

# Sampling for qEXT and MPN assays: Large-scale One-step Phage Infection of Cyanobacteria Version 3

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## Abstract

**Experiment purpose is to monitor the time-course of a large-scale infection of host cyanobacteria by phage under variable media conditions and obtain samples for proteomic and transcriptomic analysis.**

**15 Hourly Timepoints: 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14**

*Sampling is for qPCR assay to determine total extracellular phage (qEXT), and for MPN (most probable number) assay to determine % phage infectivity.*

For qEXT and MPN sampling, **250 µL of each sample in duplicates** were collected and filtered through a 0.2 µm, 96-well filter plate, for each time point. The flow through filtrate containing phage was stored at 4°C as the sample to analyze.

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[dx.doi.org/10.17504/protocols.io.fi3bkgn](https://dx.doi.org/10.17504/protocols.io.fi3bkgn)

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## Materials

- ✓ 96-well filter plates (0.2µm) Millipore #MSGVN2210 by Contributed by users
- ✓ 96 well plates for filtrate Fisher #087722C by Contributed by users
- ✓ Eppendorf centrifuge with plate adapters and a balance plate by Contributed by users
- ✓ P1000 micropipet and Filter Tips by Contributed by users
- ✓ Parafilm by Contributed by users

## Protocol

### Step 1.

Stack filter plate onto bottom 96-well plate without touching the bottom of the sterile filter plate.

## Plate-Setup Template:

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

### Step 2.

Transfer 250 uL of sample from each experiment bottle into the filter plate in duplicates. Only sample from the bottles containing test phage and not the controls.

### Step 3.

Centrifuge at 1000 x g for 3 min.

### Step 4.

Between sampling, store stacked filter/filtrate plates at 4°C, covered with a plate lid.

### Step 5.

After filling the plate, remove the filter plate and store the filtrate plate at 4°C, covered with a plate lid and wrapped well with parafilm.