

# Fixation of Planktonic Samples

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## Abstract

Modified after Glöckner et al. 1999

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## Guidelines

Needed:

- PFA Fixative
- Water sample
- Formaldehyde (optional)
- Moistened support filter (0.45 µm pore size, cellulose nitrate, 47 mm diameter)
- Membrane filter (0.2 µm pore size, white polycarbonate (GTTP), 47 mm diameter)
- Filtration tower
- Vacuum
- Moistened support filter (0.45 µm pore size, cellulose nitrate, 25 mm diameter)
- Membrane filter (0.2 µm pore size, white polycarbonate (GTTP), 25 mm diameter)
- Sterile H<sub>2</sub>O
- Petri dish lid
- Parafilm

## Protocol

### Step 1.

Add freshly prepared PFA fixative (see '[Fixing Cells with PFA Protocol](#)') to water sample to a final concentration of 1-2%.

Alternatively, use Formaldehyde, and fix for 12-24 hours at 4°C.

 **DURATION**

12:00:00

### Step 2.

**Place a moistened support filter** (0.45 µm pore size, cellulose nitrate, 47 mm diameter) **and a membrane filter** (0.2 µm pore size, white polycarbonate (GTTP), 47 mm diameter) **into a filtration tower**

### Step 3.

Filter an appropriate volume of the fixed sample by applying gentle vacuum ( 5 inch)

 **NOTES**

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Support filters may be utilized for several samples; for cell numbers of around  $10^6$  per ml, 10 ml of sample is generally sufficient

**Step 4.**

**Place a moistened support filter** (0.45  $\mu$ m pore size, cellulose nitrate, 25 mm diameter; Sartorius, Germany) **and a membrane filter** (0.2  $\mu$ m pore size, white polycarbonate (GTTP), 25 mm diameter; Millipore, Eschborn, Germany) **into a filtration tower**

**Step 5.**

Filter 1 ml of the fixed sample by applying gentle vacuum (5 inch)

 **AMOUNT**

1 ml Additional info:

 **NOTES**

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The support filter may be utilized for both samples

**Step 6.**

Filter 2 ml of the fixed sample on a second filter.

 **AMOUNT**

2 ml Additional info:

 **NOTES**

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The support filter may be utilized for both samples

**Step 7.**

After complete sample filtration, wash with 10-20 ml of sterile H<sub>2</sub>O

 **AMOUNT**

15 ml Additional info:

**Step 8.**

Remove H<sub>2</sub>O by filtration

**Step 9.**

Put membrane filter on blotting paper for drying and cover, e.g. with the lid of a cryo box or a Petri dish.

 **NOTES**

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If this is not available use kim wipes

**Step 10.**

Allow air-drying

**Step 11.**

Store each filter in a separate labelled Petri dish

**Step 12.**

Place membrane filter between separator sheets that the GTTP membrane filters were provided with

 **NOTES**

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This will prevent the membrane filter from sticking to the Petri dish.

**Step 13.**

Seal Petri dish with parafilm

**Step 14.**

Store at -20°C until processing

**📌 NOTES**

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Filters can be stored frozen for several months without apparent loss of hybridization signal.