

## Degenerated PCR with GoTaq Hot Start

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Sep 20, 2018 dx.doi.org/10.17504/protocols.io.szyef7w

Working



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### ABSTRACT

Degenerated PCR to test Y-linkage of genes in several *Drosophila* species. The reactions are made separately for males and females of 400 species and subspecies of *Drosophila* and related genera.

### TAGS

Drosophila

PCR


Show tags

### PROTOCOL STATUS

**Working**

We use this protocol in our group and it is working

### MATERIALS

|   | NAME                                | CATALOG # |                            |
|---|-------------------------------------|-----------|----------------------------|
|  | GoTaq(R) Hot Start Polymerase, 500u | M5005     | by <a href="#">Promega</a> |
|  | dNTP Mix, 10mM, 1000ul              | U1515     | by <a href="#">Promega</a> |
|  | DEPC-Treated Water                  | #AM9906   | by <a href="#">Ambion</a>  |

### Pre-Mix Preparation

- 1 Usually, we performing PCR tests in large-scale, testing several DNA samples from different species at once. We prepared a pre-mix stock to economy time in PCR experiments.

| Reagent                     | 1 reaction | 1000 reactions |
|-----------------------------|------------|----------------|
| DEPC-Treated Water          | 11.6 uL    | 11.6 mL        |
| 5x Green GoTaq Flexi Buffer | 4.0 uL     | 4.0 mL         |
| MgCl <sub>2</sub> 25mM      | 2.0 uL     | 2.0 mL         |
| dNTP 10mM                   | 0.4 uL     | 0.4 mL         |
| TOTAL VOLUME                | 18 uL      | 18 mL          |

We divide the pre-mix solution in 1 mL aliquots and stocked at -20°C.

## Final Degenerated PCR preparation

- 2 Normally, the DNA template concentration is 10 ug/uL or higher.

| Reagent                         | 1 reaction (20.1 uL) |
|---------------------------------|----------------------|
| Template                        | 1 uL                 |
| Forward degenerated primer 40mM | 0.5 uL               |
| Reverse degenerated primer 40mM | 0.5 uL               |
| Premix                          | 18 uL                |
| GoTaq Hot Start Polymerase      | 0.1 uL               |

## PCR Programs

- 3 We used different thermocycle programs, according to the primers. In all programs, the GoTaq Hot Start Polymerase was previous incubated for 2 minutes to be activated. The PCRs were performed in a Applied Biosystems Veriti™ 96-Well Thermal Cycler (Cat#4375786).

**1) Degenerated PCR Program:** Differently of the normal PCR thermocycler programs, the degenerated PCR have more time for annealing.

| cycles | Denaturation   | Annealing     | Polymerization                      |
|--------|----------------|---------------|-------------------------------------|
| 1x     | 95°C, 2:00 min | ---           | ---                                 |
| 40x    | 95°, 0:30 min  | x°C, 1:30 min | 72°C, 1:00 min /1000 pb of template |
| 1x     | ---            | ---           | 72°C, 7:00 min                      |

x°C = optimal annealing temperature for the pair of primers.

**2) Degenerated Touchdown PCR (TD-PCR) Program:** In TD-PCR, we screen a range of annealing temperatures to try optimize the reaction in different species samples. So, we have a stage where the annealing temperature decrease -0.2°C by cycle, in the end of this stage, the annealing temperature decreased -4°C.

| cycles | Denaturation   | Annealing                     | Polymerization                      |
|--------|----------------|-------------------------------|-------------------------------------|
| 1x     | 95°C, 2:00 min | ---                           | ---                                 |
| 20x    | 95°, 0:30 min  | X°C, 1:30 (Δ -0.2°C by cycle) | 72°C, 1:00 min /1000 pb of template |
| 25x    | 95°, 0:30 min  | x°C, 0:30 min                 | 72°C, 1:00 min /1000 pb of template |
| 1x     | ---            | ---                           | 72°C, 7:00 min                      |

x°C = optimal annealing temperature for the pair of primers.

### References:

Sambrook, J. & Russell, D. W. (2001) 'Chapter 8. Protocol 11. Mixed Oligonucleotide-primed Amplification of cDNA' in *Molecular cloning : a laboratory manual*. Cold Spring Harbor: Cold Spring Harbor Laboratory, p. 8.66-8.71.



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