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Working

UC Davis -  $\beta$  hydroxy butyrate Protocol [↗](#)Peter Havel<sup>1</sup><sup>1</sup>University of California, Davis[dx.doi.org/10.17504/protocols.io.ywafxae](https://doi.org/10.17504/protocols.io.ywafxae)**Mouse Metabolic Phenotyping Centers**  
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## ABSTRACT

**Summary:**

When a sample is mixed with R1, AcAc in the sample is broken down to acetone by AADC. Upon addition of R2, 3-HB in the sample is oxidized in the presence of 3-HBDH and Thio-NAD. This oxidation triggers the cyclic reactions. Since the original AcAc in the sample has been removed, only 3-HB is assayed by measuring the rate of Thio-NADH production spectrophotometrically.

## EXTERNAL LINK

<https://mmpc.org/shared/document.aspx?id=90&docType=Protocol>

## MATERIALS

NAME	CATALOG #	VENDOR
Calibrator	412-73791	FUJIFILM Wako Diagnostic U.S.A.
Reagents	417-73501, 413-73601	FUJIFILM Wako Diagnostic U.S.A.
Microplate		
Platereader		

## MATERIALS TEXT

**Reagent Preparation:**

R1 – reconstitute with buffer provided

R2 – reconstitute with buffer provided

**Note:**FUJIFILM Wako [RRID:SCR\\_013651](#)

- 1 Reconstitute R1 and R2 using the buffers provided.
- 2 Add 4  $\mu$ l of calibrator and sample to each well.
- 3 Add 270  $\mu$ l of R1 to each well. Incubate at 37°C for 5 minutes.

**IMPORTANT:** Make sure not to add any bubbles to the wells when dispensing reagents, this will interfere with reading in the platereader.

- 4 Add 90 µl of R2 to each well. Incubate at 37°C for 2 minutes. Read at 405 nm. Then continue reading every 30 seconds for 2 minutes.
- 5 Calculate the slope of the reaction for each well. The assay will be linear so the unknown samples can be calculated as  $(\text{Sample } \Delta\text{OD}/\text{min} \div \text{Calibrator } \Delta\text{OD}/\text{min}) \times \text{Calibrator Concentration}$ .



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