# 100x BG-11 media

# Anna Behle, Miriam Dreesbach, Susanne Vollmer

#### **Abstract**

This protocol describes how to prepare a 100x BG-11 +N stock.

1x BG-11 media could be created from the stock and supplemental stocks and trace metals could be added afterwards.

This protocol is based on Anne Behle M.Sc. Reciepe for standard BG-11 media protocol.

Recipes for standard and alternative BG11 for culturing freshwater cyanobacteria, such as *Synechocystis* sp. PCC 6803, as described.

Media is usually not suitable for marine cyanobacteria.

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## **Guidelines**

Always work under sterile conditions when handling sterile media or stocks. Work under the clean bench.

#### **Protocol**

#### Safety first

#### Step 1.

Always work under sterile conditions.

#### Step 2.

Add  $CaCl_2 2H_2O$  (3.6 g · L<sup>-1</sup>). We did not have hydrated  $CaCl_2$ . Therefore, we used 2.718 g  $CaCl_2$  (powder) for 1L of the stock.

#### Step 3.

Add Citric acid (0.6 g · L<sup>-1</sup>)

## Step 4.

Add NaNO<sub>3</sub> (149.58 g · L<sup>-1</sup>)

## Step 5.

Add MgSO<sub>4</sub> · 7 H<sub>2</sub>O (7.49 g · L<sup>-1</sup>). We do not have hydrated MgSO<sub>4</sub>. Therefore, we used 3.659 g MgSO<sub>4</sub> (powder) for 1L of the stock.

#### Step 6.

Add 0.25 M Na<sub>2</sub>-EDTA, pH 8.0 (0.56 ml  $\cdot$  L<sup>-1</sup>). We did not have a 0.25 M Na<sub>2</sub>-EDTA stock. Therefore, we dissolved 2.32 g Na<sub>2</sub>-EDTA (powder) and filled the bottle up with MilliQ (nuclease-free) water up to 25 mL. Afterwards, the pH was adjusted with diluted NaOH to pH = 8.0.

## Step 7.

The bottle was filled with MilliQ (nuclease-free) water up to 1L and mixed a big stir bar until the powder was completely dissolved and the solution was clear.

#### Step 8.

Afterwards, the stir bar was removed and the media was directly autoclaved at 120 °C for 30 Minutes.

# **Warnings**

Wear gloves when preparing stocks!