

CITE-seq Protocols

Marlon Stoeckius, Peter Smibert

Abstract

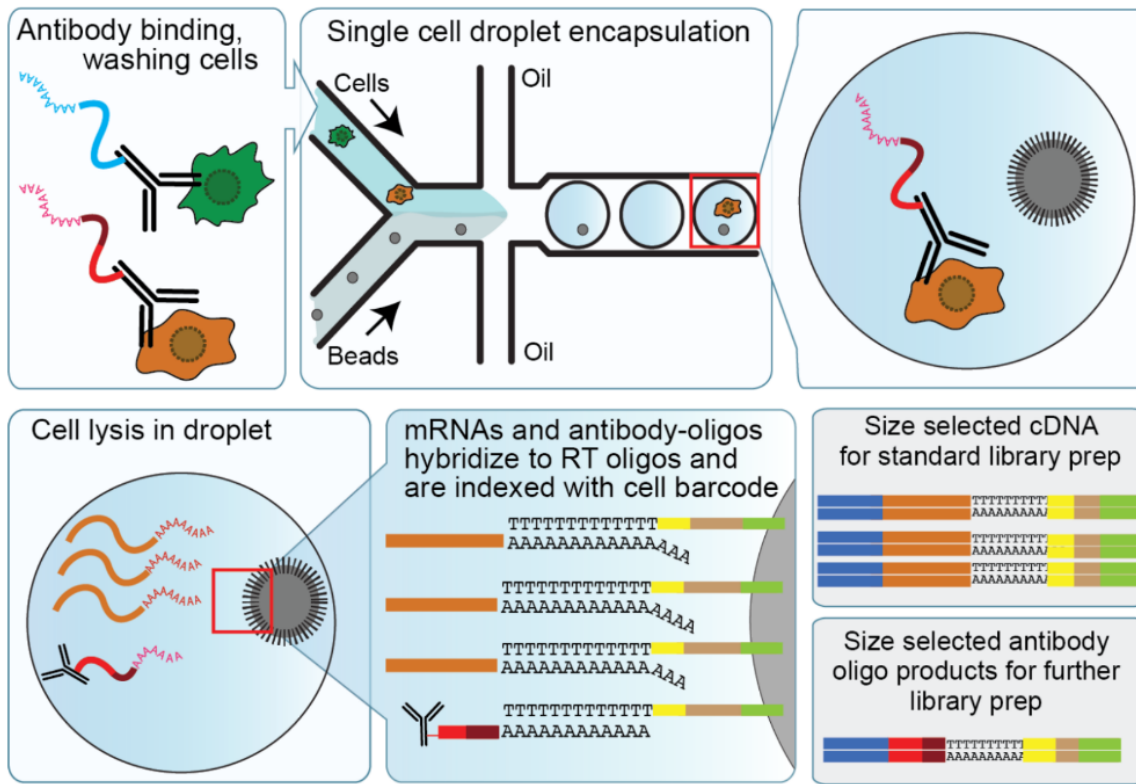
This collection contains our main protocols for performing CITE-seq and Cell Hashing, specifically on [Drop-seq](#) or [10x Genomics single cell 3' v2 chemistry](#).

CITE-seq:

Cellular Indexing of Transcriptomes and Epitopes by Sequencing ([CITE-seq](#)) is a multimodal single cell phenotyping method developed in the [Technology Innovation lab](#) at the New York Genome Center in collaboration with the Satija lab.



CITE-seq uses DNA-barcoded antibodies to convert detection of proteins into a quantitative, sequenceable readout. Antibody-bound oligos act as synthetic transcripts that are captured during most large-scale oligodT-based scRNA-seq library preparation protocols (e.g. 10x Genomics, Drop-seq, ddSeq).

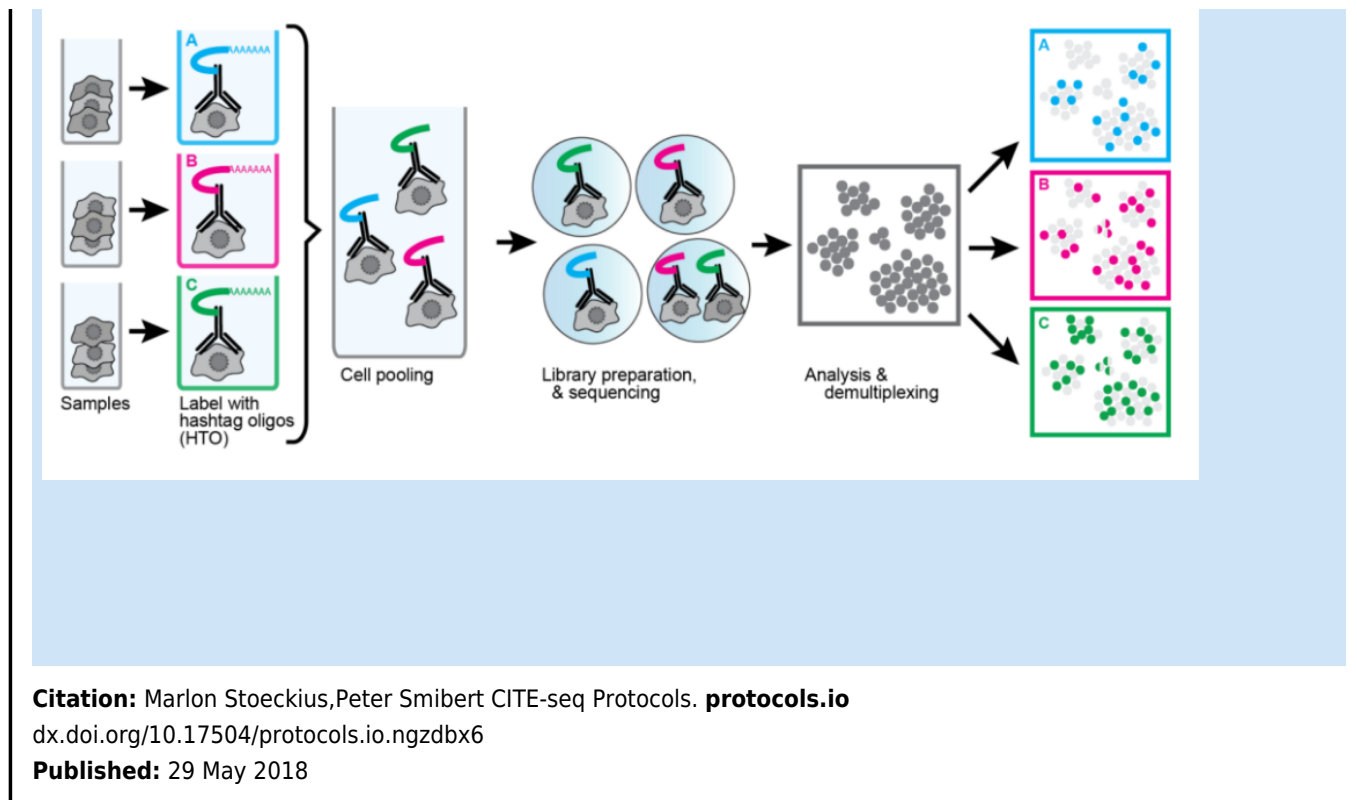


This allows for immunophenotyping of cells with a potentially limitless number of markers and unbiased transcriptome analysis using existing single-cell sequencing approaches.

Cell Hashing:

Sample multiplexing and super-loading on single cell RNA-sequencing platforms.

[Cell Hashing](#) uses a series of oligo-tagged antibodies against ubiquitously expressed surface proteins with different barcodes to uniquely label cells from distinct samples, which can be subsequently pooled in one scRNA-seq run. By sequencing these tags alongside the cellular transcriptome, we can assign each cell to its sample of origin, and robustly identify doublets originating from multiple samples.



Collection

📄 PROTOCOLS

1. [CITE-seq](#)

CONTACT: [Peter Smibert](#)

2. [Cell Hashing](#)

CONTACT: [Peter Smibert](#)

3. [CITE-seq and Cell Hashing](#)

CONTACT: [Peter Smibert](#)