

Definitive guide to western blot

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Abstract

Western blotting uses specific antibodies to identify proteins that have been separated based on size by gel electrophoresis. The immunoassay uses a membrane made of nitrocellulose or PVDF (polyvinylidene fluoride). The gel is placed next to the membrane and application of an electrical current induces the proteins to migrate from the gel to the membrane. The membrane can then be further processed with antibodies specific for the target of interest, and visualized using secondary antibodies and detection reagents.

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Guidelines

A. Sample preparation

1. Lysis buffers

To prepare samples for running on a gel, cells and tissues need to be lysed to release the proteins of interest. This disrupts the cell membrane and solubilizes intracellular proteins so they can migrate individually through the separating gel. There are many recipes for lysis buffers but a few will serve for most western blotting experiments. Broadly, they differ in their ability to solubilize proteins and buffer containing sodium dodecyl sulfate and other ionic detergents are considered to be the harshest and therefore most likely to yield high protein levels.

Most Abcam antibodies recognize reduced and denatured protein and should be used under reducing and denaturing conditions. It is important to note though that some antibodies will only recognize a protein in its native, non-denatured form and will not recognize a protein that has been extracted with a denaturing detergent (SDS, deoxycholate, and somewhat less denaturing, Triton X-100 and NP-40).

The main consideration when choosing a lysis buffer is whether the chosen antibody will recognize denatured samples. When this is not the case, it will be noted on the antibody datasheet, and buffers without detergent or with relatively mild non-ionic detergents (NP-40k, Triton X-100) should be used.

Protein location and lysis buffer choice

Protein location	Buffer recommended
Whole cell	NP-40 or RIPA
Cytoplasmic (soluble)	Tris-HCl
Cytoplasmic (cytoskeletal bound)	Tris-Triton

Membrane bound	NP-40 or RIPA
Nuclear	RIPA or use nuclear fraction protocol*
Mitochondria	RIPA or use mitochondrial fraction protocol*

* Proteins that are found exclusively or predominantly in a sub-cellular location can be enriched in a lysate of the sub-cellular fraction compared to whole cell or tissue lysates. This can be useful when trying to obtain a signal for a weakly-expressed protein. For example, a nuclear protein will be enriched in a nuclear lysate as compared to a whole cell lysate. Sub-cellular fractions are also used to remove potentially cross-reactive proteins present in the unused fractions. Please consult our separate protocol for sub-cellular fractionation.

Nonidet-P40 (NP-40) buffer

150 mM sodium chloride
1.0% NP-40 (Triton X-100 can be substituted for NP-40)
50 mM Tris, pH 8.0

This is a popular buffer studying proteins that are cytoplasmic or membrane-bound and is used for whole cell extracts. If there is concern that the protein on interest is not being completely extracted from insoluble material or aggregates, RIPA buffer may be more suitable, as it contains ionic detergents that more readily solubilize protein.

RIPA buffer (Radio Immunoprecipitation Assay buffer)

150 mM sodium chloride
1.0% NP-40 or Triton X-100
0.5% sodium deoxycholate
0.1% SDS (sodium dodecyl sulphate)
50 mM Tris, pH 8.0

RIPA buffer is also useful for whole cell extracts and membrane-bound proteins, and may be preferred for extracting nuclear proteins. It will disrupt protein-protein interactions and may therefore be problematic for immunoprecipitation / pull down assays.

The 10% sodium deoxycholate stock solution (5 g into 50 ml) must be protected from light.

In cases where it is important to preserve protein-protein interactions or to minimize denaturation (for example, when it is known that the antibody to be used will only recognize a non-denatured epitope), a buffer without ionic detergents (e.g. SDS) and ideally without non-ionic detergents (e.g. Triton X-100) should be used. Cell lysis with detergent-free buffer is achieved by mechanical shearing, often with a Dounce homogenizer or by passing cells through a syringe tip. Often a simple Tris buffer will suffice, however buffers with detergents are required to release membrane - or cytoskeleton - bound proteins.

Tris-HCl buffer

20 mM Tris-HCl, pH 7.5

Tris-Triton buffer (Cytoskeletal proteins)

10 mM Tris, pH 7.4

100 mM NaCl

1 mM EDTA

1 mM EGTA

1% Triton X-100

10% glycerol

0.1% SDS

0.5% deoxycholate

All four of these buffers will keep at 4°C for several weeks or for up to a year aliquotted and stored at -20°C.

2. Protease and phosphatase inhibitors

As soon as lysis occurs, proteolysis, dephosphorylation and denaturation begin. These events can be slowed down considerably if samples are kept on ice or at 4°C at all times and appropriate inhibitors are added fresh to the lysis buffer.

Ready-to-use cocktails of inhibitors from various suppliers are available but you can make your own cocktail.

Inhibitor	Protease/phosphatase inhibited	Final concentration in lysis buffer	Stock (store at 20°C)
Aprotinin	Trypsin, Chymotrypsin, Plasmin	2 µg/ml	Dilute in water, 10 mg/ml. Do not reuse thawed aliquots.
Leupeptin	Lysosomal	5-10 µg/ml	Dilute in water. Do not re-use once defrosted.
Pepstatin A	Aspartic proteases	1 µg/ml	Dilute in methanol, 1 mM.
PMSF	Serine, Cysteine proteases	1 mM	Dilute in ethanol. You can re-use the same aliquot.
EDTA	Metalloproteases that require MG++ and Mn++	5 mM	Dilute in dH ₂ O, 0.5 M. Adjust to pH to 8.0.
EGTA	Metalloproteases that require Ca++	1 mM	Dilute in dH ₂ O, 0.5 M. Adjust to pH to 8.0.
Na Fluoride	Serine/Threonine phosphatases	5-10 mM	Dilute in water. Do not re-use once defrosted.
Na Orthovanadate	Tyrosine phosphatases	1 mM	Dilute in water. Do not re-use once defrosted.

Sodium orthovanadate preparation

All steps to be performed in a fume hood.

1. Prepare a 100 mM solution in double distilled water.
2. Set pH to 9.0 with HCl.
3. Boil until colorless. Minimize volume change due to evaporation by covering loosely.
4. Cool to room temperature.
5. Set pH to 9.0 again.
6. Boil again until colorless.
7. Repeat this cycle until the solution remains at pH 9.0 after boiling and cooling.
8. Bring up to the initial volume with water.
9. Store in aliquots at -20°C. Discard if samples turn yellow.

3. Preparation of lysate from cell culture

1. Place the cell culture dish in ice and wash the cells with ice-cold PBS.
2. Drain the PBS, then add ice-cold lysis buffer (1 ml per 10^7 cells/100 mm dish/150 cm² flask; 0.5 ml per 5×10^6 cells/60 mm dish/75 cm² flask).
3. Scrape adherent cells off the dish using a cold plastic cell scraper, then gently transfer the cell suspension into a pre-cooled microfuge tube.
4. Maintain constant agitation for 30 minutes at 4°C.
5. Centrifuge in a microcentrifuge at 4°C. You may have to vary the centrifugation force and time depending on the cell type; a guideline is 20 minutes at 12,000 rpm but this must be determined by the end-user (e.g. leukocytes need a very light centrifugation).
6. Gently remove the tubes from the centrifuge and place on ice, aspirate the supernatant and place in a fresh tube kept on ice, and discard the pellet.

4. Preparation of lysate from tissues

1. Dissect the tissue of interest with clean tool, on ice preferably, and as quickly as possible to prevent degradation by proteases.
2. Place the tissue in round bottom microfuge tubes or Eppendorf tubes and immerse in liquid nitrogen to “snap freeze”. Store samples at -80°C for later use or keep on ice for immediate homogenization. For a ~5 mg piece of tissue, add ~300 µl lysis buffer rapidly to the tube, homogenize with an electric homogenizer, rinse the blade twice with another 2x300 µl lysis buffer, then maintain constant agitation for 2 hours at 4°C (e.g. place on an orbital shaker in the fridge). Volumes of lysis buffer must be determined in relation to the amount of tissue present (protein extract should not be too diluted to avoid loss of protein and large volumes of samples to be loaded onto gels. The minimum concentration is 0.1 mg/ml, optimal concentration is 1-5 mg/ml).
3. Centrifuge for 20 minutes at 12,000 rpm at 4°C in a microcentrifuge. Gently remove the tubes from the centrifuge and place on ice, aspirate the supernatant and place in a fresh tube kept on ice; discard the pellet.

The buffer (with inhibitors) should be ice-cold prior to homogenization.

5. Determination of protein concentration

Perform a Bradford assay, a Lowry assay, or a BCA assay. Bovine serum albumin (BSA) is a frequently-used protein standard.

Once you have determined the concentration of each sample, you can freeze them at -20°C or -80°C for later use or prepare for immunoprecipitation or for loading onto a gel.

6. Preparation of samples for loading into gels: denatured and native, reduces and non-reduced.

A denatured, reduced samples.

Antibodies typically recognize a small portion of the protein of interest (referred to as the epitope) and this domain may reside within the 3D conformation of the protein. To enable access of the antibody to this portion it is necessary to unfold the protein, i.e. denature it.

To denature, use a loading buffer with the anionic denaturing detergent sodium dodecyl sulfate (SDS), and boil the mixture at 95-100°C for 5 minutes. Heating at 70°C for 5-10 minutes is also acceptable and may be preferable when studying multi-pass membrane proteins. These tend to aggregate when boiled and the aggregates may not enter the gel efficiently.

The standard loading buffer is called 2X Laemmli buffer, first described in Nature, 1970 Aug 15; 227 (5259):680- 5. It can also be made at 4X and 6X strength to minimize dilution of the samples. The 2X is to be mixed in a 1:1 ratio with the sample.

Laemmli 2X buffer

4% SDS
10% 2-mercaptoethanol
20% glycerol
0.004% bromophenol blue 0.125 M Tris HCl

Check the pH and bring it to pH 6.8.

When SDS is used with proteins, all of the proteins become negatively charged by their attachment to the SDS anions. SDS denatures proteins by “wrapping around” the polypeptide backbone. SDS binds to proteins fairly specifically in a mass ratio of 1.4:1. In doing so, SDS confers a negative charge to the polypeptide in proportion to its length – i.e. the denatured polypeptides become “rods” of negative charge clouds with equal charge densities per unit length.

In denaturing SDS-PAGE separations, therefore, migration is determined not by intrinsic electrical charge of the polypeptide, but by molecular weight. The quality of SDS is of utmost importance: old or poor quality SDS can lead to artifacts such as a smear of protein along the entire gel track. It is usually necessary to reduce disulphide bridges in proteins by using β-mercaptoethanol or dithiothreitol (DTT). This allows proteins to adopt the randomcoil configuration necessary for separation by size.

Glycerol is added to the loading buffer to increase the density of the sample to be loaded and hence maintain the sample at the bottom of the well, restricting overflow and uneven gel loading.

To enable visualization of the migration of proteins it is common to include in the loading buffer a small anionic dye molecule (e.g. bromophenol blue). Since the dye is anionic and small, it will migrate the fastest of any component in the mixture to be separated and provide a migration front to monitor

the separation progress.

During protein sample treatment the sample should be mixed by vortexing before and after the heating step for best resolution.

B Native and non-reduced samples.

Alternatively, an antibody may recognize an epitope made up of non-contiguous amino acids. Although the amino acids of the epitope are separated from one another in the primary sequence, they are close to each other in the folded three-dimensional structure of the protein, and the antibody will only recognize the epitope as it exists on the the surface of the folded structure.

It is imperative, in these circumstances, to run a western blot in non-denaturing conditions, and this will be noted on the datasheet in the applications section. In general, a non-denaturing condition simply means leaving SDS out of the sample and migration buffers and not heating the samples.

Certain antibodies only recognize protein in its non-reduced form i.e. in an oxidized form (particularly on cysteine residues) and the reducing agents β -mercaptoethanol and DTT must be left out of the loading buffer and migration buffer (non-reducing conditions).

Protein state	Gel condition	Loading buffer	Migration buffer
Reduced - denatured	Reducing & denaturing	With β - mercaptoethanol or DTT, with SDS	With SDS
Reduced - native	Reducing & non-denaturing	With β - mercaptoethanol or DTT, no SDS	No SDS
Oxidized - denatured	Non-reducing & denaturing	No β - mercaptoethanol or DTT, with SDS	With SDS
Oxidized - native	Non-reducing & native	No β - mercaptoethanol or DTT, no SDS	No SDS

Rule of thumb: Reduce and denature unless the datasheet specifies otherwise.

B. Electrophoresis

Electrophoresis can be one dimensional (i.e. one plane of separation) or two dimensional. One dimensional electrophoresis is used for most routine protein and nucleic acid separations. Two dimensional separation of proteins is used for finger-printing, and when properly constructed can be extremely accurate in resolving all of the proteins present within a sample.

Here we will be describing techniques for one dimensional electrophoresis. We recommend Gel Electrophoresis of Proteins: A Practical Approach (3rd Edition, B.D. Hames and D. Rickwood, The Practical Approach Series, Oxford University Press, 1998) as a reference for basic understanding of 2D electrophoresis protocols.

1. Preparation of PAGE gels

When separated on a polyacrylamide gel, the procedure is abbreviated as SDS-PAGE (Sodium Dodecyl Sulfate PolyAcrylamide Gel Electrophoresis). The technique is a standard means for separating

proteins according to their molecular weight.

Polyacrylamide gels are formed from the polymerization of two compounds, acrylamide and N, Nmethylenebisacrylamide (Bis for short). Bis is a cross-linking agent for the gels. The polymerization is initiated by the addition of ammonium persulfate along with either DMAP or TEMED. The gels are neutral, hydrophilic, three-dimensional networks of long hydrocarbons cross-linked by methylene groups.

The separation of molecules within a gel is determined by the relative size of the pores formed within the gel. The pore size of a gel is determined by two factors: the total amount of acrylamide present (designated as %T) and the amount of cross-linker (%C). As the total amount of acrylamide increases, the pore size decreases. With cross-linking, 5%C gives the smallest pore size. Any increase or decrease in %C increases the pore size. Gels are designated as percent solutions and will have two necessary parameters. The total acrylamide is given as a percentage (w/v) of the acrylamide plus the bis-acrylamide. Thus, a 7.5%T would indicate that there is a total of 7.5 g of acrylamide and bis per 100 ml of gel. A gel designated as 7.5%T:5%C would have a total of 7.5% (w/v) acrylamide + bis, and the bis would be 5% of the total (with pure acrylamide composing the remaining 2.5%).

Gels can be purchased ready-made or produced in the laboratory (recipes can be found in laboratory handbooks). The Abcam laboratory uses gels from our Optiblot range. Either way, choose carefully the percentage of you gel as this will determine the rate of migration and degree of separation between proteins.

Rule of thumb: The smaller the size of the protein of interest, the higher the percentage of mono/bis. The bigger the size of the protein of interest, the lower the percentage of mono/bis.

Protein size (kDA)	Gel percentage (%)
4-40	20
12-45	15
10-70	12.5
15-100	10
25-200	8

Acrylamide is a potent cumulative neurotoxin: wear gloves at all times.

Place gels in the electrophoresis tank as instructed by the manufacturer and bathe in migration buffer.

2. Positive controls

Positive control lysate is used to demonstrate that the protocol is efficient and correct and that the antibody recognizes the target protein which may not be present in the experimental samples.

We strongly recommend the use of a positive control lysate when setting up a new experiment; this will give you immediate confidence in the protocol.

3. Molecular weight markers

A range of molecular weight markers will enable the determination of the protein size (see below) and also to monitor the progress of an electrophoretic run. A range of MW markers are commercially available.

Molecular Weight Marker (ab48854)

Prism Protein Ladder (10-175 kDa) (ab115832)

Prism Ultra Protein Ladder (10-180 kDa) (ab116027)

Prism Ultra Protein Ladder (10-245 kDa) (ab116028)

Prism Ultra Protein Ladder (3.5-245 kDa) (ab116029)

4. Loading samples and running the gel

Use special gel loading tips or a micro-syringe to load the complete sample in a narrow well. Take care not to poke the well bottom with the tip as this will create a distorted band.

Never overfill wells. This could lead to poor data if samples spill into adjacent wells, and poorly resolved bands.

Load 20-40 µg total protein per mini-gel well.

The gels will be submerged in migration buffer which normally contains SDS, except in native gel electrophoresis.

A standard migration buffer (also called running bugger) for PAGE is 1X Tris-glycine:

25 mM Tris base

190 mM glycine

0.1% SDS

Check the pH; it should be around 8.3.

Run the gel for the recommended time as instructed by the manufacturer; this can vary from machine to machine (1 hour to overnight depending on the voltage).

When the dye molecule (the “migration front”) reached the bottom of the gel, the power is turned off. Proteins will slowly elute from the gel at this point, so do not store the gel; proceed immediately to transfer.

5. Use of loading controls

Loading controls are required to check that the lanes in your gel have been evenly loaded with sample, especially when a comparison must be made between the expression levels of a protein in different samples. They are also useful to check for even transfer have not occurred, the loading control bands can be used to quantify the protein amounts in each lane. For publication-quality work, use of a loading control is absolutely essential.

Loading control	Sample type	kDa	Caution
Vinculin	Whole cell	125	

Cyclophilin B	Whole cell	24	
GAPDH	Whole cell	35	Some physiological factors, such as hypoxia and diabetes, increase GAPDH expression in certain cell types.
Cofilin	Whole cell / Nuclear / Membrane / Cytoskeleton	19	
Alpha tubulin	Whole cell / Cytoskeleton	50	Tubulin expression may vary according to resistance to antimicrobial and antimitotic drugs (Sangrajrang S, et al, 1998, Prasad V et al, 2000).
Beta tubulin	Whole cell / Cytoskeleton	50	Tubulin expression may vary according to resistance to antimicrobial and antimitotic drugs (Sangrajrang S, et al, 1998, Prasad V et al, 2000).
Actin	Whole cell / Cytoskeleton	42	
Beta actin	Whole cell / Cytoskeleton	40	Not suitable for skeletal muscle samples. Changes in cell-growth conditions and interactions with extracellular matrix components may affect actin protein synthesis (Farmer et al, 1983).
VDAC1/Porin	Mitochondrial	30	
Cox IV	Mitochondrial	20	Many proteins run at the same 16 kDa size as COX IV.
HSP60	Mitochondrial/Membrane	60	
Lamin B1	Nuclear	66	Not suitable for samples where the nuclear envelope is removed
HDAC1	Nuclear	55	
YY1	Nuclear	45	
TBP	Nuclear	35	Not suitable for samples where DNA is removed
PCNA	Nuclear	30	
cdk4	Nuclear/Membrane	34	
Nak ATPase	Membrane	110	
Transferrin	Serum	75	

C. Transfer of proteins and staining (Western blotting)

SEE STEPS FOR PROTOCOL

1. Visualization of proteins in gels

This visualization of proteins within the SDS-PAGE gel is useful to determine if proteins have electrophoresed uniformly and evenly. Use the copper stain if you plan to transfer the separated proteins to a membrane, as the Coomassie stain is not reversible. Only use the Coomassie stain on gels post-transfer to check the efficiency of the transfer, or if you have no plans to transfer and just

want to observe the results of the SDS-PAGE separation.

2. Transfer

Detailed instructions for the transfer process can be found on the websites of the manufacturers of transfer apparatus, and will vary depending on the system. The principle is the same in each case though. Just as proteins with an electrical charge (conferred by the SDS bound to them) can be induced to travel through a gel in an electrical field, so too can the proteins be transferred in an electrical field from the gel onto a sturdy support, a membrane that “blots” the proteins from the gel. Early methods relied on diffusion; blotting in an electrical field is now standard.

Transfer can be done in wet or semi-dry conditions. Semi-dry transfer is generally faster but wet transfer is less prone to failure due to drying of the membrane and is especially recommended for proteins greater than 100 kDa. For both kinds of transfer, the membrane is placed next to the gel. The two are sandwiched between absorbent materials, and the sandwich is clamped between solid supports to maintain tight contact between the gel and membrane.

In wet transfer, the gel and membrane are sandwiched between sponge and paper (sponge/paper/gel/membrane/paper/sponge) and all are clamped tightly together after ensuring no air bubbles have formed between the gel and membrane. The sandwich is submerged in transfer buffer to which an electrical field is applied. The negatively-charged proteins travel towards the positively-charged electrode, but the membrane stops them, binds them, and prevents them from continuing on.

A standard buffer for wet transfer is the same as the 1X Tris-glycine buffer used for the migration/running buffer without SDS but with the addition of methanol to a final concentration of 20%. For proteins larger than 80 kD, it is recommended that SDS is included at a final concentration of 0.1%.

In semi-dry transfer, a sandwich of paper/gel/membrane/paper wetted in transfer buffer is placed directly between positive and negative electrodes (cathode and anode respectively). As for wet transfer, it is important that the membrane is closest to the positive electrode and the gel closest to the negative electrode. The proportion of Tris and glycine in the transfer buffer is not necessarily the same as for wet transfer; consult the apparatus manufacturer's protocol. A standard recipe is 48 mM Tris, 39 mM glycine, 0.04% SDS, 20% methanol.

Two types of membranes are available: nitrocellulose and PVDF (positively-charged nylon). The choice is personal and both work very well. PVDF membranes require careful pre-treatment: cut the membrane to the appropriate size then soak it in methanol for 1-2 minutes. Incubate in ice cold transfer buffer for 5 minutes. The gel needs to equilibrate for 3-5 minutes in ice cold transfer buffer. Failure to do so will cause shrinking while transferring, and a distorted pattern of transfer.

Note on transfer of large and small proteins: The amount of SDS and methanol in the transfer buffer, protein size, and gel percentage can affect transfer efficiency. The modifications in steps 10-14 and 15-17 will encourage efficient transfer.

The following reference discusses a gel and buffer system that allows transfer of proteins as large as 500 kD: Bolt and Mahoney, High-efficiency blotting of proteins of diverse sizes following sodium dodecyl sulfatepolyacrylamide, gel electrophoresis. *Analytical Biochemistry* 247, 185-192 (1997).

Avoid touching the membrane with your fingers; use tweezers instead. Oils and proteins on the fingers will block efficient transfer and create dirty blots.

After sandwiching the gel and membrane between paper, air bubbles between the gel and membrane can be removed by rolling them out with a pipet or 15 ml tube, or by assembling the sandwich in a dish of transfer buffer to prevent formation of bubbles in the first place. Wear gloves!

Make sure the paper and membrane are cut to the same size as the gel. Large overhangs may prevent a current from passing through the membrane in semi-dry transfers.

Chicken antibodies tend to bind to PVDF and other nylon-based membranes, leading to high background. Switching to a nitrocellulose membrane should help reduce background staining.

3. Visualization of proteins in membranes: Ponceau Red

TBS 10x (concentrated TBS)

24.23 g Trizma HCl/
80.06 g NaCl Mix in 800 ml ultra pure water
pH to 7.6 with pure HCl
Top up to 1L

TBST

For 1 L: 100 ml of TBS 10x + 900 ml ultra pure water + 1 ml Tween20

Tween20 is very viscous and will stick to the tip of your measuring pipettes. Be sure you add the right amount of the detergent to the Tris buffer. A 10% solution is easier to dispense than undiluted Tween20.

4. Blocking the membrane

Blocking the membrane prevents non-specific background binding of the primary and/or secondary antibodies to the membrane (which has a high capacity for binding proteins and therefore antibodies).

Two blocking solutions are traditionally used: non-fat milk or BSA (Cohn fraction V). Milk is cheaper but is not recommended for studies of phospho-proteins (milk contains casein which is a phospho-protein; it causes high background because the phosphor-specific antibody detects the casein present in the milk).

Some antibodies give a stronger signal on membranes blocked with BSA as opposed to milk for unknown reasons. Check the application notes on the datasheet in case there are specific instructions on how to block the membrane.

5. Incubation with the primary antibody

6. Incubation with the secondary antibody

7. Development methods

Detection kits

For HRP-conjugated antibodies: ECL and ECL+ (home made or commercially available) are the traditional kits used and we recommend EDL+. For the new generation detection machines such as Genegnome, use the detection kit recommended by the manufacturer of the machine.

We do not recommend ECL or BCIP/NBT detection kits as they are not as sensitive.

X-ray films

Manual film development is traditionally used and enables the scientist to control the incubation time of the x-ray film in the developing agent and fixation agent.

Automated x-ray film developers are also widely used and easy to use.

Remember that an over-exposed film is not suitable for analysis as determination of the relative amount of protein is not possible. Over-exposed films show totally black bands with no contrast, and/or numerous non-specific bands.

Digital images

The new generation of film developers are units with a camera inside an enclosure, removing the need for a darkroom. The camera detects the chemiluminescence emanating from the membrane, transforming the signal into a digital image for rapid analysis with software provided with the detection machine.

A range of machines are now commercially available. At the front of the next generation are systems which do not use HRP-conjugated antibodies (i.e. chemiluminescence): for example, STORM Analyzers detect fluorescence from fluorochrome-conjugated secondary antibodies. The Odyssey Infrared Imaging System detects infrared fluorescence.

D. References

- Harlow, Ed, and David Lane. Using antibodies. Cold Spring Harbor, New York: Cold Spring Harbor Laboratory Press, 1999.
- B.D. Hames and D. Rickwood. Gel Electrophoresis of Proteins: A Practical Approach 3rd Edition, The Practical approach series, Oxford University Press, 1998.
- Bold and Mahoney, High-efficiency blotting of proteins of diverse sizes following sodium dodecyl sulfatepolyacrylamide, gel electrophoresis. Analytical Biochemistry 257, 185-192, 1997

Protocol

Visualization of proteins in gels: Coomassie stain

Step 1.

As soon as the power is turned off the separated protein bands will begin to diffuse (they are freely soluble in aqueous solution). To prevent diffusion of proteins treat the gel with a 40% distilled water, 10% acetic acid, and 50% methanol solution which causes almost all proteins to precipitate (become insoluble).

Visualization of proteins in gels: Coomassie stain

Step 2.

To visualize the fixed proteins place the gel in the same mixture of water/acetic acid/methanol but with the addition of 0.25% by weight Coomassie Brilliant Blue R-259.

Visualization of proteins in gels: Coomassie stain

Step 3.

Incubate 4 hours to overnight at room temperature on a shaker.

🕒 **DURATION**

04:00:00

Visualization of proteins in gels: Coomassie stain

Step 4.

Transfer the gel (save the dye mixture; it can be re-used many times) to a mixture of 67.5% distilled water, 7.5% acetic acid, and 25% methanol, place on shaker, and replace with fresh rinse mixture until the excess dye has been removed.

Visualization of proteins in gels: Coomassie stain

Step 5.

The stain will not bind to the acrylamide, and it will wash out (leaving a clear gel). However, it remains strongly bound to the proteins in the gel, and these take on a deep blue color.

Visualization of proteins in gels: Copper stain

Step 6.

Briefly rinse freshly-electrophoresed gels in distilled water (30 seconds maximum) and then transfer to 0.3 M CuCl_2 for 5 to 15 minutes.

🕒 **DURATION**

00:15:00

Visualization of proteins in gels: Copper stain

Step 7.

Wash the gels briefly in de-ionized water, and view them against a dark-field background.

Visualization of proteins in gels: Copper stain

Step 8.

Proteins come up as clear zones in a translucent blue background. Gels may be destained completely by repeated washing in 0.1-0.25 M Tris/0.25 M EDTA pH 8.0.

Visualization of proteins in gels: Copper stain

Step 9.

Move the gel to a dish of transfer buffer before proceeding with transfer according to the transfer apparatus manufacturer's instructions.

Transfer: Large proteins (>100 kD)

Step 10.

For large proteins, transfer out of the gel may be very slow, just as they run slowly within the gel during separation. If blotting a large protein, be sure to run your samples in a low-concentration gel, 8% or less. These will be very fragile, so handle carefully.

Transfer: Large proteins (>100 kD)

Step 11.

Large proteins will tend to precipitate in the gel, hindering transfer. Adding SDS to a final concentration of 0.1% in the transfer buffer will discourage this. Methanol tends to remove SDS from proteins, so reducing the methanol percentage to 10% or less will also guard against precipitation.

Transfer: Large proteins (>100 kD)

Step 12.

Lowering methanol in the transfer buffer also promotes swelling of the gel, allowing large proteins to transfer more easily.

Transfer: Large proteins (>100 kD)

Step 13.

Methanol is only necessary if using nitrocellulose. If using PVDF, methanol can be removed from the transfer buffer altogether, and is only needed to activate the PVDF before assembling the gel/membrane sandwich.

Transfer: Large proteins (>100 kD)

Step 14.

Choose wet transfer overnight at 4°C instead of semi-dry transfer.

Transfer: Small proteins (<100 kD)

Step 15.

All proteins are hindered from binding to membranes by SDS but small proteins more so than large proteins. If your protein of interest is small, consider removing SDS from the transfer buffer.

Transfer: Small proteins (<100 kD)

Step 16.

Keep the methanol concentration at 20%.

Transfer: Small proteins (<100 kD)

Step 17.

See guidelines for more discussion on transfer.

Visualization of proteins in membranes: Ponceau Red

Step 18.

To check for success of transfer, wash the membrane in TBST (for a TBST recipe, see guidelines).

Visualization of proteins in membranes: Ponceau Red

Step 19.

Dilute the stock Ponceau Red 1:10. The stock is made of 2% Ponceau S in 30% trichloroacetic acid and 30% sulfosalicylic acid.

Visualization of proteins in membranes: Ponceau Red

Step 20.

Incubate on an agitator for 5 minutes.

 DURATION

00:05:00

Visualization of proteins in membranes: Ponceau Red

Step 21.

Wash extensively in water until the water is clear and the protein bands are well defined.

Visualization of proteins in membranes: Ponceau Red

Step 22.

The membrane may be destained completely by repeated washing in TBST or water. When using a PVDF membrane, re-activate the membrane with methanol then wash again in TBST.

Blocking the membrane

Step 23.

To prepare a 5% milk or BSA solution, weigh 5 g per 100 ml of Tris Buffer Saline Tween20 (TBST) buffer.

Blocking the membrane

Step 24.

Mix well and filter. Failure to filter can lead to “spotting” where tiny dark grains will contaminate the blot during development.

Blocking the membrane

Step 25.

Incubate for 1 hour at 4°C under agitation.

🕒 DURATION

01:00:00

Blocking the membrane

Step 26.

Rinse for 5 seconds in TBST after the incubation.

🕒 DURATION

00:00:05

Incubation with the primary antibody

Step 27.

Dilute the antibody in TBST at the suggested dilution, if the datasheet does not have a recommended dilution try a range of dilutions (1:100-1:3000) and optimize the dilution according to the results. Too much antibody will result in non-specific bands.

📌 NOTES

Irina Agarkova 22 Mar 2016

It is traditional in certain laboratories to incubate the antibody in blocking buffer, while other laboratories incubate the antibody in TBST without a blocking agent. The results are variable from antibody to antibody and you may find it makes a difference to either use no blocking agent in the antibody buffer or the same agent as the blocking buffer.

Irina Agarkova 22 Mar 2016

If high background is not an issue, some antibodies produce a much stronger signal if diluted in buffer with low concentrations (0.5-0.25%) of milk or BSA, or none at all.

Incubation with the primary antibody

Step 28.

The incubation time can vary between a few hours and overnight (rarely more than 18 hours), and is dependent on the binding affinity of the antibody for the protein and the abundance of protein. We recommend a more dilute antibody and a prolonged incubation to ensure specific binding.

Incubation with the primary antibody

Step 29.

Incubation temperature is preferably cold. In incubating in blocking buffer overnight, it is imperative to incubate at 4°C or contamination will incur and thus destruction of the protein (especially phosphor groups). Agitation of the antibody is recommended to enable adequate homogenous covering of the membrane and prevent uneven binding.

Incubation with the secondary antibody

Step 30.

Wash the membrane several times in TBST while agitating, 5 minutes or more per wash, to remove residual primary antibody.

🕒 DURATION

00:05:00

Incubation with the secondary antibody

Step 31.

Dilute the antibody in TBST at the suggested dilution. If the datasheet does not have a recommended dilution, try a range of dilutions (1:1000 - 1:20,000) and optimize the dilution according to the results. Too much antibody will result in non-specific bands.

Incubation with the secondary antibody

Step 32.

You may incubate the secondary antibody (and primary antibody) in blocking buffer, but a reduction

in background may come at the cost of a weaker specific signal, presumably because the blocking protein hinders binding of the antibody to the target protein.

Incubation with the secondary antibody

Step 33.

Incubate for 1-2 hours, room temperature, with agitation.

 **DURATION**

02:00:00

Incubation with the secondary antibody

Step 34.

We recommend HRP-conjugated secondary antibodies. ALP-conjugated secondary antibodies (alkaline phosphatase) are not recommended as they are not sensitive enough.