

# Whole genome sequencing of respiratory syncytial (RSV) virus from clinical samples with low viral load

Stephanie Goya, Gabriel L. Rojo, Mercedes S. Nabaes Jordar, Laura E. Valinotto, Alicia S Mistchenko, Mariana Viegas

## Abstract

Here is a description of a protocol for whole genome sequencing of RSV from clinical samples (nasopharyngeal aspirates -NPA-). The protocol was tested with samples with viral loads as low as  $10^{+03}$  viral copies/ml NPA.

The RNA is amplified by RT-PCR in five overlapped fragments of around 2300-4500 nt in length by using specific primers which anneal in conserved regions of the genome. Briefly the protocol includes: viral RNA extraction from NPA done with silica membrane columns and fragment amplification performed in five independent reaction tubes with OneStep RT-PCR Kit (Qiagen). Each fragment size check performed in an agarose gel electrophoresis, clean-up step done with silica membrane columns and the quantification step performed by Qubit. Finally, amplicons pooled equimolarly and library prep done with Nextera XT Kit.

**Citation:** Stephanie Goya, Gabriel L. Rojo, Mercedes S. Nabaes Jordar, Laura E. Valinotto, Alicia S Mistchenko, Mariana Viegas Whole genome sequencing of respiratory syncytial (RSV) virus from clinical samples with low viral load. **protocols.io**

dx.doi.org/10.17504/protocols.io.qsedwbe

**Published:** 06 Jun 2018

## Materials

- ✓ Agarose [A5304](#) by Contributed by users
- DNA Clean & Concentrator™-5 [D4003](#) by [Zymo Research](#)
- Qubit™ dsDNA HS Assay Kit [Q32851](#) by [Invitrogen - Thermo Fisher](#)
- Ethidium bromide [EB, EtBr] [EB0195.SIZE.25g](#) by [Bio Basic Inc.](#)
- Agencourt Ampure XP [A63880](#) by [Beckman Coulter](#)
- RiboLock RNase Inhibitor #EO0381 by [Thermo Fisher Scientific](#)
- Nextera XT DNA Library Preparation Kit [FC-131-1096](#) by [illumina](#)
- OneStep RT-PCR Kit [210210](#) by [Qiagen](#)
- PureLink viral RNA/DNA mini kit [12280050](#) by [Invitrogen - Thermo Fisher](#)
- QIAamp Viral RNA Mini Kit [52904](#) by [Qiagen](#)

## Protocol

### Viral RNA extraction

### Step 1.

Centrifuge the nasopharyngeal aspirate (NPA) at 5,000 g for 5 min to remove cellular debris.

#### Viral RNA extraction

### Step 2.

Extract viral RNA from 200 µl of NPA by following the protocol PureLink viral RNA/DNA mini kit (Thermo Fisher Scientific) according the manufacturer's instructions except for 2.8 µl of carrier RNA instead of 5.6 µl as recommended to reduce the presence of tRNA in the extracted RNA. Nevertheless, 140 µl of NPA should be used if QIAamp Viral RNA Mini Kit (Qiagen) is used instead of PureLink kit. In this case, no changes from the protocol should be done.

#### Viral RNA extraction

### Step 3.

Elute viral-extracted RNA in 40 µl of DNase/RNase-free water. Add 1.25 µl of 40 U/µL RiboLock RNase Inhibitor (Thermo Fisher Scientific) to preserve the extracted RNA.

#### Viral RNA extraction

### Step 4.

Extracted RNA can be storage at -80°C.

#### RT-PCR

### Step 5.

#### PRIMERS:

#### Fragment 1: 2384 bp

Forward 1f: 5'-ACGCGAAAAAATGCGTACwAC-3'

Reverse 2384r: 5'-GCrTCTTCTCCATGrAATTC-3'

#### Fragment 2: 2731 bp

Forward 2124f: 5'- GCwGGyCTAGGCATAATG-3'

Reverse 4875r: 5'- GTTGTTTrGTGTrACTTTGT-3'

#### Fragment 3: 4351 bp

Forward 3314f: 5'-AyCCyGCATCACTwACAAT-3'

Reverse 7665r: 5'-CAGGAAACAGCTATGACCyAAGCA-3'

#### Fragment 4: 4114 bp

Forward 7094f: 5'-TGATGCATCAATATCTCAAGTC-3'

Reverse 11208r: 5'-CTCCTGTGTTAAGCTACCTATAG-3'

#### Fragment 5: 4331 bp

Forward 10438f: 5'-AGTyTkACAAGATATGGTGATCT-3'

Reverse 15180r: 5'-AAGTGTCAAAAATAATATCTCGT-3'

All the procedure should be done in ice.

Amplify each fragment in an independent RT-PCR reaction with OneStep RT-PCR kit of Qiagen by mixing:

primer f (25  $\mu$ M)--- 0.6  $\mu$ l

primer r (25  $\mu$ M)--- 0.6  $\mu$ l

RNA--- 6  $\mu$ l

Incubate at 65 °C for 5 min, then place 2 min in ice.

Add (master mix can be done):

H2O DNase/RNase free--- 5  $\mu$ l

Buffer 5X--- 10.55  $\mu$ l

RNAse Inhibitor (40 U/ $\mu$ l)---0.25  $\mu$ l

dNTP mix (10 mM)--- 1  $\mu$ l

Enzyme Mix--- 1  $\mu$ l

(Final volume: 25  $\mu$ l)

Incubate:

45 °C 30 min

95 °C 15 min

40 times: 94 °C 10 sec

55 °C 30 sec

68 °C 4.5 min

68 °C 10 min

4 °C hold

Fragment verification, quantification and pool

## Step 6.

Run 4 µl of RT-PCR product on a 1.8% agarose gel stained with Ethidium Bromide.

Fragment verification, quantification and pool

#### **Step 7.**

Perform a clean-up of RT-PCR product with DNA Clean & Concentrator kit (Zymo Research) following the manufacturer's instructions. Elute with water PCR-grade, avoid elution with buffer containing EDTA.

Fragment verification, quantification and pool

#### **Step 8.**

Run 2 µl of each amplicon in a Qubit dsDNA HS Assay.

Fragment verification, quantification and pool

#### **Step 9.**

Pool equimolarly the 5 amplicons

NGS

#### **Step 10.**

Follow NexteraXT kit (Illumina) to perform library preparation.

After index addition by amplification and clean-up with Agencourt Ampure XP, quantify the libraries with the Qubit dsDNA HS Assay Kit.

Perform a standard normalization instead of beads normalization. Never normalize with beads when libraries have a concentration lower than 15 nM

If you are not familiar with bioinformatic analysis in NGS, we recommend using UGENE or Geneious.

## **Warnings**

Ethidium bromide is a potent mutagen. Ethidium bromide solution must be handled with extreme caution and decontaminated prior to disposal.