AMPure purification protocol

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Abstract

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Guidelines

The protocol below works for 200 µl PCR tubes or plates. If working with 1.5 ml tubes, use larger volumes of EtOH for the washes (step 6 and 7) and Tris/water for the final elution (step 9).

Protocol

Step 1.
Remove beads from the fridge, equilibrate to the room temperature, mix thoroughly but do not vortex.

Step 2.
Add desired ratio of AMPure beads to the purified sample and mix well by pipetting.

Step 3.
Incubate 5 minutes.

Step 4.
Place on the magnetic rack.

Step 5.
Let it stand for 5 minutes on the rack, aspirate and discard supernatant.

Step 6.
Add 200 µl of freshly prepared 70% EtOH, incubate 30 seconds, aspirate and discard EtOH.

Step 7.
Repeat the wash: add 200 µl of freshly prepared 70% EtOH, incubate 30 seconds, aspirate and discard EtOH.

Step 8.
Wait until the pellet dries out completely. DO NOT remove the plate/tubes from the magnetic rack, as the dried AMPure might pop out from the tubes!

Step 9.

Add desired volume of Tris 10 mM or water, wetting the dried AMPure pellets (add 1 µl to the final volume to avoid pipetting out the beads)

Step 10.
Remove from the magnetic rack.

Step 11.
Resuspend by pipetting or vortexing.

Step 12.
Incubate 10 minutes, incubating in 37°C can improve DNA yield.

Step 13.
Place on the magnetic rack.

Step 14.
Let it stand for 5 minutes, pippete out and save supernatant. The eluted DNA is in the supernatant, do not discard it!