

Dec 05, 2019

## HuBMAP Tissue Sectioning for FFPE Specimens

Leigh Propper<sup>1</sup>, Marda Jorgensen<sup>1</sup>

<sup>1</sup>University of Florida

1

Works for me

[dx.doi.org/10.17504/protocols.io.93qh8mw](https://doi.org/10.17504/protocols.io.93qh8mw)

Leigh Propper  
University of Florida



### ABSTRACT

This method details our microtomy (sectioning) process for research specimens involved with HuBMAP.

This process follows the microtomy process that follows after receiving, processing, and embedding of research specimens into formalin fixed, paraffin embedded tissue blocks.

### GUIDELINES

- Managers and supervisors - are responsible for making sure that technicians are properly trained and equipment and facility are maintained in good working order.
- Laboratory personnel - are responsible for reading and understanding this SOP and related documents and to perform these tasks in accordance with the SOPs.

### MATERIALS

NAME ▾	CATALOG # ▾	VENDOR ▾
Water		
KimWipes		Fischer Scientific
Shandon™ Cartilage Curved Thumb Forceps, Curved, Fine Point, standard, 5 in. (12.7cm)	1631TS	Thermo Fisher
Flotation Bath	View	Fisher Scientific
Microtome Blades (Thermo Scientific Ultra)	3053835	Thermo Scientific
Gauze 4x4 Non-Sterile Squares	MSD-1400249	Fisher Scientific
Microtomy Brush	SLK1000	Cancer Diagnostics
Microtome	HM 315 / HM 325	Diagnostic Pathology
Fisherbrand™ Superfrost™ Plus Stain Slides	22-034979	Fisher Scientific
EZ-QUIK SLIDE STAINING RACK	NC0103846	Fisher Scientific
Tissue-Tek® Cold Plate	25608-942	VWR Scientific
Moist Mark Plus Slide/Cassette Marker	SKU: MP2100	Cancer Diagnostics

### SAFETY WARNINGS

- Use extreme care when working with microtome blades. They are extremely sharp.
- Use physical safety precautions when working with sharps (disposable blades).
- As these specimens are fixed in formaldehyde, gloves and other PPE are optional / personal preference.

### BEFORE STARTING

- Ensure you have proper slides, blades, forceps, and your personal preference of gauzes/wipes
- Ensure your water bath has enough water and is the correct temperature.

### Tissue Block Preparation

- 1 5m
  - Place the blocks face down on a cold plate, container of ice or specialized histological ice tray.
  - The best practice for most sectioning is to place a few paper towels or Wypall on top of the tray and dampen it with water.



Cold Plate Example: Sakura Tissue Tek Poly Cold Plate



- Ensure the tissue blocks you have chosen to section have been correctly embedded, and all wax has been scraped/cleaned off the sides of the block.
- Ensure the tissue blocks have been correctly labeled with proper identification (case #, type, etc)

### Microtome Preparation

- 2 5m

Locate your tissue flotation bath / water bath and fill it completely with de-ionized or purified water.

- 3 1m 30s

Turn the flotation bath on, and set the water temperature to 42°C.

#### **42 °C Flotation Bath**



At this step, also inspect the microtome you are planning to use; Ensure it is clean, well maintained, and set at the correct angle.

## Slide Preparation

- 4 ■ Using a pencil or specialized marker for microscope slides, label each slide you plan to use ahead of time with it's relevant information. (case #, type, test, etc) <sup>5m</sup>



- Be sure to locate the glass charged microscope slides that are appropriate for the tissue type or project you are about to perform microtomy for.
- Wear gloves for this step to keep the slides clean of debris/prints, and to prevent ink/graphite from smearing.

## Tissue Block Facing

- 5 ■ After locating the appropriate blades to use for your microtome, carefully select a blade from it's container, and place it inside the microtome's knife holder area. Be sure to secure it in place with the knife clamp/lock spanner. <sup>2m</sup>

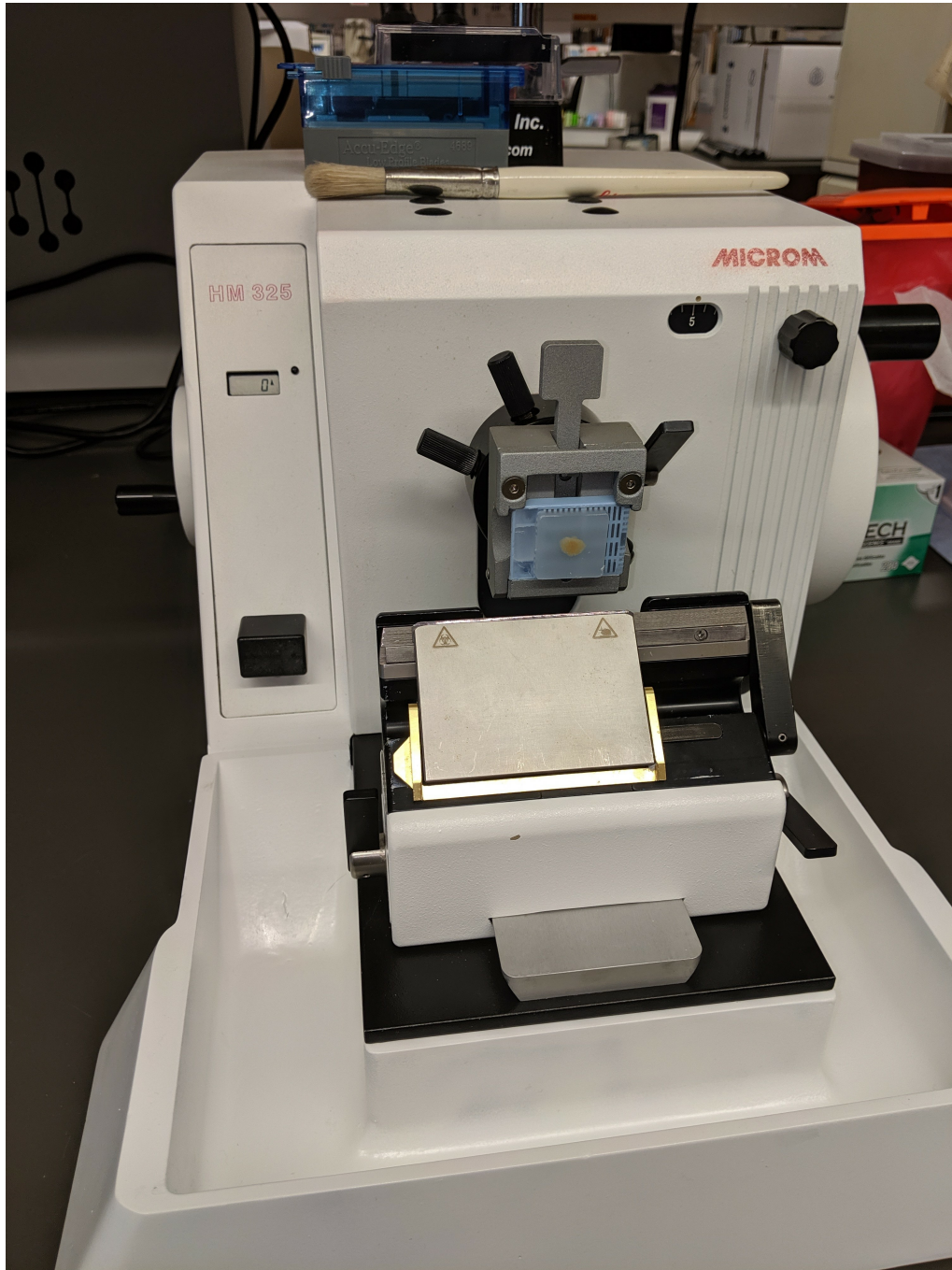


Microtome blade dispensing from storage box.



Before putting a microtome blade into the knife holder, BE SURE to utilize the locking lever under the flywheel.

- 6 After securing your knife and double checking your microtome settings one final time, retrieve a block from your ice tray and secure it in the microtome's chuck (block holder). <sup>3m</sup>



A set up microtome, with a secured knife in the knife holder and tissue block in the microtome chuck.

- 7 After the block is secured, use the coarse advance wheel on the left side of the microtome to carefully, approach the block with the blade and cut a few thin sections to ensure the positioning is correct. Adjust if necessary.
- 8 Trim gently into the block to expose the tissue surface to a level where a complete representative section can be cut.



Trimming is normally done at a thickness of 10-30  $\mu\text{m}$ . This can be performed with the advance flywheel alone, or a combination of both the advance and coarse advance wheels using the rocking method.

- 9 After each tissue block has been trimmed to expose a representative surface of the specimen, place each block back on your ice tray. Let each block chill for approximately 10-15 minutes.



Cold wax allows thinner sections to be obtained by providing support for harder elements within the tissue specimen. The small amount of moisture that penetrates the block from the melting ice pieces or ice water will also make the tissue easier to cut.

## Performing Microtomy 2h 30m

- 10 After your tissue blocks have chilled on the ice tray and are very cold to the touch, they should now be suitable for precision sectioning.

Remove your first block from the ice tray and secure it into the microtome's block chuck.

- 11 Using your coarse adjustment wheel on the left, adjust the block holder to be as close as possible to the edge of the knife, but not touching it.



A helpful tip: When you adjust your chuck holder towards the blade and begin to see small water droplets from the condensation from the frozen block accumulate near the knife holder, stop using the coarse advance wheel.

- 12 Very carefully, unlock and rotate the hand wheel on the right side of the microtome so the block holder is moving up and down in a steady, even manner.  
A ribbon of wax and tissue should begin to form down the front of the metal plate covering the knife holder.



- - Double check the micron thickness setting prior to this step. Normal thickness for sections is 4-5  $\mu\text{m}$ .
- - It is normal to have to scrap the first few sections that appear on your ribbon due to holes or other cutting artifacts.

- 13 After your ribbon has begun to form on the front of the knife holder plate, use a pair of forceps (or whatever your preference is for transferring ribbons from the microtome to the waterbath) to gently grip the bottom of the ribbon and another pair of forceps to gently grip the top of the ribbon nearest the blade.

Gently pull the ribbon up and away from the microtome and towards the waterbath.





A selection of tools ideal to use for securing and floating tissue ribbons during microtomy.



Lifting a ribbon of complete sections away from the knife holder plate.



Short gif video of sectioning a spleen by MJ @ University of Florida

- 14 ■ As carefully as possible, shift the ribbon of tissue sections toward your waterbath, and then quickly but gently lay the tissue out on the bath in a "dragging" type motion (either gently towards you, or gently away from you).

- Wait a few moments while the tissue sections sit on the waterbath, as the heated water will help expand any compression and remove some of the wrinkling or folding in the section.

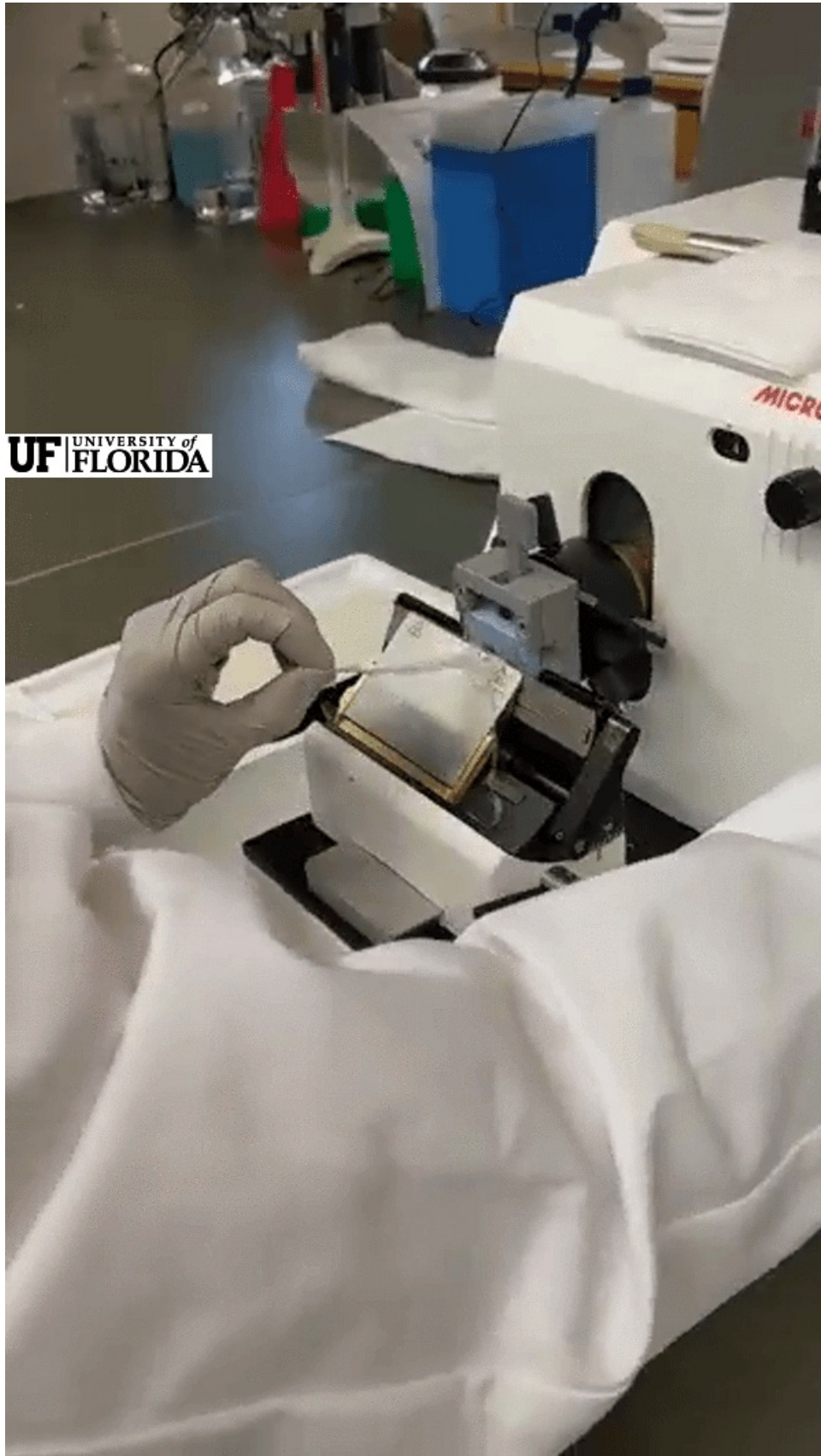


Preparing to place a paraffin ribbon of tissue sections across a waterbath prior to creating a slide.



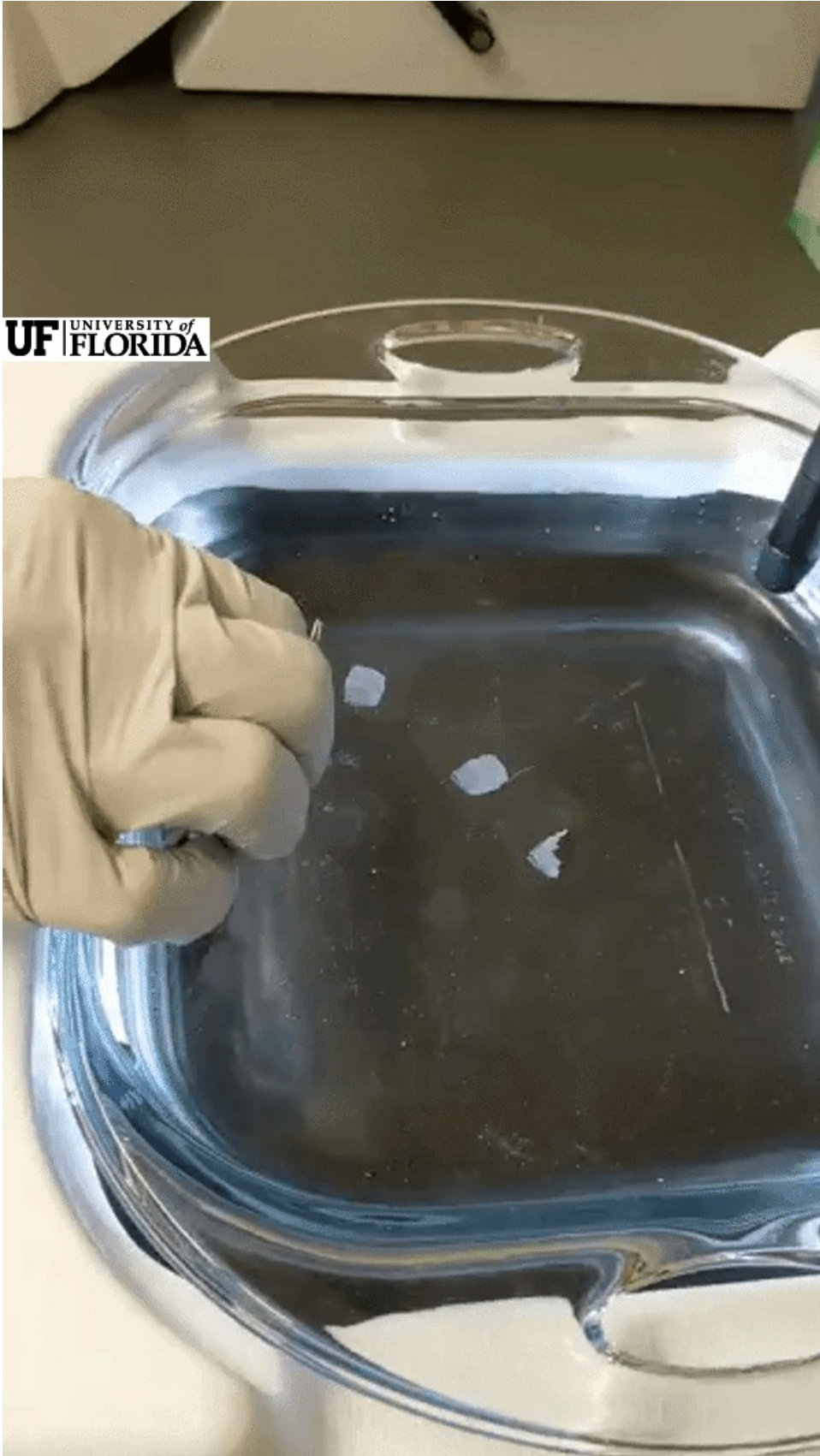
Small ribbon of tissue sections afloat on a waterbath - note they are very nice, with no wrinkling, folding, or knifelines.





Small gif video of moving a paraffin tissue ribbon to waterbath by MJ @ University of Florida

- 15 After your paraffin ribbon has floated on the waterbath for approximately 20-45 seconds, use your forceps or wooden stick applicator, select the section you wish to keep for affixing to the slide.
- 16 After selecting your section, find the appropriately labeled microscope slide that matches the tissue block you sectioned.
  - 16.1 Dip the slide into the waterbath, and place adjacent to and slightly under your chosen section.
  - 16.2 Adjust the section as desired on the slide with the forceps or wooden applicator , and lift upwards out of the waterbath.

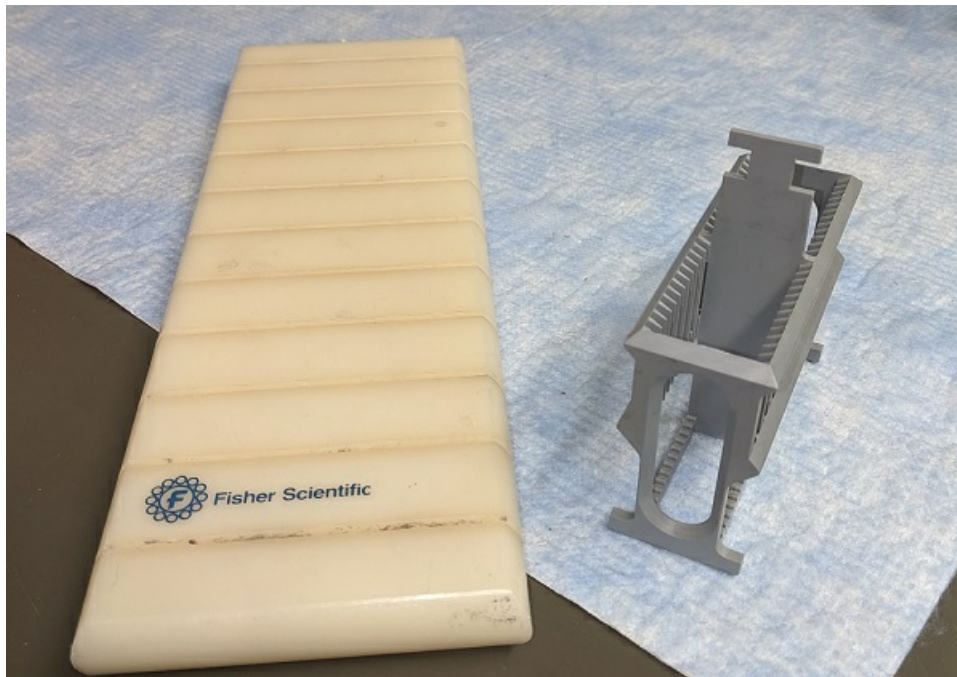


Short Gif video of putting sections on a slide - MJ @ University of Florida

16.3 Place your newly made slide into a slide drying tray or into a slide staining rack, as pictured below.

1h

After completion, let your slides air dry overnight or dry in 50°C oven for 1 hour.



A slide drying tray (left) and manual slide staining rack (right).

Cleanup Area 5m

- 17
- After you have sectioned your last tissue block, unlock the microtome knife blade holder's lever.
  - Using a magnet or forceps, carefully remove the microtome blade and place into the waste container on the blade's box, or in a sharps container.

2m



It is a good practice to clean your microtome if you are done for the day. This prevents paraffin gunk buildup on your microtome, waterbath and floor area, in addition to being ideal for safety.

- 18 Using a microtome brush, brush away all remnants of tissue and paraffin around your microtome, it's catch tray, chuck and behind the knife holder.

3m



A commercially prepared product known as "ParaGuard" can assist with the removal of wax and provide a more efficient microtome clean up. It should be lightly used and vigilantly wiped off with gauze after use.

[Example of Paraguard](#)




19 Carefully remove the glass dish from your waterbath and dump the remaining water into your nearest sink.

1m

20 Clean the emptied waterbath with warm water and dawn dish soap, then replace to the waterbath's bench unit.

1m

 This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited