

Tangential flow filtration (TFF) concentration of phytoplankton version 3

Daniel Vaultot

Abstract

Concentrate phytoplankton samples about 100-fold typically from 5L down to 20 mL. Takes about 1 hour per sample. Samples can be used for flow cytometry sorting or for cultures. Enrichment by TFF usually keep growing for a longer time than unconcentrate samples.

Reference

[Marie, D., Shi, X.L., Rigaut-Jalabert, F. & Vaultot, D. \(2010\). Use of flow cytometric sorting to better assess the diversity of small photosynthetic eukaryotes in the English Channel. *FEMS Microbiology Ecology*. 72. p.pp. 165-178.](#)

Citation: Daniel Vaultot Tangential flow filtration (TFF) concentration of phytoplankton. **protocols.io**

dx.doi.org/10.17504/protocols.io.krmcv46

Published: 10 Nov 2017

Guidelines

In order to estimate actual concentration efficiency measure *Synechoccus*, pico, and nanoeukaryote concentration before and after TFF.

Before start



Necessary equipment

- Vivaflow Cartridge 100,000 MWCO (Regenerated Cellulose - RC) VF20C4 for viruses
- Vivaflow Cartridge 0.2 μm (PES) VF20P7 for plankton
- Masterflex Pump 6-600 rpm (ref Bioblock F39671) - It is critical to have a 600 rpm pump, lower speed will not work. The rate of the pump must be adjustable.
- Rotor 3 "galets" (ref F39110)

- 1 Head high throughput (ref F40103) (Can be replaced with quick load head)
- Replace tube provided by stronger tube with two connectors (see picture)
- Bottle 6 L
- Bottle 1 L
- Conical tube 50 mL (Falcon tube)
- Masterflex Tygon tubing size 16
- Plastique pipettes (1 mL) - this is used to plunge in the sample
- Clamps with screw (to control retentate speed)

Solutions

- MilliQ water : 1L
- NaOH 0.1 N : 500 mL
- EtOH 10% : 500 mL

Materials

- ✓✓ MilliQ water by Contributed by users
- ✓✓ Filtered Seawater (0.2 µm) by Contributed by users
- ✓✓ 0.1 M NaOH by Contributed by users
- ✓✓ Ethanol 10% by Contributed by users

Protocol

Rinsing cartridge

Step 1.

Get Vivaflow cartridge out of storage

Rinsing cartridge

Step 2.

Mount Vivaflow cartridge as Fig. 1 (image de C. Brussaard) in open circuit.

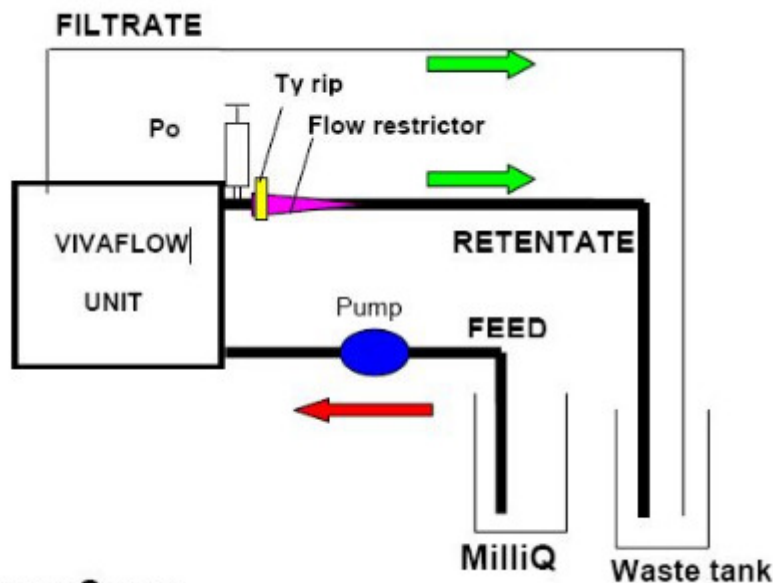


FIG 1. FLUSHING SET UP

Rinsing cartridge

Step 3.

Remove the clamps

Rinsing cartridge

Step 4.

Set the pump to maximum speed. Manometer should be at about 2.5 bars (with a new cassette sometimes the manometer get stuck, if the value is too low there is a leak in the system)

Rinsing cartridge

Step 5.

Rinse the cartridge with about 250 mL of MilliQ water (longer is cartridge has been stored in ethanol)

AMOUNT

250 ml Additional info:

REAGENTS

✓ MilliQ water by Contributed by users

DURATION

00:10:00

Rinse cartridge

Step 6.

Replace MilliQ water by sample in 6 L bottle

Rinse cartridge

Step 7.

Take sample for flow cytometry to compute concentration factor.

Rinse cartridge

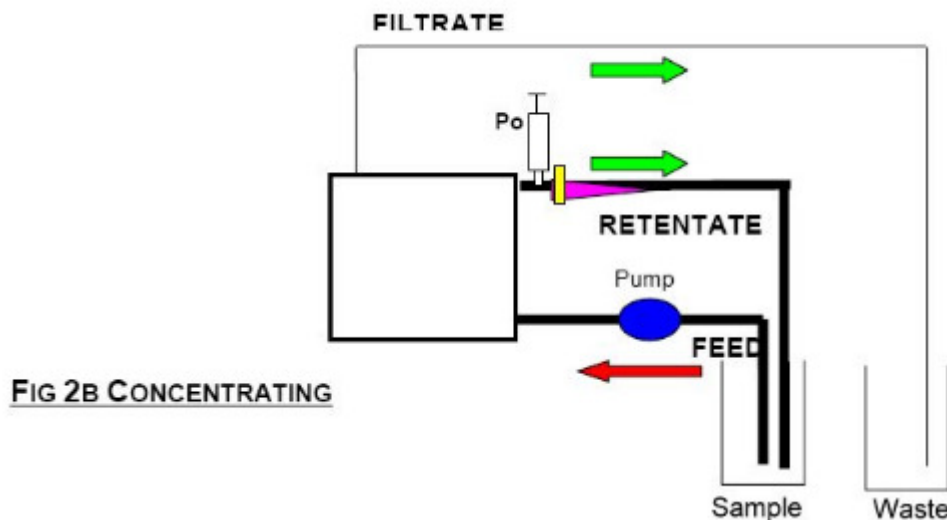
Step 8.

Rinse cartridge with about 250 mL of sample

Concentrate

Step 9.

Put the retentate line into the sample bottle (Fig. 2B - Recirculation mode). Record sample volume and start pump increasing to maximum speed.



Concentrate

Step 10.

Clamp the retentate line to increase filtrate flow so that Manometer gets up to 2.5 bars.

Concentrate

Step 11.

Concentrate sample until about 250 mL remains (6 L takes about one hour)

⌚ DURATION

01:00:00

Concentrate

Step 12.

Transfer sample to a smaller bottle (250 mL) then finally to a 50 mL tube with conical bottom

⌚ DURATION

00:10:00

Concentrate

Step 13.

Continue to concentrate very carefully, lowering the pump speed until the sample volume is reduced to 15-20 mL

📌 NOTES

Daniel Vaultot 08 Dec 2016

It is very important to lower the pump speed in order to avoid losing the sample.

Recirculation

Step 14.

When final volume is about 10 mL, clamp filtrate tube and recirculate slowly (no change of volume should take place)

Recirculation

Step 15.

Leaving the filtrate tube clamped, get the feed line out of the sample in order to get back the total volume of concentrated sample

Recirculation

Step 16.

Take sample for flow cytometry count (to compare with original sample concentration and estimate concentration factor)

Recirculation

Step 17.

Store concentrated sample for later use (e.g. flow cytometry sorting, culture etc...)

Rinsing

Step 18.

Go back to Fig. 1 configuration (Open circuit)

Rinsing

Step 19.

Rinse 1 min with filtered sea water



REAGENTS

✓ Filtered Seawater (0.2 μ m) by Contributed by users



DURATION

00:01:00

Rinsing

Step 20.

Rinse 1 min with distilled water



REAGENTS

✓ Distilled Water by Contributed by users



DURATION

00:01:00

Rinsing

Step 21.

Rinse with 50 mL NaOH 0.1 M



AMOUNT

50 ml Additional info:



REAGENTS

✓ 0.1 M NaOH by Contributed by users

Rinsing

Step 22.

Put all three tubes (feed, retentate, filtrate) in bottle containing NaOH 0.1 M



REAGENTS

✓ 0.1 M NaOH by Contributed by users

Rinsing

Step 23.

Recirculate for 20 min (to get rid of everything on the cartridge filter)



DURATION

00:20:00

Rinsing

Step 24.

Rinse with 250 mL of MilliQ water (Fig. 1)



AMOUNT

250 μ l Additional info:



REAGENTS

✓ MilliQ water by Contributed by users

Storage

Step 25.

Stop the pump and clamp all three tubes

Storage**Step 26.**

Store at 4°C. For a storage beyond 1 day, store with Ethanol 10%

**REAGENTS**

- ✓ Ethanol 10% by Contributed by users