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Working

## UC Davis - HDL Protocol [↗](#)

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[dx.doi.org/10.17504/protocols.io.ynsfvee](https://doi.org/10.17504/protocols.io.ynsfvee)

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### ABSTRACT

#### Summary:

LDL and VLDL are separated from HDL using a precipitation reagent. Then the HDL fraction is measured for either TC or TG using the same reagents for total cholesterol or triglyceride.

### EXTERNAL LINK

<https://mmpc.org/shared/document.aspx?id=93&docType=Protocol>

### MATERIALS

NAME	CATALOG #	VENDOR	CAS NUMBER	RRID
Calibrator	<a href="#">TR43002</a>	<a href="#">Fisher Diagnostics</a>		
TC Reagents	<a href="#">TR13421</a>	<a href="#">Fisher Diagnostics</a>		
TG Reagents	<a href="#">TR22421</a>	<a href="#">Fisher Diagnostics</a>		
2X LDL/VLDL Precipitation Buffer	<a href="#">ab105138</a>	<a href="#">Abcam</a>		
PBS				
Microplate				
Platereader				

### MATERIALS TEXT

#### Note:

Fisher Scientific, [RRID:SCR\\_008452](#)

Abcam, [RRID:SCR\\_012931](#)

- 1 Add 25µl 2X precipitation buffer to 25µl of sample using a positive displacement pipet.
- 2 Vortex and let sit at RT for 10 minutes.
- 3 Centrifuge at 2000×g for 10 minutes at 4°C.

4 Pipet supernatant into new tube, this is the HDL fraction.

5 Add 5 µl of calibrator and sample to each well.

**IMPORTANT: Make sure not to add any bubbles to the wells when dispensing reagents, this will interfere with reading in the platereader.**

6 Add 300 µl of TC or TG reagent to each well. Incubate at 37°C for 5 minutes. Read at 540 nm.

**IMPORTANT: If samples are hemolyzed, pipet a blank well with 5µl sample and 300µl PBS**

7 Subtract blank readings from final readings. The assay will be linear so the unknown samples can be calculated as (Sample Absorbance ÷ Calibrator Absorbance) × Calibrator Concentration.

8 HDL samples are diluted ½ so multiply these by 2 to get the final value. Subtract this from the total triglyceride or cholesterol value to get the LDL/VLDL value.



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