

# Influenza A(H1)pdm09 virus TaqMan (SwFluH1) assay

Ian Mackay, Judy Northill, Alyssa Pyke, Bruce Harrower

#### **Abstract**

This assay was designed and developed by Alyssa Pyke and Bruce Harrower at this laboratory aided by design software in use at the time.

The assay specifically targets the haemagglutinin (HA) region of influenza A(H1)pdm09 virus strains and is designed as a qualitative screening test for human cases of infection, but not for infection due to other known influenza viruses.

The assay aims to detect any circulating seasonal H1N1 influenza strains, not only influenza A(H1)pdm09 virus.

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## **Before start**

- If using a different brand or model of real-time thermocycler, check the concentration of ROX is adequate.
- Method assumes the user is familiar with the thermocycler and software used to run the protocol and with PCR in general.

#### **Materials**

SuperScript™ III Platinum™ One-Step qRT-PCR Kit 11732088 by Life Technologies

#### **Protocol**

## Oligonucleotide seguences

# Step 1.

Name	Sequence 5'-3'
SwFluH1for	CCCCATTGCATTTGGGTAAA
SwFluH1rev	TGGAGAGTGATTCACACTCTGGAT
SwFluH1prob	6FAM - TAACATTGCTGGCTGGATCCTGGGA- BHQ-1

#### Reagents

## Step 2.



SuperScript™ III Platinum™ One-Step qRT-PCR Kit 11732088 by Life Technologies

## Reaction set-up

## Step 3.

The assay has been used on both a Rotor-Gene 6000 and a Rotor-Gene Q real-time thermocycler

Prepare sufficient mix for the number of reactions.

Include a suitable 'dead volume' as necessary if using a robotic dispenser.

# **MIX PREPARATION**

Reagent	Volume (µl) x1	Final reaction concentration
Nuclease-free water	4.37	N/A
SwFluH1fwd 200pmol/μl	0.09	900nM
SwFluH1rev 200pmol/μl	0.07	700nM
SwFluH1Prb 100pmol/μl	0.03	150nM
2X Reaction Mix <sup>1</sup>	10	1X
SuperScript® III/Platinum® <i>Taq</i> Mix <sup>1</sup>	0.4	1X
ROX Reference Dye (25μM)	0.04	0.05μΜ
Template	5	N/A
TOTAL	20	

¹Superscript™III Platinum™ One-step qRT-PCR kit

- Dispense 15µL to each reaction well.
- Add  $5\mu L$  of template (extracted RNA, controls or NTC [nuclease-free water] ).
- Total reaction volume is 20μL

#### **Amplification**

## Step 4.

# **CYCLING CONDITIONS**

50°C	5min	1X
95°C	2min	1X
95°C	3sec	40X
60°C	30sec <sup>1</sup>	

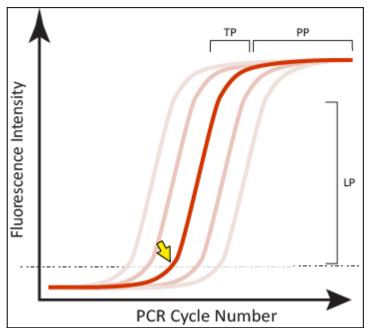
<sup>&</sup>lt;sup>1</sup>Florescence acquisition step

# **Result Analysis**

## Step 5.

The definition used for a satisfactory positive result from a real-time fluorogenic PCR should include each of the following:

- 1. A **sigmoidal curve** the trace travels horizontally, curves upward, continues in an exponential rise and followed by a curve towards a horizontal plateau phase
- 2. A **suitable level of fluorescence** intensity as measured in comparison to a positive control (y-axis)
- 3. A **defined threshold (C\_T) value** which the fluorescent curve has clearly exceeded (Fig.1 arrow), which sits early in the log-linear phase and is <40 cycles
- 4. A flat or non-sigmoidal curve or a curve that crosses the threshold with a  $C_T > 40$  cycles is considered a negative result.
- 5. NTCs should not produce a curve



**Figure 1**. Examples of satisfactory sigmoidal amplification curve shape when considering an assay's fluorescent signal output. The crossing point or threshold cycle ( $C_T$ ) is indicated (yellow arrow); it is the value at which fluorescence levels surpass a predefined (usually set during validation, or arbitrary) threshold level as shown in this normalized linear scale depiction. LP-log-linear phase of signal generated during the exponential part of the PCR amplification; TP-a slowing of the amplification and accompanying fluorescence signal marks the transition phase; PP-the plateau phase is reached when there is little or no increase in fluorescent signal despite continued cycling.