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Top2 Chromatin Accessibility by Etoposide Cross-linking

[Jacob Kirkland¹](#)¹Stanford University

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Works for me

[dx.doi.org/10.17504/protocols.io.2rngd5e](https://doi.org/10.17504/protocols.io.2rngd5e)

Jacob G. Kirkland
Stanford University



ABSTRACT

The best way to determine TOP2 specific accessibility to chromatin is via etoposide treatment, which leads to a covalent cross-link between TOP2 proteins and DNA, that can only occur where TOP2 proteins have access to DNA. TOP2 cross-linking to DNA has previously been shown to require DNA accessibility and chromatin regulators that alter DNA accessibility (Dykhuisen and Hargreaves et al. Nature 2013, Miller et al. NSMB 2017).

GUIDELINES

Buffer A

- 25mM HEPES pH 7.0
- 25mM KCl
- 0.05mM EDTA
- 5mM MgCl₂
- 10% Glycerol
- 0.1% NP-40

High Salt RIP

- 50mM TRIS pH 7.8
- 500mM NaCl
- 1% NP-40
- 0.1% DOC
- 0.1% SDS

1% SDS RIP

- 50mM TRIS pH 7.8
- 150mM NaCl
- 1% NP-40
- 0.1% DOC
- 1% SDS

Protease Inhibitors (1000x stock in DMSO all from Calbiochem)

- Chymostatin #230790 (10mg/ml)
- Leupeptin, Hemisulfate #108975 (10mg/ml)
- Pepstatin A, Synthetic #516481 (10mg/ml)

15m

- 1 Cell Culture:
Trypsinize and count cells
Transfer 0.75e6-1e6 cells to (4) different eppendorfs
Save (1) eppendorf for your input sample for step 4
Bring volumes to 1ml with media (If volume is already more than 1ml you can transfer to a 15ml conical and bring volume up to known amount where all cell lines are at the same volume)

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|---|--|-----|
| 2 | Etoposide Cross-linking
Add Etoposide (1:1000; final concentration of 100uM) for 0, 15 and 60 minutes
Add to 60' sample first, and then 15' sample 45' later
Rotate at RT for 60' | 1h |
| 3 | Washes
Spin at 500g for 4 minutes at 4°C
Wash with PBS
Spin at 500g for 4 minutes at 4°C
Remove supernatant | 10m |
| 4 | Buffer A -- Making Nuclei
For all samples + Input
Resuspend in 1 ml Buffer A + Protease Inhibitors (1:1000) + DTT (1:1000; final concentration:)
Incubate on ice for 10'
Spin at 300g for 4 minutes
Aspirate out supernatant
Spin briefly and remove remaining supernatant | 15m |
| 5 | High salt RIPA extraction
Resuspend nuclei in 50-100ul High salt RIPA with 500mM NaCl
Incubate on ice for 20'
Spin 12,000g for 5' at 4°C
Remove supernatant (can save if you want)
Spin briefly and remove remaining supernatant | 25m |
| 6 | Solubilize chromatin pellet
Resuspend in 50ul 1% SDS RIPA + PIs + DTT + Benzonase (1:200)
Be sure to fully resuspend pellet by pipetting up and down ~20 times
Incubate at room temp for 10'
Incubate on ice for 20'
Spin 12,000g for 3 minutes and transfer supernatant to a new tube (there should not be much if any of a visible chromatin pellet after spinning. If there is then try resuspending in a larger volume) | 33m |
| 7 | Proceed to Western Blot adding equal volumes of each sample | |



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