

Euplotes crassus transfection using Lipofectamine 2000 as vehicle Version 5

RACHELE CESARONI

Abstract

Citation: RACHELE CESARONI Euplotes crassus transfection using Lipofectamine 2000 as vehicle. protocols.io

dx.doi.org/10.17504/protocols.io.hhhb336

Published: 30 Mar 2017

Protocol

Step 1.

Collect 4×10^4 well-fed Euplotes crassus cells (we used E. coli as the only food source) by centrifugation at 400 rcf for 3 minutes.

Step 2.

Wash the cells twice with artificial sea water (see attachment for the recipe) and once with 500 mM sorbitol, 0.5 mM Tris-HCl pH 7.0 (400 rcf for 3 minutes each time). Then resuspend Euplotes crassus cells in 50 μ l of the medium (500 mM sorbitol, 0.5 mM Tris-HCl pH 7.0). To get this small volume you may require an additional minute of centrifugation.

NOTES

Rachele Cesaroni 30 Mar 2017

Recipe for complete sea water (1 L):

36 g Reef Crystals 1 ml Walne's solution 1 ml of 10 μg/ml FeSO₄

0.2 ml of 2 mg/ml thiamine (light sensîtive; store at 4 $^{\circ}$ C) Add distilled water up to 1 L

Step 3.

Transfer the cells into wells within a 96-well plate for transfection.

Step 4.

Dilute 2.5 μ l of Lipofectamine 2000 Reagent in 25 μ l of the same medium of the cells (500 mM sorbitol, 0.5 mM Tris-HCl pH 7.0).

Step 5.

Dilute 5 μ g of DNA dissolved in MilliQ H₂O (0.5-5 μ g/ μ l) in 125 μ l of the same medium of the cells (500 mM sorbitol, 0.5 mM Tris-HCl pH 7.0).

Step 6.

Add 25 μ l of the diluted DNA to 25 μ l of the diluted Lipofectamine 2000 Reagent (1:1 ratio) and incubate them for 10 minutes at room temperature (before incubation mix them by pipetting up and down 5 times or vortex them for 10 seconds).

Step 7.

Add 10 μ l of the transfection complexes drop-wise to the 50 μ l of Euplotes crassus cells in medium (gently swirl the dish to ensure uniform distribution of the transfection complexes).

Step 8.

One hour after addition of Lipofectamine 2000 complexes, add 50 μ l of artificial sea water to the cells.

Step 9.

After another hour, add other 50 µl of artificial sea water to the cells.

Step 10.

An hour later harvest the cells (400 rcf for 3 minutes) and wash them twice with artificial sea water (400 rcf for 3 min each time). Then resuspend them in 400 μ l of the artificial sea water.

Step 11.

Transfer the cells into depression wells.

Step 12.

Incubate the cells at 24 °C, then analyze them by fluorescence microscopy to determine gene expression.