

# Th2 Polarization of Mouse CD4+ Cells

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## Abstract

**Citation:** BioLegend, Inc. Th2 Polarization of Mouse CD4+ Cells. [protocols.io](https://doi.org/10.17504/protocols.io.ezqbf5w)

[dx.doi.org/10.17504/protocols.io.ezqbf5w](https://doi.org/10.17504/protocols.io.ezqbf5w)

**Published:** 26 May 2016

## Guidelines

### Reagent List:

- Sterile PBS
- Cell culture medium (RPMI 1640 supplemented with 10% FBS)
- Sterile plastic petri dishes
- Sterile T-75 culture flask
- RBC Lysis Buffer (Cat. No. 420301)
- Concanavalin A (Con A) (Sigma, Cat. No. C5275)
- Anti-mouse CD3 $\epsilon$ , clone 145-2C11 (LEAF™ format, Cat. No. 100314)
- Anti-mouse CD28, clone 37.51, (LEAF™ format, Cat. No. 102112)
- Recombinant mouse IL-2 (carrier-free) (Cat. No. 575402)
- Recombinant mouse IL-4 (carrier-free) (Cat. No. 574302)
- Monensin Solution (Cat. No. 420701)

## Materials

RBC Lysis Buffer [420301](#) by [BioLegend](#)

Anti-mouse CD3 $\epsilon$ , clone 145-2C11 (LEAF™ format) [100314](#) by [BioLegend](#)

Anti-mouse CD28, clone 37.51, (LEAF™ format) [102112](#) by [BioLegend](#)

Recombinant mouse IL-2 (carrier-free) [575402](#) by [BioLegend](#)

Monensin Solution (1,000X) [420701](#) by [BioLegend](#)

Recombinant Mouse IL-4 (carrier-free) [574302](#) by [BioLegend](#)

Concanavalin A from Canavalia ensiformis (Jack bean) [C5275](#) by [Sigma Aldrich](#)

## Protocol

### Isolation of CD4+ Cells From Lymph Nodes

#### Step 1.

Harvest lymph nodes (superficial cervical, mandibular, axillary, inguinal, and mesenteric) from mice.

#### Isolation of CD4+ Cells From Lymph Nodes

##### Step 2.

Tease lymph nodes through a sterile 70-µm nylon cell strainer to obtain single-cell suspensions incomplete RPMI containing 10% FCS (complete medium).

#### Isolation of CD4+ Cells From Lymph Nodes

##### Step 3.

Resuspend cells in complete medium and use your favorite method to isolate CD4+ cells. Checkout [Biocompare.com](https://www.biocompare.com) to find useful kits.

#### Th2 Polarization of CD4+ Cells

##### Step 4.

On day 0, plate CD4<sup>+</sup> cells at 30 x10<sup>6</sup> /30 ml/T-75 flask.

#### Th2 Polarization of CD4+ Cells

##### Step 5.

Culture cells for 3 days in complete RPMI containing 10% FCS, Con A (5 µg/mL), recombinant mouse IL-2 (20 ng/ml), and recombinant mouse IL-4 (50 ng/ml).



DURATION

12:00:00

#### Th2 Polarization of CD4+ Cells

##### Step 6.

On day 3, harvest the cells and wash cells once.

#### Th2 Polarization of CD4+ Cells

##### Step 7.

Seed 15 x10<sup>6</sup> cells/30 ml/T-75 flask along with recombinant mouse IL-2 (20 ng/ml) and recombinant mouse IL-4 (50 ng/ml).

#### Th2 Polarization of CD4+ Cells

##### Step 8.

On day 5, coat a 60 x 15 mm tissue culture petri dish with anti-mouse CD3ε, clone 145-2C11, 10 µg/mL in PBS, 5 ml/dish. Incubate in a 4°C refrigerator overnight.



DURATION

16:00:00

#### Th2 Polarization of CD4+ Cells

##### Step 9.

On day 6, wash the anti-mouse CD3ε pre-coated tissue culture petri dish with PBS.

#### Th2 Polarization of CD4+ Cells

##### Step 10.

Harvest the cells from the flask (that were seeded on Day 5).

#### Th2 Polarization of CD4+ Cells

##### Step 11.

Wash the cells. (wash 1/2)

#### Th2 Polarization of CD4+ Cells

##### Step 12.

Wash the cells. (wash 2/2)

#### Th2 Polarization of CD4+ Cells

##### Step 13.

Seed at  $20 \times 10^6$  /10 ml/petri dish along with 10  $\mu$ l of monensin (1000x) and anti-mouse CD28, clone 37.51 (5  $\mu$ g/ml).

#### Th2 Polarization of CD4+ Cells

##### Step 14.

Incubate for 6 hours at 37°C in a CO<sub>2</sub> incubator.

 DURATION

06:00:00

#### Th2 Polarization of CD4+ Cells

##### Step 15.

After harvesting, the cells are ready for staining.