DNA Precipitation Protocol

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Abstract

Citation: Matthew Sullivan DNA Precipitation Protocol. **protocols.io**
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Guidelines

This protocol comes from a group of other protocols.This protocol is (3) of (4):
1. 'Large Volume Marine Cyanophage Phage Protocols'
2. 'DNA Extraction Protocol'
3. 'DNA Precipitation Protocol'
4. 'Checking DNA Concentration with Agarose Gel'

Needed:

- Ethanol
- Ice
- Microfuge
- 70% ethanol made from 100% with autoclaved milli-Q-water)
- Centrifuge
- TE Buffer

Protocol

Step 1.
Add 2 volumes of ice cold ethanol, mix well.

Step 2.
Let sit on ice 15-30 minutes

DURATION
00:15:00

NOTES
VERVE Team 21 Jan 2016
Let stand longer if expecting significantly low DNA yields (<µg). (I leave overnight at -20°C)

Step 3.
Microfuge at max speed, 20 minutes.

DURATION
00:20:00

Step 4.

Carefully remove supernatant and fill tube halfway with 70% ethanol

NOTES

VERVE Team 24 Jun 2015

The 70% ethanol is made from 100% with autoclaved milli-Q-water.

Step 5.

Spin at max speed for 2 minutes.

DURATION

00:02:00

Step 6.

Repeat the above step one time (2nd 70% wash).

Step 7.

Remove as much ethanol as possible without disturbing the pellet

NOTES

VERVE Team 17 Jun 2015

Good idea to hold onto the supernatant until confirmed DNA.

Step 8.

Leave tube open on bench 15 minutes

DURATION

00:15:00

NOTES

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This lets the ethanol disperse.

Step 9.

Dissolve in TE buffer (pH 7.6).