



## Immunofluorescence staining protocol for co-staining of fetuin-A and GFAP in older human autopsy tissue via Tyramide Signal Amplification [↗](#)

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### ABSTRACT

This staining was performed to detect fetuin-A in astrocytes. Fetuin-A and astrocytes were detected in paraffin sections (1  $\mu$ m thickness) of formalin-fixed human brain tissue. Glial Fibrillary Acid Antigen (GFAP) was marked by a polyclonal rabbit-anti-human antibody (Spring Bioscience Cat# E18320, RRID:AB\_1661177, dilution 1:50) and a polyclonal goat-anti-rabbit Alexa Fluor® 488 conjugated secondary antibody (Thermo Fisher Scientific Cat# A-11070, RRID:AB\_2534114, dilution 1:300). Fetuin-A was detected by using a monoclonal IgG2a mouse-anti-human antibody (clone MAHS-1, dilution 1.0  $\mu$ g/mL), raised against purified human fetuin-A in our laboratories. Antibody binding was detected by tyramide signal amplification using a secondary biotinylated polyclonal goat-anti-mouse antibody (Dako Cat# E0433, RRID:AB\_2687905, dilution 1:300) and a Tyramide Signal Amplification Kit (Life Technologies, Carlsbad, USA, T-20933). To minimize lipofuscin autofluorescence, sections were counterstained with Sudan Black (Sigma-Aldrich, Munich, Germany, 199664, dilution 0.3% in 70% ethanol, 5 minutes). Nuclei were stained with DAPI (Sigma-Aldrich, Munich, Germany D9542, dilution 0.25  $\mu$ g/mL, 5 minutes). Sections were mounted with Immumount (Thermo Scientific, Waltham, USA, 9990402) and stored at 8°C in the dark.

### EXTERNAL LINK

<https://doi.org/10.1371/journal.pone.0206597>

### THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

Heinen MC, Babler A, Weis J, Elsas J, Nolte K, Kipp M, Jahnen-Dechent W, Häusler M (2018) Fetuin-A protein distribution in mature inflamed and ischemic brain tissue. PLoS ONE 13(11): e0206597. doi: [10.1371/journal.pone.0206597](https://doi.org/10.1371/journal.pone.0206597)

### PROTOCOL STATUS

**Working**

### GUIDELINES

use with Tyramide Signal Amplification Kit (life technologies, T-20933)

### BEFORE STARTING

Prepare the following components of the Tyramide Signal Amplification Kit according to the manufacturers protocol:

1. Prepare tyramide stock solution by dissolving Component A in 150  $\mu$ L of Component B (DMSO). (Store unused portions of this stock solution in small aliquots at  $\leq -20^{\circ}\text{C}$ , protected from light).
2. Prepare a 1% (10 mg/mL) solution of Component D (BSA) in phosphate-buffered saline (PBS).
3. Prepare the horse radish peroxidase (HRP) conjugate stock solution by reconstituting Component C in 200  $\mu$ L PBS. (This solution may be stored at  $2-8^{\circ}\text{C}$  for up to 3 months).
4. Prepare amplification buffer/0.0015% H<sub>2</sub>O<sub>2</sub> by adding 30% hydrogen peroxide (Component F) to amplification buffer (Component E) to obtain a final concentration of 0.0015% H<sub>2</sub>O<sub>2</sub>. For example, add 1  $\mu$ L of 30% H<sub>2</sub>O<sub>2</sub> to 200  $\mu$ L of amplification buffer and then add 1  $\mu$ L of this intermediate dilution (0.15% H<sub>2</sub>O<sub>2</sub>) to a further 100  $\mu$ L of amplification buffer.

### See Guidelines

- 1 Prepare the components of the Tyramide Signal Amplification Kit

### Clear slides

- 2 Place slides in hybridization oven: 37°C overnight, then 1 hour at 65°C
- 3 Deparaffination in xylene 3x20 minutes (different containers) on a shaker
- 4 Rehydration in graded ethanol: 3x2 minutes in 100% ethanol, followed by 2x2 minutes in 96% ethanol and at last 2 minutes in 70% ethanol
- 5 Wash in PBS for 5 minutes

#### Antigen retrieval

- 6 Antigen retrieval in citrate buffer (10 mM, pH 6) in a heat steamer for 30 minutes. Afterwards let the slides cool down in PBS for 30 minutes
- 7 Wash in PBS for 5 minutes

#### Immunostaining

- 8 Encircle tissue section with wax pen
- 9 To block non-specific binding, apply a solution of 10% goat serum in PBS, incubation for 1 hour in a moisture chamber
- 10 Tip off excess solution and apply the primary antibodies MAHS-1 (dilution 1 µg/ml) and Anti-GFAP (dilution 1:50) in 1% BSA in PBS, 100 µl per section (negative controls: only 1% dilution), incubation overnight in a moisture chamber
- 11 Tip off excess solution and rinse in demineralized water
- 12 Wash in PBS for 3x5 minutes
- 13 Apply the secondary antibodies: biotinylated goat-anti-mouse antibody (dilution 1:300) and polyclonal goat-anti-rabbit Alexa Fluor® 488 conjugated secondary antibody (dilution 1:300), both in 1% of BSA in PBS, 100 µl per section, incubation for 90 minutes at room temperature in a moisture chamber.  
  
*keep the tissue samples from now on light protected!*
- 14 Tip off excess solution and rinse in demineralized water
- 15 Wash in PBS for 3x5 minutes
- 16 Apply the Tyramide Signal Amplification Kit as follows: Dilute the prior prepared HRP solution 1:100 in 1% BSA in PBS, apply 100 µl per section, incubation for 1 hour at room temperature in a moisture chamber.

- 17 Tip off excess solution and rinse in demineralized water
- 18 Wash in PBS for 3x5 minutes
- 19 Prepare a tyramide working solution following the manufacturer's protocol: dilute the prior prepared tyramide stock solution 1:100 in the prepared 0.0015% hydrogen peroxid solution
- 20 Apply 100 µl per section, incubation for 10 minutes
- 21 Tip off excess solution and rinse in demineralized water
- 22 Wash in PBS for 3x5 minutes
- 23 To minimize lipofuscin autofluorescence, perform counterstaining with Sudan Black (dilution 0.3% in 70% ethanol) for 5 minutes
- 24 Rinse the container with demineralized water until the solution becomes clear and colorless
- 25 Wash in demineralized water for 3x5 minutes
- 26 Counterstain with DAPI (dilution 0.25 µg/ml) for 5 minutes
- 27 Wash in PBS for 3x5 minutes
- 28 Coverslip using mounting medium
- 29 store at 8°C in the dark



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