# LB Media

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ABSTRACT

LB (Luria-Bertani) media is commonly used for bacterial culture. We use it for overnight cultures after transformations.

THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

Taken from Benchling Protocol

MATERIALS

| NAME | CATALOG # | VENDOR |
|---|---|---|
| Yeast Extract | Y1625 | |
| Sodium Chloride | PubChem CID: 5234 | |
| Tryptone | BP1421-500 | Fisher Scientific |

MATERIALS TEXT

- 1 Liter Glass Bottle
- Autoclave
- Stirring plate and magnetic stirrer

## Creating the Broth

2m

1 Put a stir bar into a 1L glass bottle and fill the glass bottle with 1L of DI water (using a graduated cylinder). Mark the level of the water with a thin Sharpie or label tape. Pour out ~50mL of the water.

We do this step because QSing the media in a graduated cylinder gets messy - it's often difficult to transfer the solute back and forth.

10m

2 In a large glass bottle (at least 1L), add the following:

| *Component* | *Amount* |
|---|---|
| Deoinized Water | 950 ml |
| Tryptone | 10 g |
| Yeast Extract | 5 g |

NaCl 10 g

(You do not have to add the exact amount of water, just make sure it is close to 950mL).

3 Shake or stir to dissolve all the solutes. 20m

4 Optional: We have never done this step, but it is recommended by the Sambrook Molecular Cloning manual. 10m

Adjust the pH of the solution to 7.0 using 5M NaOH (~0.2mL).

5 QS the solution to the mark you made in Step 1 (1L) using DI water. 1m

## Sterlizing the Broth 1h

6 Autoclave bottle follwing Autoclave protocol on the liquid cycle. Choose the appropriate cycle for the amount of liquid you have. Make sure the bottle cap is on loosely

7 Let bottle cool on the lab bench with loose cap 30m

8 Keep bottle in the cold room for storage, with the cap tightly closed. Make sure you keep the bottle STERILE. We want a clear solution and nothing growing in our broth