

Stranded Transcript Count Table Generation from Long Reads

Forked from Transcript Coverage Analysis from Long Reads

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dx.doi.org/10.17504/protocols.io.xu4fnyw





ABSTRACT

This protocol is for comparing different samples at the transcript level, using long reads that are mapped to transcripts.

Input(s): demultiplexed fastq files (see protocol Demultiplexing Nanopore reads with LAST), transcript reference fasta file, annotation file

Output(s): transcript table, sorted by differential coverage, annotated with gene name / description / location

PROTOCOL STATUS

In development

We are still developing and optimizing this protocol

BEFORE STARTING

Obtain a transcript fasta file, and an annotation file. For the mouse genome, I use the following files:

- 1. Transcript [CDS] sequences from Ensembl; this file was the most current when I last checked.
- 2. Annotation file obtained from Ensembl BioMart (Ensembl Genes -> Mouse Genes) as a compressed TSV file with the following attribute columns:
- Transcript stable ID
- Gene description
- Gene start (bp)
- Gene end (bp)
- Strand
- Gene name
- Chromosome/scaffold name

Barcode Demultiplexing

Demultiplex reads as per protocol <u>Demultiplexing Nanopore reads with LAST</u>.

If this has been done, then the following command should produce output without errors:

for bc in \$(awk '{print \$2}' barcode_counts.txt); do Is reads_\${bc}.fastq.gz; done

Example output:

reads_BC03.fastq.gz reads_BC04.fastq.gz reads_BC05.fastq.gz reads_BC06.fastq.gz reads_BC07.fastq.gz

reads_BC08.fastq.gz

✓ protocols.io 02/07/2019 If the barcode_counts.txt file is missing, the output will look like this:

awk: fatal: cannot open file `barcode_counts.txt' for reading (No such file or directory)

If one or more of the barcode-demultiplexed files are missing, the output will look something like this:

reads_BC03.fastq.gz
reads_BC04.fastq.gz
reads_BC05.fastq.gz
ls: cannot access 'reads_BC06.fastq.gz': No such file or directory
ls: cannot access 'reads_BC07.fastq.gz': No such file or directory
reads_BC08.fastq.gz

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