

Transform Stratagene's XL-10 Gold Ultracompetent cells

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Abstract

Protocol for transforming XL-10 GOld Ultracompetent cells from Stratagene (now Agilent). Protocol adopted from manufacturer's instructions.

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Guidelines

Use of 14-ml BD Falcon polypropylene round-bottom tubes: It is important that 14-ml BD Falcon polypropylene round-bottom tubes (BD Biosciences Catalog #352059) are used for the transformation protocol, since other tubes may be degraded by \hat{l}^2 -mercaptoethanol. In addition, the duration of the heat pulse has been optimized using these tubes.

Aliquoting Cells: Keep the cells on ice at all times during aliquoting. It is essential that the polypropylene tubes are placed on ice before the cells are thawed and that the cells are aliquoted directly into pre-chilled tubes. It is also important to use 100 $\hat{l}\frac{1}{4}$ of cells per transformation. Decreasing the volume will reduce efficiency.

Use of Î²-Mercaptoethanol (**Î²-ME**): **Î²-ME** has been shown to increase transformation efficiency. The **Î²-ME** mixture provided is diluted and ready to use. Stratagene cannot guarantee results with **Î²-ME** from other sources.

Use of NZY+ Broth: Transformation of the supplied ultracompetent cells has been optimized using NZY+ as the medium for outgrowth following the heat pulse. Substitution with another outgrowth medium may result in a loss of efficiency.

Quantity and Volume of DNA: The greatest efficiency is obtained from the transformation of $1 \hat{1}\frac{1}{4}$ of 0.01 ng/ $\hat{1}\frac{1}{4}$ supercoiled pUC18 DNA per 100 $\hat{1}\frac{1}{4}$ of cells. When transforming a ligation mixture, add $2 \hat{1}\frac{1}{4}$ of the ligation mixtureper 100 $\hat{1}\frac{1}{4}$ of cells. A greater number of colonies may be obtained by transforming up to 50 ng DNA, although the resulting efficiency (cfu/ $\hat{1}\frac{1}{4}$ g) may be lower. The volume of the DNA solution added to the reaction may be increased to up to 10% of the reaction volume, but the transformation efficiency may be reduced.

Heat Pulse Duration and Temperature: Optimal transformation efficiency is observed when cells are heat-pulsed at $42\hat{A}$ °C for 30 seconds. Efficiency decreases sharply when cells are heat-pulsed for <30 seconds or for >40 seconds. Do not exceed $42\hat{A}$ °C.

Plating the Transformation Mixture: If plating <100 $\hat{i}\frac{1}{4}$ I of cells, pipet the cells into a 200 $\hat{i}\frac{1}{4}$ I pool of medium and then spread the mixture with a sterile spreader. If plating $\hat{a}_{\parallel} \neq 100 \hat{i}\frac{1}{4}$ I, the cells can be spread on the plates directly. Tilt and tap the spreader to remove the last drop of cells. If desired, cells may be concentrated prior to plating by centrifugation at1000 rpm for 10 minutes followed by resuspension in 200 $\hat{i}\frac{1}{4}$ I of NZY+ medium.

Protocol

Step 1.

Pre-chill 14mL sterile culture tubes on ice

Step 2.

Pre-heat 0.9mL of NZY+ broth to 42°C

■ AMOUNT

1 ml Additional info:



NZY broth by Contributed by users

Step 3.

Thaw XL-10 Gold Ultracompetent cells on ice then mix gently after completely thawed



XL-10 Gold Ultracompetent cells 200314 by Agilent Technologies

Step 4.

Aliquot cells into pre-chilled sterile culture tubes

■ AMOUNT

100 μl Additional info:



XL-10 Gold Ultracompetent cells 200314 by Agilent Technologies

Step 5.

Add beta-mercaptoethanol

AMOUNT

4 μl Additional info:



XL-10 Gold Beta-mercaptoethanol mix by Agilent Technologies

Step 6

Swirl tube gently then incubate on ice for 10 minutes swirling gently every 2 minutes

O DURATION

00:10:00

Step 7.

Add 0.1-50ng of DNA or 2uL of ligation product

■ AMOUNT

2 μl Additional info:

Step 8.

Swirl gently then incubate on ice for 30 minutes

O DURATION

00:30:00

Step 9.

Heat pulse tube at 42°C for exactly 30 seconds. The duration of the heat pulse is critical.

O DURATION

00:00:30

Step 10.

Incubate cells on ice for 2 minutes

© DURATION

00:02:00

Step 11.

Add 0.9mL of pre-heated NZY+ broth to each tube



1 ml Additional info:

Step 12.

Incubate at 37°C for 1 hour with shaking at 225-250 rpm

© DURATION

01:00:00

Step 13.

Plate no more than 200uL of transformation mixture per LB-agar plate with antibiotic selection

Step 14.

Incubate plates at 37°C overnight.

© DURATION

15:00:00