

FABP4 ELISA Protocol

Boya Li, Huixia Yang*, Wanyi Zhang, Shengtang Qin, Yundi Shi, Yumei Wei, Yingdong He, Wenshuang Yang, Shiju Jiang, Hongyan Jin

Abstract

Plasma FABP4 concentrations were analyzed blinded to their preeclampsia status with BioVendor Human AFABP ELISA (Cat. No: RD191036200R, Biovendor, Modrice, Czech Republic) in our study. The manufacturer reports a normal range of 19.58 ± 16.32 ng/mL (mean \pm 2 SD) for 35- to 52-year-old women. The limit of detection of the FABP4 ELISA assay is 0.05 ng/ml. The antibodies used in this ELISA are specific for human AFABP according to the manufacturer.

Citation: Boya Li, Huixia Yang*, Wanyi Zhang, Shengtang Qin, Yundi Shi, Yumei Wei, Yingdong He, Wenshuang Yang, Shiju Jiang, Hongyan Jin FABP4 ELISA Protocol. [protocols.io](https://doi.org/10.17504/protocols.io.md9c296)
dx.doi.org/10.17504/protocols.io.md9c296

Published: 23 Dec 2017

Materials

✓ BioVendor Human AFABP ELISA [Cat. No: RD191036200R](#) by Contributed by users

Protocol

Reconstitute Master Standard and prepare set of standards

Step 1.

Reconstitute the lyophilized Master Standard with Dilution Buffer just prior to the assay. Let it dissolve at least 15 minutes with occasional gentle shaking. The resulting concentration of the human AFABP in the stock solution is 25 ng/ml.

Prepare set of standards using Dilution Buffer as follows:

Volume of Standard Dilution Buffer Concentration

| | | |
|---------------------|--------|-----------|
| Stock | - | 25 ng/ml |
| 200 ml of stock | 300 ml | 10 ng/ml |
| 250 ml of 10 ng/ml | 250 ml | 5 ng/ml |
| 250 ml of 5 ng/ml | 250 ml | 2.5 ng/ml |
| 200 ml of 2.5 ng/ml | 300 ml | 1 ng/ml |
| 250 ml of 1 ng/ml | 250 ml | 0.5 ng/ml |

Prepare Quality Controls (QCs)

Step 2.

Dilute reconstituted Quality Controls 10x with Dilution Buffer.

Prepare the working Biotin Labelled Antibody solution

Step 3.

Adding 1 part Biotin Labelled Antibody Concentrate (100x) with 99 parts Biotin-Ab Diluent.

Prepare Dilute Wash Solution Concentrate (10x)

Step 4.

Dilute Wash Solution Concentrate (10x) ten-fold in distilled water to prepare a 1x working solution.

Prepare samples

Step 5.

Dilute plasma samples 10x with Dilution Buffer just prior to the assay

Work sheet

Step 6.

Pipet 100 ul of diluted Standards, Quality Controls, Dilution Buffer (=Blank) and samples, preferably in duplicates, into the appropriate wells.

| | strip 1+2 | strip 3+4 | strip 5+6 | strip 7+8 | strip 9+10 | strip 11+12 |
|---|--------------|-----------|-----------|-----------|------------|-------------|
| A | Standard 25 | QC HIGH | Sample 8 | Sample 16 | Sample 24 | Sample 32 |
| B | Standard 10 | Sample 1 | Sample 9 | Sample 17 | Sample 25 | Sample 33 |
| C | Standard 5 | Sample 2 | Sample 10 | Sample 18 | Sample 26 | Sample 34 |
| D | Standard 2.5 | Sample 3 | Sample 11 | Sample 19 | Sample 27 | Sample 35 |
| E | Standard 1 | Sample 4 | Sample 12 | Sample 20 | Sample 28 | Sample 36 |
| F | Standard 0.5 | Sample 5 | Sample 13 | Sample 21 | Sample 29 | Sample 37 |
| G | Blank | Sample 6 | Sample 14 | Sample 22 | Sample 30 | Sample 38 |
| H | QC LOW | Sample 7 | Sample 15 | Sample 23 | Sample 31 | Sample 39 |

Assay procedure

Step 7.

1. Incubate the plate at room temperature (ca. 25°C) for 1 hour, shaking at ca. 100 rpm.
2. Wash the wells 5-times with Wash Solution (0.35 ml per well).
3. Add 100 ul of Biotin Labelled Antibody into each well.
4. Incubate the plate at room temperature (ca. 25°C) for 1 hour, shaking at ca. 100 rpm.
5. Wash the wells 5-times with Wash Solution (0.35 ml per well).
6. Add 100 ul of Streptavidin-HRP Conjugate into each well.
7. Incubate the plate at room temperature (ca. 25°C) for 30 minutes, shaking at ca. 100 rpm.
8. Wash the wells 5-times with Wash Solution (0.35 ml per well).
9. Add 100 ul of Substrate Solution into each well. .
10. Incubate the plate for 10 minutes at room temperature. No shaking
11. Stop the colour development by adding 100 ul of Stop Solution.
12. Determine the absorbance of each well using a microplate reader set to 450 nm, preferably with the reference wavelength set to 630 nm. Subtract readings at 630 nm (550-650 nm)

from the readings at 450 nm.