

# Measles Vaccine Virus Taqman-MGB

Mitchell Finger, Judy Northill, Ian Mackay

## Abstract

This previously unpublished protocol aims to amplify genotype A measles virus (MeV) strains but not non-measles viruses.

Mitchell Finger and Michael Lyon developed this in-house test in 2010.

The assay targets the intergenic region between the M (matrix) and F (fusion) genes, designed as a qualitative test for investigating measles vaccine virus (MVV) strains.

Numbering indicates the oligonucleotide location on the sequence with MeV strain Edmonston (Moraten vaccine), complete genome, GenBank accession number AF266287.

**Citation:** Mitchell Finger, Judy Northill, Ian Mackay Measles Vaccine Virus Taqman-MGB. **protocols.io**

dx.doi.org/10.17504/protocols.io.rs2d6ge

**Published:** 23 Aug 2018

## Before start

- If using a different brand or model of real-time thermocycler, check the concentration of ROX is adequate.
- Method assumes the user is familiar with the thermocycler and software used to run the protocol and with PCR in general.

## Protocol

### Oligonucleotide sequences

#### Step 1.

Name	Sequence 5'-3'
Measles F 4729 Vac	AAACCCCCAGCAATTGGAA
Measles R 4795 Vac	GGTCACCTCGGTCGCTTGT
Measles Probe 4757	FAM - CCCTCTTCCTCAACACA - MGBNFQ

### Reagents

#### Step 2.



## Reaction set-up

### Step 3.

The assay has been used on both a Rotor-Gene 6000 and a Rotor-Gene Q real-time thermocycler

Prepare sufficient mix for the number of reactions.

Include a suitable 'dead volume' as necessary if using a robotic dispenser.

### MIX PREPARATION

Reagent	Volume (µl) x1	Final reaction concentration
Nuclease-free water	4.42	N/A
Measles F 4729 Vac 150pmol/µl	0.04	300nM
Measles R 4795 Vac 150pmol/µl	0.04	300nM
Measles Probe FAM 100pmol/µl	0.06	300nM
2X Reaction Mix <sup>1</sup>	10	1X
SuperScript® III/Platinum® Taq Mix <sup>1</sup>	0.4	1X
ROX Reference Dye (25µM)	0.04	0.05µM
Template	5	N/A
<b>TOTAL</b>	<b>20</b>	

<sup>1</sup>Superscript™III Platinum™ One-step qRT-PCR kit

- Dispense 15µL to each reaction well.
- Add 5µL of template (extracted RNA, controls or NTC [nuclease-free water] ).
- Total reaction volume is 20µL

## Amplification

### Step 4.

### CYCLING CONDITIONS

50°C	5min	1X
95°C	2min	1X
95°C	3sec	40X
60°C	30sec <sup>1</sup>	

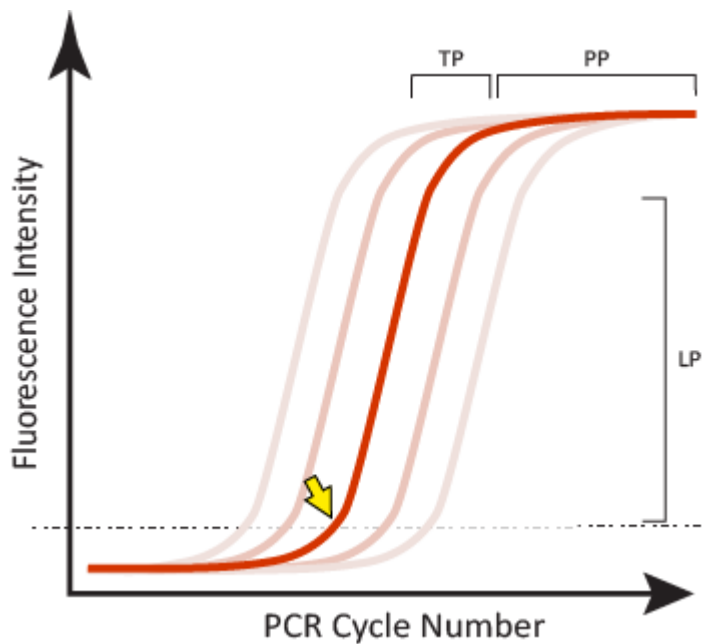
<sup>1</sup>Fluorescence acquisition step

## Result Analysis

### Step 5.

The definition used for a satisfactory positive result from a real-time fluorogenic PCR should include each of the following:

1. A **sigmoidal curve** – the trace travels horizontally, curves upward, continues in an exponential rise and followed by a curve towards a horizontal plateau phase
2. A **suitable level of fluorescence** intensity as measured in comparison to a positive control (y-axis)
3. A **defined threshold ( $C_T$ ) value** which the fluorescent curve has clearly exceeded (Fig.1 arrow), which sits early in the log-linear phase and is  $<40$  cycles
4. A flat or non-sigmoidal curve or a curve that crosses the threshold with a  $C_T >40$  cycles is considered a negative result.
5. NTCs should not produce a curve



**Figure 1.** Examples of satisfactory sigmoidal amplification curve shape when considering an assay's fluorescent signal output. The crossing point or threshold cycle ( $C_T$ ) is indicated (yellow arrow); it is the value at which fluorescence levels surpass a predefined (usually set during validation, or arbitrary) threshold level as shown in this normalized linear scale depiction. LP-log-linear phase of signal generated during the exponential part of the PCR amplification; TP-a slowing of the amplification and accompanying fluorescence signal marks the transition phase; PP-the plateau phase is reached when there is little or no increase in fluorescent signal despite continued cycling.