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Working

## Loop and uLoop assembly

Version 5

Bernardo Pollak<sup>1</sup>

<sup>1</sup>J. Craig Venter Institute/PUC

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Protist Research to Optimize Tools in Genetics (PROT-G)



Bernardo Pollak  
J. Craig Venter Institute/PUC



### ABSTRACT

This protocol is used for performing Type IIS assembly by either *Bsa*I or *Sap*I-mediated restriction/ligation using Loop assembly with either Loop or uLoop plasmids.

Loop assembly comprises 8 receiver plasmids in odd and even levels (4 per level), which contain directional overhangs for higher level assembly. Odd and even level receiver plasmids are used in a recursive schema for assembly in iterative 'loops'. Any assembly (except for L0 part composition into a L1 transcriptional unit) requires the usage of 4 donor plasmids (each in a specific position) and one receiver plasmid. *Bsa*I is used for assembly into odd levels using kanamycin selection, and *Sap*I is used in even levels using spectinomycin selection. Loop odd receivers use the Common Syntax standard for L0 part assembly into TUs (Patron, *et al.* 2015 [10.1111/nph.13532]). Then, 4 L1 plasmids (positions 1-4) are assembled into an even receiver plasmid to provide a L2 assembly (4 TUs). For higher level assemblies the same assembly structure is followed, 4 L2 plasmids (positions 1-4) with an odd receiver to yield a L3 assembly (16 TUs).

The Loop type IIS assembly protocol is based on:

"Patron, NJ (2016) DNA Assembly for Plant Biology. Current Protocols in Plant Biology 1:1-13 [doi: 10.1002/cppb.20038]", but with slight modifications for DNA concentrations:

**This protocol uses a target amount of each donor DNA of 15 fmol and 7.5 fmol for receiver plasmid DNA in a 10  $\mu$ L reaction. In order to perform accurate dispensing, the plasmid DNA is diluted to their corresponding concentration of 15 fmol/ $\mu$ L for donor parts and of 7.5 fmol/ $\mu$ L for the receiver plasmid, and then 1  $\mu$ L of each plasmid is added to the DNA mix. The plasmid part target concentration (in ng/ $\mu$ L) equals to each donor plasmid length / 100, and of the receiver plasmid length / 200.**

**Volumes CAN be reduced by half, but expect more variability from aliquoting.**

THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

<https://nph.onlinelibrary.wiley.com/doi/full/10.1111/nph.15625>

### PROTOCOL STATUS

#### Working

We use this protocol in our group and it is working

### GUIDELINES

Each level (odd and even) contains four 'receiver' plasmids:

pOdd-1, 2, 3, 4

pEven-1, 2, 3, 4

For the assembly of L0 parts into a L1 TU, a variable number of parts can be used for the assembly of the TU, however, for higher level assemblies only 4 donor modules can be assembled into any receiver.

Use only high-purity plasmid preparations. Genomic contamination of plasmid DNA will interfere with assembly reactions.

## MATERIALS

NAME	CATALOG #	VENDOR
T4 DNA Ligase Reaction Buffer - 6.0 ml	B0202S	New England Biolabs
BSA-Molecular Biology Grade - 12 mg	B9000S	New England Biolabs
BsaI - 1,000 units	R0535S	New England Biolabs
SapI - 250 units	R0569S	New England Biolabs
Ultrapure Distilled, Nuclease Free Water		
X-Gal	B4252	Sigma-aldrich
T4 DNA Ligase	M0202	New England Biolabs

## SAFETY WARNINGS

Avoid several freeze/thaw cycles and replace 10X T4 ligase buffer every month or so. Replace enzymes every 1 or 2 months. Protect X-Gal from light. DMF and DMSO are toxic, be careful when handling X-Gal.

## BEFORE STARTING

**Do not use BsaI-HF, that enzyme will NOT work with this reaction. Use either BsaI or BsaI-HFv2.**

**Thaw 10X T4 ligase buffer at RT and then leave on ice. Keep BSA on ice. Briefly vortex 10X ligase buffer before using. Use aliquoted volumes of T4 ligase Buffer since ATP and DTT will degrade over short amounts of time (less than a month) affecting the efficiency of the reaction.**

### Calculate plasmid target concentrations

- 1 The required concentration for each donor part in ng/μL is plasmid length / 100. That will yield a concentration of 15 fmol/μL. For the receiver plasmid the target concentration is length / 200, yielding 7.5 fmol/μL.

### Prepare DNA mixture

- 2 Add into a PCR tube:  
1 μL of each donor part at 15 fmol/μL.  
1 μL of receiver plasmid at 7.5 fmol/μL.

Add water up to 5 μL (assemblies from L1 and above do not need adjustment since 5 plasmids are used)

### Master mixes

- 3 Odd and even level assemblies use different formulations due to specific requirements of the restriction enzymes used.

For odd levels (BsaI or BsaI-HFv2), 10x T4 DNA ligase buffer is used.

For even levels (SapI), 10x Cutsmart buffer is used mixed with 10x T4 DNA ligase buffer in a 1:1 ratio.

For BSA (bovine serum albumin), check stock concentration. NEB 'molecular biology grade BSA' comes at 20 mg/mL, deprecated NEB purified BSA comes at 10 mg/mL. Prepare 20 μl of 1 mg/mL diluted BSA in water solution.

Thaw buffers at RT, then place on ice.

### IMPORTANT:

**Do not use BsaI-HF, that enzyme will NOT work with this reaction. Use either BsaI or BsaI-HFv2.**

Use aliquoted volumes of T4 Buffer since ATP and DTT will degrade over short amounts of time (less than a month) affecting the efficiency of the reaction.

If low efficiency is seen, use double the restriction enzyme volumes.

#### Prepare reaction master mix

- 4 Calculate the number of reactions to be performed and include an extra reaction to account for pipetting errors.

Per reaction:

Odd level BsaI-Ligase 2x master mix:

Reagent	Concentration	Volume (μL)
HPLC H <sub>2</sub> O		3
T4 DNA ligase buffer	10X	1
Diluted BSA (Bovine Serum Albumin)	1 mg/mL	0.5
T4 DNA ligase	400 U/μL	0.25
BsaI (or BsaI-HFv2)	10 U/μL	0.25

Even level SapI-Ligase 2x master mix:

Reagent	Concentration	Volume (μL)
HPLC H <sub>2</sub> O		3.5
Cutsmart buffer	10X	0.5
T4 DNA ligase buffer	10X	0.5
T4 DNA ligase	400 u/μL	0.25
SapI	10 U/μL	0.25

Mix by pipetting (enzymes in glycerol will be in the bottom of the tube). Then pipette from the top for accurate dispensing.

#### Mix DNA and reaction master mix

- 5 Add the 5 μL of the corresponding reaction master mix to the 5 μL of DNA mix and mix thoroughly by pipetting.

#### Incubate reaction

- 6 Place the reaction on a thermocycler (PCR machine). And run the following program:

Step 1 - 37 °C for 3 min  
Step 2 - 16 °C for 4 min  
Repeat 25x step 1 and 2  
Step 3 - 50 °C - 5 min  
Step 4 - 80 °C - 10 min

Note: number of cycles and incubation times can be adjusted but the reaction efficiency may decrease.

#### Transform and plate

- 7 Use 1 μL of reaction to transform chemically competent cells.

Recover in SOC for 1 hour incubating at 37 °C in a shaking incubator.

Prepare plates with respective antibiotics with 50 ug/mL of X-GAL and 1 mM IPTG.

#### Pick colony grow overnight and validate

- 8 Pick 2 white colonies for each construction and grow in 10 mL of LB + respective antibiotic and perform plasmid purification. Validate assembly by means of restriction digest. Suggested enzymes: ThermoFischer XbaI and PstI FastDigest enzymes. If higher concentrations are required, increase inoculation volume and perform Midi prep as directed by manufacturer's instructions.



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