

# Tangential flow filtration (TFF) concentration of phytoplankton version 2

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## Abstract

Concentrate phytoplankton samples about 100-fold typically from 5L down to 20 mL. Takes about 1 hour per sample. Samples can be used for flow cytometry sorting or for cultures. Enrichment by TFF usually keep growing for a longer time than unconcentrate samples.

## Reference

[Marie, D., Shi, X.L., Rigaut-Jalabert, F. & Vaultot, D. \(2010\). Use of flow cytometric sorting to better assess the diversity of small photosynthetic eukaryotes in the English Channel. \*FEMS Microbiology Ecology\*. 72. p.pp. 165-178.](#)

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## Guidelines

In order to estimate actual concentration efficiency measure *Synechoccus*, pico, and nanoeukaryote concentration before and after TFF.

## Before start



## Necessary equipment

- Vivaflow Cartridge 100,000 MWCO (Regenerated Cellulose - RC) VF20C4 for viruses
- Vivaflow Cartridge 0.2  $\mu$ m (PES) VF20P7 for plankton
- Masterflex Pump 6-600 rpm (ref Bioblock F39671)
- Rotor 3 "galets" (ref F39110)
- 1 Head high throughput (ref F40103) (Can be replaced with quick load head)
- Replace tube provided by stronger tube with two connectors (see picture)

- Bottle 6 L
- Bottle 1 L
- Conical tube 50 mL
- Masterflex Tygon tubing size 16
- Plastique pipettes (1 mL)
- Clamps with screw (to control retentate speed)
- Clamps

## Materials

- ✓ MilliQ water by Contributed by users
- ✓ Filtered Seawater (0.2  $\mu\text{m}$ ) by Contributed by users
- ✓ 0.1 M NaOH by Contributed by users
- ✓ Ethanol 10% by Contributed by users

## Protocol

### Rinsing cartridge

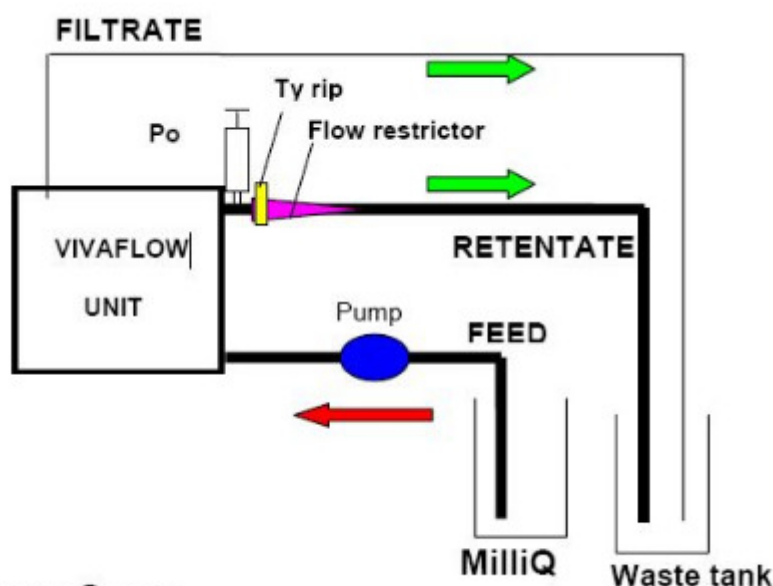
#### Step 1.

Get Vivaflow cartridge out of storage

### Rinsing cartridge

#### Step 2.

Mount Vivaflow cartridge as Fig. 1 (image de C. Brussaard) in open circuit.



**FIG 1. FLUSHING SET UP**

### Rinsing cartridge

#### Step 3.

Remove the clamps

### Rinsing cartridge

#### Step 4.

Set the pump to maximum speed. Manometer should be at about 2.5 bars (with a new cassette

sometimes the manometer get stuck, if the value is too low there is a leak in the system)

### Rinsing cartridge

#### Step 5.

Rinse the cartridge with about 250 mL of MilliQ water (longer is cartridge has been stored in ethanol)

##### AMOUNT

250 ml Additional info:

##### REAGENTS

✓ MilliQ water by Contributed by users

##### DURATION

00:10:00

### Rinse cartridge

#### Step 6.

Replace MilliQ water by sample in 6 L bottle

### Rinse cartridge

#### Step 7.

Take sample for flow cytometry to compute concentration factor.

### Rinse cartridge

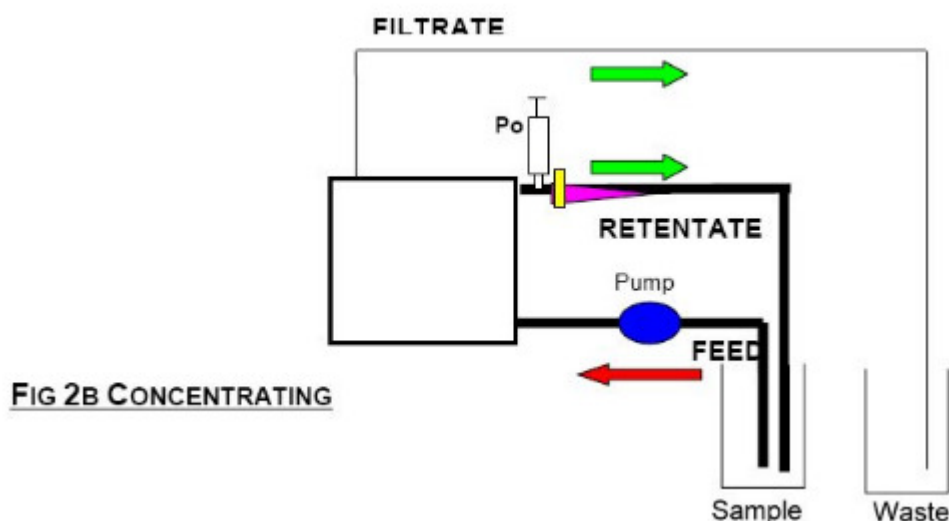
#### Step 8.

Rinse cartridge with about 250 mL of sample

### Concentrate

#### Step 9.

Put the retentate line into the sample bottle (Fig. 2B - Recirculation mode). Record sample volume and start pump increasing to maximum speed.



### Concentrate

#### Step 10.

Clamp the retentate line to increase filtrate flow so that Manometer gets up to 2.5 bars.

### Concentrate

#### Step 11.

Concentrate sample until about 250 mL remains (6 L takes about one hour)

##### DURATION

01:00:00

## Concentrate

### Step 12.

Transfer sample to a smaller bottle (250 mL) then finally to a 50 mL tube with conical bottom

 **DURATION**

00:10:00

## Concentrate

### Step 13.

Continue to concentrate very carefully, lowering the pump speed until the sample volume is reduced to 15-20 mL

 **NOTES**

**Daniel Vaultot** 08 Dec 2016

It is very important to lower the pump speed in order to avoid losing the sample.

## Recirculation

### Step 14.

When final volume is about 10 mL, clamp filtrate tube and recirculate slowly (no change of volume should take place)

## Recirculation

### Step 15.

Leaving the filtrate tube clamped, get the feed line out of the sample in order to get back the total volume of concentrated sample

## Recirculation

### Step 16.

Take sample for flow cytometry count (to compare with original sample concentration and estimate concentration factor)

## Recirculation

### Step 17.

Store concentrated sample for later use (e.g. flow cytometry sorting, culture etc...)

## Rinsing

### Step 18.

Go back to Fig. 1 configuration (Open circuit)

## Rinsing

### Step 19.

Rinse 1 min with filtered sea water

 **REAGENTS**

✓ Filtered Seawater (0.2 µm) by Contributed by users

 **DURATION**

00:01:00

## Rinsing

### Step 20.

Rinse 1 min with distilled water

 **REAGENTS**

✓ Distilled Water by Contributed by users

 **DURATION**

00:01:00

## Rinsing

### Step 21.

Rinse with 50 mL NaOH 0.1 M

 [AMOUNT](#)

50 ml Additional info:

 [REAGENTS](#)

✓ 0.1 M NaOH by Contributed by users

Rinsing

### Step 22.

Put all three tubes (feed, retentate, filtrate) in bottle containing NaOH 0.1 M

 [REAGENTS](#)

✓ 0.1 M NaOH by Contributed by users

Rinsing

### Step 23.

Recirculate for 20 min (to get rid of everything on the cartridge filter)

 [DURATION](#)

00:20:00

Rinsing

### Step 24.

Rinse with 250 mL of MilliQ water (Fig. 1)

 [AMOUNT](#)

250 µl Additional info:

 [REAGENTS](#)

✓ MilliQ water by Contributed by users

Storage

### Step 25.

Stop the pump and clamp all three tubes

Storage

### Step 26.

Store at 4°C. For a storage beyond 1 day, store with Ethanol 10%

 [REAGENTS](#)

✓ Ethanol 10% by Contributed by users