Option 2: Bead Cleanup – Alternatively a 2X AMPure XP bead clean up for NEXTflex™ mtDNA-Seq Kit

Nina Orellana

Abstract

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Protocol

Step 1.
Add 260 µL of AMPure XP beads to each sample Mix thoroughly.

AMOUNT
250 µl Additional info:

REAGENTS
Agencourt AMPure XP A63880 by Beckman Coulter

Step 2.
Incubate sample at room temperature for 5 minutes.

DURATION
00:05:00

Step 3.
Place the tube on the magnetic rack at room temperature for 5 minutes or until the supernatant appears clear.

DURATION
00:05:00

Step 4.
Remove and discard clear supernatant taking care not to disturb beads. Some liquid may remain in the tube.

Step 5.
Wash #1: With the tubes on the rack, gently add 500 µL of freshly prepared 80% ethanol to each magnetic bead pellet and incubate at room temperature for 30 seconds. Carefully, remove ethanol by pipette.

DURATION
00:00:30

Step 6.
Wash #2: With the tubes on the rack, gently add 500 µL of freshly prepared 80% ethanol to each magnetic bead pellet and incubate at room temperature for 30 seconds. Carefully, remove ethanol by pipette. **Ensure all ethanol has been removed.**

Step 7.

Remove the tube from the magnetic rack and let dry at room temperature for 3 minutes. Do not overdry the beads.

DURATION
00:03:00

Step 8.
Resuspend the dried beads with 42 µL Nuclease-free Water. Mix well by pipetting. Ensure beads are no longer attached to the side of the well.

Step 9.
Incubate resuspended beads at room temperature for 2 minutes.

DURATION
00:02:00

Step 10.
Place the tube on magnetic rack for 5 minutes or until the sample appears clear.

DURATION
00:05:00

Step 11.
Gently transfer 40 µL of clear sample to a fresh microcentrifuge tube.