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Gel Electrophoresis

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ABSTRACT

How to prepare and run gel electrophoresis using agarose gel for the verification of nucleic acid amplification products.

GUIDELINES

Gloves should be worn at all times.

MATERIALS TEXT

- 50x TAE Buffer
- Agarose
- DNA ladder
- Loading dye (optional)
- Nucleic acid amplification solution (e.g. PCR or LAMP)
- Ethidium bromide
- Deionized water

SAFETY WARNINGS

Gel bottle will be hot from microwave, use a hot mitt and be cautious to not get hot gel on your skin. Ethidium bromide may be a mutagen; skin contact should be avoided.

Make TAE buffer

- 1 In a large glass bottle, add **750 ml** DI water.
- 2 Add 15 ml 50x TAE buffer to the deionized water.
- 3 Cap the bottle and mix by swirling

Make gel

- 4 Pour **□300 ml** TAE buffer into microwavable glass bottle.
- Add the following amount of agarose powder to TAE buffer for different gel concentrations:

Concentration (w/v)	Agarose (g)
2.0%	6
2.5%	7.5
3%	9

Swirl solution lightly to mix agarose into buffer. Swirling too vigorously will introduce bubbles that will make cas difficult.	ting
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- 6 Microwave in © 00:00:40 intervals until completely dissolved, swirling bottle between heating intervals.
- Pour into gel mold with well insert. Add to black line (~ \(\subseteq 100 ml \))
- 8 Allow gel to set for approximately **© 00:20:00**.

Load and run gel

- Q Transfer gel boat with gel into electrophoresis chamber.
- 10 Add buffer to line marked on chamber, completely covering the gel.
- 11 Carefully load the wells with ladder (green) and amplification product solutions at volumes between $\frac{1}{2}$ 1 μ 1 and $\frac{1}{2}$ 5 μ 1.
 - Optionally, you can mix 1 µl loading dye with 4 µl amplification product solution in a separate microcentrifuge tube to make the loading process easier to visualize.
- 12 Put the lid on the chamber, turn on the power supply and press 'RUN'. Run for \bigcirc 00:50:00.
 - The power supply does not turn off automatically, so set a timer and shut off after © 00:50:00 . This time may change based on factors such as gel concentration and amplicon length.

Stain the gel

Transfer the gel into the dying tray. 13 Check buffer level and add more if necessary. 14 15 Add 38 µl ethidium bromide to dying tray. 16 Turn on stage, and allow to stain for **© 00:20:00**. Image gel 17 Place gel in imaging machine. Turn on the machine by the switch on left side of machine. Press 'Trans UV'. 18 Take a picture with the camera. 19 (cc) BY ■ This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited