

Working

## DNA extraction and Nested PCR 👄

PLOS Neglected Tropical Diseases

Baiyan Gong1 ¶1, Yaming Yang2 ¶2, Xiaohua Liu1, Jianping Cao3, 4, 5, 6, 7, 8, Meng Xu3, 4, 5, 6, 7, 8, Ning Xu3, 4, 5, 6, 7, 8 8, Fengkun Yang<sup>1</sup>, Fangwei Wu<sup>2</sup>, Benfu Li<sup>2</sup>, Aiqin Liu \*<sup>1</sup>, Yujuan Shen \*<sup>3, 4, 5, 6, 7, 8</sup>

<sup>1</sup>Department of Parasitology, Harbin Medical University, Harbin, Heilongjiang, China, <sup>2</sup>Yunnan Institute of Parasitic Diseases, Puer, Yunnan, China, <sup>3</sup>National Institute of Parasitic Diseases, Chinese Center for Disease Control and Prevention, 4Chinese Center for Tropical Diseases Research, 5WHO Collaborating Centre for Tropical Diseases, <sup>6</sup>National Center for International Research on Tropical Diseases, Ministry of Science and Technology, <sup>7</sup>Key Laboratory of Parasite and Vector Biology, MOH, <sup>8</sup>Shanghai, China

dx.doi.org/10.17504/protocols.io.yvjfw4n





ABSTRACT

This is the protocol for the Nested PCR assay

**EXTERNAL LINK** 

https://doi.org/10.1371/journal.pntd.0007356



MITSM (M) 410bp.pdf

**GUIDELINES** 

All primer sequences are listed below:

The first round of PCR was carried out using EbGeno-fe (5'-TTC AGA TGG TCA TAG GGA TG-3') as the forward primer and EbGeno-re (5'-ATT AGA GCA TTC CGT GAGG-3') as the reverse primer, which together amplify a 465-bp specific fragment.

A second round of PCR was performed using the forward primer EbGeno-fi (5'-TCG GCT CTG AAT ATC TAT GG-3') and the reverse primer EbGeno-ri (5'-ATT CTT TCG CGC TCG TC-3') in order to amplify a 410-bp internal fragment used for genotype specification.

MATERIALS TEXT

DNase/RNase-Free Deionized Water, primer, TaKaRa Taq DNA Polymerase (TaKaRa Bio Inc., Tokyo, Japan), 10×PCR Buffer, 10 mmol dNTP, 1.5 % agarose gel, GelStrain

## **DNA** extraction

Genomic DNA of E. bieneusi was extracted directly from 180 to 200 mg of each fecal specimen using the recommended procedures and the provided reagents by the manufacture of QIAamp DNA Stool Mini Kit (QIAGEN, Hilden, Germany). To obtain high yield of DNA, the lysis temperature was increased to 95 °C according to the manufacturer's suggestion. DNA was eluted in 200 µL of AE and stored at -20 °C before it was used for PCR analysis.

## PCR amplification

Amplification was accomplished in final volume of 25 μL, containing 17 μL of DNase/RNase-Free Deionized Water, 0.5 μL of each primer,  $0.5\,\mu\text{L}$  of TaKaRa Taq DNA Polymerase (TaKaRa Bio Inc., Tokyo, Japan),  $2.5\,\mu\text{L}$  of  $10\,\times$  PCR Buffer,  $2\,\mu\text{L}$  of  $10\,\text{mmol}$  dNTP and  $2\,\mu\text{L}$  of DNA .

The target DNA undergoes the first run of PCR with the first set of primers.

© 00:05:00 Initial denaturation

```
8 95 °C
      © 00:00:40 Denaturation
       8 94 °C
 5
      © 00:00:45 Annealing
       8 53 °C
      © 00:00:45 Extension/elongation
       8 72 °C

☆ go to step #4 35 cycles

      © 00:04:00 Final extension
       8 72 °C
      The product from the first reaction undergoes a second run of PCR with the second set of primers.
       © 00:05:00 Initial denaturation
       8 95 °C
10
       © 00:00:35 Denaturation
       8 94 °C
11
       © 00:00:40 Annealing
       ₫ 55 °C
12
      © 00:00:40 Extension/elongation
       8 72 °C
13

☆ go to step #10 30 cycles

14
       © 00:05:00 Final extension
       8 72 °C
```

Final hold

15

The final step cools the reaction chamber to 4°C for an indefinite time, and may be employed for short-term storage of the PCR products.

All secondary PCR products were subjected to electrophoresis in a 1.5 % agarose gel and visualized by staining the gel with GelStrain (TransGen Biotech., Beijing, China).

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited