

# Ligation Protocol WITH T4 DNA Ligase (M0202)

## New England Biolabs

### Abstract

Please see the NEB website for more information.

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## Materials


 T4 DNA Ligase - 20,000 units [M0202S](#) by [New England Biolabs](#)

## Protocol

### Step 1.

Thaw the T4 DNA Ligase Buffer and resuspended at room temperature.

#### REAGENTS

 T4 DNA Ligase Reaction Buffer - 6.0 ml [B0202S](#) by [New England Biolabs](#)

#### ANNOTATIONS

**Francisco Maresca** 26 Sep 2015

A useful thing to do is to aliquote the 10x buffer less concentrated so when thawing the DTT gets soluble more easily.

### Step 2.

Set up the reaction in a microcentrifuge tube on ice.

#### PROTOCOL

#### . [T4 DNA Ligase Reaction](#)

CONTACT: [New England Biolabs](#)

#### NOTES

**New England Biolabs** 24 Sep 2014

Note that these are instructions for a ligation using a molar ratio of 1:3 vector to insert for the indicated DNA sizes. Use [NEBioCalculator](#) to calculate molar ratios.

#### Step 2.1.

##### AMOUNT

2 µl Additional info:

##### REAGENTS

 T4 DNA Ligase - 20,000 units [M0202S](#) by [New England Biolabs](#)

### Step 2.2.

Vector DNA (4 kb) 50 ng (0.020 pmol)

#### AMOUNT

50 ng Additional info:

#### ANNOTATIONS

**Ben Claywell** 17 Jul 2015

Use NEBioCalculator to determine concentration

**Low Sin Yee** 23 Jul 2015

recommended vector concentration  $50\text{ng}=0.05\mu\text{g}$

my linearized vector concentration =  $5\mu\text{g}/\text{ml}=0.005\mu\text{g}/\mu\text{l}$

$1\mu\text{l vector}=0.005\mu\text{g}$

how many  $\mu\text{l}$  of vector to make up  $0.05\mu\text{g}$ ?

$(0.05\mu\text{g}\times 1\mu\text{l})/0.005\mu\text{g}=10\mu\text{l}$

thus,  $10\mu\text{l}$  of vector should be added into ligation reaction.

### Step 2.3.

Insert DNA (1 kb) 37.5 ng (0.060 pmol)

#### AMOUNT

38 ng Additional info:

#### ANNOTATIONS

**Ben Claywell** 17 Jul 2015

Use NEBioCalculator to determine concentration

### Step 2.4.

Nuclease-free water to  $20\mu\text{l}$

### Step 2.5.

T4 DNA Ligase,  $1\mu\text{l}$

#### AMOUNT

$1\mu\text{l}$  Additional info:

#### REAGENTS

 T4 DNA Ligase - 20,000 units [M0202S](#) by [New England Biolabs](#)

#### ANNOTATIONS

**mehrdad alirezaei** 17 Jul 2015

T4 DNA Ligase - 100,000 units

Catalog #: [M0202M](#)

### Step 3.

Gently mix the reaction by pipetting up and down and microfuge briefly.

### Step 4.

For cohesive (sticky) ends, incubate at  $16^{\circ}\text{C}$  overnight or room temperature for 10 minutes. For blunt ends or single base overhangs, incubate at  $16^{\circ}\text{C}$  overnight or room temperature for 2 hours.

#### NOTES

**New England Biolabs** 23 Sep 2014

Alternatively, high concentration of T4 DNA Ligase can be used in a 10-minute ligation for blunt ends.

#### ■ ANNOTATIONS

**Ben Claywell** 17 Jul 2015

We are using sticky ends, so incubate at room temperature for 10 minutes.

#### **Step 5.**

Heat inactivate at 65 degrees C for 10 minutes.

#### 🕒 DURATION

00:10:00

#### ■ ANNOTATIONS

**Maohan Su** 07 Dec 2016

Why? No heat inactivation will interference with transformation?

#### **Step 6.**

Chill on ice and transform 1-5 µl of the reaction into 50 µl competent cells.

#### ■ ANNOTATIONS

**Ben Claywell** 17 Jul 2015

Use 25 uL DH5α cells, and add 2 uL of reaction mixture.