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## The EV71 neutralizing antibody test [↗](#)

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### ABSTRACT

This protocol describes the process for the neutralizing antibody test of EV71.

### EXTERNAL LINK

<https://doi.org/10.1371/journal.pone.0224110>

### THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

Lee J, Yen T, Shih W, Lu C, Liu D, Huang Y, Chang L, Huang L, Lin T (2019) Enterovirus 71 seroepidemiology in Taiwan in 2017 and comparison of those rates in 1997, 1999 and 2007. PLoS ONE 14(10): e0224110. doi: [10.1371/journal.pone.0224110](https://doi.org/10.1371/journal.pone.0224110)

### GUIDELINES

The serum neutralizing antibody response is the major indicator of EV71 protective immunity. The neutralizing antibody test of EV71 followed the standard protocol of a neutralization test. Serum samples were heat-treated for 30 minutes at 56°C, serially diluted and mixed with 100 50% tissue culture-infective doses (TCID<sub>50</sub>) of the EV71 TW/2272/98 strain (GenBank accession number AF119795), and incubated for 2 hours at 37°C. Thereafter, rhabdomyosarcoma cells were added into each reaction well and incubated at 37°C in 5% CO<sub>2</sub> incubator. Each plate included a cell control, serum control, and virus back-titration. The cytopathic effect was monitored from 5 to 6 days after incubation, and the serotiter was determined when the cytopathic effect was observed in one TCID<sub>50</sub> of the virus back-titration. Seropositivity was defined as a serotiter ≥ 8.

### MATERIALS TEXT

Serum or plasma

96 well microtiterplate

1x PBS

DMEM with 2% FBS medium

Multichannel Pipette (8 or 12)

1.5 ml eppendrof

EV71 TW/2272/98 strain (GenBank accession number AF119795)

### SAFETY WARNINGS

EV71 needs to operate in BSL-2 lab

BEFORE STARTING

Set water bath to 56°C

Confirm that the RD cells are healthy

- 1 Inactivate serum (300 µl) at 56°C for 30 min

Sample volumn (µl)	1x PBS (µl)	Dilute factor
Ex. 75	225	1:4

- 2 Make 2 fold serial dilution of serum from 1:4 to 1:256 with 2% FBS DMEM

sample	1-1	1-2	2-1	2-2	3-1	3-2	4-1	4-2	5-1	5-2	6-1	6-2
A 1:4	●	●	○	○	○	○	○	○	○	○	○	○
B 1:8	●	●	○	○	○	○	○	○	○	○	○	○
C 1:16	○	○	○	○	○	○	○	○	○	○	○	○
D 1:32	○	○	○	○	○	○	○	○	○	○	○	○
E 1:64	○	○	○	○	○	○	○	○	○	○	○	○
F 1:128	○	○	○	○	○	○	○	○	○	○	○	○
G 1:256	○	○	○	○	○	○	○	○	○	○	○	○
H control	●	○	○	○	○	○	○	○	○	○	○	○

a. Add 50 µl 2% FBS DMEM in well B to H

b. ● means add 50µl serum sample

c. After dilution, every well has 50µl sample

d. ex. sample1-1 well H: serum control; sample 1-2 well H: cell control; 1-1 and 1-2 are duplicate

- 3 Add 50 µl of adjusted virus suspension (100TCID<sub>50</sub>EV71) onto the well A to well G of microtiterplate, shake reaction well slightly

- 4 Prepare control wells including virus control, serum control and cell control.

Virus control well↯back titration from 100, 10, 1 to 1/10 100TCID<sub>50</sub>

Serum control↯add serum and cell only, no virus

Cell control↯add cell only, no serum and no virus

- 5 Incubate above reaction wells at 37°C for 2 hours in 5% CO<sub>2</sub> incubator

- 6 Prepare RD cells suspension and adjust cell concentration to 1×10<sup>5</sup>/ml

- 7 Add 100µl of adjusted RD cell into each reaction well after 2 hours in step5
- 8 Incubate wells at 37°C in 5% CO<sub>2</sub> incubator for 5-6 days until virus control well reach 100TCID<sub>50</sub>
- 9 Observe CPE in each well under reverse microscope after 5-6 days incubation
- 10 Check CPE in control wells  
  
Virus control well 100% CPE in 100TCID<sub>50</sub> and 10TCID<sub>50</sub>; 50% CPE in 1TCID<sub>50</sub>, < 10% CPE in 1/10TCID<sub>50</sub>  
  
Serum control well no CPE  
  
Cell control well no CPE
- 11 The NT titer was calculated as the reciprocal value of the last dilution of the antiserum that completely inhibited the CPE after incubation.
- 12 Seropositivity was defined as a reciprocal of the serotiter  $\geq 8$ .



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