Protoplast Isolation - Enzyme Buffer

Steven Burgess

Abstract

Enzyme solution for the isolation of protoplasts suitable for *Arabidopsis thaliana* and *Nicotiana* benthamiana. Adapted from Yoo et al.

2007 http://www.nature.com/nprot/journal/v2/n7/full/nprot.2007.199.html

And excellent video from the Sheen Lab is

available: https://www.youtube.com/watch?v=5-xm1EoLrW4

Citation: Steven Burgess Protoplast Isolation - Enzyme Buffer. protocols.io

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Protocol

Step 1.

500mM MES, pH 5.6



1 ml Additional info:

NOTES

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Final concentration is 1.5% (w/v)

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Final concentration is 10mM. Yoo et al. 2007 mention that MES is preheated to 70°C for 3-5 minutes prior to addition of the enzyme powder.

Step 2.

Mannitol

■ AMOUNT

5 g Additional info:

NOTES

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Final Concentration is 0.3% (w/v)

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Final concentration is 1.5% (w/v)

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Final concentration is 0.6M

Step 3.

1M Potassium Chloride

KCI

AMOUNT

1 ml Additional info:

NOTES

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Final Concentration is 0.3% (w/v)

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Final concentration is 1.5% (w/v)

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Final concentration is 20µM

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Final concentration is 0.6M

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Final concentration is 10mM. Yoo et al. 2007 mention that MES is preheated to 70° C for 3-5 minutes prior to addition of the enzyme powder.

Step 4.

Add dH₂O up to 50mL

P NOTES

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Final Concentration is 0.3% (w/v)

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Final concentration is 1.5% (w/v)

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Final concentration is 20µM

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Final concentration is 0.6M

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Final concentration is 10mM. Yoo et al. 2007 mention that MES is preheated to 70°C for 3-5 minutes prior to addition of the enzyme powder.

Step 5.

Cellulase R10

NOTES

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Final Concentration is 0.3% (w/v)

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Final concentration is 1.5% (w/v)

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Final concentration is 20µM

Step 6.

Macerozyme R10

₽ NOTES

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Final Concentration is 0.3% (w/v)

Step 7.

Heat the enzyme solution at 55°C for 10 min, then allow to cool to room temperature.

Step 8.

1M Calcium Chloride

CaCl₂

■ AMOUNT

50 μl Additional info:

Step 9.

Step 10.

Filter final solution through a 0.45-µm syringe filter.

Warnings

Solution must be made fresh.