

# A General Protocol for Western Blotting Mammalian Cell Lysates

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## Abstract

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## Protocol

### Harvesting and Lysis

#### Step 1.

For adherent cell lines, wash with 1 x PBS and detach cells by incubating in 0.25% trypsin. Once cells have detached at an equal volume of complete media and transfer to a microfuge tube. Suspension cells can be transferred directly to microfuge tube from culture. Spin cells at 500 x G for 5 min to pellet. Wash pellet with 1 x PBS and repeat centrifugation followed by removal of supernatant.

### Harvesting and Lysis

#### Step 2.

Resuspend cell pellet in 50  $\mu$ L - 100  $\mu$ L lysis buffer. Incubate on ice for 10 minutes and then add SDS to 1% final.

#### NOTES

**David Dilworth** 13 May 2018

#### Lysis Buffer:

- \* 20 mM Tris-HCl pH8
- \* 150 mM NaCl
- \* 10 mM MgCl<sub>2</sub>
- \* 1mM EDTA
- \* 0.5 % Triton X-100

Add fresh protease Inhibitors (100x) & benzoase (10 000x) prior to lysis.

### Harvesting and Lysis

#### Step 3.

Quantify and normalize protein concentration between samples using the bicinchoninic acid assay (BCA assay) - Pierce BCA Protein Assay Kit (Cat# 23225)

### Western Blotting

#### Step 4.

To each sample add SDS Loading buffer to 1 x and boil samples for 5 min.

#### 🔥 TEMPERATURE

98 °C Additional info:

#### Western Blotting

##### Step 5.

Load SDS page with 50 - 100 µg total protein and run in an appropriate buffer at 100 V for 2 hours, until the dye front runs off. For NuPAGE 4-12% Bis-Tris Protein Gel (NP0322BOX), we run in 1x MOPS Running Buffer.

#### Western Blotting

##### Step 6.

Transfer proteins using appropriate transfer apparatus, for 1.5 hrs at 80 volts in 1 x Tris-Glycine transfer buffer to a 0.2 µm PVDF membrane. For PVDF membranes, pre-soak in 100% methanol followed by transfer buffer.

#### Western Blotting

##### Step 7.

Block membrane in 5% milk in PBS-T (1x PBS - 0.1% Tween-20) for 30 min at RT.

#### Western Blotting

##### Step 8.

Cut membrane and probe with desired antibodies diluted in 5% BSA in PBS-T overnight at 4°C.

#### Western Blotting

##### Step 9.

Wash membranes 3 x - 10min in PBS-T.

#### Western Blotting

##### Step 10.

Incubate membranes in secondary LiCor antibodies to mouse and/or rabbit (diluted - 1:5000) in Licor Odyssey Blocking buffer (927-40000) diluted 1 in 5 in PBS-T.

#### Western Blotting

##### Step 11.

Wash membranes 3 x - 10min in PBS-T.

#### Western Blotting

##### Step 12.

Image blots on Licor Odyssey CLx Imaging System.