

# **Chlorella Virus Plaque Assay**

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## **Abstract**

Adapted from: Van Etten, J. (n.d.). Titering of *Chlorella* Viruses. Retrieved from

http://ncv.unl.edu/vanettenlab/

Citation: Steven Wilhelm Chlorella Virus Plaque Assay. protocols.io

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## **Protocol**

#### MBBM Soft Agar

## Step 1.

Melt MBBM soft agar and dispense in 3.0 mL aliquots and hold at 45°C - 50°C in a water bath

#### Concentrate Chlorella

#### Step 2.

Concentrate Chlorella to  $4.0*10^8$  cells/mL at 5,000 rpm for 5 min at  $4^{\circ}$ C and resuspend with fresh MBBM

\*Concentrate so that 0.3 mL can be used per plate

#### Dilute Chlorella Virus

## Step 3.

Dilute virus with 50 mM Tris-HCl, pH 7.8 in 1/10 serial dilutions

\*Fresh lysate contains approximately 10<sup>6</sup>-10<sup>7</sup> PFU/mL (check the titer prior to plaque assay)

\*Dilute sample to have 20-200 plaque forming units (PFU) per plate

## Titering

### Step 4.

To each 3.0 mL soft agar aliquot, add 0.1 mL Chlorella virus and 0.3 mL Chlorella

\*Mix by rolling between palms and pour onto MBBM plate

\*Tilt the plate gently to allow the entire surface to be covered

\*Allow plate to solidify

# Incubation

# Step 5.

Incubate at 25°C in continuous light for 3-4 days or until plaques are visible to count

 ${}^*$ Use a sharpie to count plaques once there is a visible contrast