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Intracellular Staining With True-Phos™ Perm Buffer in Cell Suspensions V.2 [↗](#)Sam Li<sup>1</sup><sup>1</sup>BioLegend

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 Works for me [dx.doi.org/10.17504/protocols.io.bac4iayw](https://doi.org/10.17504/protocols.io.bac4iayw)

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## EXTERNAL LINK

<https://www.biolegend.com/protocols/intracellular-staining-with-true-phos-perm-buffer-in-cell-suspensions-protocol/4262/>

## MATERIALS

NAME <span>▼</span>	CATALOG # <span>▼</span>	VENDOR <span>▼</span>
Cell Staining Buffer	420201	BioLegend
Fixation Buffer	420801	BioLegend

## Buffer Preparation:

- 1 Warm Fixation Buffer (BioLegend Cat [420801](#)). For each  $1 \times 10^6$  cells aliquot 0.5mL of buffer and warm to 37°C.
- 2 Chill True-Phos™ Perm Buffer to -20°C. For each  $1 \times 10^6$  cells aliquot 1.0ml of True-Phos™ Perm Buffer and chill to -20°C.

## Sample Preparation:

- 3 Prepare a single cell suspension with the sample of interest (Human PBMC, splenocytes, cell lines, etc).
 

Tips:
 
  - Prepare two aliquots, Negative control: untreated, Positive control: treated with stimuli.
  - Incubate the cells with the appropriate stimuli, at the suitable temperature and time.
- 4 Fix the cells immediately after treatment by adding an equal volume of pre-warmed Fixation Buffer. Gently pipette to ensure thorough mixing.
- 5 Incubate at 37°C for 15 minutes to ensure cells are properly fixed.
- 6 Centrifuge cells at 350xg at room temperature for 5 minutes, decant supernatant, vortex to resuspend cell pellet.

#### Staining with Specific Antibodies:

- 7 Add sufficient Cell Staining Buffer to wash the cells (approximately 2ml for each  $1 \times 10^6$  cells, BioLegend Cell Staining Buffer recommended, Cat [420201](#)), centrifuge at 350xg at room temperature for 5 minutes and decant supernatant. Repeat, for a total of two washes.
- 8 Gently pipette cells using residual volume to resuspend cell pellet. Note: if cells are not fully resuspended, True-Phos™ Perm Buffer addition will cause significant cell loss.
- 9 While vortexing, permeabilize cells by adding pre-chilled True-Phos™ Perm Buffer. Example:  $10 \times 10^6$  cells should be permeabilized with 10mL of pre-chilled True-Phos™ Perm Buffer.
- 10 Incubate at  $-20^{\circ}\text{C}$  for at least 60 minutes to ensure cells are properly permeabilized. Note: cells can be stored in the True-Phos™ Perm Buffer overnight at  $-20^{\circ}\text{C}$ .
- 11 Centrifuge cells at 1000xg at room temperature for 5 minutes, decant supernatant, vortex to resuspend cell pellet.
- 12 Add sufficient Cell Staining Buffer to wash the cells, centrifuge cells at 1000xg at room temperature for 5 minutes, decant supernatant. Repeat, for a total of two washes.
- 13 Resuspend the cells in Cell Staining Buffer at a concentration of  $10 \times 10^6$  cells/ml.
- 14 Transfer 100 $\mu\text{L}$  (or  $1 \times 10^6$  cells) to a 12 x 75mm tube.
- 15 Add antibody cocktail(s) to appropriate tubes, vortex to mix, and incubate for 30 minutes at room temperature in the dark.
- 16 Add 2mL of Cell Staining Buffer, centrifuge cells at 1000xg at room temperature for 5 minutes, decant supernatant. Repeat, for a total of two washes.
- 17 Resuspend cells in approximately 500 $\mu\text{L}$  of Cell Staining Buffer and analyze with a flow cytometer.



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