

# Preparation of chemical competent E. coli cells (w/ calcium chloride)

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## Abstract


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## Materials

✓ Glycerol G5516 by Contributed by users

 Calcium Chloride Dihydrate c79 by Fisher Scientific

 Manganese(II) chloride tetrahydrate M3634 by Sigma Aldrich

✓ 1.5 mL Eppendorf tubes by Contributed by users

 Magnesium chloride hexahydrate view by Sigma Aldrich

## Protocol

### Preparation of buffer CCMB 80

#### Step 1.

- 10 mL of 1M K-Acetate stock
  - 11.8 g  $\text{CaCl}_2 \cdot \text{H}_2\text{O}$
  - 2 g  $\text{MgCl}_2 \cdot 6 \text{H}_2\text{O}$
  - 4 g  $\text{MnCl}_2 \cdot 4 \text{H}_2\text{O}$
  - 100 mL glycerol
  - adjust pH to 6.4 w/ 0.1M HCl
  - add ddH<sub>2</sub>O to 1 L
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- filter buffer through syringe filter (e.g. Acrodisc Syringe Filter, 0.2  $\mu\text{M}$  Supor Membrane, Pall Life

Science)


--> work in the Clean Bench

#### AMOUNT

1000 ml Additional info:

Buffer CCMB 80

#### SAFETY INFORMATION

**HCl is corrosive! Wear goggles, gloves and don't inhale! Work in the hood!** 

### Autoclave material

#### Step 2.

- 255 mL LB medium
  - >100 x 1.5 mL Eppendorf tubes (e.g. in a big flask or beaker)
  - 1 mL Erlenmeyer flask (closed w/ aluminium foil)
  - 1 x > 10 mL test tube (if not using disposables)
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- store sterile Eppendorf tubes at -20°C

### Overnight (o/n) culture

#### Step 3.

- Inoculate 5 mL of LB medium w/ seed cells (e.g. DH5α) in a test tube
- Incubate at 37 °C and >200 rpm

### Overday (o/d) culture

#### Step 4.

- Inoculate 250 mL fresh LB medium w/ 5 mL o/n culture (1:50) in 1L Erlenmeyer flask
- Incubate at 37 °C and >200 rpm until OD<sub>600</sub> 0.3

#### NOTES

Work in the Clean Bench!

### CaCl<sub>2</sub> treatment

#### Step 5.

- Cool down centrifuge including inlets.
- transfer o/d cultures to five sterile 50 mL tubes (Falcon)
- Centrifuge at 3000 \*rcf and 4 °C for 10 min
- gently resuspend each pellet in 16 mL **ice cold** CCMB
- incubate on ice for 20 min
- Centrifuge o/d culture at 3000 \*rcf and 4 °C for 10 min
- gently resuspend each pellet in 2 mL **ice cold** CCMB

- incubate on ice for 20 min

 [TEMPERATURE](#)

4 °C Additional info: keep  
cells on ice

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### Prepare aliquots

#### **Step 6.**

- transfer each 100 µL of cell suspension in 1.5 mL Eppendorf tube
- store at -80°C

 [TEMPERATURE](#)

4 °C Additional info: keep  
tubes on ice

#### [NOTES](#)

**Option:** Quick-freeze aliquots instantly in liquid nitrogen

This yields slightly more than 100 1,5 ml Eppendorf tubes. Tip: mark the tubes beforehand. This makes the tubes easier to find in the ice when transforming the cells afterwards. To reduce workload, mark the whole freezer box with the type of competent cells and only make a line or dot on the Eppendorf tubes, preferably with a thick marker.

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