Protocol for STO Cell Transfection by FuGENE HD Version 2

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Abstract

Protocol for Transfection Mouse Embryonic Stem Cells.

Citation: Misha Gurevich, V. Katerov Protocol for STO Cell Transfection by FuGENE HD. protocols.io

dx.doi.org/10.17504/protocols.io.e8sbhwe

Published: 23 Jun 2016

Protocol

Cell plating

Step 1.

STO cells were seeded the day before transfection with the density 15,000 cells per well in 100 μ l complete growth medium DMEM+10% Fetal Bovine Serum.

Complex preparation (per 20 wells)

Step 2.

Prepare 0.02μg/μl pCMVβ plasmid DNA solution in OptiMEM®.

NOTES

Misha Gurevich 23 Jun 2016

Tissue culture 96-round bottom well plates were used for complex preparation.

Complex preparation (per 20 wells)

Step 3.

Add 6µl of reagent to 100 µl of OptiMEM® /DNA solution.

Complex preparation (per 20 wells)

Step 4.

Mix carefully by pipetting (10-15 times).

Complex preparation (per 20 wells)

Step 5.

Incubate 5 min at room temperature.

O DURATION

00:05:00

Complex preparation (per 20 wells)

Step 6.

Add 5µl complex per well to the cells, and mix thoroughly.

P NOTES

Misha Gurevich 23 Jun 2016

Optimal ratio reagent/DNA may vary in range 0.2µl/0.1µg to 0.35µl/0.1µg.

Incubation

Step 7.

Place the cells into CO2 incubator for 26-28 hours.

© DURATION

26:00:00

Detection of β-gal expression

Step 8.

Remove the medium from the well and wash the cells once with 100µl per well PBS.

Detection of β-gal expression

Step 9.

Fix the cells in the well with 50µl solution of 4% formaldehyde in PBS for 5min at room temperature.

O DURATION

00:05:00

Detection of β-gal expression

Step 10.

Wash each well with 100µl PBS. (1/2)

Detection of β-gal expression

Step 11.

Wash each well with 100µl PBS. (2/2)

Detection of β-gal expression

Step 12.

Add 50µl per well of substrate/stain solution and incubate the plate overnight at 37°C.

O DURATION

16:00:00

Detection of β-gal expression

Step 13.

Observe the cells under microscope and evaluate the proportion of blue (β -gal-positive) cells.