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Working

## aCGH Hybridization &amp; wash




Hendrik F van Essen<sup>1</sup><sup>1</sup>Amsterdam UMC

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Hendrik F HFvE. van Essen  
Amsterdam UMC

## aCGH Hybridization

- 1 Turn on hybridization oven ⚡ 65 °C  
Turn on heat block ⚡ 95 °C  
Turn on heat block ⚡ 37 °C
- 2 Mix Cy3 and Cy5 samples
- 3 Mix the following components in 1.5 centrifuge tubes
  - \* Cy3/Cy5 gDNA mixture 39 µl
  - \* Cot-1 DNA (1.3 µg/µl) 5 µl
  - \* Agilent 10x blocking agent 11 µl
  - \* Agilent 2x Hybridization buffer 55 µl
  - Total 110 µl
- 4 Mix sample by pipetting, by careful not to create any bubbles.
- 5 Spin down with maximum speed for a few seconds.
- 6 Heat for ⌚ 00:03:00 at ⚡ 95 °C and incubate immediately thereafter for ⌚ 00:30:00 at ⚡ 37 °C .
- 7 Leave samples in heat block, spin down for ⌚ 00:01:00 at maximum speed.
- 8 Load gasket slide in hybridization chamber and dispense 100 µl hyb mix in a drag and drop manner (use a 200 µl pipette, be careful not to create any bubbles)
- 9 Place microarray slide – active side down – and assemble chamber.

- 10 Check for immobile air bubbles by rotating the assembled hybridization chamber. If necessary, slam side of the chamber on table to loosen the air bubbles.
- 11 Place assembled chamber in the hybridization oven (  65 °C at 20 rpm) and make sure to balance with empty chambers.
- 12 Hybridize for  24:00:00 ,  65 °C at 20 rpm.

#### aCGH wash

- 13 Turn on Agilent Slide Scanner
  - 14 Wash buffer dishes with distilled water and blow dry with pressurized air.
  - 15 Add buffers to dishes and have tin foil covering ready.-  
Wash a maximum of 5 Agilent slides at the same time.
- | Dish | Method        | Buffer       | Temp (°C) | Time (min)              |
|------|---------------|--------------|-----------|-------------------------|
| #1   | Disassembly   | Buffer 1     | RT        | -                       |
| #2   | 1st wash step | Buffer 1     | RT        | 5 (stirrer)             |
| #3   | 2nd wash step | Buffer 2     | 37        | 1 (stirrer & hot plate) |
| #4   | Dry step      | Acetonitrile | RT        | 1                       |
- 16 Separate gasket slide from microarray slide in dish #1 (buffer 1)
  - 17 Place slides in holding rack and was in dish #2 for  00:05:00 , rotation of stirrer is switched ON!
  - 18 Cover dish with tin foil.
  - 19 Start the "*Scan Control Software*"  
For Agilent 180k, select the "AgilentG3\_CGH" Option
  - 20 Move holding rack to dish #3 (buffer 2) and wash for  00:01:00 , rotation of stirrer and heating is switched ON!
  - 21 Move holding rack to dish #4 (acetonitrile) leave for  00:01:00 .
  - 22 Remove slide gently with tweezers, thus preventing the formation of droplets.
  - 23 Place slide in holder and load into the carousel of the scanner  
Agilent Slide Scanner scans through the back of the slide (this side must contain the numeric barcode)

- 24 Click on "Set Values" to confirm all settings

- 25 Click Scan Slot m-n on the Scan Control main window  
The letter m represents the Start slot where your first slide is located, and the letter n represents the End slot where your last slide is located.
- 26 Start the scan
- 27 Open “feature extraction” and check TIFF file



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