OmniPrep™ For High Quality Genomic DNA Extraction From Plant Tissue (Fresh or Frozen)

G-Biosciences

Abstract

The OmniPrep™ kit isolates high quality genomic DNA from many different species and tissue types including animal, plant, bacteria, yeast, fungi, whole blood, and cells in culture. DNA can be isolated from samples high in polysaccharides or other contaminants that are difficult to remove from the DNA preparations.

This protocol is for use with **plant tissue**. Please refer to the appropriate protocol depending on your application.

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Guidelines

INTRODUCTION

The OmniPrep™ kit isolates high quality genomic DNA from many different species and tissue types including animal, plant, bacteria, yeast, fungi, whole blood, and cells in culture. DNA can be isolated from samples high in polysaccharides or other contaminants that are difficult to remove from the DNA preparations.

Several unique features separate the OmniPrep™ kit from other methods:

• A unique and proprietary formulation of detergents and salts. DNA can be extracted and purified from almost any tissue without the use of toxic agents such as phenol.

• A quick protocol, which isolates extremely clean genomic DNA. On an average, the DNA is 100kb in size and generally hydrates in minutes.

• Provides an option to modify protocol for difficult to handle samples.

• Suitable for 200 Preps (10mg/prep). The kit is adaptable for larger tissue volumes.

ITEM(S) SUPPLIED

Description	Cat. # 786-136	Cat. # 786-136S
Genomic Lysis Buffer	100ml	2 x 2ml
Nuclei Isolation Buffer	2 x 30ml	-
DNA Stripping Solution	10ml	0.5ml
Precipitation Solution	30ml	2ml
Mussel Glycogen (10mg/ml)	1ml	25µl
TE Buffer	20ml	0.5ml
Longlife™ RNase (5mg/ml; >60U/mg)	0.5ml	50µl
Longlife™ Proteinase K (5mg/ml)	2 x 0.5ml	50µl

NOTE: Cat. # 786-136S is a trial/sample size and does not contain enough reagents for all the protocols. For regular size kit, order Cat. # 786-136.

STORAGE CONDITIONS

The kit is shipped at ambient temperature. Upon arrival, store the kit components as recommended on the reagent label.

ADDITIONAL REAGENTS REQUIRED

• For all samples: Isopropanol, 70% Ethanol, and Chloroform

• For Gram positive Bacteria: Lysozyme, 0.5M EDTA. Longlife™ Lysozyme, (Cat# 786-037) available.

• For Yeast: Zymolyase® , β-mercaptoethanol, phosphate buffered saline (PBS). Longlife™ Zymolyase® (Cat# 786-036) available.

PREPARATION BEFORE USE

Proteinase K Solution: To avoid repeated freezing-thaw, dispense the Proteinase K solution into aliquots of 30µl/tube and freeze at -20°C.

Genomic Lysis Buffer & DNA Stripping Solution: If a precipitate forms due to cold storage allow to

warm to room temperature until precipitate dissolves.

PROTOCOLS

The protocols contained in this manual are listed below and are based on the Extraction from Solid Tissue protocol. Thoroughly review the appropriate protocols before commencing isolation of genomic DNA.

TROUBLESHOOTING

• POOR DNA RECOVERY:

o Increase volumes of Genomic Lysis Buffer, DNA Stripping Solution and Precipitation Solution proportionally.

o Introduce optimal Proteinase K step after step 4.

o Improve grinding technique.

• For efficient grinding of small samples we offer Molecular Grinding Resin™ (Cat. # 786-138), high tensile micro-particles that do not bind nucleic acids and are recommended for grinding and isolation of genomic DNA.

• DNA/RNA free matching pestles and microfuge tubes (1.5ml) for grinding small samples are also available (Cat. # 786-138P).

• FOR >100kb GENOMIC DNA

o Recommend using MegaLong™ DNA isolation kit (Cat. # 786-146, 786-147). Isolates nuclei under mild condition, which are then transferred to Tube-ODIALYZER™ , a new device for DNA isolation. Nuclei are digested with protease followed by dialysis to remove protein and other contaminants. For more information call our Technical Department.

Typical Yield: 0.5-3µg/mg plant tissue.

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Before start

Proteinase K Solution: To avoid repeated freezing-thaw, dispense the Proteinase K solution into aliquots of 30µl/tube and freeze at -20°C.

Genomic Lysis Buffer & DNA Stripping Solution: If a precipitate forms due to cold storage allow to warm to room temperature until precipitate dissolves.

Materials

OmniPrep™ 786-136 by G-Biosciences

Protocol

Step 1.

Most plant tissues are best prepared by freezing in liquid nitrogen. Grind samples in liquid nitrogen to a fine powder and quickly add to an appropriate volume of Lysis Buffer.

Step 2.

Add 50-100mg finely ground dried tissue, frozen tissue or fresh leave tissue to a microcentrifuge tube containing 500µl Genomic Lysis Buffer.

Step 3.

If ground, vortex for 5 seconds; if unground, homogenize the sample with a microfuge pestle until a homogenous suspension is acquired, approximately 30-60 strokes.

Step 4.

Incubate the sample at 65°C for 60 minutes with periodic inversions.

DURATION
00:00:00

Step 5.

Allow the sample to cool to room temperature.

Step 6.

Add 200µl chloroform and mix by inverting the tube several times.

Step 7.

Centrifuge for 10 minutes at 14,000xg and carefully remove the upper phase to a clean microcentrifuge tube.

DURATION
00:10:00

Step 8.

Add 50µl DNA Stripping Solution to the sample and invert several times to mix.

Step 9.

Incubate the sample for 5-10 minutes at 60°C.

DURATION
00:05:00

Step 10.

Add 100µl Precipitation Solution and mix by inverting the tube several times.

NOTES
Colin Heath 17 Jun 2016

A white precipitate should be produced, if not add 50µl aliquots of Precipitation Solution until a white precipitate forms.

Step 11.
Centrifuge the sample at 14,000xg for 5 minutes.

DURATION
00:05:00

Step 12.

Transfer the supernatant to a clean tube and precipitate the genomic DNA with 500µl isopropanol.

Step 13.

Invert the tubes 10 times to precipitate the DNA.

Step 14.

OPTIONAL: For increased DNA recovery, add 2µl Mussel Glycogen as a DNA carrier.

Step 15.

Centrifuge at 14,000xg for 5 minutes to pellet genomic DNA.

DURATION
00:05:00

Step 16.

Remove the supernatant.

Step 17.

Add 700µl 70% ethanol to the tube and invert several times to wash the DNA pellet.

Step 18.

Centrifuge for 1 minute at 14,000xg.

DURATION
00:01:00

NOTES
Colin Heath 17 Jun 2016

In some samples, the pellet may be hard to see at this point and will be loosely attached to the tube.

Step 19.

Decant or pipette off the ethanol wash.

Step 20.

Invert the tube on a clean absorbent surface for several minutes to allow any excess ethanol to drain away.

NOTES
Colin Heath 17 Jun 2016

Do not let the pellet dry completely or it will be difficult to rehydrate.

Step 21.

Add 50µl TE Buffer to the pellet.

Step 22.

Incubate at room temperature for at least 15 minutes to rehydrate.

DURATION
00:15:00

NOTES
Colin Heath 17 Jun 2016

Incubating the tube at 55-60°C will speed up rehydration. Incubate for 5-60 minutes.

Step 23.

OPTIONAL: 1µl LongLife™ RNase for every 100µl TE Buffer can be added at this stage.

Step 24.

Store DNA at 4°C, for long term storage store at -20°C or -80°C.

NOTES
Colin Heath 22 Jun 2016

Typical Yield: 0.5-3µg/mg plant tissue.