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Working

## U Michigan - Retinal Vascular Permeability [↗](#)

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[dx.doi.org/10.17504/protocols.io.yagfsbw](https://doi.org/10.17504/protocols.io.yagfsbw)

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### ABSTRACT

#### Summary:

The vascular permeability is quantified by measuring albumin leakage from blood vessels into the retina. Fluorescent dye (FITC-BSA) is used to measure the breakdown in blood-retinal barrier to be detected when increased vessel leakage is extravasated into the interstitial space.

### EXTERNAL LINK

<https://mmpc.org/shared/document.aspx?id=309&docType=Protocol>

### MATERIALS

NAME	CATALOG #	VENDOR	CAS NUMBER  RRID
Ketamine	0409-2051-05	Hospira(Pfizer)	
Xylazine	510004	VetOne	
Heparinized micro-cuvette	Microvette 300 LH 20.1309.100	Sarstedt	
0.3 c.c. insulin syringe (31-gauge x 5/16")	328440	BD Biosciences	
1 c.c. syringe with ½" 27-gauge needle	309623	BD Biosciences	
FITC-BSA (albumin-fluorescein isothiocyanate conjugate bovine)	A9771	Sigma-aldrich	
EZ Clip wound closure	EZC KIT	Braintree Scientific	
Blunt end needle (22-gauge x ½")	Kahnetics KDS2212P	Weller	
Saline	2B1324X	Baxter	
Triton-PBS (1%)	X100	Sigma Aldrich	
Microcentrifuge tubes	C2170	Denville Scientific Inc.	
384-well microplate	Bio-One 781096	greiner bio-one	

### MATERIALS TEXT

#### Reagent Preparation:

##### Reagent 1:

Ketamine/xylazine (90 and 10 mg/ml)

Procedure: Add 0.5 ml xylazine (100 mg/ml) to 4.5 ml ketamine (100 mg/ml)

##### Reagent 2:

FITC-BSA (100 mg/ml)

Procedure: Dissolve 1 g FITC-BSA in 10 ml sterile PBS, aliquots stored at -80°C and warmed to 37°C before use

**Reagent 3:**

Triton-PBS (1%)

Procedure: Dissolve 1 ml Triton X-100 in 100 ml PBS

**Note:**

Hospira, [RRID:SCR\\_003985](#)

BD Biosciences, [RRID:SCR\\_013311](#)

Baxter [RRID:SCR\\_003974](#)

Sigma-Aldrich, [RRID:SCR\\_008988](#)

- 1 Weigh animal and record body weight for anesthetic and dye injections
- 2 Anesthetize animal with ketamine/xylazine mixture
- 3 Make an incision on skin inside of the hind leg and carefully tear away the membranes to isolate the femoral vein
- 4 Inject FITC-BSA into the femoral vein at 2 µl/g body weight (equal to 200 mg/kg body weight) using a 31-gauge 0.3 c.c. insulin syringe (vortex FITC-BSA before use)
- 5 Apply pressure on the injection site with a sterile gauze or cotton swab to stop bleeding
- 6 Staple the incision site and allow FITC-BSA to circulate for 2 hours
- 7 Anesthetize the animal again with ketamine/xylazine
- 8 Open the abdomen and draw 0.3 ml blood from the vena cava with a 27-gauge 1 c.c. syringe
- 9 Remove needle and expel blood into a heparinized micro-cuvette
- 10 Mix blood sample by gently reversing the tube several times and keep on ice
- 11 Open the chest cavity, cannulate the heart with a blunt end 22-gauge needle into the left ventricle and incise right atrium to release pressure
- 12 Perfuse with saline (warmed to 37°C) at 20 ml/min via the left ventricle for 2 minutes

- 13 Harvest the retina and place in a pre-weighed microcentrifuge tube. Rinse the harvest tools between samples to avoid cross contamination
- 14 Centrifuge the blood sample at 2,000 x g at 20°C for 15 minutes to separate plasma
- 15 Transfer the plasma to a new microcentrifuge tube and store at -80°C
- 16 Dry the retina samples with a Speed-Vac overnight
- 17 Weigh the microcentrifuge tube containing the dry retina to obtain the dry retina weight
- 18 Add 100 µl of 1% Triton-PBS to each retina and shake overnight to extract FITC-BSA
- 19 Vortex briefly, centrifuge the microcentrifuge tubes at 17,000 x g for 30 minutes and transfer supernatant to a new microcentrifuge tube
- 20 Dilute the plasma samples with 1% Triton-PBS
- 21 Dilute the stock FITC-BSA (100 mg/ml) with 1% Triton-PBS and make serial dilutions to obtain the FITC-BSA standards
- 22 Measure the standards (0.156 to 10 µg/ml), retina extract and diluted (1:900) plasma samples (20 µl/well, triplicate) in a 384-well black/clear bottom plate with a fluorescent plate reader (excitation 488 nm, emission 520 nm)
- 23 Calculate the FITC-BSA concentration of each sample with the standard curve

$$\text{Permeability } (\mu\text{l/g/h}) = \frac{(\text{Retina FITC-BSA } (\mu\text{g}) - \text{Autofluorescence}) / \text{Retina Weight (g)}}{\text{Plasma FITC-BSA Concentration } (\mu\text{g}/\mu\text{l}) \times \text{Circulation Time (h)}}$$

**NOTE:** The auto-fluorescence background of the retina from animal without FITC-BSA injection should be subtracted for the permeability calculation. Additional tip that improves efficiency.



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