



Aug 16, 2019

Hiseq 2000 Library Construction and Sequencing for RNA Seq

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CATALOG #

VENDOD



ABSTRACT

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THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0050226

STEPS MATERIALS

NAME

NAME	CATALOG #	VENDOR
DNasel		NEB
Dynabeads mRNA purification kit		Life Technologies
fragmentation buffer		Life Technologies
SuperScript II reverse transcription kit		Life Technologies
RNase H		Life Technologies
DNA polymerase		Enzymatics
QIAquick PCR purification kit		Qiagen
Agencourt AMPure beads		Beckman Coulter
T4 DNA polymerase and T4 polynucleotide kinase		Enzymatics
Klenow (3' to 5' exo-)		Enzymatics
dATP		Ge Healthcare
Phusion DNA polymerase		NEB

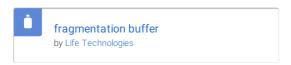


using



.It is best to use up to 50 μ g as the use of a lower mass (typically 20 μ g) has been insufficient for successful library construction. This can be assessed by running final PCR products on an agarose gel; the library construction is considered to have failed when there was no visible band. It is possible to use less than 20 μ g of total RNA when isolation of an important sample yielded low RNA mass but library construction was successful.

2 Purified polyA RNA is fragmented in a

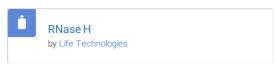


at § 70 °C for © 00:01:30 to 200-300 nt fragment sizes.

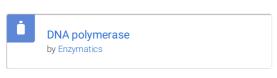
3 The first cDNA strand is then synthesized with random hexamer primers using the



⚠ The second-strand synthesis is performed by incubation with

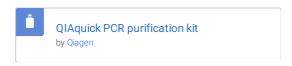


and



5 Short double-stranded cDNA fragments are then purified using one of two methods.

6 Our standard procedure was to use the



, whereas for samples with low RNA mass it is better to use



7 Both methods are then followed by end-repair with Klenow polymerase,



A single 3' adenosine (A base) was added to the double-stranded cDNA using



and



9 The Illumina PE Adapter oligo mix is ligated onto the A base on repaired double-stranded cDNA ends and DNA fragments of a selected size are then gel-purified to make sure the insert size is 200 bp (±10% deviation).

10 Thereafter, libraries were amplified by 15 cycles of PCR with



and "indexed" paired-end PCR primers; the prepared libraries were 322 bp long.

- The amplified libraries were denatured with sodium hydroxide and diluted to [M]2.5 Picomolar (pM) in hybridization buffer for loading into a HiSeq flowcell.
- 12 Read lengths viewed on the HiSeq platform were predominately 90 bp with a small number of sequences in the 84–87 bp range.

Samples are sequenced with paired-end reads, and up to eleven samples can be multiplexed into a single lane of the Illumina Hiseq flow cell. With average run time of three to twelve days depending on read length.

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