

DNA Extraction Procedure Using SDS

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Abstract

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Materials

- Ethyl alcohol, Pure 200 proof, for molecular biology [E7023](#) by [Sigma Aldrich](#)
- ✓ Liquid Nitrogen by Contributed by users
- 2-Mercaptoethanol [View](#) by [Sigma Aldrich](#)
- Buffer EB [19086](#) by [Qiagen](#)
- Chloroform:Isoamyl alcohol 24:1 [C0549](#) by [Sigma Aldrich](#)
- 2 x 0.5ml LongLife™ Proteinase K [5mg/ml] [786-038](#) by [G-Biosciences](#)
- 2 x 0.5ml LongLife™ RNase [10U/?l] [786-040](#) by [G-Biosciences](#)
- 1 Liter STE Buffer [10X] (100mM Tris.HCl (pH8.0), 10mM EDTA, 1M NaCl) [786-569](#) by [G-Biosciences](#)
- 100g SDS (Sodium dodecyl sulfate) [DG092](#) by [G-Biosciences](#)
- 1kg Tris Base [RC-106](#) by [G-Biosciences](#)
- ✓ isopropyl alcohol by Contributed by users

Protocol

Cut tissue and grind by liquid nitrogen.

Step 1.

Transfer grinded tissue to 15 ml tube, Add 6ml TEN, 700ul 20% SDS (56°C preheat), 150ul beta mercaptoethanol and 150 ul proteinase K, rapidly mix.

Step 2.

Incubate homogenate for 2 h at 56 °C , gently blending for every 5 ~ 10 min.

Step 3.

Cool down to room temperature then add equal volume of Tris saturated phenol and mix, centrifuge at room temperature (16 °C) with 14000 RPM for 10 min, then save supernatant.

Step 4.

Add chloroform and isoamyl alcohol (24:1) to supernatant, then mix, centrifuge with 14000 RPM for 10 min, save supernatant.

Step 5.

Resuspend nuclei pellet with 0.8 X volume of frozen isopropyl alcohol, wash twice with cold ethanol

75%, then dry.

Step 6.

Add 200 ul TEN and 2 ul RNase (100 mg/ml), 37 °C for 30 min, incubation period precipitation with Tip dolly, precipitate dissolve completely.

Step 7.

Add buffer (TEN and 200 ul 20% SDS) and 40 ul Protease K up to 2 ml, then incubate for 30 min at 56 °C, repeat steps 5 and 6.

Step 8.

Add 300 ul EB (pH8.0) to dissolve.

Step 9.