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Working

Long staining procedure of nuclei in Euplotes crassus using DAPI

Forked from [Long staining procedure of nuclei in Euplotes crassus using DAPI](#)Rachele Cesaroni¹¹Universität Bern[dx.doi.org/10.17504/protocols.io.2akgacw](https://doi.org/10.17504/protocols.io.2akgacw)

Protist Research to Optimize Tools in Genetics (PROT-G)

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- 1 Pellet Euplotes crassus cells at 400 rcf for 3 minutes, and remove as much supernatant as possible by pipetting.
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- Both algae and bacteria are autofluorescent. Better to have a completely starved Euplotes crassus culture.
- 2 Add 1 ml of 2% PFA in 1X PHEM or 4% PFA in 1X PBS to the cells, and incubate them for 10 minutes at room temperature.
 - 3 Pellet Euplotes crassus cells by centrifugation at 400 rcf for 3 minutes, and remove as much supernatant as possible by pipetting.
 - 4 Wash cells twice with 1X PBS (400 rcf for 3 minutes each time).
 - 5 Add 1 ml of TBSTEM - 3% BSA and 0.5 µl of DAPI (0.1 mg/ml) to the cells, and stain for 10 minutes at room temperature.
 - 6 Pellet Euplotes crassus cells by centrifugation at 400 rcf for 3 minutes.
 - 7 Add 50 µl of Prolong medium.
 - 8 Place an approx. 10 µl droplet of Euplotes crassus cells on a slide for observation by fluorescence microscopy.



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