

# **Rat ACTB one-step RT-qPCR**

# **Daniel Groelz, Nadine Dettmann**

### **Abstract**

RNA from matched FFPE, PFPE and cryo preserved rat tissues, stored for up to nine years at 22°C, 4°C, -20°C or -80°C, examined for integrity and usability in quantitative RT-PCR

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### **Protocol**

# Step 1.

Experimental design	RNA from matched FFPE, PFPE and cryo preserved rat tissues, stored for up to nine years at 22°C, 4°C, -20°C or -80°C, examined for integrity and usability in quantitative RT-PCR			
		• •	Provider/ manufacturer	
Sample	Species	Rats (rattus norvegicus) raised to a weight of approximately 500 g		
		Sacrificed by CO2 asphyxiation		
		Organs removed within 5 min of sacrifice		
	Tissue types	Adjacent, equally sized tissues no larger than $15 \times 15 \times 4$ mm		
	Tissue types	grossed from Liver, Kidney, Spleen, Intestine, Lung		
	Fixation	Snap frozen in liquid nitrogen or placed into standard tissue cassettes and completely submerged in a container filled with fixative with fixation solutions in a ratio of at least 20 parts fixative		
		to one part of tissue (v/v)		
	Fixative	NBF (neutral buffered formalin) for 24 hours at room temperature	Merck KGaA Darmstadt Germany	
		PAXgene Tissue Fix for 2-4 hours at room temperature, afterwards transfer into PAXgene Tissue Stabilizer for 24-72 hours at room temperature	PreAnalytiX Gmbh, Hombrechtikon, CH	
		Snap-frozen in liquid nitrogen, transported on dry ice, and stored at -80°C.		
	Processing and paraffine embedding	Samples fixed with formalin or PAXgene Tissue were processed in separate runs on an automated tissue processor TP1020	Leica-microsystems, Wetzlar, German	
		Incubation at 80%, 90%, 99% ethanol (2x), followed by isopropano (2x), xylene (2x) for no longer than $1h$ at each position		
		Low-melting point paraffin was used for infiltration and embedding	Surgipath Paraplast-XTRA, Carl Roth GmbH, Karlsruhe, Germany	
		For infiltration of tissue with paraffin, samples were incubated (3 x 1 h) under vacuum at $56^{\circ}\text{C}$		
		Within 30 min after infiltration, samples were, embedded		
	Storage	PFPE and FFPE blocks stored in the dark at 22°C, 4°C, -20°C and -80°C		
		Cyro preserved tissue stored for up to one year at -80°C, RNA extracted and stored at -20°C		
RNA extraction	Cryo	10 mg of frozen tissue		
	,	RNeasy Mini kit	QIAGEN GmbH, Hilden, Germany	
		Elution in 40µl RNAse free water, storage at -20°C	quiezn emen, mach, cernany	
	FFPE	Three sections, each 10 µm thick		
		miRNeasy FFPE kit	QIAGEN GmbH, Hilden, Germany	
		Elution in 40µl RNAse free water, storage at -20°C	quiezn emen, mach, eerman,	
	PFPE	Three sections, each 10 µm thick		
		PAXgene Tissue RNA kit	PreAnalytiX Gmbh, Hombrechtikon, CH	
		Elution in 40µl Buffer TR4, storage at -20°C	,	
	Replicates	All preparations were done in triplicate		
	RNA analyses	RNA yield and purity was determined by spectrophotometric absorbance on Nanodrop ND-1000 spectrophotometer at 260nm	Nanodrop Technologies, Wilmington, USA	
		RNA integrity was assessed by microcapillary electrophoresis on an Agilent 2100 Bioanalyzer and analyzed with the Agilent 2100 expert software	Agilent Technologies, Waldbronn, Germany	
Reverse transcription	Template	10 ng of total RNA		
	Assay	one-step RT-PCR assay, 25µl assay volume		
	Kit	QuantiTect® SYBR Green RT-PCR	QIAGEN GmbH, Hilden, Germany	
	NIL	Quantificity 31DN GIERII NI-FCN	QIAGEN GITIDE, FINGER, GETTIANY	

RT-qPCR target information	Gene	PCR primers specific to the rat beta-actin gene (NC_005111)	
	Amplicons	One common forward primer and six different reverse primers to amplify six different transcript sequences with lengths between 109 and 610 nucleotides	
RT-qPCR	Oligonucleotides	All oligonucleotides ordered lyophilized and HPLC-purified.	Metabion GmbH, Planegg/Steinkirchen, German
		Lyophilized oligonucleotides dissolved to 100 µM (stock solution)	,,,,,
		Stored at -15°C to -30°C until use.	
		Name and amplicon length	Seguence 5′ - 3′
		Rn actB for1	CCACACTGTGCCCATCTATGA
		Rn actB rev109 - 109 bp	ACGCTCGGTCAGGATCTTCATG
		Rn actB rev189 - 189 bp	AAGTCTAGGGCAACATAGCAC
		Rn actB rev287 - 287 bp	GGAACCGCTCATTGCCGATAG
		Rn actB rev331 - 331 bp	TTCCATACCCAGGAAGGAAGG
		Rn actB rev438 - 438 bp	TACATGGTGGTGCCACCAGAC
		Rn actB rev465 - 465 bp	TTCTGCATCCTGTCAGCAATG
	Reaction Mix	10ng RNA	TICIOCATCCIOTCAGCAATO
	Reaction Mix	12.5µl 2x Quantitect SYBR Green Master Mix	
		1.25µl forward primer (working solution 10µM) - 0.5µM	
		1.25µl revers primer (working solution 10µM) - 0.5µM	
		1.25µl revers primer (working solution тоµм) - 0.5µм 0.25µl RT Mix	
		Water ad 25µl	
	Cycle condition:	water au 25µi	
		Time - Temperature	Cyclo
	Stage	30min - 50°C - 1x	Cycle 1
	1 - Reverse Transcription Stage 2 - Hold Stage	15min - 95°C - 1x	1
	z - noiù stage		1
		15sec - 94°C - 40x	
	3 - PCR Stage	30sec - 60°C - 40x	40
		30sec - 70°C - 40x	
		15sec - 95°C - 1x	
	4 - Melt Curve Stage	30sec - 60°C -1x	_1
		15sec - 95°C - 1x	
	Tubes and Caps	Rotorgene 72-Well Rotor Strip Tubes and Caps, 0.1mL	QIAGEN GmbH, Hilden, Germany
	Replicates	One amplification per triplicate RNA preparation	
	Instrument	Rotor-Gene Q series II	QIAGEN GmbH, Hilden, Germany
	Software	Rotor Gene Q Series Software 2.3.1	
qPCR validation	Specificity	Examination of amplicon length by agarose gel electrophoresis	
	Specificity	Melting curve analysis	
	Run validity	No template controls no amplifiable, i.e. no Ct determined	
	Acceptance criteria for single reactions	Ct <40	
	Acceptance criteria for single reactions		
		Melting curves had to be free of extraneous peaks or peaks which indicated non-specific amplification to be accepted as valid. All data not meeting these acceptance criteria were excluded from data analysis	