



Jul 12, 2019

## Isolation of total DNA from *Synechocystis* sp. PCC 6803

Anna Behle<sup>1</sup><sup>1</sup>Institute for Synthetic Microbiology, Heinrich-Heine University, Düsseldorf**1** Works for me [dx.doi.org/10.17504/protocols.io.gufbwtn](https://doi.org/10.17504/protocols.io.gufbwtn)[Axmann Lab](#) [CyanoWorld](#) [view 1 more group](#)Anna Behle  
Institute for Synthetic Microbiology ⚡ 🌞 🌱

### ABSTRACT

This protocol can be used for extraction of total genomic DNA from *Synechocystis* sp. PCC 6803.

### SAFETY WARNINGS

Wear goggles, a lab coat and gloves when handling chloroform and phenol. Work under the fume hood.

Wear goggles when handling liquid nitrogen.

#### Buffers required:

- 1 TE-Puffer : 10 mM Tris/HCl ; 1 mM EDTA ; pH 8.0  
TE+S-Puffer : 25% (w/v) Sucrose; 50 mM Tris/HCl ; 1 mM EDTA, pH 8.0

#### Culturing

- 2 Grow 50 mL *Synechocystis* culture to the end of the logarithmic phase ( $OD_{750} \approx 1.0$ ).  
Centrifuge at 4800 rpm and 4°C for 7 minutes. Remove supernatant.

#### Wash steps:

- 3 Resuspend cells in 10 mL TE-buffer, centrifuge at 4800 rpm and 4°C for 7 minutes. Remove supernatant.
- 4 Repeat washing step. Resuspend pellet in 1 mL TE-buffer. Flash freeze in liquid nitrogen or dry ice.

#### Cell lysis

- 5 Add  $5 \text{ mg} \cdot \text{mL}^{-1}$  lysozyme (optional: + 100 mM EDTA).  
Incubate 1 hour at 37°C.  
🕒 01:00:00
- 6 Add 14.4  $\mu\text{L}$  Proteinase K (20 mg/mL stock) + 100  $\mu\text{L}$  20 % SDS.  
Incubate 1 hour at 60 °C. (Alternatively, incubation can be carried out at 37 °C for 16 hours).  
🕒 01:00:00



#### DNA extraction:

- 7 Add 1 volume of Phenol/Chloroform (1:1 v/v). Mix well.



Centrifuge for 10 min at 13000 rpm and 4° C.

- 8 Transfer upper, aqueous phase to a fresh tube.  
Add 1 volume of Phenol/Chloroform (1:1 v/v). Mix well.  
Centrifuge for 10 min at 13000 rpm and 4° C.
- 9 Transfer upper, aqueous phase to a fresh tube.  
Add 1 volume of Chloroform in order to remove residual phenol. Mix well.  
Centrifuge for 10 minutes at 13000 rpm and 4° C.

#### DNA precipitation

- 10 Add 0.7 volumes of isopropanol to the sample. Incubate for 5 minutes at RT.  
 00:05:00
- 11 Centrifuge for 30 minutes at 13000 rpm and 4° C.
- 12 Wash pellet with 1 mL 70 % EtOH.  
Centrifuge for 10 minutes at 13000 rpm and 4°C.
- 13 Air-dry pellet for 1 hour.  
 01:00:00

#### RNase treatment:

- 14 Resuspend DNA pellet in 30-100 µL TE-buffer.  
Add 2 µL RNase A  
Incubate 30 minutes at 37°C.  
 00:30:00
- 15 Heat inactivate RNase A by incubating for 10 minutes at 72 °C.  
 00:10:00



This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited