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Th1 Polarization of Mouse CD4⁺ Cells V.3 [↗](#)Sam Li¹¹BioLegend

1

Works for me

[dx.doi.org/10.17504/protocols.io.795hr86](https://doi.org/10.17504/protocols.io.795hr86)

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EXTERNAL LINK

<https://www.biolegend.com/protocols/th1-polarization-of-mouse-cd4--cells-protocol/4242/>

MATERIALS

NAME	CATALOG #	VENDOR
Phorbol 12-myristate 13-acetate (PMA)	P8139	Sigma Aldrich
Ionomycin calcium salt from Streptomyces conglobatus	I0634	Sigma Aldrich
MojoSort™ Mouse CD4 T Cell Isolation Kit	480005, 480006, 480033	BioLegend
RBC Lysis Buffer	420301	BioLegend
Anti-mouse CD3ε clone 145-2C11 (Ultra-LEAF™ format)	100339	BioLegend
Anti-mouse CD28 clone 37.51 (Ultra-LEAF™ format)	102116	BioLegend
Anti-mouse IL-4 clone 11B11 (Ultra-LEAF™ format)	504122	BioLegend
Monensin Solution	420701	BioLegend
Recombinant mouse IL-2 (carrier-free)	575402	BioLegend
Recombinant mouse IL-12 (p70) (carrier-free)	577002	BioLegend



MATERIALS TEXT

- Sterile PBS
- Cell culture medium (RPMI 1640 supplemented with 10% FBS)
- Sterile 12-well plate
- Sterile 6-well plate

Isolation of CD4⁺ Cells From Lymph Nodes

- 1 Harvest lymph nodes (superficial cervical, mandibular, axillary, inguinal, and mesenteric) from mice.
- 2 Tease lymph nodes through a sterile 70-μm nylon cell strainer to obtain single-cell suspensions incomplete RPMI containing 10% FCS (complete medium).
- 3 Resuspend cells in complete medium and use your favorite method to isolate CD4⁺ cells. Consider using our Mojosort™ Mouse CD4 T Cell Isolation Kit.

Th1 Polarization of CD4+ Cells

- 4 On day 0, coat 12-well plate with anti-mouse CD3 ϵ , clone 145-2C11 (3 μ g/ml). Incubate at 37°C for 2 hours or 4°C overnight. Aseptically decant antibody solution from the plate. Wash plate 3 times with sterile PBS. Discard liquid.
 02:00:00
- 5 Plate CD4⁺ cells at 1.0×10^6 /1ml/well. Culture cells for 5 days at 37°C, 5% CO₂, in the presence of anti-mouse CD28, clone 37.51 (3 μ g/mL), anti-mouse IL-4, clone 11B11 (10 μ g/mL), recombinant mouse IL-2 (5 ng/mL), and recombinant mouse IL-12 (10 ng/ml).
- 6 On day 3, if media is yellow, add 2 ml/well of fresh media.
- 7 On day 5, wash cells once and then restimulate in complete media with 50 ng/ml PMA, 1 μ g/ml ionomycin and 10 μ l monensin (1000x), in a 6-well plate in incubator at 37°C for 5 hours.
 05:00:00
- 8 After harvesting, the cells are ready for staining.



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