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## Growth curve analysis

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Works for me

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## ABSTRACT

To observe the potential of defense mechanisms of either native or synthetic systems in *Escherichia coli* (and more) when incubated with a bacteriophage stock.

## MATERIALS

NAME ▾	CATALOG # ▾	VENDOR ▾
96-well plate, flat bottom, tissue culture treated, black wall with clear bottom	3904	Fisher Scientific
Microplate Reader Synergy Mx	View	

## STEPS MATERIALS

NAME ▾	CATALOG # ▾	VENDOR ▾
96-well plate, flat bottom, tissue culture treated, black wall with clear bottom	3904	Fisher Scientific

## Preparations

- Media and bacteriophage stock solutions :
  - 1L Luria-Bertani (LB) media (with antibiotics)
  - Desired Bacteriophage stock solution in LB media (with known Plaque Forming Units (PFU) ml<sup>-1</sup>)
- Fill in plate reader protocol as follows:**
  - Set temperature: 37°C  
preheat before moving to next step
  - Start kinetics:  
Runtime 15:00:00 (HH:MM:SS), Interval 0:04:00
  - Shake:  
medium, 0:30 (MM:SS)
  - Read:  
Absorbance Endpoint, Full Plate  
Wavelengths: 600  
Read Speed: Normal, Delay: 100 msec
  - End kinetics
- Prepare overnight cultures of desired samples (with associated antibiotics).

4 Measure OD600 of overnight cultures and dilute cultures to an OD600 of 0.02

5 Load  **180 µl of diluted overnight culture** into a



96-well plate, flat bottom, tissue culture treated, black wall with clear bottom

by Fisher Scientific

Catalog #: 3904

Include a serie of LB (without bacteria) as a control and as zero point for the OD600 measurements!

6 Start plate reader protocol  **go to step #2** and let the bacteria grow to an OD600 of 0.11.

7 Prepare Bacteriophage PFU dilutions (with associated antibiotics) for;

MOI  $10^1$  :  $4.0 \times 10^{10}$  PFU ml<sup>-1</sup>

MOI  $10^0$  :  $4.0 \times 10^9$  PFU ml<sup>-1</sup>



MOI  $10^{-1}$  :  $8.0 \times 10^8$  PFU ml<sup>-1</sup>

MOI  $10^{-2}$  :  $8.0 \times 10^7$  PFU ml<sup>-1</sup>

MOI  $10^{-3}$  :  $8.0 \times 10^6$  PFU ml<sup>-1</sup>

An OD600 of 0.10 correlates to  $8.0 \times 10^8$  cells per ml.

The above concentrations are required when 20 µl of bacteriophage dilution is added into 180 µl of cell culture with an OD600 of 0.11 (1:10 dilution).

8 At the moment an OD600 of 0.11 is reached, the plate reader must be stopped and  **20 µl of bacteriophage dilution\*** must be added to a final volume of  **200 µl** to both the samples and the LB controls.

\* include as a control, a serie without bacteriophages and only LB media (with antibiotics)

9 Restart the plate reader protocol and measure over 15 hours the growth of the samples.



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