

Short insert size WGS libraries preparation for assembly of the *Lateolabrax maculatus* genome

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Abstract

This protocol is used to clarify the process of the short insert size WGS libraries preparation for the *L. maculatus*.

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Protocol

Genomic DNA interruption

Step 1.

The extracted DNA was sheared into fragments between 50 bp and 800 bp in size using a Covaris E220 ultrasonicator (Covaris, Brighton, UK). Treat time 20s, Acoustic Duty Factor 25%, Peak Incident Power 500W, Cycles Per Burst 500, 24 cycles.

DURATION


00:00:20 :

Fragment selection

Step 2.

Fragments between 150 bp and 250 bp or 200 to 500 bp were selected using AMPure XP beads (Agencourt, Beverly, the U.S.).

REAGENTS

 AMPure XP beads by [Beckman Coulter](#)

End-repair

Step 3.

Repaired using T4 DNA polymerase, (ENZYMATICS, Beverly, the U.S.) 30 min. at 20 °C to obtain blunt ends which were then 3'-adenylated to create sticky ends.

TEMPERATURE

20 °C :

REAGENTS

 T4 DNA polymerase by [Enzymatics](#)

DURATION

00:30:00 :

Add adapter

Step 4.

These DNA fragments were ligated at both ends to T-tailed adapters and amplified.

PCR amplification

Step 5.

The temperature profile was 3 min. at 95 °C followed by 8 cycles of 20 sec. at 98 °C, 15 sec. at 60 °C, 30 sec. at 72 °C, and more 10 min. at 72 °C for further elongation.

Library purification

Step 6.

AMPure XP beads (Agencourt, Beverly, the U.S.) was used to purify the PCR production.