



# Neuron-astrocyte culture preparation

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### ABSTRACT

Primary cortical neuronal-astrocyte cell culture preparation. Dissociation, preparation and plating of mice cortex neurons and glia cells on MFA.

**PROTOCOL STATUS** 

### Working

We use this protocol in our group and it is working

### Sample collection

1 Dissect cortices from pups and place them on ice.

NOTE

Cortical cultures were prepared from post-natal day 0 or 1 mice.

# Cell lysis

- 2 1. Chop with scissors in a papain-based dissociation buffer [2.5 mM CaCl2, 0.83 mM EDTA, 137 U papain (Sigma-Aldrich)], 100 μl DNAse (Sigma-Aldrich), 3-5 crystals of L-Cysteine (Sigma-Aldrich), HBSS with 20 mM HEPES (pH 7.4);
  - 2. Place on a rotating shaker at 50 rpm for 15 min **© 00:15:00** at room temperature.

# Cell preparation

- 3 1. After centrifuging (1000 rpm,1 min), discard the supernatant, resuspend the pallet in modified essential medium (MEM) without L-glutamine with essential amino acids (Beit Haemek, 06-1025-01-1A), 5% heat-inactivated fetal calf serum (Biological Industries), heat-inactivated 5% horse serum (Beith Haemek, 04-004-1), 2 mM glutamine (Beit Haemek, 03-020-1c), 3 mg/ml glucose, 2% B-27 (Gibco, 17504-044), 0.5% Pen/Strep (100 U/ml penicillin, 100 μg/ml streptomycin; Beit Haemek, 03-031-1B);
  - 2. Triturate seven times by slowly passing the tissue through a 10 mL serological pipette until the tissues are smaller than the opening of the 1 mL pipette tip.

# Cell cult ure

- 4. 1. Plate the cells on poly-D-lysine (PDL, Sigma-Aldrich, P7405-5MG) coated micro-electrode arrays (MEAs; 200/30iR-Ti-gr and 500/30iR-Ti-pr; Multichannel Systems) with a cell density of 2000-2500 cells/mm2 (~106 cells per dish);
  - 2. Cultures were maintained at 37 °C with 5% CO2. Partially replace growth medium every 3-4 days [MEM-EAGLE (without L-glutamine with essential amino acid), 5 mg/ml glucose, 5% heat-inactivated fetal calf serum, 0.8% GlutaMAX (100X; Gibco, 35050-038), 0.5% Pen/Strep, 2 mM glutamine, 2% B-27].

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