



Electroporation protocol for *Vibrio natriegens*

joosef Hoff¹

¹Phillips University

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joosef Hoff ⚡

ABSTRACT

Electroporation protocol

Weinstock paper:

[Matthew T Weinstock](#), [Eric D Heseke](#), [Christopher M Wilson](#), [Daniel G Gibson](#)

Vibrio natriegens as a fast-growing host for molecular biology

Nature Methods volume 13, pages 849–851 (2016)

To prepare before:

recovery medium

(BHI + v2 salts (204 mM NaCl, 4.2 mM KCl, 23.14 mM MgCl₂), and 680 mM sucrose) sterile filtration

* i have a box of aliquots in the freezer and pre heat them before use

-A vial of competent cells is retrieved from storage at –80 °C and allowed to thaw on ice.

-Plasmid DNA and electrocompetent cells are combined and gently mixed in a chilled 1.5-mL microcentrifuge tube.

-The cell–DNA suspension is transferred to a **chilled** electroporation cuvette with a 0.1-cm gap size.

-Cells are electroporated with 900V (in our Electroportor we cant set other parameters)

* in the Weinstock paper they recommend depending on the strain 700-900V, 25 µF and 200 Ω

-Cells are **immediately** recovered in 500 µL **preheated** (50°C) recovery medium and transferred to a 1,5-mL tube.

* we preheat the media to 50°C because the recovery media is cooled down by pipetting and up taking the chilled cells from the cold cuvettes

-The cells are recovered by incubating at 37 °C for 1.5h.

(also put the agar plates for preheating in the incubator at 37°C)

-The cells are centrifuged down for one minute at 3000g. The supernatant is then discarded.

- the pellet is resuspended in the leftover of the media and plated out on

warm agar plates containing appropriate antibiotic.

* for Chloramphenicol 2µg/mL; for Kanamycine 200µg/mL; for Carbenicillin 200µg/mL

-The plates are incubated for several hours or overnight at 37 °C for colonies to appear.

TAGS

electroporation

Vibrio natriegens



Electroporation
protocol.pdf

PROTOCOL STATUS

Working

We use this protocol in our group and it is working



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