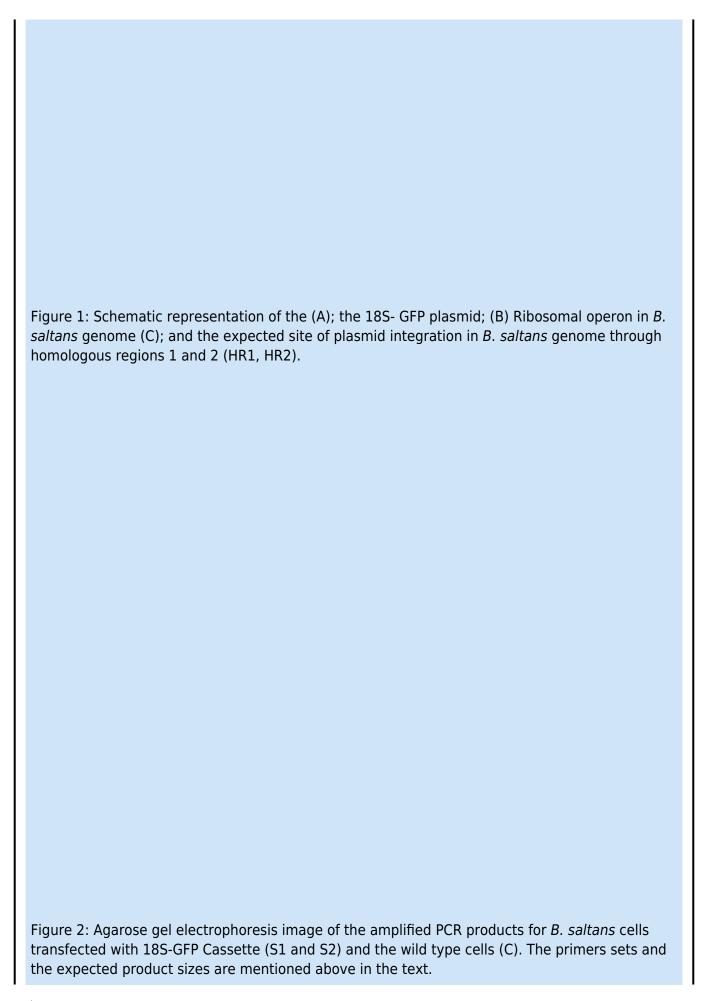
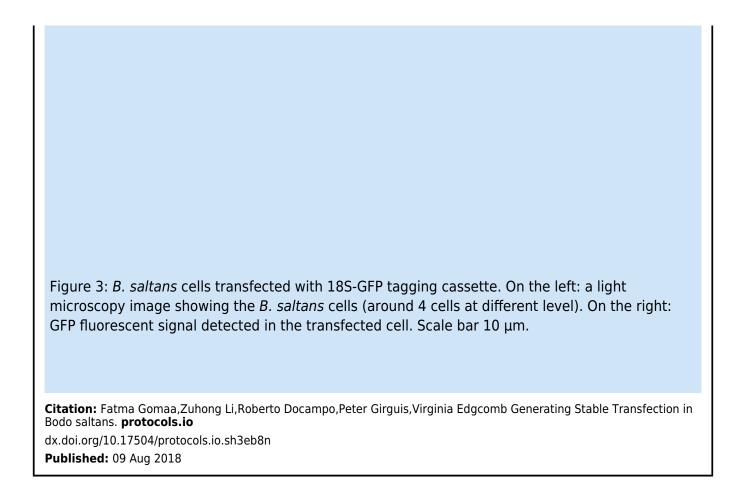
Generating Stable Transfection in Bodo saltans

Fatma Gomaa, Zuhong Li, Roberto Docampo, Peter Girguis, Virginia Edgcomb

Abstract

- B. saltans cells were electroporated using a square-wave electroporator (Nepa21, Bulldog Bio, Inc.) using one poring pulse of 200 volts with a pulse duration of 25 ms and five transfer pulses of 60 volts with a pulse duration of 99 ms, with plasmid targeting the 18S region (18S-GFP). A schematic representation of the plasmid, the target locus and the expected site of integration into the B. saltans genome is shown in Figure 1.
- Electroporated cells were selected with 1 μg/ml of G418, added 24 hours after electroporation. Cells were washed and subcultured into fresh selection medium every 3-4 days. G418 resistant cells started to emerge 7-9 days post-electroporation.
- Cells were processed for genotyping analysis to confirm plasmid integration 3 weeks postelectroporation. DNA was extracted from pools of transfected and wild cells using the Qiagen DNeasy Blood & Tissue kit.
- PCR analyses were used to characterize the 18S-GFP tagging using 6 sets of PCR primers, as shown in Figure 1 C.
- Gel electrophoresis image (Figure 2) showing the amplified PCR products at the expected sizes.
- Amplified PCR#1 with primer sets Ribo tag forward & GFP reverse (800 bp)
- Amplified PCR #2 with primer sets Neo forward & Ribo tag reverse (1000 bp)
- Amplified PCR #3 with primer sets TubR & IG forward (3 bands)
- Amplified PCR #4 with primer sets Tub forward & IG reverse (3 bands)
- Amplified PCR #5 with primer sets Ribo_tag_forward & Ribo_tag_reverse (Wild (C) cells band at 350 bp, transfected cells S1 and S2 two bands, 350 bp and 2800 bp)





Protocol