Sequential smFISH (Allen Institute) Version 2

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Abstract

We have developed a multiplexed single molecule FISH protocol for use at the Institute. This protocol was optimized on human tissue, but will work on mouse tissue as well. It was adapted from Lyubimova et. al., Nature Protocols, 2013.

Citation: Jennie Close, Zoe Maltzer Sequential smFISH (Allen Institute). protocols.io

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Guidelines

Ensure that all reagents are in recombinant and RNAse-free format, as we have noticed RNA degradation in solutions that contain enzymes derived from whole organisms.

We filter every solution with a 0.2um syringe filter prior to use. This reduces background spots and dust that interfere with imaging of diffraction limited spots.

For the SDS treatment after fixation and permeabilization, be gentle when dropping SDS onto the section, as well as during washes. This treatment is relatively harsh and the tissue must be treated somewhat delicately.

Before start

Ensure all incubators and ovens are at the appropriate temperature prior to experiment.

Protocol

Tissue and Sectioning

Step 1.

10-14 um cryosections are taken from fresh-frozen tissue, which are collected on poly-lysinetreated

#1 coverslips at room temperature (RT). After 5-10 min at RT, sections are placed at 4°C until sectioning is complete. At that point, proceed immediately to fixation and permeabilization.

© DURATION 00:05:00 : RT

Fixation/Permeabilization

Step 2.

Post-fix sections for 15 min with 4% PFA @ 4 °C.

■ TEMPERATURE

4 °C: Post-fixing



O DURATION

00:15:00 : Post-fixing Fixation/Permeabilization

Step 3.

Wash with PBS (1/3)



✓ PBS by Contributed by users

Fixation/Permeabilization

Step 4.

Wash with PBS (2/3)



✓ PBS by Contributed by users

Fixation/Permeabilization

Step 5.

Wash with PBS (3/3)



✓ PBS by Contributed by users

Fixation/Permeabilization

Step 6.

Permeabilize with cold methanol at -20 C for 10 min.

■ TEMPERATURE

-20 °C: Permeabilizing

O DURATION

00:10:00 : Permeabilizing

Fixation/Permeabilization

Step 7.

Air dry for 30 min in fume hood (Stopping point: store coverslips at -80C)

O DURATION

00:30:00 : Air drying Fixation/Permeabilization

Step 8.

Optional: Treat sections with 8% SDS/PBS for 10 minutes, followed by 3 - 5 rinses with PBSor 2XSSC

© DURATION 00:10:00:

Fixation/Permeabilization

Step 9.

Add 2ml 2X SSC

AMOUNT
2 ml : 2X SSC

REAGENTS

✓ 2X SSC by Contributed by users

Hybridization

Step 10.

Pre-heat hyb oven to 37 °C

Hybridization

Step 11.

Place sections in hyb buffer without probes.

Hybridization

Step 12.

Add 4 ul probe 400ul hyb buffer.

AMOUNT
4 μl : probe
AMOUNT

400 μl : hyb buffer

NOTES

Specific to 6-well plate format – if using perfusion chamber, this volume can be reduced.

Hybridization

Step 13.

√ protocols.io

3 Published: 15 Aug 2018

Incubate at 37 C for 2H.

♣ TEMPERATURE
37 °C : Incubation
♦ DURATION

02:00:00 : Incubation

Wash

Step 14.

Add 2 ml wash buffer to each well.



2 ml: wash buffer

Wash

Step 15.

Incubate at 37 C for 15 min.

▼ TEMPERATURE

37 °C : Incubation © DURATION

00:15:00 : Incubation

Wash

Step 16.

Remove wash buffer.

Wash

Step 17.

Add 2 ml fresh wash buffer and incubate at 37 C for 15 min.

■ AMOUNT

2 ml : wash buffer ↓ TEMPERATURE 37 °C : Incubation © DURATION

00:15:00 : Incubation

Wash

Step 18.

Replace wash buffer with fresh wash buffer + DAPI (final 5ug/mL) and incubate at 37 C for 15 min.

↓ TEMPERATURE 37 °C : Incubation

O DURATION

00:15:00 : Incubation

Wash

Step 19.

Wash

Step 20.

Mount and image or store at 4 C in 2XSSC until imaging session

▮ TEMPERATURE

4 °C:



✓ 2X SSC by Contributed by users

Stripping

Step 21.

65% formamide/2X SSC, 10 min X 3, 30 C

▮ TEMPERATURE

30 °C:



O DURATION

00:10:00:

Stripping

Step 22.

Wash in 2XSSC (1/3)

Stripping

Step 23.

Wash in 2XSSC (2/3)

Stripping

Step 24.

Wash in 2XSSC (3/3)

Stripping

Step 25.

Following stripping, proceed to hybridization step.



Hybridization step -> go to step #10

Warnings

Please refer to the SDS (Safety Data Sheet) for hazard information and safety warnings.

Avoid exposure to formamide, DAPI