



Illumina (post-MR DNA) processing pipeline : mothur

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ABSTRACT

This pipeline is for datasets generated from Illumina (2x300) sequencing by MR DNA. MR DNA runs sequences through a proprietary pipeline for quality control of sequences and OTU clustering. This pipeline begins with the OTU abundance table and utilizes the representative sequence set generated by the MR DNA pipeline. **Through trial and error, I have discovered the easiest and best practice for my lab is to use a combination of QIIME (full installation) and mothur. This protocol is the mothur portion.** This protocol does not include data visualization.

As of 4/5/17, the following versions were used: QIIME v.1.9.1-amd64.vdi and mothur v.1.39.5

PROTOCOL STATUS

Working

Working for mothur

Before you start this protocol

- 1 **Begin with Illumina (post-MR DNA) processing pipeline : QIIME** steps 1-7 to rarefy the OTU abundance table. Complete step 12 to convert the rarefied OTU abundance table to a text file.

Install the executable version of mothur in Windows

- 2 "Downloading an executable version of mothur is the easiest and fastest way to get started." -[mothur download page](#)

The best way to use mothur is in Windows or Mac where you can download an executable version of mothur that runs through a terminal window. All scripts are written in C++, a wrapped language, so you can execute a command within the mothur terminal just like you would execute a QIIME command in the QIIME terminal window. [Here is the latest release of mothur on github](#). Follow instructions of the [installation page](#). If you download the executable version of mothur, it will simply show up as an icon that opens to a terminal window when double-clicked. That is mothur! It is simpler than it appears.

Assign taxonomy

- 3 To compare the taxonomy assigned by the sequencing facility (MR DNA in our case) through a proprietary pipeline and curated database, you can use the [classify_seqs](#) command in mothur.

COMMAND

```
classify_seqs(fasta=CCWT_16S_Jan_Feb_Apr_rep_set.fa, template=gg_13_8_99.fasta, taxonomy=gg_13_8_99.tax)
```

fasta = is your unaligned representative sequences (1 OTU = 1 sequence)

template = can be either Greengenes or SILVA database template, unaligned

taxonomy = needs to be from the same database as the template file



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