

# Quick Protocol for Monarch® Total RNA Miniprep Kit (NEB #T2010)

New England Biolabs

## Abstract

Quick Protocol for Monarch® Total RNA Miniprep Kit ([NEB #T2010](#)).

Quickly and easily purify up to 100 µg of high-quality total RNA from multiple sample types – all with one kit!

- For use with blood, cells and tissues
- Also works with tough to lyse samples (bacteria, yeast, plant)
- Effectively purifies total RNA of all sizes, including small RNAs >20 nt
- Efficient genomic DNA removal (column and DNase I-based)
- Contains Proteinase K for processing of tissues and blood samples
- Includes RNA Protection Reagent for sample preservation
- Excellent value
- Kit components available separately

**Citation:** New England Biolabs Quick Protocol for Monarch® Total RNA Miniprep Kit (NEB #T2010). [protocols.io](#)  
[dx.doi.org/10.17504/protocols.io.nyadfse](https://doi.org/10.17504/protocols.io.nyadfse)

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## Guidelines

*We recommend that first-time users of this kit review [the product manual](#) before starting. The manual provides additional relevant information to consider at various steps. This quick protocol is meant for experienced users.*

### RNA Purification Consists of Two Stages:

PART 1: Sample Disruption and Homogenization (Differs depending on starting material)

PART 2: RNA Binding and Elution

### Have any questions?

Our tech support scientists would be happy to help. Email us at [info@neb.com](mailto:info@neb.com).

## Before start

- Addition of RNA Lysis Buffer and all subsequent steps should be performed at room temperature to prevent formation of precipitate. If samples are accidentally placed on ice and precipitate forms, allow the samples to return to room temperature to resolubilize before loading onto the column.
- Protocols are also available for [RNA reaction cleanup](#), [RNA fractionation](#), and [extraction of RNA from other sample types](#) (including those in preservation reagents or TRIzol®).

## For the 50 prep kit:

- Reconstitute DNase I by adding 275 µl nuclease-free water. Gently invert to mix. Aliquot for storage at -20°C to minimize freeze-thaw cycles.
- Reconstitute Proteinase K (Prot K) by adding 1040 µl of Proteinase K Resuspension Buffer. Vortex and store at -20°C.
- Add 100 ml ethanol (≥ 95%) to the 25 ml RNA Wash Buffer concentrate.

## Materials

 Monarch Total RNA Miniprep Kit [T2010S](#) by [New England Biolabs](#)

## Protocol

### PART 1: Sample Disruption and Homogenization

#### Step 1.

Please select your starting material of the following:

- **Cultured Mammalian Cells**
- **Mammalian Whole Blood (Fresh or Frozen)**
- **Tissue or Leukocytes**
- **Tough-to-Lyse Samples (bacteria, yeast, plant, etc.) using Mechanical Lysis**

### PART 1: Sample Disruption and Homogenization

#### Step 2 - Cultured Mammalian Cells.

Pellet cells by centrifugation (500 x g) for 1 min.

### PART 1: Sample Disruption and Homogenization

#### Step 3 - Cultured Mammalian Cells.

Discard supernatant.

#### PART 1: Sample Disruption and Homogenization

##### Step 2 - Mammalian Whole Blood (Fresh or Frozen).

Add an equal volume (up to 200  $\mu$ l) of DNA/RNA Protection Reagent (2X concentrate) to an aliquot of whole blood and vortex briefly.

#### NOTES

**Danielle Freedman** 21 Mar 2018

Do not place samples on ice. For frozen samples, quickly thaw in the presence of 2X DNA/RNA Protection Reagent while vortexing or shaking.

#### PART 1: Sample Disruption and Homogenization

##### Step 3 - Mammalian Whole Blood (Fresh or Frozen).

For every 400  $\mu$ l of DNA/RNA Protection Reagent/blood mixture, add 10  $\mu$ l of Prot K. Vortex briefly and incubate at room temperature for 30 min.

#### PART 1: Sample Disruption and Homogenization

##### Step 2 - Tissue or Leukocytes.

Determine the amount of 1X DNA/RNA Protection Reagent that you will need according to the table below. Prepare the 1X aliquot by diluting the 2X stock with nuclease-free water (not included for this step).

Sample Input Amount	Volume 1X DNA/RNA Protection Reagent
Tissue (up to 10 mg)	300 $\mu$ l
Tissue (10–30 mg)	300–600 $\mu$ l
Tissue (30–50 mg)	$\geq$ 600 $\mu$ l
Leukocytes (up to $3 \times 10^6$ )	300 $\mu$ l
Leukocytes ( $3 \times 10^6$ to $1 \times 10^7$ )	$\geq$ 600 $\mu$ l

#### PART 1: Sample Disruption and Homogenization

##### Step 3 - Tissue or Leukocytes.

Add 1X DNA/RNA Protection Reagent to sample.

#### NOTES

**Danielle Freedman** 21 Mar 2018

Solid tissue samples should be submerged in protection reagent, not to exceed 10% (w/v). For maximal RNA recovery, tissues can be mechanically homogenized using a bead mill or similar device.

#### PART 1: Sample Disruption and Homogenization

## Step 2 - Tough-to-Lyse Samples.

Determine the amount of 1X DNA/RNA Protection Reagent that you will need according to the table below. Prepare the 1X aliquot by diluting the 2X stock with nuclease-free water (not included for this step).

Sample Input Amount			Volume 1X DNA/RNA Protection Reagent
Bacteria	Yeast	Plant	
$\leq 5 \times 10^7$	$\leq 5 \times 10^6$		400 $\mu$ l
$\leq 5 \times 10^7$ - $1 \times 10^9$	$\leq 5 \times 10^6$ - $5 \times 10^7$	$\leq 100$ mg	800 $\mu$ l

## PART 1: Sample Disruption and Homogenization

### Step 3 - Tough-to-Lyse Samples.

Add 1X DNA/RNA Protection Reagent to sample.

## Warnings

Please refer to the SDS for safety warnings.