

Nuclear Factor Fixation and Permeabilization Staining Protocol

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Abstract

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Guidelines

Prior to following the staining procedure, dilute one (1) part Nuclear Factor Fixation Buffer (4X) with three (3) parts PBS, and dilute one (1) part Nuclear Factor Permeabilization Buffer with nine (9) parts PBS.

Protocol

Step 1.

Prepare target cells of interest and perform surface staining as described in BioLegend's [Cell Surface Immunofluorescence Staining Protocol](#)

Step 2.

Fix the cells with 1 ml/tube BioLegend's Nuclear Factor Fixation Buffer, at room temperature in the dark for 20 minutes.

 DURATION

00:20:00

Step 3.

Centrifuge at 250 x g for 5 minutes; discard supernatant.

 DURATION

00:05:00

Step 4.

Wash once with 1 ml BioLegend's Nuclear Factor Permeabilization Buffer (1x).

Step 5.

Resuspend cells in 1 ml BioLegend's Nuclear Factor Permabilization Buffer (1x), incubate at room temperature in the dark for 20 minutes, spin down cells and discard the supernatant; then resuspend the pellet in 100 ul of BioLegend's Nuclear Factor Permeabilization Buffer (1x).

 DURATION

00:20:00

Step 6.

Add appropriate amount of flurochrome-conjugated antibodies for nuclear target of interest, and incubate at room temperature in the dark for 30 minutes.

 DURATION

00:30:00

Step 7.

Wash twice with Cell Staining Buffer, and resuspend in 0.5 ml Cell Staining Buffer, then analyze with flow cytometer with appropriate instrument settings.