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## Stranded Mapping from Oriented Long Reads V.5 👄

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#### **ABSTRACT**

This protocol demonstrates how to map strand-oriented long reads to a genome, and visualise them in a genome browser.

The general idea is to use minimap 2 to create stranded BAM files, which are split for forward/reverse orientation then converted into BigWig format for display in a genome browser.

### Input(s):

- stranded fastq files (see protocol <u>Preparing Reads for Stranded Mapping</u>)
- a FASTA file containing the genome / sequence of interest.

# Output(s):

- Genome-mapped stranded BAM files
- Genome-mapped stranded BigWig files

**EXTERNAL LINK** 

https://bioinformatics.stackexchange.com/a/3922/73

### BEFORE STARTING

You will need access to the following free and open-source software program(s):

- minimap2
- samtools

And the following additional data file(s):

• a FASTA file containing the genome / sequence of interest.

# Orient Reads

1 Orient reads as per protocol Preparing Reads for Stranded Mapping.

If this has been done, then the following command should produce output without errors:

```
for bc in $(awk '{print $2}' barcode_counts.txt);
  do ls oriented/${bc}_reads_dirAdjusted.fq.gz;
done
```

## Example output:

```
oriented/BCO3_reads_dirAdjusted.fq.gz
oriented/BCO4_reads_dirAdjusted.fq.gz
oriented/BCO5_reads_dirAdjusted.fq.gz
oriented/BCO6_reads_dirAdjusted.fq.gz
oriented/BCO7_reads_dirAdjusted.fq.gz
oriented/BCO8_reads_dirAdjusted.fq.gz
```

**Index Preparation** 

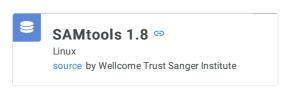
2 Prepare genome index for spliced alignment



```
minimap2 -d mmus_ucsc_all-splice.idx -Q -t 10 -x splice mmus_ucsc_all.fa
```

### Read Mapping

3 Map the long reads to the genome using minimap2, using samtools to covert to a sorted BAM format. This is where the reverse complementing done during demultiplexing gives a big saving of effort.



# Creating BigWig Coverage Files

4 mpileupDC.pl

A bedGraph of coverage is created using samtools mpileup and mpileupDC.pl, excluding any skipped intronic sequence. When 'mpileupDC.pl' is provided with a single file, it will output a bedGraph file with a header line starting with '##'; this header line is removed. The particular JBrowse plugin that I use for stranded display requires that the reverse strand have negative coverage values, so that file needs to be changed:

```
for bc in $(awk '{print $2}' barcode_counts.txt);
  do echo ${bc};
  samtools view -b -F 0x10 mapped/mm2_called_${bc}_vs_MmusG.bam | \
     samtools mpileup -A -B -Q 0 -q 0 -I -q 0 -Q 0 - | \
     mpileupDC.pl | tail -n +2 > mapped/mm2_called_${bc}_vs_MmusG.bg.plus
  samtools view -b -f 0x10 mapped/mm2_called_${bc}_vs_MmusG.bam | \
     samtools mpileup -A -B -Q 0 -q 0 -I -q 0 -Q 0 - | \
     mpileupDC.pl | tail -n +2 > mapped/mm2_called_${bc}_vs_MmusG.bg.minus
     perl -i -pe 's/([0-9]+)$/-$1/' mapped/mm2_called_${bc}_vs_MmusG.bg.minus
```

done;

5 Stranded bedgraph files are converted to bigwig. This requires BEDTools and a genome information file containing chromosome lengths (one for Mmus/mm10 is attached to this step).



### BEDTools 2.26.0 ©

source by Quinlan laboratory, University of Utah

```
for bc in $(awk '{print $2}' barcode_counts.txt);
  do echo ${bc};
  basename="mapped/mm2_called_${bc}_vs_MmusG"
  bedGraphToBigWig ${basename}.bg.plus Mmus_genome.chrInfo.txt ${basename}.bw.plus
  bedGraphToBigWig ${basename}.bg.minus Mmus_genome.chrInfo.txt ${basename}.bw.minus
  done
```

Mmus\_genome.chrlnfo.txt

# JBrowse Configuration

6 Each track should have its own JBrowse configuration section using the StrangedBigWig class and StrandedXYPlot type. An example is shown here:

```
[tracks.BWCG004-4T1-BC04-both-track]
storeClass
              = StrandedPlotPlugin/Store/SeqFeature/StrandedBigWig
urlTemplate
               = bw/mm2_called_CG004_BC04_vs_MmusG.bw
               = MinION - Coverage
category
               = StrandedPlotPlugin/View/Track/Wiggle/StrandedXYPlot
type
key
               = MinION minimap2 coverage from CG004-4T1-WT (combined strands)
               = log
scale
scoreType
               = maxScore
               = global
autoscale
style.pos_color = darkred
style.neg_color = darkgreen
```

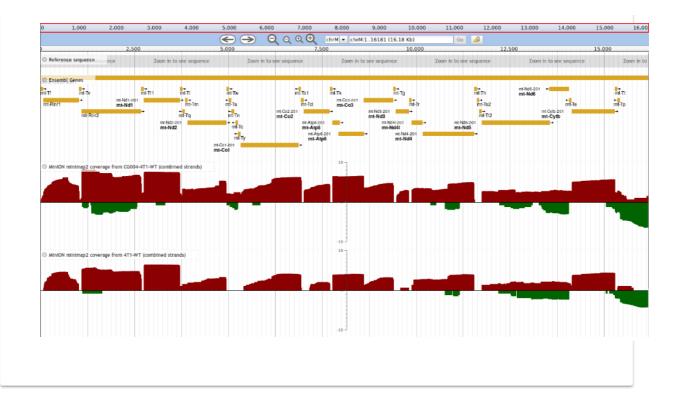
# Sanity Check

7 If this has worked properly, then mapping human or mouse to the mitochondrial genome should show most expression appearing on the positive strand, with a small scattering of negative-strand expression, a bit like the Expected Results shown here.

If not, check for the following issues:

- Tracks not displaying at all in JBrowse -- make sure track IDs inside square brackets are of the form [ tracks.<unique-id-without-dots>-track]
- JBrowse track is reflected in the X axis -- make sure that the reverse bedgraph file is orientated the correct way; it should be created with the '-f 0x10' flag (no capitalisation).
- JBrowse track only shows one direction -- make sure that the reverse bedgraph file has negative values, and re-generate the bigwig file





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