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Handling of genomics samples

In 1 collection

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ABSTRACT

This protocol describes the handling of genomics samples for the *Tara* Oceans expedition and is part of [Viral to metazoan marine plankton nucleotide sequences from the *Tara* Oceans expedition](#).

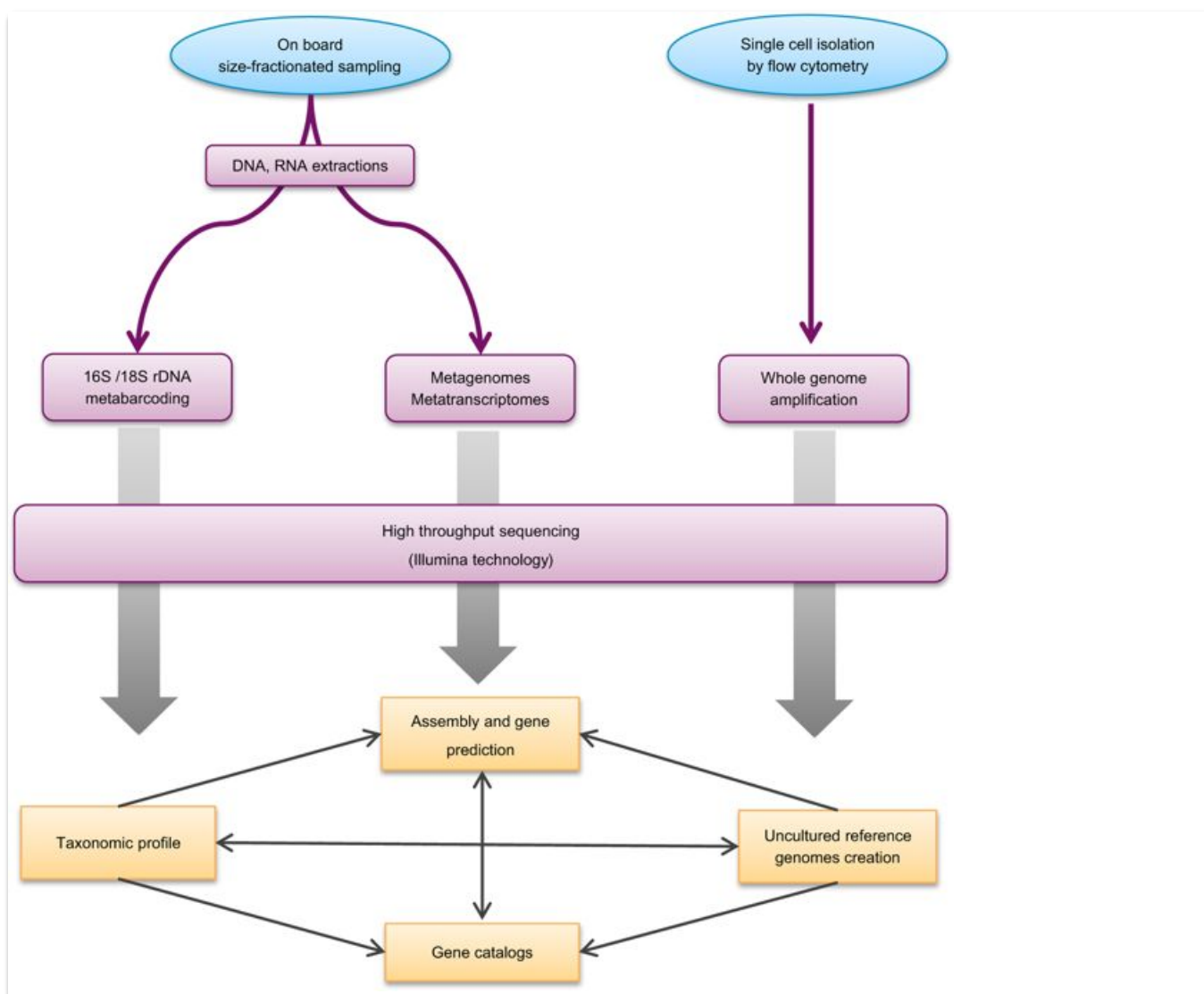


Figure 1: Overview of -omics analysis strategy applied on Tara Oceans samples.

EXTERNAL LINK

<https://www.nature.com/articles/sdata201793#methods>

THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

Alberti, A. (2017). Viral to metazoan marine plankton nucleotide sequences from the Tara Oceans expedition. *Scientific Data***4**, 170093 (2017)
doi: [10.1038/sdata.2017.93](https://doi.org/10.1038/sdata.2017.93)

ATTACHMENTS

[Viral to metazoan marine plankton nucleotide sequences from the Tara Oceans expedition.pdf](#)

GUIDELINES

Genomics samples were transferred on average every 6 weeks from a port of call to the European Molecular Biology Laboratory (EMBL, Heidelberg) in Germany. Transportation was organized by experts from World Courier (www.worldcourier.com) who ensured that the chain of cold was never broken. At EMBL, samples were sorted, repackaged according to their final destination, and transported again by World Courier to the different laboratories responsible for their analysis (Table 1).

Table 1: Summary of libraries generated from Tara Oceans DNA and RNA samples and sequencing experiments performed on each type of library.

Size fractions (μm)	Mainly targeted organisms	Targeted genomic analysis	Sample storage laboratory	Sequencing laboratory	Method ID Nucleic acids preparation (Section)	Method ID Amplicons generation (Section)	Method ID Library preparation (Section)
< 0.2 μM	Viruses	Metagenomics	M. Sullivan lab (University of Arizona, AZ, US)	CEA, Genoscope, France	Virus_DNA_ext (2.4)		MetaG_virus (4.2)
0.2–1.6, 0.1–0.2, 0.45–0.8, 0.2–0.45	Giruses	Metagenomics	N. Grimsley lab (CNRS, Banyuls-sur -Mer, France)	CEA, Genoscope, France	Girus_DNA_ext (2.5)		MetaG (4.1)
0.2–1.6, 0.2–3	Viruses, Giruses, Prokaryotes, small Eucaryotes	16S metabarcoding	S.G. Acinas lab (ICM-CSIC, Barcelona, Spain)	CEA, Genoscope, France	Acinas_Prok_DNA_ext (2.2)	16S_PCR (3.2)	MetaBar_16S (4.6)
		Metagenomics			Acinas_Prok_DNA_ext (2.2)		MetaG (4.1)
		Metatranscriptomics by random priming			Acinas_Prok_RNA_ext Genoscope_Prok_RNA_ext (2.2)		RiboZero_SMART_strand (4.4)
0.8-inf, 3-inf, 0.8–5 (0.8–3), 5–20 (3–20), 20–180, 180–2,000	Protists and metazoa	18S metabarcoding	C. De Vargas lab (CNRS/UPMC, Roscoff, France)	CEA, Genoscope, France		18S_PCR (3.1)	MetaBar_18S (4.5)
		16S metabarcoding				16S_PCR (3.2)	MetaBar_16S (4.6)
		Metagenomics	P. Wincker lab (CEA, Genoscope, France)		Euk_ DNA_RNA_ext (2.1)		MetaG (4.1)
		Metatranscriptomics on poly(A) ⁺ RNA			Euk_ DNA_RNA_ext (2.1)		TS_RNA (4.4) TS_strand (4.4) SMART_dT (4.4)
Samples for SAGs	Protists	De novo sequencing	N. Poulton lab (Bigelow lab, ME, US)	CEA, Genoscope, France	SAGs_amplif (2.6)		MetaG_SAGs (4.3)

*Number of libraries with available readsets in public databases at the date of publication of the paper

In the respective laboratories, samples were immediately identified by scanning/reading their barcode label and were stored in cryo boxes or in –80 °C freezers. During all these steps, samples were manipulated on dry ice. Each laboratory used its own sample management system to record the storage location and to monitor sample usage.



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