

Feb 02, 2020

# Novel coronavirus (2019-nCoV) real-time RT-PCR ORF1ab 2020 (Wuhan-ORF1ab; nCoV-specific test) V.2

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*In Development* [dx.doi.org/10.17504/protocols.io.bb3qiqmw](https://doi.org/10.17504/protocols.io.bb3qiqmw)

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## ABSTRACT

- A real-time RT-PCR to detect the "novel Wuhan" betacoronavirus or Wuhan seafood market pneumonia virus. Based on sequence [MN908947](#) made available by Professor Yong-Zhen Zhang, Fudan University, Shanghai, China.
- The target region is within the ORF1ab sequence.
- Not tested on wild-type virus (as of 25Jan2020), it is expected to be capable of only detecting the Wuhan coronavirus.
- Limit of detection not yet determined.

## Notes

1. Assay is fully optimised (as of 24Jan2020).
2. A final name for this virus has not been decided (as of 25Jan2020).

## GUIDELINES

- If using a different brand or model of real-time thermocycler, check the concentration of ROX is adequate.
- Method assumes the user is familiar with the thermocycler and software used to run the protocol.

## STEPS MATERIALS

NAME	CATALOG #	VENDOR
SuperScript™ III Platinum™ One-Step qRT-PCR Kit	11732088	Life Technologies

## Mix

### 1 Oligonucleotides

Oligo Name	Sequence 5'-3'	Location based on NC_045512*
WuhanORF1ab-F	AATCCACCTGCTCTACAAGATG	5455-5476
WuhanORF1ab-R	CATCACCTAACTCACCTACTGTC	5566-5544
WuhanORF1ab-P	6FAM-AGCTTCACCAGCCCTTGCTCT-BHQ1	5505-5485

\*GenBank accession NC\_045512 Wuhan seafood market pneumonia virus isolate Wuhan-Hu-1

## 2 Reagents



### SuperScript™ III Platinum™ One-Step qRT-PCR Kit

by Life Technologies

Catalog #: 11732088

## 3 Synthetic controls

Synthetic controls are produced using the [binary synthetic template oligonucleotide positive control for in-house diagnostic real-time RT-PCR method](#).

The oligonucleotide sequences required to make controls for this assay are:

Probe control:

AAAAATAATACGACTCACTATAGGGTGAAGAGAATCCACAAGGAATTGAAAGCTTCACCAGCCCTTGCTCTACAGTGTCAGCAGGTC  
CTGTTGAAAA

Primer control:

AAAAATAATACGACTCACTATAGGGAATCCACCTGCTCTACAAGATGATGATCTGGCACGGGACCCTCCAAGACAGTAGGTGAGTTAG  
GTGATGAAAA

## 4 Reaction Set-up

- Assay has been designed to be used on both a Rotor-Gene 6000 / Rotor-Gene Q 5-plex using 100-place rotor discs and a ABI 7500 Fast real-time machine.
- Total reaction volume is 20µL.
- Prepare sufficient for number of reaction plus a 'dead volume' usually 2 extra. Adjust as necessary if using a robotic dispenser.

Reagent	Volume (ul) X1	Final reaction concentration
Nuclease free water	4.37	
WuhanORF1ab-F (200uM)	0.07	700nM
WuhanORF1ab-R (200uM)	0.09	900nM
WuhanORF1ab-P (100uM)	0.03	150nM
2 X Reaction mix*	10	1X
Superscript III/Platinum Taq enzyme mix*	0.4	
ROX reference dye (25uM)*	0.04	50nM
TOTAL VOLUME	15	

\*Superscript®III Platinum® One-Step qRT-PCR kit

Dispense 15µl to each reaction well.

Add 5µl of template, extracted RNA, controls or NTC (nuclease-free water).

Total reaction volume is 20µl.

## Amplification

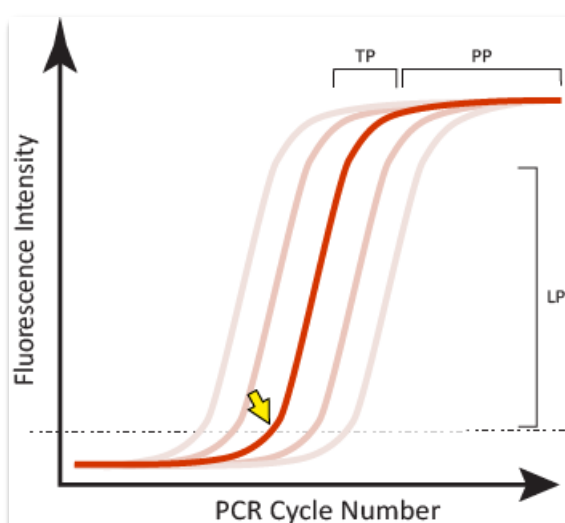
### 5 PCR amplification

1 cycle	40 cycles
50°C 5min	95°C 3 seconds
95°C 2min	60°C 30 seconds*

\*Fluorescence acquisition step

## Result Analysis

- The definition used for a satisfactory positive result from a real-time fluorogenic PCR should include each of the following:
  - A sigmoidal curve – the trace travels horizontally, curves upward, continues in an exponential rise and followed by a curve towards a horizontal plateau phase
  - A suitable level of fluorescence intensity as measured in comparison to a positive control (y-axis)
  - A defined threshold ( $C_T$ ) value which the fluorescent curve has clearly exceeded (Fig.1 arrow) and which sits early in the log-linear phase and is <40 cycles
  - A flat or non-sigmoidal curve or a curve that crosses the threshold with a  $C_T$  value >40 cycles is considered a negative result
  - NTCs should not produce a curve



**Figure 1.** Examples of satisfactory sigmoidal amplification curve shape when considering an assay's fluorescent signal output. The crossing point or threshold cycle ( $C_T$ ) is indicated (yellow arrow); it is the value at which fluorescence levels surpass a predefined (usually set during validation, or arbitrary) threshold level as shown in this normalized linear scale depiction. LP-log-linear phase of signal generated during the exponential part of the PCR amplification; TP-a slowing of the amplification and accompanying fluorescence signal marks the transition phase; PP-the plateau phase is reached when there is little or no increase in fluorescent signal despite continued cycling.



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