

# untitled protocol

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#### **Abstract**

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#### **Protocol**

## Step 1.

Incubate E.coli cells 1:100 in 10 ml LB medium with antibiotics.

## Step 2.

OD600=0.4 , collect all cells at 4  $^{\circ}$ C, 8000 rpm for 10 min.

## Step 3.

Wash with 1 ml 1\*PBS one time.

#### Step 4.

Resuspend in 500 ul 1 \* PBS and sonicate.

#### Step 5.

Spin at 10000 rpm for 15 min.

## Step 6.

Transfer supernatant into a new 1.5 ml tube and sit on ice.

#### Step 7.

Load into 120% SDS-PAGE gel, run at 200v for 45 min.

## Step 8.

Get the PVDF membrane and filter paperPVDF: Put in methanol (5s) then in transfer bufferFilter: transfer bufferGel: transfer buffer

#### Step 9.

Sit on Room Temperature for 20 min.

## Step 10.

Transfer membrane at 25 V for 30 min.

#### Step 11.

Blook woth 5% skim milk RT for 1 hour.

# Step 12.

1:1000 Ab1 DnaA Ab.

# Step 13.

4 °C over night rotation.

# Step 14.

Wash the membrane with 1\* PBST for 3 times (15 min, 10 min, 5 min).

#### Step 15.

Dillute AB2 1:10000 in 50 ml PBST.

# Step 16.

Incubate with Ab2 for 1 h.

# Step 17.

Wash the membrane with 1\* PBST for 3 times (15 min, 10 min, 5 min).

# Step 18.

Mix solution 1 and 2 (1:1).

#### Step 19.

Incubate the membrane with Mix solution for 5 s.

#### Step 20.

Develop the membrane with ChemiDoc MP Imaging system (BIO-RAD).