

# Megabase DNA Extraction from Animal Blood

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## Abstract

This protocol provides an efficient and reliable technique for obtaining HMW DNA(>1Mb) from animal blood. It accompanies:

Huang Zhihai, Xu Jiang, Xiao Shuiming, Liao Baosheng, Gao Yuan, Zhai Chaochao, Qiu Xiaohui, Xu Wen, Chen Shilin

(2016): Supporting data for 'Comparative optical genome analysis of two Pangolin species *Manis pentadactyla* and *Manis javanica*'. GigaScience Database.

**Citation:** Huang Zhihai, Xu Jiang, Xiao Shuiming, Liao Baosheng, Gao Yuan, Zhai Chaochao, Qiu Xiaohui, Xu Wen, Chen Shilin Megabase DNA Extraction from Animal Blood. **protocols.io**

[dx.doi.org/10.17504/protocols.io.gagbsbw](https://dx.doi.org/10.17504/protocols.io.gagbsbw)

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## Protocol

### sample collection

#### Step 1.

Collect 3mL animal's blood in 5 mL EDTA anticoagulative tube, and store at 4 °C

### sample collection

#### Step 2.

Mix blood by gently rocking for 10min at room temperature to ensure a homogenous WBC distribution.

⌚ DURATION

00:10:00

### sample collection

#### Step 3.

Transfer 3ml blood to 15ml screw cap tube containing 9ml RBC lysis solution (3 volumes), and mix by gently inverting 10x.

### sample collection

#### Step 4.

Incubate 5 min at room temperature. Gently mix by inverting at least once during incubation.

⌚ DURATION

00:05:00

### sample collection

#### Step 5.

Spin 5 min at 2000xg at 4°C and carefully pipet out supernatant while not disturbing the pellet.

 DURATION

00:05:00

sample collection

**Step 6.**

Resuspend cell pellet in 3ml PBS (1 blood volume):

sample collection

**Step 7.**

Add 9ml rbc lysis sol (3 volumes), invert to mix and incubate 5 min at room temperature.

 DURATION

00:05:00

sample collection

**Step 8.**

Spin cell suspension for 5 min at 2000xg at 4°C and carefully pipet out supernatant while not disturbing the pellet. Remove the last drop of liquid by tilting the tube and removing liquid with pipet tip.

 DURATION

00:05:00

sample collection

**Step 9.**

Resuspend cell pellet in 580ul cell suspension buffer (Bio Rad plug lysis kit) until homogenous suspension.

sample collection

**Step 10.**

Transfer 375ul to a new microfuge tube and label (400). Transfer 187.5ul to another tube labeled (200); add 187.5ul cell suspension buffer, and pipet mix gently 5x. Keep both tubes on ice until ready to embed in agarose (section II).

sample collection

**Step 11.**

Melt 2% agarose (Bio Rad kit) by immersing in microwave-boiled water for 10-15min until agarose is completely melted.

 DURATION

00:10:00

sample collection

**Step 12.**

Equilibrate both tubes from step 11 section I in 43°C water bath/heat block for at least 5 min before use.

 DURATION

00:05:00

sample collection

**Step 13.**

After 5 min, prepare 5 plugs from one tube by performing the next steps rapidly to avoid solidification of the cell-agarose mixture before pipeting into plug mold:

sample collection

**Step 14.**

Place plug cast on ice for at least 45min or until agarose has solidified. Alternatively, can place plug cast on inverted metal microfuge ice block on ice for 45min, to avoid potential contact of agarose with ice

 DURATION

00:45:00

sample collection

**Step 15.**

Wash away hemoglobin and cellular material with Proteinase K and WashBuffer.