

# Isolated astrocyte culture preparation - protocol 1

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## ABSTRACT

Isolated astrocyte cell culture preparation. Dissociation, preparation and plating of mice cortex neurons and glia cells on MEA. Isolatioin process of the culture from neurons, to obtain an isolated astrocyte culture.

## PROTOCOL STATUS


### Working

We use this protocol in our group and it is working

### Sample collection

- 1 Dissect and place on ice cortices from pups (post natal day 0 or 1 mice).

### Cell lysis

- 2
  1. Chop with scissors in a papain-based dissociation buffer (2.5 mM CaCl<sub>2</sub>, 0.83 mM EDTA, 137 U papain (Sigma-Aldrich)), 100 µl DNase (Sigma-Aldrich), 3–5 crystals of L-Cysteine (Sigma-Aldrich), HBSS with 20 mM HEPES (pH 7.4);
  2. Place on a rotating shaker for 15 min  00:15:00 at room temperature.

### Cell preparation

- 3
  1. After centrifuging, discard the supernatant, resuspend the pellet in modified essential medium (MEM) without L-glutamine with essential amino acids (Beit Haemek, 06-1025-01-1A), 5% heat-inactivated fetal calf serum (Biological Industries), heat-inactivated 5% horse serum (Beith Haemek, 04-004-1), 2 mM glutamine (Beit Haemek, 03-020-1c), 3 mg/ml glucose, 2% B-27 (Gibco, 17504-044), 0.5% Pen/Strep (100 U/ml penicillin, 100 µg/ml streptomycin; Beit Haemek, 03-031-1B);
  2. Triturate seven times.

### Cell culture

- 4
  1. Plate the cells on poly-D-lysine (PDL, Sigma-Aldrich, P7405-5MG) coated micro-electrode arrays (MEAs; 200/30iR-Ti-gr and 500/30iR-Ti-pr; Multichannel Systems) with a cell density of 2000-2500 cells/mm<sup>2</sup> (~106 cells per dish);
  2. Maintain cultures at 37 °C with 5% CO<sub>2</sub>. Replace growth medium every 3-4 days to astrocyte enriched growth medium: MEM-EAGLE (without L-glutamine, with essential amino acids) (Beit Haemek), 3 mg/ml glucose, 10% heat-inactivated fetal calf serum (Biological Industries), 0.8% GlutaMAX (100X; Gibco), 0.5% Pen/Strep, 2 mM glutamine (Beit Haemek).



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