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Working

## UC Davis - IN-VIVO Glucose-stimulates Insulin Secretion Test 👄

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**ABSTRACT** 

## Summary:

An in-vivo glucose-stimulates insulin secretion test is designated to determine alterations in insulin secretion by the pancreas upon a bolus IP Glucose injection.

**EXTERNAL LINK** 

https://mmpc.org/shared/document.aspx?id=86&docType=Protocol

## MATERIALS

NAME ~	CATALOG #	VENDOR ~	CAS NUMBER $\vee$ RRID $\vee$
45% Glucose solution	NC0025179(50-165-7017 replaced)	Fisher Scientific	
Insulin Syringes	14-826-79	Fisher Scientific	
Saline Solution	L97753	Fisher Scientific	
Ultra Sensitive Mouse Insulin ELISA kit	90080	Crystal chem	AB_2783626
Heparin Sodium	401586B	Abraxis	
Easy Check Glucose test strips	00-101( new SKU 88982400)	JRS Medical	
Easy Check Glucose monitor	Y4209 (new SKU 88972401)	JRS Medical	

MATERIALS TEXT

Dilute the glucose stock solution (45%) with saline to 20% by adding 20ml stock to 25ml 0.9% (w/v) sterile saline.

## Note:

Fisher Scientific, RRID:SCR\_008452

Ultra Sensitive Mouse Insulin ELISA kit, Citethis, (Crystal Chem Cat# 90080, RRID:AB\_2783626)

- Fast mice for 16 hours by taking away food the day before (3:00pm)
- The following day, Calibrate the glucose meter according to the manufacturer's instructions.

Deprive mice from water then measure blood glucose level using a glucometer and remove immediately approximately 50 µl of blood from

- the tail via a tail tip cut and transfer directly onto a sterile 0.5ml microcentrifuge tube containing 2ul Heparin.
  Centrifuge at 8000rpm for 5min then transfer plasma (supernatant) to a new 0.5ml microcentrifuge tube and freeze at -80°C.
  Give the mouse an intraperitoneal injection of Glucose (2g/kg) with a 27 G needle
- 7 NOTE:

At the end of the experiment, wipe tail with 70% alcohol and allow drying. Ensure that blood loss from the tail stopped before placing the animal back to its cage.

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Continue to take blood samples from the initial tail cut at 2, 5, 15 and 30 min flowing injection and repeat step3 and 4.