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## gRNA design and cloning with Sapl into Loop plasmid L2\_lacZgRNA-Cas9-CsA

Forked from [gRNA design and cloning into Loop L2 plasmids \(L2\\_gRNA-Cas9-CsA and L2\\_gRNA-CsA plasmids\)](#)

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1 Works for me dx.doi.org/10.17504/protocols.io.93wh8pe

OpenPlant Project



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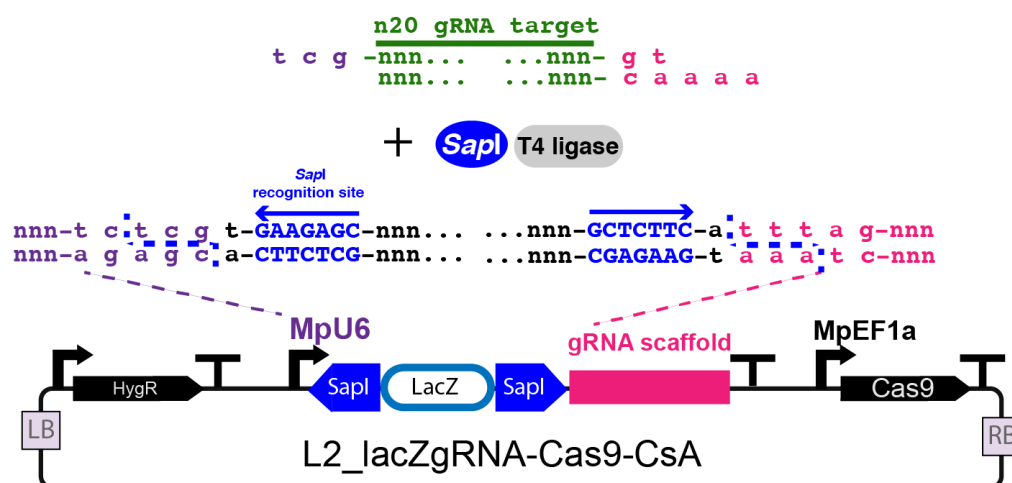


### ABSTRACT

This protocol explains how to design and clone the guide RNA target sequence into a L2 plasmid ready to accept the gRNA by cloning with Sapl (L2 plasmid also contains a cassette to express Cas9)

### Summary of design of gRNA and cloning into L2 with Sapl

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Design of oligos for gRNA Sapl mediated cloning into L2\_lacZgRNA-Cas9-CsA vector. The L2\_lacZgRNA-Cas9-CsA Sapl digested vector has AGC and TTT overhangs. Therefore, oligos for gRNA should be designed such that the forward strand has a 5' overhang of TCG and the reverse strand has a 5' overhang of gt-AAA (addition of "gt" nucleotides is necessary to reconstitute the full sequence of the gRNA scaffold in pink). Blue arrows: Sapl recognition site. Blue dashed lines: Sapl cleavage site. LacZ: lacZα cassette for blue-white screening of colonies.

## 2 **gRNA oligo design**

Order two oligos that contain the forward and reverse guide sequence plus the overhangs necessary for ligation (highlighted with bold) into L2\_lacZgRNA-Cas9-CsA:

**oligo F:** 5'- TCG-NNNNNNNNNNNNNNNNNNNNNN-gt 3'

**oligo R:** 5'-AAAac-NNNNNNNNNNNNNNNNNNNNNN-3'

Note: Standard de-salted oligos are ok

## 3 **Oligo annealing**

Mix oligos with water as follow:

oligo F (100μM) 1μl

oligo R (100μM) 1μl

water 8μl

Total volume 10μl

Anneal in a thermocycler using the following parameters: 37°C for 30 min, 95°C for 5 min and then ramp down to 25°C at 5°C per min. After annealing the gRNA can be directly cloned into L2\_lacZgRNACas9-CsA plasmid without the need of any further processing (step 4).

#### 4 Cloning into backbone vector

In a 0.2 mL tube set up the following reaction:

Component	Volume (μL)
Sterile water	5
10x Tango buffer (Thermo Fisher)	1
1 mg/mL bovine serum albumin (NEB)	0.5
T4 DNA ligase (5 U/μL) (Thermo Fisher)	0.25
10mM ATP (SIGMA)	1
SapI (LgI) (5 U/μL) (Thermo Fisher)	0.25
L2_lacZgRNACas9-Csa (25-50 ng)	1
annealed oligo	1
Final volume	10

- Place samples on the thermocycler and incubated using the following program:

Assembly: 15 cycles: 3 minutes at 37°C and 4 minutes at 16°C

Termination: 5 minutes at 50°C and 10 minutes at 80°C

- Transform chemically competent using 1 μL of reaction and plate on LB agar plates with 100 μg/mL spec and X-gal 40. Incubate at 37 oC for 16 h.
- Confirm with sequencing



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