



Feb 27, 2019

Working

Characterization of iPSC

In 1 collection

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Neurodegeneration Method Development Community

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Comprehensive Genomic
Editing and Screening
Protocol Updated
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PROTOCOL STATUS

Working

We use this protocol in our group and it is working

GUIDELINES

This protocol is part of the [Screening Edited iPSC Clones collection](#).

SAFETY WARNINGS

Please refer to the SDS (Safety Data Sheet) for information about hazards, and to obtain advice on safety precautions.

BEFORE STARTING

Starting Material: 1 confluent 6 well plate.

Use 2 wells to characterize the iPSC and 4 wells for freezing the iPSC.

1 Coat T25 flask, chamber slide and plates with Matrigel ⌚ 01:00:00 prior to passaging.

2 Aspirate media from cell culture.

3 Wash with 🧴 1 ml PBS per well and aspirate.

4 Add 🧴 1 ml Accutase per well of 6 well plate.

5 Incubate in 🌡 37 °C for 3-4 minutes. ⌚ 00:03:00

6 Collect cells from 2 wells with  3 ml DMEM/F12 per well and transfer into 15mL conical A.

7 Collect cells from 4 wells with  3 ml DMEM/F12 per well and transfer into 15mL conical B.




8 Spin cells at 750 rpm for  00:03:00 .




9 Aspirate media from each tube.




Tube A

10 To 15mL conical tube A, add  2 ml mTesR1 and distribute cells.



11 Karyotype: Add  2 ml of mTesR1 to T25 flask, then add  500 µl cells .

12 gDNA pellet:  500 µl cells in 1.7 mL tube, spin down at max speed for  00:00:15 , aspirate media, store in  -80 °C .

13 RNA pellet:  900 µl cells in 1.7 mL tube, spin down at max speed for  00:00:15 , aspirate media, store in  -80 °C

14 ICC: Dilute  100 µl cells in  750 µl mTesR1. Plate  200 µl cells per well in 4 wells of a chamber slide.

Tube B

15 To 15mL conical tube B, add  4 ml mTesR1 and  4 ml of 2x Freezing Media (20% DMSO, FBS).

16 Add  1 ml cell suspension to each vial (1 well will freeze down into approximately 2 vials).



Cryovials need to be labeled with the following before freezing down:

- Cell Type
- Line Name
- Passage #
- Date
- Your Name



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