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Intracellular Staining With True-Phos™ Perm Buffer in Cell Suspensions

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Abstract

Citation: Kelsey Miller Intracellular Staining With True-Phos™ Perm Buffer in Cell Suspensions. **protocols.io**

dx.doi.org/10.17504/protocols.io.e2fbgbn

Published: 03 Jun 2016

Protocol

Buffer preparation

Step 1.

Warm Fixation Buffer (BioLegend Cat#420801). For each 1 x 10^6 cells aliquot 0.5 mL of buffer and warm to 37°C.

Buffer preparation

Step 2.

Chill True-Phos™ Perm Buffer to -20°C. For each 1 x 10⁶ cells aliquot 1.0 ml of True-Phos™ Perm Buffer and chill to -20°C.

Sample Preparation

Step 3.

Prepare a single cell suspension with the sample of interest (Human PBMC, splenocytes, cell lines, etc).

Tips:

- 1. Prepare two aliquots, Negative control: untreated, Positive control: treated with stimuli.
- 2. Incubate the cells with the appropriate stimuli, at the suitable temperature and time.

Sample Preparation

Step 4.

Fix the cells immediately after treatment by adding an equal volume of pre-warmed Fixation Buffer. Gently pipette to ensure thorough mixing.

Sample Preparation

Step 5.

Incubate at 37°C for 15 minutes to ensure cells are properly fixed.

O DURATION

00:15:00

Sample Preparation

Step 6.

Centrifuge cells at 350 x g at room temperature for 5 minutes, decant supernatant, vortex to resuspend cell pellet.

© DURATION

00:05:00

Staining with Specific Antibodies

Step 7.

Add sufficient Cell Staining Buffer to wash the cells (approximately 2 ml for each 1×10^6 cells, BioLegend Cell Staining Buffer recommended, Cat#420201).

Centrifuge at 350 x g at room temperature for 5 minutes and decant supernatant. Repeat, for a total of two washes.

Staining with Specific Antibodies

Step 8.

Gently pipette cells using residual volume to resuspend cell pellet.

NOTES

Kelsey Knight 01 Jun 2016

Note: if cells are not fully resuspended, True-Phos™ Perm Buffer addition will cause significant cell loss

Staining with Specific Antibodies

Step 9.

While vortexing, permeabilize cells by adding pre-chilled True-Phos™ Perm Buffer.

Example:10 x 10⁶ cells should be permeabilized with 10 mL of pre-chilled True-Phos™ Perm Buffer.

Staining with Specific Antibodies

Step 10.

Incubate at -20°C for 60 minutes to ensure cells are properly permeabilized.

O DURATION

01:00:00

Staining with Specific Antibodies

Step 11.

Centrifuge cells at 1000 x g at room temperature for 5 minutes, decant supernatant, vortex to resuspend cell pellet.

© DURATION

00:05:00

Staining with Specific Antibodies

Step 12.

Add sufficient Cell Staining Buffer to wash the cells, centrifuge cells at 1000 x g at room temperature for 5 minutes, decant supernatant. Repeat, for a total of two washes.

Staining with Specific Antibodies

Step 13.

Resuspend the cells in Cell Staining Buffer at a concentration of 10×10^6 cells/ml.

Staining with Specific Antibodies

Step 14.

Transfer 100 uL (or 1×10^6 cells) to a 12 x 75 mm tube.

Staining with Specific Antibodies

Step 15.

Add antibody cocktail(s) to appropriate tubes, vortex to mix, and incubate for 30 minutes at room temperature in the dark.

O DURATION

00:30:00

Staining with Specific Antibodies

Step 16.

Add 2 mL of Cell Staining Buffer, centrifuge cells at 1000 x g at room temperature for 5 minutes, decant supernatant. Repeat, for a total of two washes.

Staining with Specific Antibodies

Step 17.

Resuspend cells in approximately 500 ml of Cell Staining Buffer and analyze with a flow cytometer.