

Light/dark preference test for adult zebrafish (*Danio rerio*)

Version 2

Caio Maximino

Abstract

The light/dark test is increasingly being adopted as one of the main behavioral assays in zebrafish neuroscience research. It is based on the innate preference of adult zebrafish for a black vs. a white compartment. Modifications and extensions of the protocol increased its breadth, but also its heterogeneity. The protocol presented here involves confining the animal in a central compartment for a initial 3-min acclimation period, followed by free exploration of an acrylic tank with a black and a white compartment. In addition to observing the spatial distribution of the animal, "ethological" variables (risk assessment, erratic swimming, freezing, thigmotaxis) are also manually scored in the white compartment; these endpoints are selectively sensitive to drug treatments. The protocol is intended to measure anxiety-like behavior, with anxiogenic effects decreasing time on white and increasing risk assessment and thigmotaxis; some treatments also increase erratic swimming and freezing.

Citation: Caio Maximino Light/dark preference test for adult zebrafish (*Danio rerio*). **protocols.io**
dx.doi.org/10.17504/protocols.io.srfed3n

Published: 28 Aug 2018

Guidelines

Standard guidelines for the ethical care of fish in research apply. Before beginning experiments, researchers should obtain all legally binding licenses, including IACUC approval.

Before start

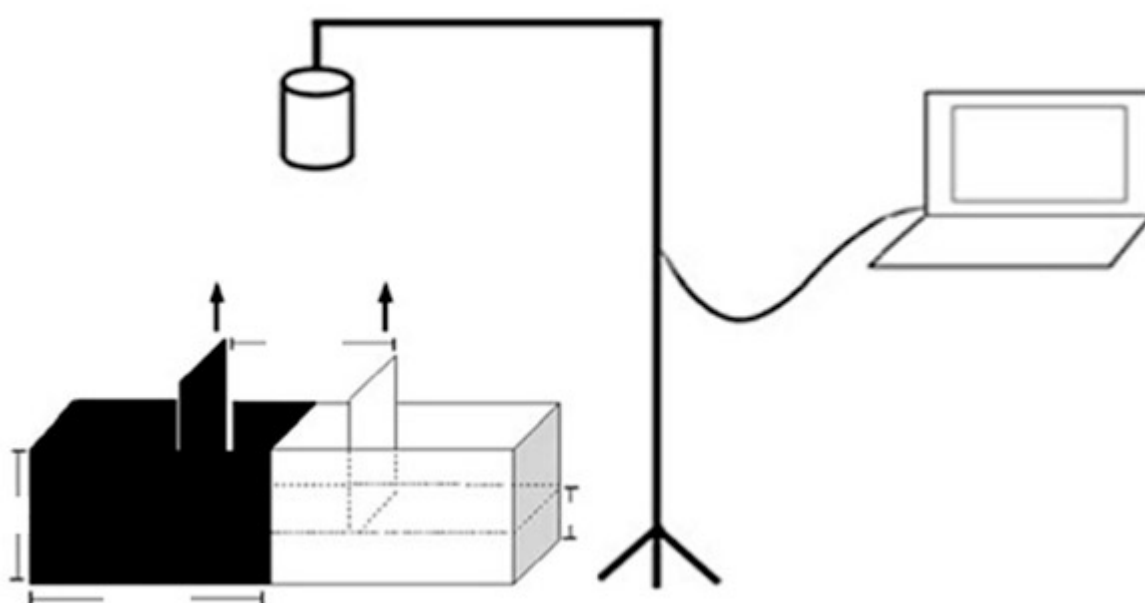
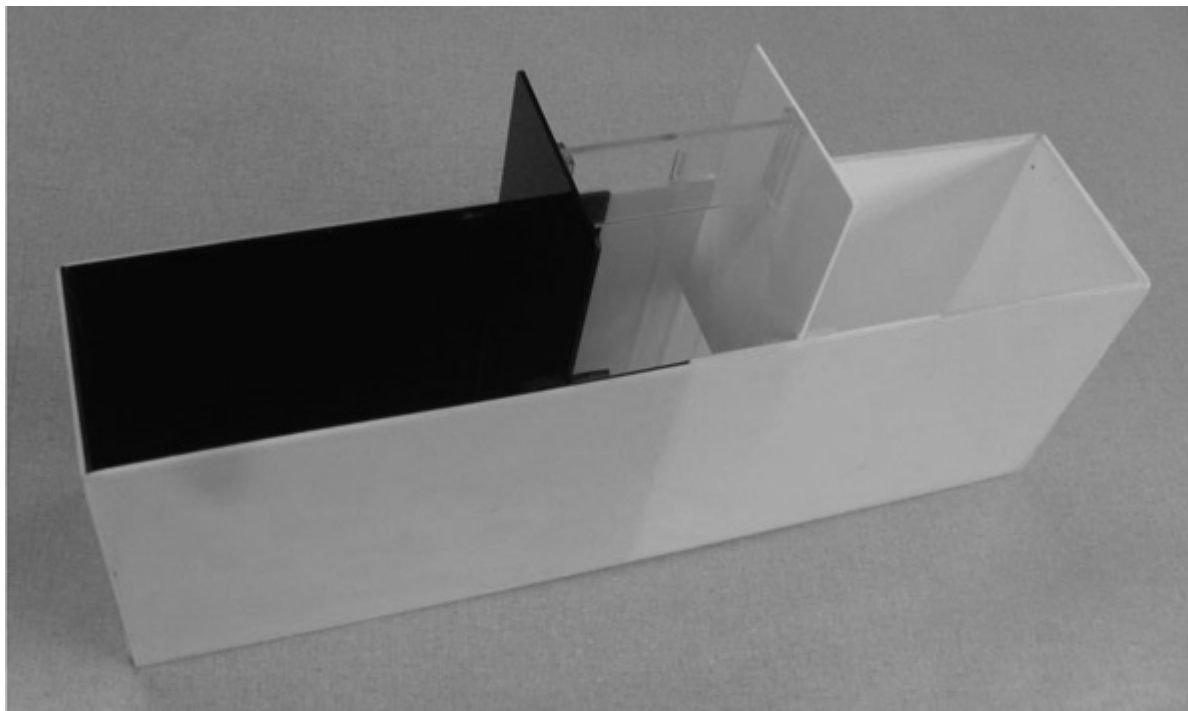
1. Room illumination is a critical variable, as it can alter fish behavior and the aversiveness of the white compartment. Before each trial, light levels should be measured; variations above or below 2 SDs from the average should be considered as too discrepant for an experiment. Ideally, lighting above the tank should be on the range of 500-600 lux.
2. Use Gaussian white noise throughout the experiment to cover unwarranted noise in the experimental room.
3. Bring the animal in a temporary transport recipient into the behavioral test room.

Protocol

Apparatus

Step 1.

- An acrylic tank (15 cm × 10 cm × 45 cm height × width × length) is used that is divided equally into one-half black and one-half white.
- Walls and bottom are either black or white, so as to warrant uniform substrata for each compartment.
- The water column is kept at 10 cm, yielding a final volume of 4.5 liters.
- Water should be de-chlorinated.
- The colored material chosen should not be reflective, to avoid the tendency of those animals that present shoaling tendencies to behave in relation to their own reflection.
- The tank contains central sliding doors, colored with the same color of the aquarium side, thereby defining an uncolored central compartment measuring 15 cm × 10 cm × 10 cm.



Preparation steps

Step 2.

Room illumination is a critical variable, as it can alter fish behavior and the aversiveness of the white compartment. 2. Before each trial, light levels should be measured; variations above or below 2 SDs from the average should be considered as too discrepant for an experiment. Ideally, lighting above the tank should be on the range of 500-600 lux.

Preparation steps

Step 3.

Inject drug or vehicle in the animal, using a microsyringe (10 μ L vol) and a wettened sponge (see Kinkel et al., 2010; DOI: 10.3791/2126). Injection volume should be proportional to body weight (1 μ L / 0.1 g), and the animal should be anesthetized with cold water (12 $^{\circ}$ C) or eugenol (60 ppm).

Preparation steps

Step 4.

Use Gaussian white noise throughout the experiment to cover unwarranted noise in the experimental room.

Preparation steps

Step 5.

Bring the animal in a temporary transport recipient into the behavioral test room.

Behavioral assay

Step 6.

Transfer the animal to the central compartment of the tank with a net. Allow the animal to acclimate to the tank for 3 min

 **DURATION**

00:03:00 : Duration of acclimation

Behavioral assay

Step 7.

Remove the sliding doors, allowing the fish to swim freely in the apparatus for 15 min. Video recording is recommended.

 **DURATION**

00:15:00 : Duration of observation

Step 8.

Transfer the animal to a discard tank with a net.

Endpoints

Step 9.

Videos should be analyzed blindly (i.e., the experimenter should not know the treatment/genotype of the animal) by two experienced researchers. An event/spatial distribution logger, such as X-Plo-Rat (<https://github.com/lanec-unifesspa/x-plo-rat>).

Step 10.

Variables to be recorded (and ZBC references) are:-Time spent in the white compartment (ZBC 1.137)-Latency to enter the white compartment-Number of entries in the white compartment-Duration of entries in the white compartment-"Risk assessment" events (a fast (<1 s) entry in the white compartment followed by re-entry in the black compartment, or a partial entry in the white compartment (i.e., the pectoral fin does not cross the midline))-Duration and number of erratic swimming (ZBC 1.51): a zig-zag, fast, unpredictable course of swimming of short duration.-Duration and number of freezing events (ZBC 1.68): complete cessation of movements except eye and opercular movements.-Time on thigmotaxis (ZBC 1.173): swimming in a distance of 2 cm or less from the nearest wall.

Step 11.

If X-Plo-Rat is used, risk assessment, erratic swimming, freezing, and thigmotaxis are assigned to keyboard keys (A, S, D, and F are a good choice). The I-maze pre-configuration can be used, with the "walled" squares equivalent to the black compartment.

 **SOFTWARE PACKAGE (Windows)**

X-Plo-Rat, 1.1.0

Fernando Cardenas

<https://github.com/lanec-unifesspa/x-plo-rat>

 **EXPECTED RESULTS**

Under these conditions, adult zebrafish present preference for the dark compartment, spending

about 80% of a 15 min session on that portion of the tank.

Drugs or treatments with an "anxiogenic"-like effect increase the preference, and can also increase risk assessment, thigmotaxis, erratic swimming, and/or freezing in the white compartment. Conversely, drugs with an "anxiolytic"-like effect reduce some or all of these endpoints.

Warnings

The behavioral protocol is usually very innocuous for researchers. Care must be taken while preparing drugs for injection, following standard biosecurity practices. Moreover, care must be taken while capturing fish to not expose open wounds to tank water, reducing the risk of acquiring waterborne infections (cf. Murray et al., 2016, doi: 10.1089/zeb.2015.1206, for a primer on zebrafish biosecurity).