PCR with Q5® High-Fidelity 2X Master Mix (M0492)

New England Biolabs

Abstract

This protocol is for PCR with Q5® High-Fidelity 2X Master Mix (M0492)

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Guidelines

Please note that protocols with Q5 High-Fidelity DNA Polymerase may differ from protocols with other polymerases. Conditions recommended below should be used for optimal performance.

Reaction Setup:
We recommend assembling all reaction components on ice and quickly transferring the reactions to a thermocycler preheated to the denaturation temperature (98°C). All components should be mixed prior to use.

Component	25 µl Reaction	50 µl Reaction	Final Concentration
Q5 High-Fidelity 2X Master Mix	12.5 µl	25 µl	1X
10 µM Forward Primer	1.25 µl	2.5 µl	0.5 µM
10 µM Reverse Primer	1.25 µl	2.5 µl	0.5 µM
Template DNA	variable	variable	< 1,000 ng
Nuclease-Free Water	to 25 µl	to 50 µl	

Notes: Gently mix the reaction. Collect all liquid to the bottom of the tube by a quick spin if necessary. Overlay the sample with mineral oil if using a PCR machine without a heated lid.

Transfer PCR tubes to a PCR machine and begin thermocycling.

Thermocycling Conditions for a Routine PCR:

STEP	TEMP	TIME
Initial Denaturation	98°C	30 seconds
	98°C	5–10 seconds
25–35 Cycles	*50–72°C	10–30 seconds
	72°C	20–30 seconds/kb
Final Extension	72°C	2 minutes
Hold	4–10°C	

*Use of the NEB Tm Calculator is highly recommended.

General Guidelines:

1. Template:
Use of high quality, purified DNA templates greatly enhances the success of PCR. Recommended

amounts of DNA template for a 50 µl reaction are as follows:

DNA	AMOUNT
DNA Genomic	1 ng–1 µg
Plasmid or Viral	1 pg–1 ng

2. Primers:
Oligonucleotide primers are generally 20–40 nucleotides in length and ideally have a GC content of 40–60%. Computer programs such as Primer3 can be used to design or analyze primers. The best results are typically seen when using each primer at a final concentration of 0.5 µM in the reaction.

3. Mg++ and additives:
The Q5 High-Fidelity Master Mix contains 2.0 mM Mg++ when used at a 1X concentration. This is optimal for most PCR products generated with this master mix.

4. Deoxynucleotides:
The final concentration of dNTPs is 200 µM of each deoxynucleotide in the 1X Q5 High-Fidelity Master Mix. Q5 High-Fidelity DNA Polymerase cannot incorporate dUTP and is not recommended for use with uracil-containing primers or templates.

5. Q5 High-Fidelity DNA Polymerase concentration:
The concentration of Q5 High-Fidelity DNA Polymerase in the Q5 High-Fidelity 2X Master Mix has been optimized for best results under a wide range of conditions.

6. Denaturation:
An initial denaturation of 30 seconds at 98°C is sufficient for most amplicons from pure DNA templates. Longer denaturation times can be used (up to 3 minutes) for templates that require it.

During thermocycling, the denaturation step should be kept to a minimum. Typically, a 5–10 second denaturation at 98°C is recommended for most templates.

7. Annealing:
Optimal annealing temperatures for Q5 High-Fidelity DNA Polymerase tend to be higher than for other PCR polymerases. The NEB Tm Calculator should be used to determine the annealing temperature when using this enzyme. Typically use a 10–30 second annealing step at 3°C above the Tm of the lower Tm primer. A temperature gradient can also be used to optimize the annealing temperature for each primer pair.

For high Tm primer pairs, two-step cycling without a separate annealing step can be used (see note 10).

8. Extension:
The recommended extension temperature is 72°C. Extension times are generally 20–30 seconds per kb for complex, genomic samples, but can be reduced to 10 seconds per kb for simple templates (plasmid, E. coli, etc.) or complex templates < 1 kb. Extension time can be increased to 40 seconds per kb for cDNA or long, complex templates, if necessary.

A final extension of 2 minutes at 72°C is recommended.

9. Cycle number:

Generally, 25–35 cycles yield sufficient product. For genomic amplicons, 30-35 cycles are recommended.

10. 2-step PCR:
When primers with annealing temperatures ≥ 72°C are used, a 2-step thermocycling protocol (combining annealing and extension into one step) is possible.

Amplification of long products:
When amplifying products > 6 kb, it is often helpful to increase the extension time to 40–50 seconds/kb.

11. PCR product:
The PCR products generated using Q5 High-Fidelity 2X Master Mix have blunt ends. If cloning is the next step, then blunt-end cloning is recommended. If T/A-cloning is preferred, the DNA should be purified prior to A-addition, as Q5 High-Fidelity DNA Polymerase will degrade any overhangs generated.

Addition of an untemplated -dA can be done with Taq DNA Polymerase (NEB #M0267) or Klenow exo– (NEB #M0212).

Materials

Q5 High-Fidelity 2X Master Mix - 100 rxns M0492S by New England Biolabs

Protocol

Step 1.
Set up the following reaction on ice:

Component	25 µl Reaction	50 µl Reaction	Final Concentration
Q5 High-Fidelity 2X Master Mix	12.5 µl	25 µl	1X
10 µM Forward Primer	1.25 µl	2.5 µl	0.5 µM
10 µM Reverse Primer	1.25 µl	2.5 µl	0.5 µM
Template DNA	variable	variable	< 1,000 ng
Nuclease-Free Water	to 25 µl	to 50 µl	

PROTOCOL

. Mixture for M0492 Q5 2X Master Mix

CONTACT: New England Biolabs
Step 1.1.
Q5 High-Fidelity 2X Master Mix

Step 1.2.

10 µM Forward Primer

ANNOTATIONS

Florence Servais 28 Jul 2015

Primers from 100µM stock => 1/10 dilution first

rPAP1 into secNLuc reporter plasmid: rPAP1secNLucGibsonFor

rPAP1 into CLuc reporter plasmid: rPAP1CLucGibsonFor

STAT3 binding sites + CMV into secNLuc report plasmid: STAT3allsitessecNLucFor

Step 1.3.

10 µM Reverse Primer

ANNOTATIONS

Florence Servais 28 Jul 2015

From 100µM stock => 1/10 dilution first

rPAP1 into secNLuc reporter plasmid: rPAP1secNLucGibsonRev

rPAP1 into CLuc reporter plasmid: rPAP1CLucGibsonRev

STAT3 binding sites + CMV into secNLuc report plasmid: STAT3allsitessecNLucRev

Step 1.4.

Template DNA

ANNOTATIONS

Florence Servais 28 Jul 2015

Use of 0.25ng of each DNA template (plasmids):

#582 (pXP2d2-rPAP1): cc = 0.115µg/µL => do 1/100 dilution and take **0.217µL**

#630 (pSTAT3-CLuc Reporter-MH1): cc = 0.4µg/µL => do 1/200 dilution and take **0.125µL**

Step 1.5.

Nuclease-Free Water

Step 2.

Gently mix the reaction.

Step 3.

Collect all liquid to the bottom of the tube by a quick spin if necessary and overlay the sample with mineral oil if using a PCR machine without a heated lid.

Step 4.

Quickly transfer PCR tubes to a thermocycler preheated to the denaturation temperature (98°C).

Warnings

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