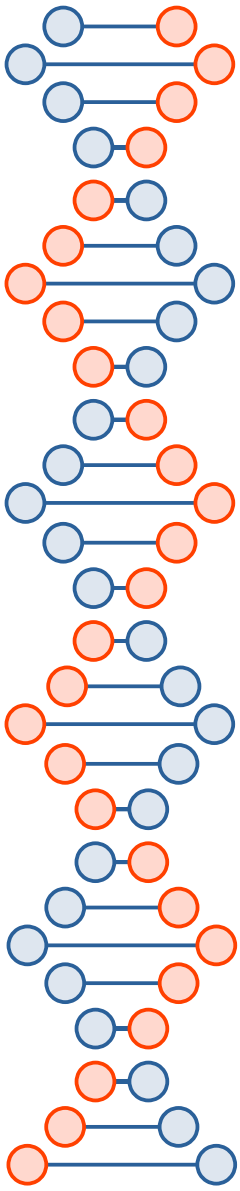
- Can assess phase switching at read level
- Error-correction methods here introduce very little phase switching overall



Wrap up

- Can assess phase switching at read level
- Error-correction methods here introduce very little phase switching overall

Peregrine-2021 may introduce more phase switching than others tested



Wrap up

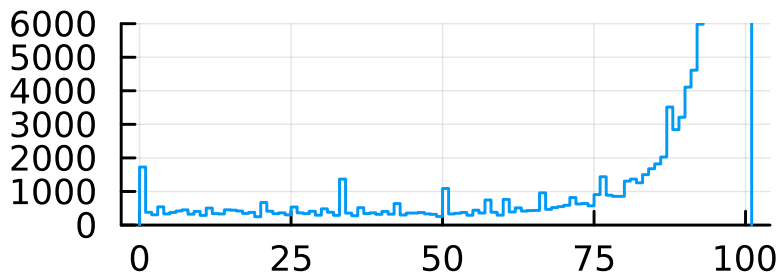
- Can assess phase switching at read level
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- change program settings?

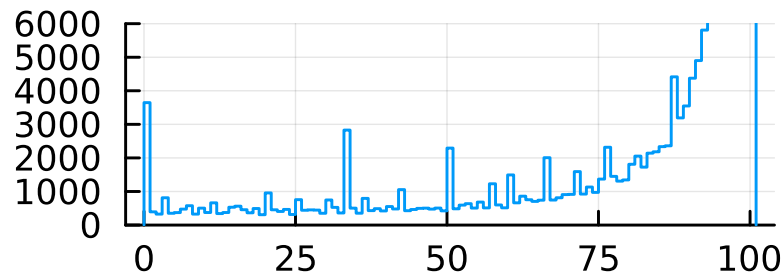
SUP

Num. Reads



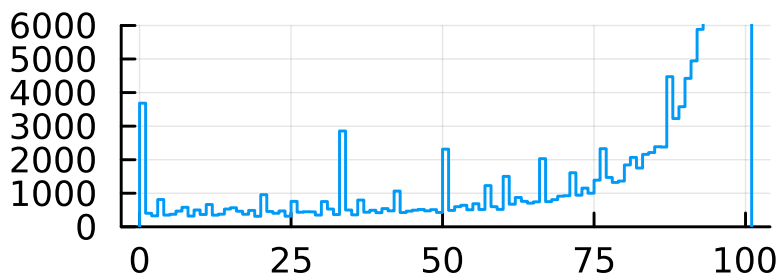
Herro

Num. Reads



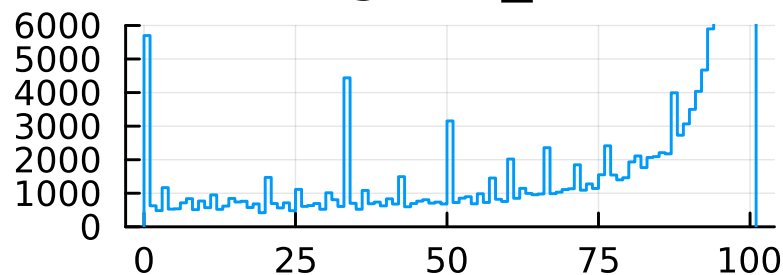
Brutal Rewrite

Num. Reads



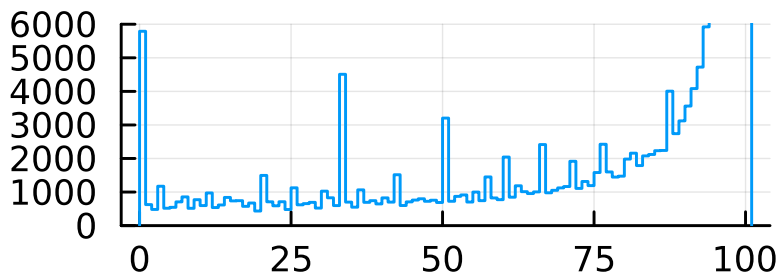
Peregrine_2021

Num. Reads

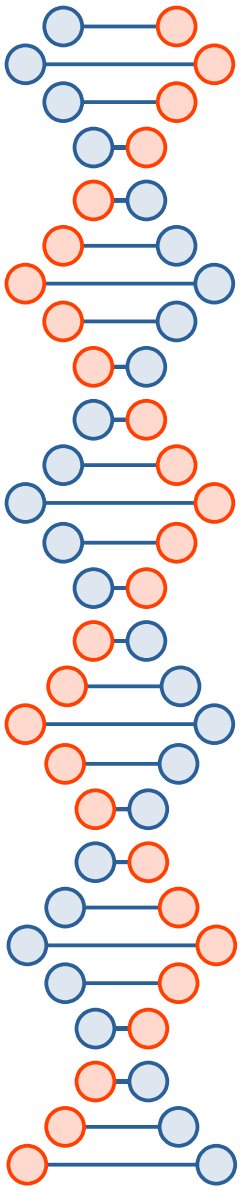


Dechat

Num. Reads

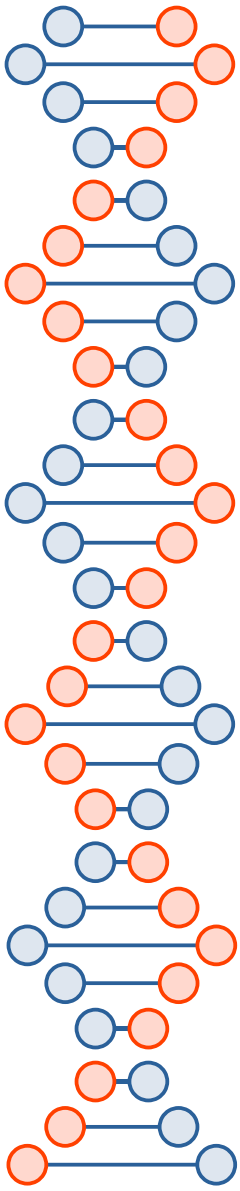


Absolute value of difference in percentage matching hapmers



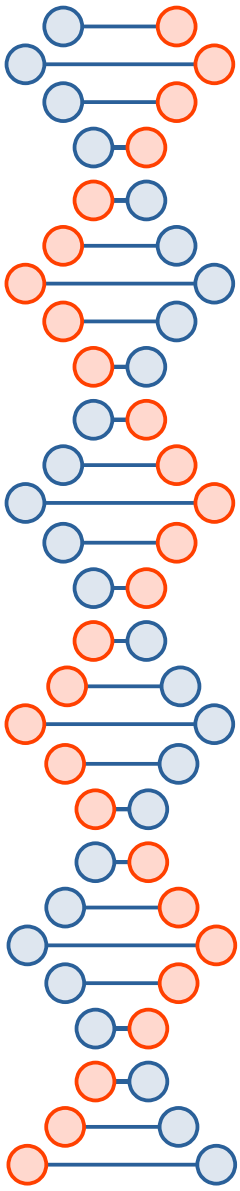
Wrap up

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- Thoughts to explore:



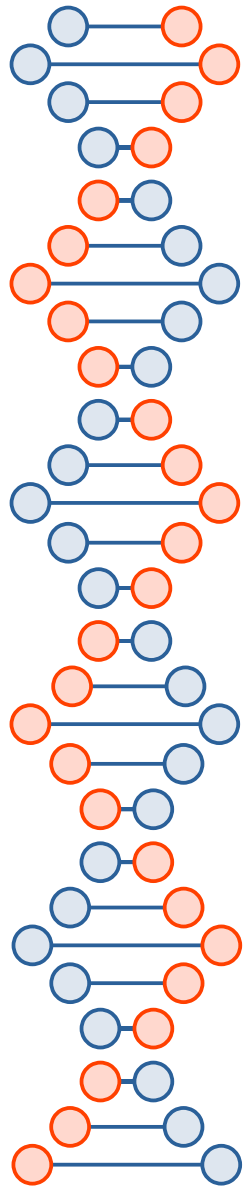
Wrap up

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Wrap up

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- Thoughts to explore:
 - What are reads with 0 matching hapmers?
 - Regions without hapmers



Wrap up

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- Thoughts to explore:
 - What are reads with 0 matching hapmers?
 - Regions without hapmers
 - Regions with few hapmers & phase switching so that readmers do not match hapmers



Acknowledgements

- MedUni Wien High Performance Computing Cluster
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Thanks for your time!

