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Staining of survival motor neuron (SMN) protein in peripheral blood mononuclear cells (PBMC) protocol

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Abstract

Functional SMN protein of peripheral blood-derived mononuclear cells was detected with an anti-SMN antibody labeled with a fluorescent dye and analyzed semiquantitatively using intracellular expression intensity and SMN spot formed in cell nucleus as an index, Consider the relationship with the clinical condition and motor function.

Briefly,

- 1) In order to identify the cell fraction, the fluorescently labeled cell surface antigen-specific antibody is added to the whole blood sample and stained.
- 2) Hemolyze peripheral blood and fix mononuclear cells.
- 3) Intracellular staining is performed with SMN protein and nuclear-specific fluorescently labeled antibody.
- 4) SMN protein expression analysis was performed on a strongly CD33 cell population detected as a cell surface marker, SMN spot detection algorithm was used to analyze the proportion of cells in which intracellular SMN expression and SMN protein aggregated in the nucleus.

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https://www.protocols.io/view/staining-of-survival-motor-neuron-smn-protein-in-p-ggkdvuw

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Materials

- Clear Back MTG-001 by Contributed by users
- BD Phosflow™ Lyse/Fix Buffer (5 x) 5558049 by BD Biosciences
- BD Phosflow™ Perm/Wash Buffer I (10 x) 557885 by BD Biosciences
- BD Pharmingen™ Stain Buffer (FBS) 554656 by BD Biosciences
- BD™ CompBeads 51-90-9001229 by BD Biosciences

- ✓ AF488-anti human SMN mAb by Contributed by users
- AF488-MOPC21 by BioLegend
- BV610-anti human CD3 mAb by BioLegend
- R-PE-anti human CD19 mAb by BioLegend
- ✓ PE-Cy5-anti human CD33 mAb by Contributed by users
- ✓ Hoechst 33342 H3570 by Contributed by users

Protocol

Staining cell surface molecules

Step 1.

- (1) Dispense 1.5 mL of heparin-treated whole blood into 15 mL_conical tube
- (2) Add 30 μL of normal human Ig solution (Clear Back, MTG-001, MBL, Nagoya, Japan)
- (3) Gently shake, leave in the dark at room temperature for 15 minutes
- (4) Add 5 μL each of PE-HIB19, PE-Cy5-WM53, BV610-UCHT1 (Biolegend)
- (5) Gently shake, leave in the dark at room temperature, for 30 minutes

Hemolysis of erythrocytes and immobilization of cell membranes

Step 2.

- (6) Warm lysis/fixation buffer and PBS (-) to 37 °C in a water bath
- (7) Add 10 mL of pre-warmed lysis/fixation buffer (37 °C) and mix immediately upside down
- (8) Incubate in a water bath at 37 °C for 10 minutes
- (9) Mix by inverting, centrifuge (900 \times q, 5 min)
- (10) Discard the supernatant using an aspirator
- (11) Resuspended in PBS (-) (37 °C)
- (12) Centrifuge (900 \times g, 5 min)
- (13) Discard supernatant

Permeabilaization of transmembrane of cells

Step 3.

- (14) Resuspend with 1.5 mL of permeabilization buffer
- (15) Incubate at room temperature in the dark for 30 min
- (17) Discard supernatant

- (18) Resuspended with 1.5 mL of permeabilization buffer
- (19) Centrifugation (900 \times g, 5 min)
- (20) Discard supernatant

Cell count and Intracellular staining

Step 4.

- (21) Cell count
- (22) Dispensing the 1 x 10^6 cells/50 μ L in Permeabilization buffer into two microcentrifuge tubes (for stained with specific Ab and isotype control)
- (23) Tube for SMN staining: Add 1 µg of AF488-2B1
- (24) Tube for isotype control staining: Add 1 test of AF488-MOPC21 (5 μL)
- (25) Incubate stained cells (both 23 and 24) at room temperature in the dark for 45 min
- (26) Add 500 μ L of permeabilization buffer and centrifuge (500 \times g, 5 min)
- (27) Repeat (26) twice, and discard supernatant

Nuclear staining

Step 5.

- (28) Suspend in 50 μL of Hoechst 33342 (final concentration: 0.25 μg/mL)
- (29) Incubate in the dark at room temperature for 15 min
- (30) Add 500 μ L of PBS (-) and centrifuged (500 \times g, 5 min)
- (31) Discard supernatant
- (32) Suspend with 50 μL of PBS (-)
- (33) Analysis