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## Drug tracking on hydra

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### ABSTRACT

Protocol for preparing worms, preparing drug plates, dispensing worms with wormsorter and tracking on Hydra rigs

#### Prepare tracking plates and worms (-2 days)

- 1 Pour low peptone NGM into 96 well plates using VIAFILL dispenser. Dispense **200  $\mu$ l** per well. Weigh 3 plates after they have cooled and store at **4 °C** lidside down.
- 2 Refeed bleach synchronised worms onto 4 x 150mm plates by pipette 4 small droplets around the plate 2.5 days prior to tracking (eg 5pm on Monday for Thursday tracking) 60h

#### Prepare drugs onto plates (-1 days)

- 3 Get low peptone 96WP out of the **4 °C** fridge, weigh three plates and then place the rest of the plates in the flow hood to dry 3h for 3 hours. Weigh plates after drying, they should have lost at least 3% of their original mass before proceeding
- 4 Label each (destination) plate with the source plate used and the location in the opentrons robot.
- 5 Prepare drug source plates in skirted 96 well plate so that each column corresponds to a single drug at a single concentration. The final concentration of DMSO should be 0.1%.



Per well:

**0.2  $\mu$ l 1000X drug in DMSO**

**2.8  $\mu$ l water**

This needs to be scaled up to account for the number of replicates along each column (Eg. total\_master\_mix\_volume = volume\_per\_well x 8 x n, where n=number of destination plates and 8 is number of wells per column.)

Run opentrons robot using prepared protocol.



NB.

- One drug source plate with each column corresponding to a drug for use with a multichannel pipette.
- Destination and source plate locations are specified.
- The opentrons robot will randomise the column locations from each source plate to a destination plate
- The date is used as the random seed

#### 6.1 Protocol parameters:

# multichannel pipette parameters and tipracks

multi\_pipette\_type = 'p10-Multi'

multi\_pipette\_mount = 'left'

tiprackdrugs\_slots = ['3']

tiprackdrugs\_type = 'opentrons-tiprack-10ul'

tiprackdrugs\_startfrom = '1'

tiprackH2O\_slot = '6'

tiprackH2O\_type = 'opentrons-tiprack-10ul'

# tiprackH2O\_startfrom = '1'

# water trough

H2O\_source\_slot = '9'

H2O\_source\_type = 'trough-12row'

H2O\_source\_well = 'A1'

H2O\_volume = 5

# drugs source

drugs\_source\_slots = ['2', '5', '8', '11']

drugs\_source\_type = '96-well-plate-pcr-thermofisher'

frombottom\_off = +0.3 # mm from bottom of src wells

drugs\_volume = 3

# destination plates

agar\_thickness = +3.7 # mm from the bottom of the well for 200ul agar per well

destination\_slots = ['1', '4', '7', '5']

destination\_type = '96-well-plate-sqfb-whatman'

n\_columns = 12

# create mapping from sources to destination.

seed = 20191031 # for reproducibility. Let's use the experimental date for the actual experiment, something else for debugging

np.random.seed(seed)

#### 6.2 Make sure all labware is loaded correctly

#### 6.3 Run protocol and monitor robot to ensure all the tips are removed

6.4 Tip racks have to be replaced after each destination plate is filled

7 Allow the plates to dry for 30 minutes

#### Seed plates

8 Use VIAFILL dispenser to dispense  5 µl 1:10 diluted OP50 into each well of each drug plate

9 Allow to dry for 30 minutes under the flow hood

10 Keep at room temperature over night (covered in the dark as some drugs are light-sensitive)

#### Dispensing worms

11 Wash worms off the 150mm plates with M9 using a pasteur pipette into 15ml falcons

12 Spin at  1500 rpm ascending 9, descending 7 for 2 minutes to pellet the worms

13 Remove supernatant and fill M9

14 Repeats steps 12-13 two more times

15 After final wash fill falcon with M9, transfer contents of 15mL falcons to 50mL and fill up to 45mL with M9.

16 Use wormsorter to dispense 2 worms per well

17 Allow liquid to dry off the plates for 30 mins under the flow hood

18 Incubate in drug for 4 hours

19 Hydra tracking : 15 mins 25fps, exposure 25000msec



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