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Working

Prediction of pneumoconiosis by serum and urinary biomarkers in workers exposed to asbestos-contaminated minerals \leftrightarrows

PLOS One

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ABSTRACT

Prediction of pneumoconiosis by serum and urinary biomarkers in workers exposed to asbestos-contaminated minerals

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EXTERNAL LINK

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PROTOCOL STATUS

Working

SAFETY WARNINGS

Required software

This protocol requires R-3.5.2 for Windows (32/64 bit) (https://cran.r-project.org/bin/windows/base/)

Dataset preparation

2 Download the dataset and store at "C:\r". PLoS One Excel Main Raw Datafile.csv

Cpoy the R script of six machine learning algorithms on R consol.

3



library(rattle) # Access the weather dataset and utilities. library(magrittr) # Utilise %>% and %<>% pipeline operators.

building <- TRUE

scoring <-! building

A pre-defined value is used to reset the random seed

so that results are repeatable.

crv\$seed <- 42

Load a dataset from file.

fname <- "file:///C:/r/PLoS One Excel Main Raw Datafile.csv" crs\$dataset <- read.csv(fname.



```
na.strings=c(".", "NA", "", "?"),
 strip.white=TRUE, encoding="UTF-8")
# Action the user selections from the Data tab.
# The following variable selections have been noted.
         <- c("Age", "Sex", "CEA", "SMRP", "Fibulin3",
          "OhdG")
crs$numeric <- c("Age", "Sex", "CEA", "SMRP", "Fibulin3",
crs$categoric <- NULL
crs$target <- "Pneumoconiosis"
crs$risk
        <- NULL
crs$ident <- NULL
crs$ignore <- c("FeNO", "FVC", "FEV1", "Smoking")
crs$weights <- NULL
# Decision Tree
# The 'rpart' package provides the 'rpart' function.
library(rpart, quietly=TRUE)
# Reset the random number seed to obtain the same results each time.
set.seed(crv$seed)
# Build the Decision Tree model.
crs$rpart <- rpart(Pneumoconiosis ~ .,</pre>
  data=crs$dataset[, c(crs$input, crs$target)],
  method="class",
  parms = list(split = "information"),\\
  control=rpart.control(usesurrogate=0,
    maxsurrogate=0),
  model=TRUE)
# Generate a textual view of the Decision Tree model.
print(crs$rpart)
printcp(crs$rpart)
cat("\n")
# Extreme Boost
# The `xgboost' package implements the extreme gradient boost algorithm.
# Build the Extreme Boost model.
set.seed(crv$seed)
```

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crs$ada <- xgboost(Pneumoconiosis ~ .,
 data
             = crs$dataset[,c(crs$input, crs$target)],
 max_depth
                = 6,
            = 0.3,
 num_parallel_tree = 1,
 nthread
              = 2,
 nround
              = 50.
 metrics
              = 'error',
              = 'binary:logistic')
 objective
# Print the results of the modelling.
print(crs$ada)
cat('\nFinal iteration error rate:\n')
print(round(crs$ada$evaluation_log[crs$ada$niter, ], 2))
cat('\nImportance/Frequency of variables actually used:\n')
print(crs$imp <- importance(crs$ada, crs$dataset[,c(crs$input, crs$target)]))</pre>
# Build a Random Forest model using the traditional approach.
set.seed(crv$seed)
crs$rf <- randomForest::randomForest(as.factor(Pneumoconiosis) ~ .,</pre>
 data=crs$dataset[, c(crs$input, crs$target)],
 ntree=500,
 mtry=2,
 importance=TRUE,
 na.action=randomForest::na.roughfix,
 replace=FALSE)
# Generate textual output of the 'Random Forest' model.
crs$rf
# The `pROC' package implements various AUC functions.
# Calculate the Area Under the Curve (AUC).
pROC::roc(crs$rf$y, as.numeric(crs$rf$predicted))
# Calculate the AUC Confidence Interval.
pROC::ci.auc(crs$rf$y, as.numeric(crs$rf$predicted))FALSE
# List the importance of the variables.
rn <- round(randomForest::importance(crs$rf), 2)
rn[order(rn[,3], decreasing=TRUE),]
# Support vector machine.
```

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# The 'kernlab' package provides the 'ksvm' function.
library(kernlab, quietly=TRUE)
# Build a Support Vector Machine model.
set.seed(crv$seed)
crs$ksvm <- ksvm(as.factor(Pneumoconiosis) ~ .,</pre>
   data=crs$dataset[,c(crs$input, crs$target)],
   kernel="rbfdot",
   prob.model=TRUE)
# Generate a textual view of the SVM model.
crs$ksvm
# Regression model
# Build a Regression model.
crs$glm <- glm(Pneumoconiosis ~ .,
  data=crs$dataset[, c(crs$input, crs$target)],
  family=binomial(link="logit"))
# Generate a textual view of the Linear model.
print(summary(crs$glm))
cat(sprintf("Log likelihood: %.3f (%d df)\n",
       logLik(crs$glm)[1],
       attr(logLik(crs$glm), "df")))
cat(sprintf("Null/Residual deviance difference: %.3f (%d df)\n",
       crs$glm$null.deviance-crs$glm$deviance,
       crs\$glm\$df.null-crs\$glm\$df.residual))
cat(sprintf("Chi-square p-value: %.8f\n",
       dchisq(crs$glm$null.deviance-crs$glm$deviance,
           crs$glm$df.null-crs$glm$df.residual)))
cat(sprintf("Pseudo R-Square (optimistic): %.8f\n",
       cor(crs$glm$y, crs$glm$fitted.values)))
cat('\n====ANOVA=====\n\n')
print(anova(crs$glm, test="Chisq"))
cat("\n")
#_____
# Neural Network
# Build a neural network model using the nnet package.
library(nnet, quietly=TRUE)
```

```
# Build the NNet model.
set.seed(199)
crs$nnet <- nnet(as.factor(Pneumoconiosis) ~ .,</pre>
  data=crs$dataset[,c(crs$input, crs$target)],
  size=10, skip=TRUE, MaxNWts=10000, trace=FALSE, maxit=100)
# Print the results of the modelling.
cat(sprintf("A %s network with %d weights.\n",
  paste(crs$nnet$n, collapse="-"),
  length(crs$nnet$wts)))
cat(sprintf("Inputs: %s.\n",
  paste(crs$nnet$coefnames, collapse=", ")))
cat(sprintf("Output: %s.\n",
  names(attr(crs\$nnet\$terms, "dataClasses"))[1]))
cat(sprintf("Sum of Squares Residuals: %.4f.\n",
  sum(residuals(crs$nnet) ^ 2)))
cat("\n")
print(summary(crs$nnet))
cat('\n')
# Evaluate model performance on the training dataset.
# Sensitivity/Specificity Plot: requires the ROCR package
library(ROCR)
# Generate Sensitivity/Specificity Plot for rpart model on PLoS One Excel Main Raw Datafile.csv [**train**].
crs$pr <- predict(crs$rpart, newdata=crs$dataset[,c(crs$input, crs$target)])[,2]</pre>
# Remove observations with missing target.
no.miss <- na.omit(crs$dataset[,c(crs$input, crs$target)]$Pneumoconiosis)
miss.list <- attr(no.miss, "na.action")
attributes(no.miss) <- NULL
if (length(miss.list))
 pred <- prediction(crs$pr[-miss.list], no.miss)</pre>
} else
 pred <- prediction(crs$pr, no.miss)</pre>
ROCR::plot(performance(pred, "sens", "spec"), col="#CC0000FF", lty=1, add=FALSE)
# Sensitivity/Specificity Plot: requires the ROCR package
library(ROCR)
# Generate Sensitivity/Specificity Plot for xgb model on PLoS One Excel Main Raw Datafile.csv [**train**].
crs$pr <- predict(crs$ada, crs$dataset[,c(crs$input, crs$target)])</pre>
```

```
# Remove observations with missing target.
no.miss <- na.omit(crs$dataset[,c(crs$input, crs$target)]$Pneumoconiosis)
miss.list <- attr(no.miss, "na.action")
attributes(no.miss) <- NULL
if (length(miss.list))
 pred <- prediction(crs$pr[-miss.list], no.miss)</pre>
} else
 pred <- prediction(crs$pr, no.miss)</pre>
ROCR::plot(performance(pred, "sens", "spec"), col="#CCCC00FF", lty=2, add=TRUE)
# Sensitivity/Specificity Plot: requires the ROCR package
library(ROCR)
# Generate Sensitivity/Specificity Plot for rf model on PLoS One Excel Main Raw Datafile.csv [**train**].
crs$pr <- predict(crs$rf, newdata=na.omit(crs$dataset[,c(crs$input, crs$target)]),</pre>
  type = "prob")[,2]
# Remove observations with missing target.
no.miss <- na.omit(na.omit(crs$dataset[,c(crs$input, crs$target)])$Pneumoconiosis)
miss.list <- attr(no.miss, "na.action")
attributes(no.miss) <- NULL
if (length(miss.list))
 pred <- prediction(crs$pr[-miss.list], no.miss)</pre>
} else
 pred <- prediction(crs$pr, no.miss)</pre>
ROCR::plot(performance(pred, "sens", "spec"), col="#00CC00FF", lty=3, add=TRUE)
# Sensitivity/Specificity Plot: requires the ROCR package
library(ROCR)
# Generate Sensitivity/Specificity Plot for ksvm model on PLoS One Excel Main Raw Datafile.csv [**train**].
crs$pr <- kernlab::predict(crs$ksvm, newdata=na.omit(crs$dataset[,c(crs$input, crs$target)]),
  type = "probabilities")[,2]
# Remove observations with missing target.
no.miss <- na.omit(na.omit(crs$dataset[,c(crs$input, crs$target)])$Pneumoconiosis)
miss.list <- attr(no.miss, "na.action")
attributes(no.miss) <- NULL
if (length(miss.list))
```

```
pred <- prediction(crs$pr[-miss.list], no.miss)</pre>
} else
 pred <- prediction(crs$pr, no.miss)</pre>
ROCR::plot(performance(pred, "sens", "spec"), col="#00CCCCFF", lty=4, add=TRUE)
# Sensitivity/Specificity Plot: requires the ROCR package
library(ROCR)
# Generate Sensitivity/Specificity Plot for glm model on PLoS One Excel Main Raw Datafile.csv [**train**].
crs$pr <- predict(crs$glm,</pre>
  type = "response",
  newdata = crs$dataset[,c(crs$input, crs$target)])
# Remove observations with missing target.
no.miss <- na.omit(crs$dataset[,c(crs$input, crs$target)]$Pneumoconiosis)
miss.list <- attr(no.miss, "na.action")
attributes(no.miss) <- NULL
if (length(miss.list))
 pred <- prediction(crs$pr[-miss.list], no.miss)</pre>
 pred <- prediction(crs$pr, no.miss)</pre>
ROCR::plot(performance(pred, "sens", "spec"), col="#0000CCFF", lty=5, add=TRUE)
# Sensitivity/Specificity Plot: requires the ROCR package
library(ROCR)
# Generate Sensitivity/Specificity Plot for nnet model on PLoS One Excel Main Raw Datafile.csv [**train**].
crs$pr <- predict(crs$nnet, newdata=crs$dataset[,c(crs$input, crs$target)])</pre>
# Remove observations with missing target.
no.miss <- na.omit(crs$dataset[,c(crs$input, crs$target)]$Pneumoconiosis)
miss.list <- attr(no.miss, "na.action")
attributes(no.miss) <- NULL
if (length(miss.list))
 pred <- prediction(crs$pr[-miss.list], no.miss)</pre>
} else
 pred <- prediction(crs$pr, no.miss)</pre>
ROCR::plot(performance(pred, "sens", "spec"), col="#CC00CCFF", lty=6, add=TRUE)
# Add a legend to the plot.
```

```
legend("bottomleft", c("rpart", "xgb", "rf", "ksvm", "glm", "nnet"), col=rainbow(6, 1, .8), lty=1:6, title="Models", inset=c(0.05))

# Add decorations to the plot.

title(main="Sensitivity/Specificity (tpr/tnr) PLoS One Excel Main Raw Datafile.csv [**train**]", sub=paste("Rattle", format(Sys.time(), "%Y-%b-%d %H:%M:%S"), Sys.info()["user"]))
grid()
```

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