

# Beta-glucuronidase (GUS) assay (adapted for *Phaeodactylum tricornutum*)

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## Abstract

A convenient extraction and measurement protocol for GUS activity from *Phaeodactylum tricornutum*.

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## Protocol

### Spin down cells

#### Step 1.

Centrifuge cells for 10 min at 3,000 x g. We typically use 4-5 mL cells grown to  $1 \times 10^6$  cells/mL.

### Resuspend cells in extraction buffer

#### Step 2.

Pour off supernatant and resuspend cell pellets in 200  $\mu$ L extraction buffer. Transfer to 1.5 mL microfuge tubes. Freeze cell suspension in liquid N<sub>2</sub> or dry ice/ethanol bath and either 1) store at -80C or 2) proceed with extraction and assay.

GUS extraction buffer:

50 mM sodium PO<sub>4</sub> buffer pH 7

10  $\mu$ M beta-mercaptoethanol (BME)

0.1% Triton X-100

### Perform freeze-thaw cycles

#### Step 3.

Do a total of three freeze-thaw cycles. Freeze on liquid N<sub>2</sub> or with dry ice/ethanol and thaw by floating tubes in a rack in room temperature water. M

### Centrifuge

#### Step 4.

Spin down cell debris 5 min, 15,000 xg, 4 C. Transfer supernatant to a clean microfuge tube.

#### Prepare 96-well plates

##### Step 5.

Add 190 µL GUS+MUG buffer to each well in a 96 well plate. Do each *P. tricornutum* sample in duplicate or triplicate, so the total number of wells needed is equal to number of samples x 2 or 3. We standard clear, flat bottom 96 well plates for this step.

GUS+MUG buffer:

50 mL GUS extraction buffer + 17.6 mg MUG (4-Methylumbelliferyl-β-D-galactopyranoside)

#### Add samples to wells

##### Step 6.

Add 10 µL clarified sample (step 4) to wells, mix by gently tapping plate. Incubate 30-90 min at 37C.

#### Stop reaction

##### Step 7.

Prepare a stop buffer plate using a black 96-well flat bottom plate containing 180 µL stop buffer per well used in the reaction plate.

After incubation of reaction at 37 C for required time, transfer 20 µL of each reaction to the aliquotted stop buffer plate using a multichannel pipet. Mix by gentle tapping.

Stop Buffer: 0.2 M Na<sub>2</sub>CO<sub>3</sub> (sodium carbonate)

#### Read reaction on fluorescent plate reader

##### Step 8.

Settings for fluorescence detection: excitation, 360 nm, emission, 440 nm

#### Freeze lysates for total protein quantification

##### Step 9.

Perform total protein quantification by BCA assay or equivalent. Normalize GUS activities to total protein.