

# Isotopic labeling of freshwater mixotrophic algae using isotopic labeled heat-killed bacteria

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## Abstract

This protocol aims to achieve an isotopic labeling of mixotrophic algae using  $^{15}\text{N}$  and  $^{13}\text{C}$  labeled heat-killed bacteria. It was developed to be used with the mixotrophic chrysophyte *Ochromonas* to assess the sources of carbon and nitrogen of this algae when growing mixotrophically. Sources of nitrogen and carbon in the media used were reduced to 1 inorganic source ( $\text{NH}_4\text{Cl}$  for nitrogen and bicarbonate for carbon) and 1 organic source (heat-killed bacteria); media used for *Ochromonas* growth was modified accordingly to reflect this.

Downstream analysis of isotopic labeled *Ochromonas* include bulk IRMS measurements or nanoSIMS measurements.

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## Protocol

### Step 1.

## Prepare modified DY-V media

Media used for *Ochromonas* growth was a modified version of DY-V media. Modifications were:

- No MES was used, as it can be used as a source of carbon (bicarbonate and heat-killed bacteria are the only sources of carbon used in this media)
- No  $\text{Na}_2\text{SiO}_3$  was used (*Ochromonas* does not need silica)
- No  $\text{NaNO}_3$  (ammonium and heat-killed bacteria were the only sources of nitrogen used in this media)
- Sodium bicarbonate added at a final concentration of 95  $\mu\text{M}$ . Bicarbonate addition is done before algal inoculum by 0.2  $\mu\text{m}$  filtration of a stock solution (do not autoclave).

LINK:

<https://ncma.bigelow.org/media/pdf/NCMA-algal-medium-DY-V.pdf>

## Step 2.

### Media inoculation with labeled HKB

Add  $15\text{N}$   $13\text{C}$  labeled heat-killed bacteria (HKB) to the media (isolation and labeling of HKB was done [following this protocol](#)).

Add an inoculum of the mixotrophic algae (volume of inoculum should be low as to avoid dilution of the isotopic label and carry over of nutrients).

NOTE: as an example, values commonly used were  $5 \times 10^3$  *Ochromonas*  $\text{mL}^{-1}$  and  $5 \times 10^7$  HKB  $\text{mL}^{-1}$  as starting concentrations for the cultures.

## Step 3.

### Mixotrophic growth of the algae

Let the algae grow so it incorporates the isotopic signature in its biomass. Track the algae growth through microscopy (live samples and/or fixed samples) and the decline of heat-killed bacteria (fixed samples and staining with DAPI to assess HKB concentrations through epifluorescence microscopy).

## Step 4.

### Sampling for assessment of algal isotopic signature

*Ochromonas* was allowed to grow for 2-3 generations before sampling for isotopic signature. Two kind of samples can be collected:

- Bulk measurements: filter 30-50 mL of the cultures onto pre-combusted glass fiber filters and dry at  $60\text{ }^{\circ}\text{C}$  over night to stop all biological activity. Afterwards, filters can be stored in glass vials at room temperature. Further processing of the sample included an acidification step with HCl to remove inorganic carbon and the C- and N- isotopic composition of the sample was determined by an isotope ratio mass spectrometer (IRMS).
- Cell-specific measurements: collect 2 mL of sample and fix with 2X EM-grade glutaraldehyde. Sample can be stored in the fridge at  $4\text{ }^{\circ}\text{C}$ . Further processing of the sample involves the deposition of cells onto silicon wafers, wash with MQ-water and dry; map cells on the wafer

using microscopy and analysis of single cells using a Cameca NanoSIMS 50 instrument (NOTE: actual manipulation of the machine will be done by an expert user).