

Coring Technique

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Abstract

This is a technique for isolating phage from agar plates. When working with lytic phage, well-isolated, cleared areas in a lawn of bacteria (called plaques) are removed from the agar (by coring with a sterile pipette) and used to infect fresh bacteria in liquid medium. The lysed culture can be used for a new plaque assay after 0.2 μ m filtering to remove residual bacteria. New phage are "plaque-purified" a minimum of three times before they are considered to be free of other phage.

Citation: Matthew Sullivan Lab Coring Technique. **protocols.io**

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Protocol

Step 1.

Poke the plaque in the agarose with the tip of an autoclaved Pasteur pipette to core it out drawing agarose chunks into the pipette

🔗 NOTES

Bonnie Poulos 17 Jun 2015

I would pick a few (2-4?) plaques just in case one does not lyse

■ ANNOTATIONS

Chris Upton 10 Sep 2015

Can you use sterilized tooth-picks?

Would be cheaper.

Step 2.

Inoculate 2-5 ml dense cells (whatever is available) with the cyanophage plaque by pipetting up and down a few times

Step 3.

Let this sit in the incubator for 60 minutes to adsorb and then dilute with about 25 ml media

🕒 DURATION

01:00:00

Step 4.

Do this in parallel to a control tube of just cells and observe the cyanophage inoculated cultures for lysis (clearing) relative to the controls