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## Luminex Bead Conjugation

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Working

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### ABSTRACT

Protocol for amine-coupling of proteins to magnetic Luminex beads

Quantity as written:  $1 \times 10^6$  beads under typical yields (sufficient for 2000 wells assuming 500 beads/well)

### Needs a good qualification section!

### STEPS MATERIALS

NAME	CATALOG #	VENDOR
EDC (no-weigh formulation)	A35391	Thermo Scientific
sulfo-NHS (no-weigh format)	A39269	Thermo Scientific
PBS-TBN	P0210	Teknova
PBS-TBN	P0210	Teknova

### Activation

1 Resuspend beads by briefly vortexing; dispense **100  $\mu$ l** (contains approx. 1.25 million beads)

2 BEAD SEPARATION BLOCK

2.1 Collect volume by pulse centrifugation **100 x g** **00:00:03**

2.2 Place on magnetic tube rack, shielded from light **00:01:00**




#### Protocol tip

Partially open the microcentrifuge tube before placing in the rack. This allows them to be opened on-rack with minimal disruption of the bead pellet that's held in place only by the magnet.

2.3 Remove  **90 µl** of supernatant (while still on-rack) using a pipette




n.b.

This leaves a  **10 µl** excess that allows the removal of supernatant at all subsequent steps to use the same volume that was added immediately prior.

### 3 WASH BLOCK









3.1 Resuspend in  **100 µl** 1x PBS; vortex briefly to mix

3.2 Collect volume by pulse centrifugation  **100 x g**  **00:00:03**

3.3 Place on magnetic tube rack, shielded from light  **00:01:00**

3.4 Remove supernatant using a pipette

### 4 Resuspend beads in a solution containing:

-  **100 Milimolar (mM)** sodium phosphate (pH 6.2)  **80 µl**
-  **12.5 mg/mL** (approx.  **65 Milimolar (mM)**) in sodium phosphate (pH 6.2)  **5 µl**
-  **12.5 mg/mL** (approx.  **58 Milimolar (mM)**) sulfo-NHS in phosphate (pH 6.2)  **5 µl**



**EDC (no-weigh formulation)**

by Thermo Scientific

Catalog #: A35391



**sulfo-NHS (no-weigh format)**

by Thermo Scientific

Catalog #: A39269

5 Mix tubes end-over-end, shielded from light 🌡️ Room temperature ⌚ 00:20:00

#### Coupling

6 [go to step #2](#) (separation block) for removal of supernatant

7 Resuspend beads in [25 µg/mL](#) protein of interest [250 µl](#), made in 1x PBS

8 Mix tubes end-over-end, shielded from light 🌡️ Room temperature ⌚ 02:00:00

#### Blocking + Washes

9 [go to step #3](#) (wash block) for a **total of 3** washes using [400 µl](#) 1x PBS-TBN



n.b.

DO NOT use P1000 for removing supernatant as the wider tip and faster flow tends to lead to greater bead loss. Instead, use a P200 twice.



PBS-TBN

by Teknova

Catalog #: P0210

10 Perform final resuspension in [250 µl](#) 1x PBS-TBN



PBS-TBN

by Teknova

Catalog #: P0210

11 Will the beads be used immediately?

#### Counting

Dilute beads by 1:10 and 1:100 in 1x PBS

12

13 Measure concentration of each dilution by cell counter



Typical yields are approx.  $1 \times 10^6$  beads, or  $4 \times 10^6$  beads/mL



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