

Mammalian Cell Staining

Version 3

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ABSTRACT

This protocol details how to stain mammalian cells cultured on a 96-well plate. Actin filaments, focal adhesion sites (as indicated by the presence of vinculin), and nuclei will be stained.

PROTOCOL STATUS



Working

We use this protocol in our group and it is working


MATERIALS TEXT

- 4% Paraformaldehyde solution
- 0.1% Triton X-100 solution in PBS
- Blocking buffer (PBS + 1% bovine serum albumin)
- Washing buffer (PBS + 0.05% Tween-20)
- Anti-vinculin solution (1:500 in blocking buffer)
- TRITC-conjugated phalloidin and FITC-conjugated antivinculin secondary antibody solution (1:1:248, TRITC:FITC:blocking buffer) referred to as FITC:TRITC
- DAPI solution (1:249 DAPI:blocking buffer)
- Phosphate buffered saline (PBS)

Fix the cells

- 1 Remove cell culture media.
- 2 Add  100 µl of [M]4 Volume Percent paraformaldehyde solution.
- 3 Incubate for  00:05:00 .


Perforate cell membrane

- 4 Remove paraformaldehyde solution.
- 5 Wash twice with  100 µl washing buffer.



Washing buffer is PBS with the detergent Tween-20.

6 Add  100 µl of [M]0.1 Volume Percent Triton X-100.

7 Incubate for  00:05:00 .

Block unspecific binding

8 Remove Triton X-100.

9 Wash twice with  100 µl washing buffer.

10 Add  100 µl blocking buffer.



Blocking buffer is PBS with BSA (bovine serum albumin) and is used to prevent unspecific binding.

11 Incubate for  00:10:00 .

Bind anti-vinculin to vinculin

12 Remove blocking buffer.

13 Wash twice with washing buffer.

14 Add  250 µl of Anti-Vinculin and blocking buffer mixture.

15 Incubate for  00:20:00 .

Stain actin filaments and focal adhesion sites

16 Remove anti-vinculin blocking buffer mixture.

17 Wash twice.


18 Add  100 µl FITC:TRITC solution, cover in foil.

19 Incubate for  00:30:00 .

Stain nuclei

20 Remove stains.

21 Add  100 μ l of DAPI solution, cover in foil.

22 Incubate for  00:05:00

23 Remove DAPI solution.

24 Add  100 μ l PBS.

Image

25 Image your cells using UV, Blue, and Green excitation.



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