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#### **Nat Brown**

## **Abstract**

Transformation of heat-shock competent E. coli cells

Citation: Nat Brown Transformation of competent E.coli cells with plasmid DNA. protocols.io

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## **Before start**

For incubation on ice, make sure the tubes are standing in an ice-water mix, because without water, the cooling effect of ice is not reproducible due to the air between the ice fragments, especially if you have to incubate for a certain period of time.

## **Protocol**

## Step 1.

Thaw the appropriate amount of competent cells on ice.

## NOTES

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The appropriate amount refers to the amount you will need to all transformations you need to do. If you need to do one, thaw 100  $\mu$ l (or the next biggest aliquot size). If you need to do six, thaw 600  $\mu$ l (minimum, or the closest volume you can from the size of the aliquots you have).

#### Step 2.

Pre-chill the required number of empty 1.5 ml microcentrifuge tubes.

## Step 3.

Pipet 100 µl aliquots of cells into the pre-chilled tubes.

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Add 5-10 µl of a ligation reaction mix or 5 ng of pure plasmid DNA to each tube. Mix gently!

## Step 5.

Incubate the tubes of ice for 30 min

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00:30:00

## Step 6.

Heat shock the cells for 45 sec at 42°C

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00:00:45

## Step 7.

Place the tubes immediately on ice for at least 2 min

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00:02:00

## Step 8.

Add 1000 µl of SOC medium to each tube and incubate for 1 hour at 37°C with shaking.

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01:00:00

## NOTES

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LB medium can be used instead on SOC with a small loss in efficiency.

## Step 9.

Transfer the cultures to 1.5 ml microcentrifuge tubes and spin for 1 min at 10000 x g.

**O** DURATION

00:01:00

#### NOTES

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Before proceeding with this step, consider how many transformants you expect. Unless you expect a low yield (<200) plate 100  $\mu$ l (or less) before concentrating the bacteria. By plating two or more amounts of bacteria you increase your chances of having well isolated colonies on your plates the next day.

## Step 10.

Remove 800 µl of the supernatant and resuspend the pellet.

## NOTES

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This can be achieved, with just a little practice, by gently decanting the supernatant without trying to fully remove all the liquid. With a little experience, decanting and leaving 100 - 200  $\mu$ l remaining in the tube will be routinely achieved.

## **Step 11.**

Plate out the suspension on a LB agar plate containing the appropriate antibiotic.

#### **Step 12.**

Incubate the plates overnight at 37°C.

# Warnings

If you notice a significant drop in colony numbers after several transformations with plasmid DNA it's time to prepare fresh competent cells.