

# Long staining procedure of nuclei in *Euplotes crassus* using DAPI

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## Abstract

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## Protocol

### Step 1.

*Euplotes crassus* cells were pelleted at 400 rcf for 3 minutes.

### 📌 NOTES

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Both algae and bacteria are fluorescent. Better to have a completely starved *Euplotes crassus* culture.

### Step 2.

The supernatant was removed as much as possible and 1 ml of 2% PFA in 1X PHEM or 4% PFA in 1X PBS was added to the cells and incubated for 10 minutes at room temperature.

### Step 3.

*Euplotes crassus* cells were centrifuged at 400 rcf for 3 minutes.

### Step 4.

The supernatant was removed as much as possible and cells were washed twice with 1X PBS (400 rcf for 3 minutes each time).

### Step 5.

1 ml of TBSTEM - 3% BSA and 0.5 µl of DAPI (0.1 mg/ml) were added to the cells. Cells were incubated for 10 minutes at room temperature.

### Step 6.

*Euplotes crassus* cells were centrifuged at 400 rcf for 3 minutes.

### Step 7.

#### ■ ANNOTATIONS

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This step was added by mistake.

### Step 8.

50 µl of Prolong medium were added to the cells.

### Step 9.

10 µl of *Euplotes crassus* cells were put on a slide and observed under the fluorescent microscope.