

## RNA Isolation from Plant Tissue Protocol 15: Hot Acid Phenol Method for Algae

1 Works for me

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**ABSTRACT** 

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This RNA isolation method is modified from that described by Köhrer and Domdey<sup>5</sup>.

This protocol is part of a collection of eighteen protocols used to isolate total RNA from plant tissue. (RNA Isolation from Plant Tissue Collection: <a href="https://www.protocols.io/view/rna-isolation-from-plant-tissue-439gyr6">https://www.protocols.io/view/rna-isolation-from-plant-tissue-439gyr6</a>)

<sup>5</sup> Kohrer, K. & Domdey, H. Preparation of high molecular weight RNA. Methods in Enzymology 194, 398-405 (1991).

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MATERIALS TEXT

## Reagents

## Extraction Buffer:

- 1 % SDS (v/v, starting from 10 % SDS stock solution)
- 51 mM sodium acetate pH 5.5
- 10 mM EDTA
- DEPC treated water



The final reaction buffer was filter purified using Nalgene 0.22  $\mu$ M filter.

## Other reagents:

- Acid phenol (pH 4.3)
- Phenol:chloroform (5:1) acid equilibrated to pH 4.7 from Sigma
- Isopropanol
- 70 % ethanol (diluted in DEPC treated water H<sub>2</sub>0)
- 3 M Sodium acetate pH 5.5

SAFETY WARNINGS

Please see SDS (Safety Data Sheet) for hazards and safety warnings.

Preheat phenol and phenol:chloroform to 8 65 °C. Heated phenol should not be re-used.

2 Collect algae cells via centrifugation for © 00:10:00 at @16100 x g at & Room temperature. 2.1 Flash freeze pellets with liquid nitrogen and keep at 8-80 °C until extractions are carried out. 3 Re-suspend frozen pellet in  $\blacksquare 800 \mu I$  of preheated extraction buffer. Immediately add 300 µl of hot acid phenol. 4.1 Vortex the tubes for **© 00:00:15**. Incubate at 8 65 °C for © 00:10:00. Vortex every 1 min for © 00:00:10. Centrifuge at  $\textcircled{3}16100 \times g$  at 4 °C for 000:05:00. The aqueous phase was transferred to fresh 1.5 ml micro-centrifuge tube. Repeat steps 5 - 7. Repeat 3x (depending on the amount of cells used). 5 go to step #5 Extract with equal volume of phenol:chloroform (5:1). 9 Vortex for © 00:01:00 at & Room temperature. 9.2 Spin for ( 00:05:00 in a microcentrifuge at top speed. 9.3 Repeat step 9 three times. 🤚 go to step #9

Transfer aqueous phase to a new 1.5 ml microfuge tube. Volume should be  $\sim 1.5$  ml microfuge tube.

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- 11.1 Hold at δ 4 °C for © 00:30:00 or more.

  12 Spin in micro-centrifuge at δ 4 °C at top speed for © 00:20:00.

  13 Remove the supernatant without dislodging the pellet.

  14 Wash the pellet with 70 % ethanol.

  15 Invert and air dry tubes at room temperature.

  16 The pellet was re-suspended in □50 μl DEPC treated H<sub>2</sub>O.
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