Lysis Buffer

DeLong Lab

Abstract

Adapted from Steripak protocol, with addition of RNase.

Citation: DeLong Lab Lysis Buffer. **protocols.io**
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Guidelines

Required volumes: for ½ plate 20 mls is plenty - split into 15 mls plus 5 mls; for a whole plate 35 mls is good - split into 30 mls plus 5 mls.

Materials

EDTA (0.5 M), pH 8.0 AM9260G by Life Technologies

Protocol

Step 1.
Combine EDTA, Tris, and sucrose

Final Concentration	For 20 ml	For 35 ml
40 mM EDTA	1.6 ml of 0.5 M EDTA	2.8 ml of 0.5 M EDTA
50 mM Tris (pH 8.3)	1.0 ml of 1 M Tris (pH 8.3)	1.75 ml of 1 M Tris
0.73 M Sucrose	5.13 g of Sucrose	8.98 g of Sucrose

REAGENTS
EDTA (0.5 M), pH 8.0 AM9260G by Life Technologies

NOTES
Bonnie Poulos 24 Jul 2015
Make fresh because of fructose.

Step 2.
Shake vigorously to dissolve

Step 3.
Add water to appropriate final volume and shake

Step 4.
Split into two aliquots (15+5 or 30+5)

NOTES

Bonnie Poulos 19 Jun 2015

Can filter-sterilize now but will be sterilizing each aliqout separately so if proceeding immediately (as you should) then no need to double sterilize.

Warnings

Make fresh because of sucrose.