

Dec 03, 2019

# KU Leuven Exp Urology - Urodynamics in Female MInipigs 👄

PLOS One

Yodi Soebadi<sup>1,2</sup>, Marko Bakula<sup>1</sup>, Lukman Hakim<sup>2</sup>, Robert Puers<sup>1</sup>, Dirk De Ridder<sup>1</sup>

<sup>1</sup>Katholieke Universiteit Leuven, <sup>2</sup>Universitas Airlangga

1 Works for me

dx.doi.org/10.17504/protocols.io.3eggjbw



ABSTRACT

#### Summary:

This protocol provides information for the invasive urodynamic measurement of bladder pressure, by urethral catheter as well as wireless device (bladder pill).

**EXTERNAL LINK** 

https://doi.org/10.1371/journal.pone.0225821

THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

Soebadi MA, Bakula M, Hakim L, Puers R, Ridder DD (2019) Wireless intravesical device for real-time bladder pressure measurement: Study of consecutive voiding in awake minipigs. PLoS ONE 14(12): e0225821. doi: 10.1371/journal.pone.0225821

#### MATERIALS TEXT

### Equipment

- 1. Aquarius TT urodynamics machine calibrated
- 2. Calibration tube 30 cm
- 3. Scale 2 kg
- 4. Luggage scale
- 5. Speculum, forceps, needle holder/scissor combo
- 6. Guidewire
- 7. Amplatz renal dilator set (antenna, dilator, sheath)
- 8. Cystoscope, grasping forceps, light cable
- 9. Telepack/endoscopy camera

## Consumable

- 1. Ketamine 3 vials
- 2. Xylazine 6 ml
- 3. Catheter single channel & multi channel (3 new each)
- 4. NaCl 0.9% 3 L x3
- 5. Catheter gel
- 6. Suture polyglactin (vicryl) 3-0
- 7. Duct tape
  - 1 Preparation: Bring machine and all consumables to room

2	Fill calibration tube with saline
3	Check calibration with tube
4	Open charge switch AND flush baloon
5	Measurement: Isolate minipig for study, bring in transport cart & administer sedation: Ketamine 15 mg/kg (Nimatek 100 mg/ml) Xylazine 2 mg/kg (Xyl-M 2%); (at 60-80 kg, 6-8 ml of Nimatek, 1-1.5 ml of xylazine>>
6	Wait 10-15 minutes outside of cage until minipig sedated
7	Place minipig prone at caudal end,.
8	Prepare urethral silicone cath, gel, tape.
9	Insert single and triple channel catheter in urethra, insert rectal catheter
10	Put belt or frame around midsection
11	Set zero on UDS machine then charge catheter.
12	Put tape around charge switch
13	Tape plastic around posterior and tape funnel
14	Wait for animal to wake up (start eating) and start filling at 10% of voided volume (at volume more than 500 ml fill 50 ml/minute)
15	Repeat up to 5 voids
16	At end of procedure, anesthetise and extract pill.
This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits nrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited	