

Preparation of Denaturing Agarose Gel for RNA Analysis

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Abstract

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Protocol

Gel Preparation

Step 1.

Melt 1.0 g agarose in 87 ml of DEPC water, by dispersing the agarose uniformly and heating in a microwave until all particles are dissolved.

Gel Preparation

Step 2.

Bring the melted agarose to 60 °C.

Gel Preparation

Step 3.

Add 10 ml 10X MOPS Buffer and 3 ml 37% formaldehyde.

✓ PROTOCOL

. [10X MOPS Buffer](#)

CONTACT: [Irina Agarkova](#)

⊕ NOTES

Irina Agarkova 31 Mar 2016

Use DEPC treated water and RNase free reagents.

Step 3.1.

10X MOPS consists of the following (per liter):

Step 3.2.

0.2 M MOPS pH 7 with NaOH

Step 3.3.

50 mM sodium acetate

Step 3.4.

10 mM EDTA

Gel Preparation

Step 4.

FORMALDEHYDE IS VOLATILE AND TOXIC. WORK IN A HOOD FROM THIS POINT FORWARD.

Gel Preparation

Step 5.

Pour standard agarose gel. USE A FUME HOOD!

Gel Preparation

Step 6.

Allow gel to set for 1 hour.

 DURATION

01:00:00

Gel Preparation

Step 7.

Run gel in 1X MOPS Buffer.

 PROTOCOL

. 10X MOPS Buffer

CONTACT: [Irina Agarkova](#)

Step 7.1.

10X MOPS consists of the following (per liter):

Step 7.2.

0.2 M MOPS pH 7 with NaOH

Step 7.3.

50 mM sodium acetate

Step 7.4.

10 mM EDTA

Sample Preparation

Step 8.

Mix 40 µl sample buffer with 10 µl sample, heat to 55 °C 15 minutes.

 DURATION

00:15:00

 NOTES

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The sample buffer is comprised of 65% formamide, 22% formalin (37% formaldehyde) and 13% 10X MOPS.

Sample Preparation

Step 9.

Add 10µl of the following:

- 50% glycerol
- 1 mM EDTA
- 0.3% each bromophenol blue and 0.3% xylene cyanol.

Warnings

Note: RNA is subject to rapid degradation by RNase present in the environment. For optimal results, use water treated with DEPC.