

# Colony mix assays

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## Abstract

Protocol to co-culture individual colonies of different strains (e.g. transformed with different vectors) on a LB agar plate in a distributed and random manner.

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## Materials

- ✓ 14 ml round bottom culture tubes by Contributed by users
- ✓ LB agar plates with the proper antibiotic(s) (eg. Kanamycin) by Contributed by users
- ✓ Glass beads (~ 5mm diameter) or L shaped spreader by Contributed by users
- ✓ Sterile conditions (e.g. laminar flow or a flame) by Contributed by users

## Protocol

### Step 1.

Grow TOP10 E.coli cells containing the different vectors (e.g. three bacterial cultures: the first containing CyOFP plasmid, the second containing mBeRFP and the third containing sfGFP). Cells should be grown overnight at 37 °C with agitation in a 14 ml round bottom culture tube with 1-5 ml of LB medium including the proper antibiotics.

#### NOTES

**Tamara Matute** 08 Sep 2017

If you want to work with a single culture go to step 3.

### Step 2.

Mix all cultures proportionally (eg, 100 µl of each one).

#### NOTES

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If you want to obtain more or less colonies of one of the strains (e.g. mBeRFP) you can simply tune the mixture proportion accordingly.

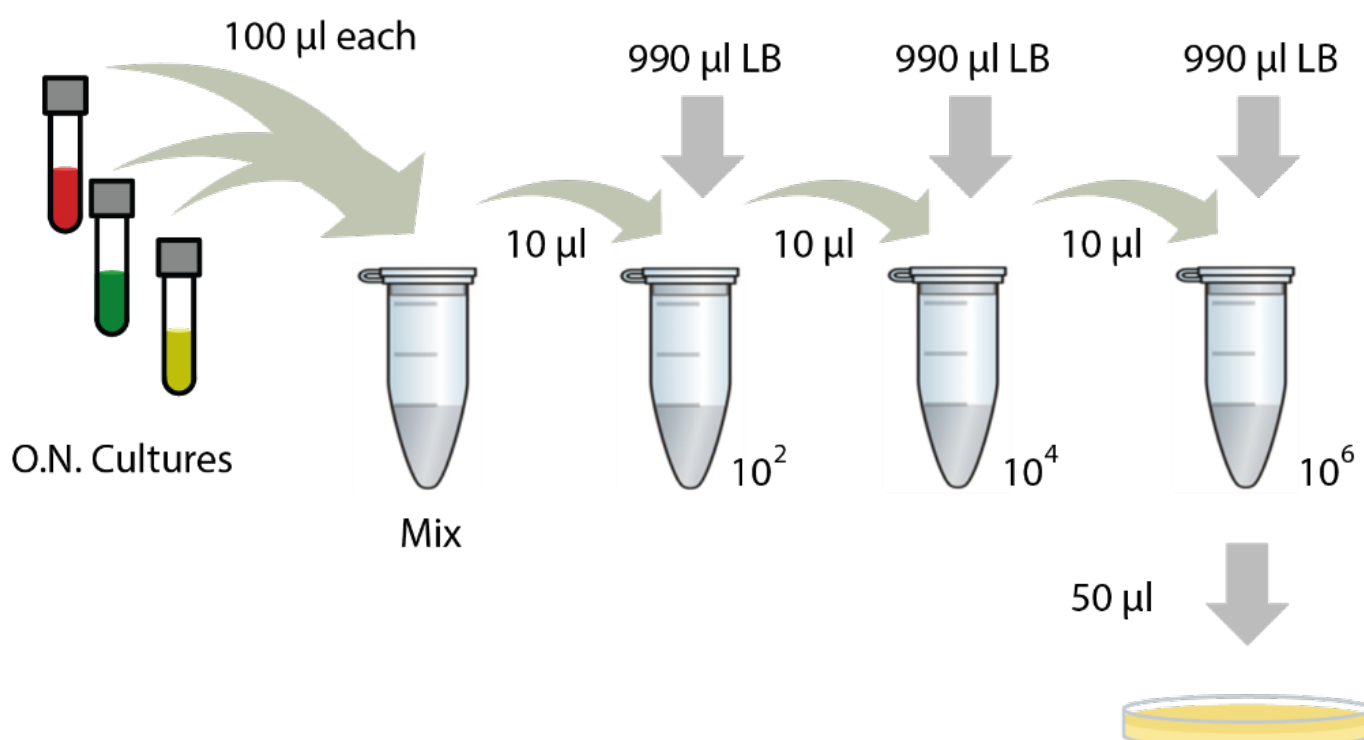
### Step 3.

Three consecutive dilutions of the culture (pure or mixture) have to be made:

(1)  $10^2$  dilution  $\rightarrow$  1:100 (culture:LB)

(2)  $10^4$  dilution  $\rightarrow$  1:100 ((1):LB)

(3)  $10^6$  dilution  $\rightarrow$  1:100 ((2):LB)



### Step 4.

Spread 50  $\mu$ l of the dilution (3), performed in the previous step, on an LB agar plate with the proper antibiotic(s) (e.g. Kanamycin). To obtain a homogeneous distribution of colonies on the plate it is recommended to use glass beads to spread the dilution over it. However the above, it is possible to use other methods to this aim (e.g. to use an L shaped spreader employing a good technique).

#### 📌 NOTES

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It is important to seal the plates with parafilm to avoid dehydration across the experiment or if is desired to preserve the plate for a long time (e.g. weeks). The above does not apply if you want to do timelapse of the plate, in which case it is advisable not to seal it to reduce lid condensation that ruins the images.

### Step 5.

Incubate the plate at 37°C for the first 8 hours to accelerate the colony emergence (this is an optional step).

### Step 6.

Incubate at the desired temperature (e.g. room temperature or 37°C) for a period of 1 to 4 days.

#### NOTES

**Tamara Matute** 08 Sep 2017

To accelerate colony growth, it is recommended to incubate the plates at 37 ° C (1-2 days for maximum size). Plates can be left at room temperature if necessary, but growth will take approximately twice as long.