



 $m{\ell}$  Addition of the adaptor to RNA substrates for 3 $^{\prime}$  RACE (mapping P ends)

Forked from Addition of the adaptor to RNA substrates for 3' RACE (mapping OH ends)

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**ABSTRACT** 

Simple protocol for mapping 3'-P RNA termini by RT-PCR after the addition of a 3' adaptor using E. coli RtcB RNA ligase.

PROTOCOL STATUS

## Working

We use this protocol in our group and it is working

MATERIALS

NAME

CATALOG #

**VENDOR** 

RtcB Ligase - 25 rxns111212

M0458S

**New England** 

SAFETY WARNINGS

## BEFORE STARTING

Prepare samples, including controls, according to the aim of the experiment. Use the chart below to decide which enzyme is appropriate for pre-treatments of the RNA. For example, if only interested in mapping 5' RNA ends, use T4 polynucleotide kinase with ATP and without ATP to infer the original phosphorylation state of the end of interest.

Setup of the phosphorylation reaction can be found here: dx.doi.org/10.17504/protocols.io.cpdvi5 However, avoid heat denaturation of the enzyme and rather purify the RNA using a trizol extraction (e.g., http://dx.doi.org/10.17504/protocols.io.eiebcbe) or a column clean-up (e.g., Monarch RNA Cleanup Kits from NEB is optimal when interested in small RNA molecules < 200 nt, which is the usual exclusion limit in other products).

| Original | Original | T4PNK  | T4PNK  | T4PNK (- | T4PNK (- | T4PNK-  | T4PNK-  | T4PNK-     | T4PNK-     |
|----------|----------|--------|--------|----------|----------|---------|---------|------------|------------|
|          |          | (+ATP) | (+ATP) | ATP)     | ATP)     | 3′Pase⊖ | 3′Pase⊖ | 3′Pase⊖ (- | 3′Pase⊖ (- |
|          |          |        |        |          |          | (+ATP)  | (+ATP)  | ATP)       | ATP)       |
| 5′       | 3′       | 5′     | 3′     | 5′       | 3′       | 5′      | 3′      | 5′         | 3′         |
| Р        | Р        | Р      | ОН     | Р        | ОН       | Р       | Р       | Р          | Р          |
| Р        | ОН       | Р      | ОН     | Р        | ОН       | Р       | ОН      | Р          | ОН         |
| ОН       | Р        | Р      | ОН     | ОН       | ОН       | Р       | Р       | ОН         | Р          |
| ОН       | ОН       | Р      | ОН     | ОН       | ОН       | Р       | ОН      | ОН         | ОН         |

Mix the following components (9  $\mu$ L):

| Component                                  | Amount [μL] | Final concentration |
|--|-------------|---------------------|
| RNA [1 μg]                                 | 4           | 100 ng/μL           |
| 5'-hydroxylated 3' RACE RNA oligo [100 μM] | 0.4         |                     |



| RNase-free water (ddH20) 4.6 | .6 |  |
|------------------------------|----|--|
|------------------------------|----|--|

- Blocking the 3½ end of the 3½ RACE oligo is not necessary, as 3½-OH ends are not a substrate for EcRtcB RNA ligase. 5½-end has to be hydroxylated. For practical purposes, a 5½ RACE RNA oligo, which usually has both ends hydroxylated, can be used for as the 3½ RACE oligo when mapping 3½-P ends by EcRtcB.
- 2 Denature for 2 min at 70 °C, place on ice.
- 3 Mix the following components (20 μL):

| Component                  | Amount [μL] | Final concentration |
|----------------------------|-------------|---------------------|
| RNA + oligo mix (step 2)   | 9           |                     |
| 10× RtcB RNA ligase buffer | 2           | 1×                  |
| 50% PEG-8000               | 4           | 10%                 |
| GTP [1 mM]                 | 2           | 0.1 mM              |
| MnCl2 [10 mM]              | 2           | 1 mM                |
| EcRtcB RNA ligase [15 μM]  | 1           | 0.75 μΜ             |

- ▲ Incubate for 120 min at 37 °C.
- $5 \qquad \hbox{Purify the RNA from the RNA ligase reaction (e.g., trizol extraction or column clean-up)}.$
- 6 Proceed to RT-PCR. For the RT reaction, use 250-500 ng of the purified adaptor-ligated RNA and a primer reverse complementary to the 3' adaptor. For the PCR, use an upstream (forward) primer binding to the RNA of interest and a downstream (reverse) primer binding to the 3' adaptor.

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