

# Dengue virus type 4 (DENV-4) TaqMan (DENV4-TM2017) assay

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## Abstract

This protocol was designed and developed at this laboratory.

The assay specifically targets the 3' UTR region of DENV-4 strains and is designed as a qualitative screening test for human cases of DENV-4 infection, but not for infection due to other known DENVs.

**Citation:** Ian Mackay, Judy Northill Dengue virus type 4 (DENV-4) TaqMan (DENV4-TM2017) assay. **protocols.io** dx.doi.org/10.17504/protocols.io.n7pdhmn

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## Before start

- If using a different brand or model of real-time thermocycler, check the concentration of ROX is adequate.
- Method assumes the user is familiar with the thermocycler and software used to run the protocol and with PCR in general.

## Materials

SuperScript® III First-Strand Synthesis System [18080-051](#) by [Thermo Scientific](#)

## Protocol

### Oligonucleotide sequences

#### Step 1.

Name	Sequences (5'-3')
D4-3UTR-for	CTGTAGCTCCGCCAAHAAYGG
D4-3UTR-rev	CTCCTCTAACCRCCTAGTCCAATA
D4-3UTR-Prb	6FAM - TGCGCCACGGAAGCTGTACGCGTGG - BHQ

## Reagents

## Step 2.



### REAGENTS

SuperScript™ III Platinum™ One-Step qRT-PCR Kit [11732088](#) by [Life Technologies](#)

## Reaction set-up

### Step 3.

Reagent	Vol (μl) x1	Final reaction concentration
Nuclease-free water	4.37	N/A
D4-3UTR-for (200μM)	0.07	700nM
D4-3UTR-rev (200μM)	0.09	900nM
D4-3UTR-Prb (100μM)	0.03	150nM
2X Reaction Mix <sup>1</sup>	10	1X
ROX reference dye (25μM)	0.04	50nM
SuperScript™ III/Platinum™ Taq Mix <sup>1</sup>	0.4	1X
TOTAL	15	

<sup>1</sup>Superscript™III Platinum™ One-step qRT-PCR kit

- Dispense 15μL to each reaction well.
- Add 5μL of template (extracted RNA, controls or NTC [nuclease-free water] ).
- Total reaction volume is 20μL

## Amplification

### Step 4.

50°C	5min	1X
95°C	2min	1X
95°C	3sec	40X
60°C	30sec <sup>1</sup>	

<sup>1</sup>Fluorescence acquisition step

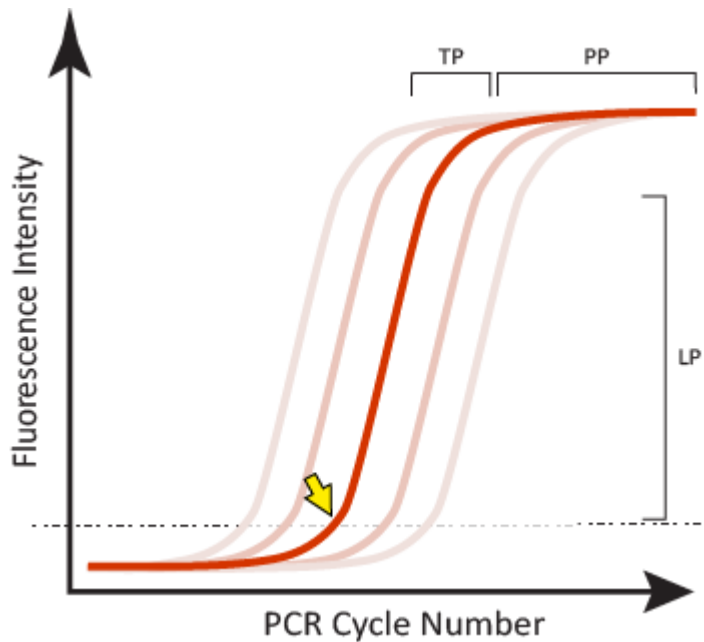
## Result analysis

### Step 5.

The definition used for a satisfactory positive result from a real-time fluorogenic PCR should include each of the following:

1. A **sigmoidal curve** – the trace travels horizontally, curves upward, continues in an exponential rise and followed by a curve towards a horizontal plateau phase
2. A **suitable level of fluorescence** intensity as measured in comparison to a positive control (y-axis)
3. A **defined threshold (C<sub>T</sub>) value** which the fluorescent curve has clearly exceeded (Fig.1 arrow), which sits early in the log-linear phase and is <40 cycles

A flat or non-sigmoidal curve or a curve that crosses the threshold with a  $C_T > 40$  cycles is considered a negative result. NTCs should not produce a curve.



**Figure 1.** Examples of satisfactory sigmoidal amplification curve shape when considering an assay's fluorescent signal output. The crossing point or threshold cycle ( $C_T$ ) is indicated (yellow arrow); it is the value at which fluorescence levels surpass a predefined (usually set during validation, or arbitrary) threshold level as shown in this normalized linear scale depiction. LP-log-linear phase of signal generated during the exponential part of the PCR amplification; TP-a slowing of the amplification and accompanying fluorescence signal marks the transition phase; PP-the plateau phase is reached when there is little or no increase in fluorescent signal despite continued cycling.