ATAC-seq on nuclei from frozen, sliced, Drosophila melanogaster embryo halves

Jenna Haines

Abstract

Protocol for performing ATAC-seq on nuclei isolated from *Drosophila melanogaster* stage 5 embryos that were flash frozen and then cut in half along the anterior-posterior midline. Data from this protocol are presented in the following paper: https://www.biorxiv.org/content/early/2017/09/27/195073.

This protocol is based adapted from the following sources: https://www.nature.com/nmeth/journal/v10/n12/full/nmeth.2688.html and http://journals.plos.org/plosgenetics/article?

Citation: Jenna Haines ATAC-seq on nuclei from frozen, sliced, Drosophila melanogaster embryo halves. **protocols.io** dx.doi.org/10.17504/protocols.io.j9zcr76
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Protocol

Solutions

Step 1.

Lysis Buffer NO detergent

- 10mM Tris-Hcl, pH 7.4
- 10mM NaC1
- 3mM MgCl2

Store at 4dC- make fresh weekly

Lysis Buffer Freezing Media

- 10ml Lysis Buffer No Detergent
- 1ml of 50% glycerol
- 1ul of bromoblue dye

Lysis Buffer + Spermine

- 10ml Lysis Buffer No Detergent
- 0.15mM spermine

Embryo Collection

Step 2.

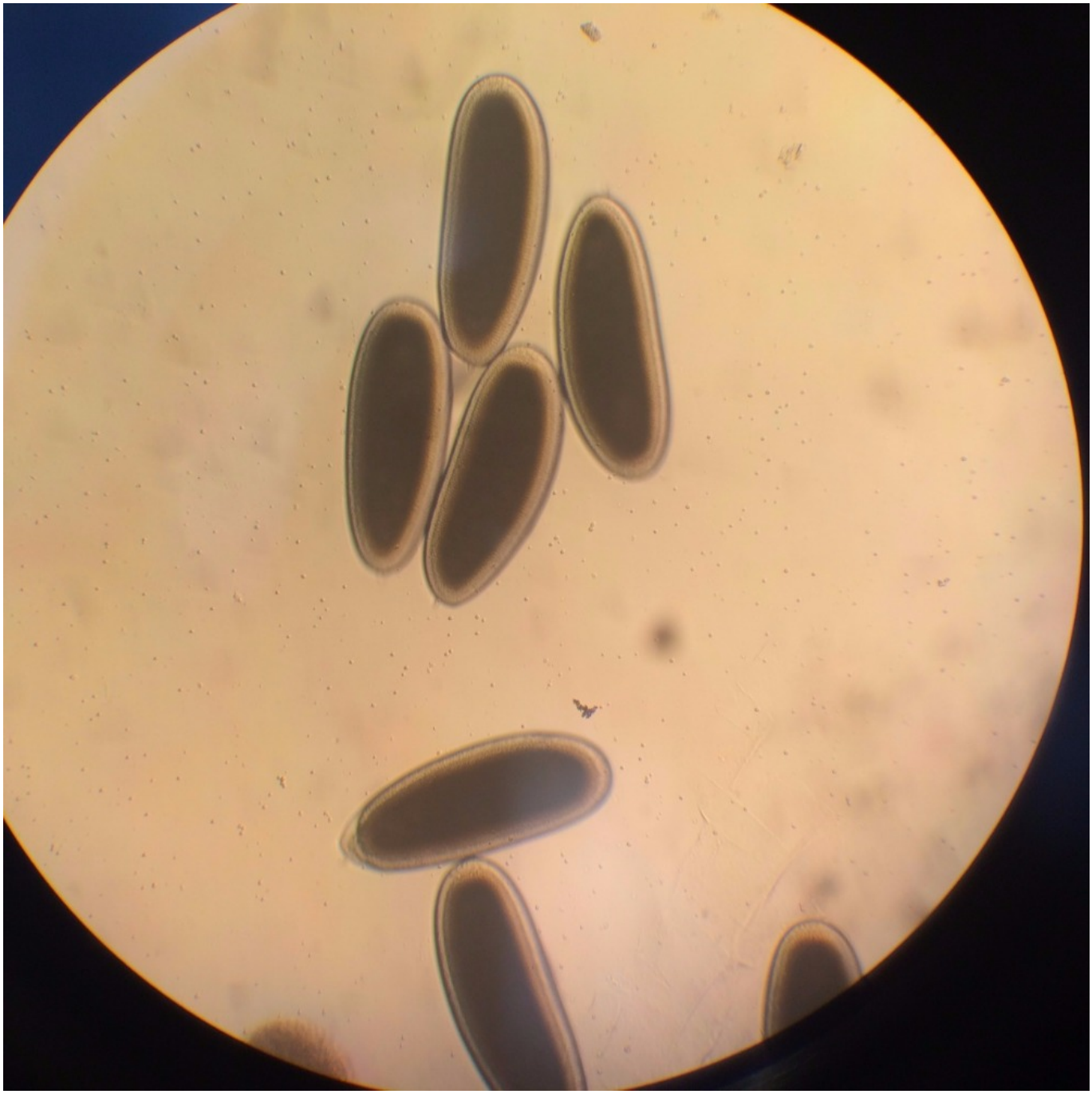
Several hundred adult OregonR *Drosophila melanogaster* (http://flybase.org/reports/FBsn0000276.html) were put into a fly cage at 25°C for three days. Flies are fed with standard molasses plates a spread of yeast paste. Embryos were collected for 2 hours and then aged at 25°C for an hour and thirty minutes. This should yield embryos between 2:00 and 3:30 hours old.

Step 3.

Bleach embryos for 3 minutes in 30%-50% bleach to remove the chorion. Rinse well with water. Pat dry on towel. Rinse with 1xPBS + 0.5% Triton. Brush embryos into an 1.5ml tube filled with 1xPBS + 0.5% Triton. Let embryos settle. Then use a pipette tip to transfer a drop of embryos to a slide.

Step 4.

Stage the embryos to the desired age. We usually go for Stage 5.



Freezing and Slicing embryos

Step 5.

Chill a cold block in the -80 for at least 30 minutes. Once it's cold, place in a box surrounded by dry ice to keep it cool while you slice

REAGENTS

Corning® CoolRack CF15, Holds 15 Cryovial or FACS Tubes (Product #432049) 432049 by Corning

Freezing and Slicing Embryos

Step 6.

Place staged embryos into ependorf tubes containing Lysis buffer freezing media.

Step 7.

Keep the slides, poker tool, and scalpel on dry ice to keep from melting.

REAGENTS

Shandon™ Straight Point Teasing Needles, Straight Point, Wooden Handle, 5.5 in. (14.0cm) 19010 by Thermo Scientific

Fisherbrand™ High Precision #10 Style Scalpel Blade 12-000-162 by Fisher Scientific

Step 8.

Underneath a dissection scope, space out staged embryos on the glass slide such that they are evenly separated and can be cut easily with the blade. I affix a grid pattern to the bottom side of the glass slide to offer contrast to the white embryos.

Step 9.

Once the embryos are lined up on a slide, gently place the slide on dry ice for 2-5 minutes while you prepare the next slide. The embryos will turn from clear to completely white when frozen. I usually line up 10 embryos / slide and slice 5-6 slides at a time.

Step 10.

Once embryos are frozen, immediately transfer the slide to a cold block that has been chilled. I surround the block with dry ice to keep it cool. Additionally it is necessary to cool down the scalpel on dry ice before cutting. I leave the scalpel and 2 poker tools in dry ice while cutting to keep them cold. Be sure to wear gloves and keep the scalpal handle away from the dry ice to keep hands warm.

Step 11.

Cut embryos along the midline with the dry ice chilled blade or scalpel. Move each half to a 1.5ml tube filled with 50ul of ATAC Lysis buffer + Spermine with the chilled poker tool.

- The embryos will thaw and dissolve as soon as they are transfered. To reduce contamination, we use 2 poker tools, one to handle anterior halves and one to handle posterior halves. Additionally, we wash pokers in 70% EtOH between slices

Step 12.

Once your tube contains the desired number of embryo halves, move on to isolate the nuclei.

Embryo Lysing

Step 13.

Crush using a plastic pestle upwards of 20 times vigorously.

REAGENTS

Pellet pestles Z359947 by Sigma

Step 14.

After homogenization, rinse the pestle with 50ul of lysis buffer to make sure not to lose any material. This will bring the total volume up to 100ul.

Step 15.

Vortex slightly at low speed and spin down on a table top centrifuge.

Step 16.
Add 1ul of 10% IGEPAL CA-630 drop-wise to a final concentration of 0.1%. Mix well by flicking tube with finger. **Let sit for about 10 minutes**

Step 17.
Spin in microcentrifuge at 800xg for 10 minutes at 4dC. Remove as much supernatant as possible to to avoid nuclei loss.

Step 18.

Resuspend nuclei in enough DNAse and RNAse free water for the transposition reaction (below).

Tagmentation

Step 19.

Make sure the cell pellet is set on ice.

To make the transposition reaction mix, combine the following:

Sample	20 halves-10 embryos
2x TD Buffer-	25
Tn5 Enzyme	7.5
H20/ DNA	17.5
Total	50

REAGENTS

- Nextera DNA library preparation kit 24 samples FC-121-1030 by Contributed by users

Step 20.
Gently pipette to resuspend nuclei in the transposition reaction mix.

Step 21.
Incubate the transposition reaction at **37°C for 30 min**. Immediately following transposition, purify

using a **Qiagen MinElute Kit**.

REAGENTS

MinElute Reaction Cleanup Kit 28204 by Qiagen

Step 22.

Purified DNA can be stored at -20°C

I quantify DNA at this point with the qubit.

PCR amplification

Step 23.

To amplify transposed DNA fragments, combine the following in a PCR tube:

- 10 µL Transposed DNA
- 5 ul 25uM Primer 1
- 5 ul 25uM Primer 2
- 25ul NebNext 2x Master Mix
- 5ul of PPC

50 µL Total

Run PCR as follows:

(1) 72°C, 5 min

(2) 98°C, 30 sec

(3) 98°C, 10 sec

(4) 63°C, 30 sec

(5) 72°C, 1 min

(6) Repeat steps 3-5, **5x**

(7) Hold at 4°C

Optional - QPCR

Step 24.

You can stop at this point and run a small qPCR reaction to calculate the number of additional cycles to use to avoid overamplification.

Take out 5 ul of PCR reaction and run the qPCR reaction with the same conditions.

5 µL Transposed DNA

0.25 ul 25uM Primer 1

0.25 ul 25uM Primer 2

5ul NebNext 2x Master Mix

3.9 ul of H20

0.09 100x Sybr Green I Dye

_____

15 µL Total

Put in the QPCR machine and run:

1 cycle: 30 sec 98° C

20 cycles: 10 sec 98° C

30 sec 63° C

1 min 72° C.

To calculate the additional number of cycles needed, plot linear Rn (fluorescence) versus cycle and determine the cycle number that corresponds to one-third of the maximum fluorescent intensity.

Step 25.

Once you decide how many additional cycles to run, run the rest of the PCR :

(2) 98°C, 30 sec

(3) 98°C, 10 sec

(4) **63°C**, 30 sec

(5) 72°C, 1 min **For X cycles**

(7) Hold at 4dC

Ampure (or Spri) bead Cleanup

Step 26.

1) Bring Beads to RT

2) add 1.25x Beads, Mix well, let stand for 5 minutes

3) Put on magnet, let stand for 2 minutes

4) Wash 2x with 70% EtOH

5) Let dry for 10 minutes

7) Resuspend in 20ul H20

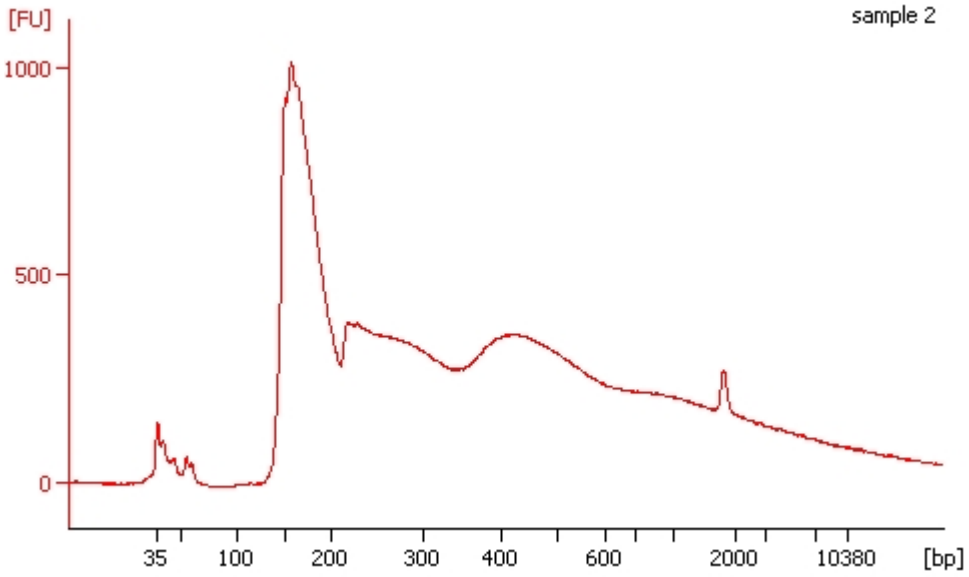
8) Let sit for one minute, put on magnet, take of super

Library Validation

Step 27.

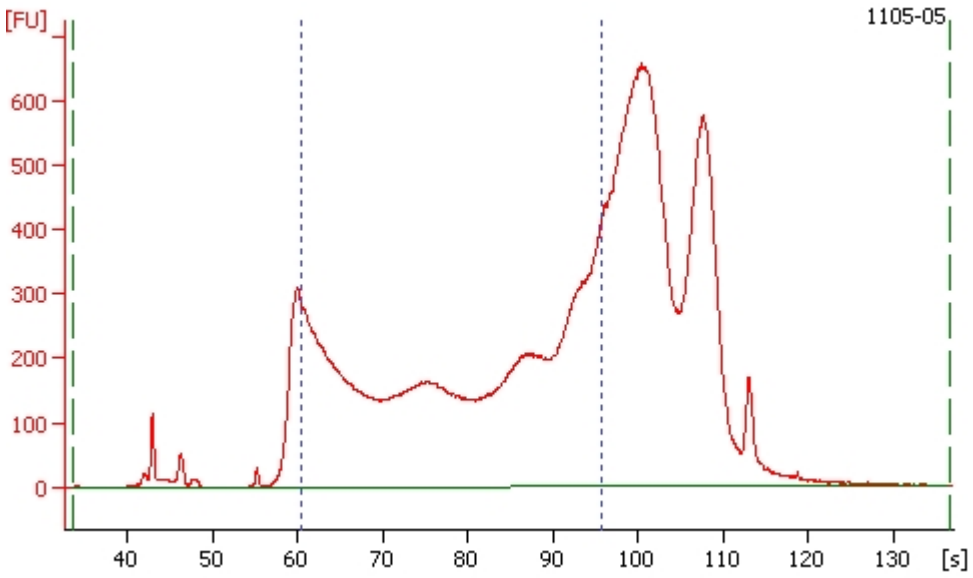
To validate my libraries, I quantify them with qubit HS DNA and I evaluate them with the Bioanalyzer High Sensitivity DNA assay.

Sometimes they look like this:



Sometimes they look like this: I usually see this when I am working with smaller amounts of embryos. Below is from one embryo half. I am not 100% sure what this means (whether it's actually undertagmented or what) but when I sequenced this sample the data was comprable to my other samples. If you have any ideas let me know! I usually try not to sequence these samples but they

could be perfectly fine.



Warnings

- Always wear proper PPE
- Be careful to not get frostbite when working with dry ice. Double glove when handling the chilled scalpel.