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## $holdsymbol{lpha}$ Total DNA extraction from plant tissue using CTAB method

Forked from Total DNA extraction from microalgae using CTAB method

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## MATERIALS TEXT

**2X CTAB Buffer:** 100 Mm Tris pH 8.0, 1.4 M NaCl, 20 mM EDTA, 2% CTAB, 1% PVP40, and 0.5% (v/v) beta-mercaptoethanol (added daily before the extraction)

- 1 Using a pre-chilled mortar and pestle, grind frozen plant tissue (1-5g grams) into a fine powder
- 2 Add 500 ul of pre-heated 2 x CTAB buffer (with beta-mercaptoethanol added) to the pelleted cells. Vortex and re-suspend by gently pipetti

ng up and down. § 60 °C

- Incubate at 60 C for 30 min. Periodically, mix gently during the incubation. § 60 °C © 00:30:00
- 4 Add an equal volume of chloroform:isoamyl alcohol (24:1), mix very well at RT for 5 min to generate a monophasic solution.
- 5 Centrifuge for 10 min in 5,000 x g
- 6 Transfer aqueous phase (upper layer) to new tube.
- 7 (Optional) Repeat chloroform:isoamyl alcohol extraction (steps 3 and 4) until the interface is clean.
- 8 Add 2/3 volumes of isopropanol, mix gently for 5 min at RT.
- 9 Centrifuge for 15 min at 10,000 x g. Discard supernatant.
- 10 Add 500 uL of 70% ethanol to the DNA pellet, and briefly vortex to dislodge the pellet.
- 11 Air dry pellet for 30 min. Resuspend in 100ul of TE Buffer.
- 12 Add 1 ug/mL DNase-free RNase (final concentration) and incubate at 37C for 30 min.
- Extract the DNA once with an equal volume of phenol:chloroform:isoamyl alcohol (25:24:1). Centrifuge at 10,000x g for 10 min.

  Transfer the aqueous phase to a clean tube.

- 14 Precipitate DNA with 1/3 volume of 7.5 M ammonium acetate and 2.5 volumes of 100% ethanol.
- 15 Centrifuge at 10,000x g for 10 min, wash with 70% ethanol, and air dry for 30 min at RT. Resuspend in 50 ul of TE Buffer. Allow the DNA to dissolve overnight at 4C

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