



Bradford protein assay in 96-Well plate

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ARSTRACT

Assay for quantification of protein by comparing measured Absorbance at 595 nm to bovine serum albumine standard

- Bovine serum albumin (concentrated >200 μg/ml)
- Roti-Quant 5x (ROTH)
- 96 Well Plate
- Plate Reader

Standard measurements

Dilute your BSA stock solution to 200µg/ml in the same buffer used for your solution of interest.

Dilute the Roti-Quant 5x reagent at a rate of 2:7,5. For 15 ml solution, this means 4 ml Roti-Quant added to 11 ml of Water.

Create 200 µl each of standard concentrations according to the following table:

Concentration [µg/ml]	Volume of stock needed	Volume of solution buffer needed
0	0	200
20	20	180
40	40	160
60	60	140
80	80	120
100	100	100

Work in Triplicates. Pipette 50µl of the standards into the 96-Well plate

Add 200 µl of the diluted Bradford reagent to each well.

Incubate for 5 minutes at room temperature

Measure the OD₅₉₅ for each well. Create a regression line from the averages of each concentrations' measurements

Protein concentration measurements

Dilute your solution of interest 1:20 or 1:40, depending on expected protein levels.

Work in triplicates and use blanks. Pipette 50 µl of each dilution into a well on the 96-Well plate.

When working with solutions containing pigments, prepare additional wells to which NO Bradford reagent will be added

Add 200 µl of the diluted Bradford reagent to each well. For the pigment controls, add 200 µl of solution buffer instead. Optimally, use a multi-pipette to reduce the time needed for adding the reagent.

Incubate for 5 minutes at room temperature

Measure the OD₅₉₅ for each well.

When processing the data, correct the measurements for the blanks. When pigments are present in the solution, correct the OD₅₉₅ by subtracting the ${\rm OD}_{595}$ of the pigment controls from the appropriate measurements.

Determine the amount of protein by putting the achieved OD₅₉₅ of your solution of interest into the regression formula and multiplying by the dilution factor.

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