

# RNA Extraction Protocol from RNA-SIP Experiments

Julie Huber

## Abstract

This is the protocol to extract RNA from Sterivex filters from RNA-SIP experiments carried out with seawater, vent fluids, etc.

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## Protocol

### Step 1.

Wipe bench top, pipets, racks, with Nucleo-clean, 70% ethanol, and RNAzap wipes.

### Step 2.

Prepare 15ml tube, add 750ul Lysis Buffer and 0.5ml of RNA Powersoil beads (1/2 of tube).

### Step 3.

Thaw Sterivex on ice.

### Step 4.

Using sterile pliers or pipe cutters (both ethanol-flamed) crack Sterivex cartridge.

### Step 5.

Pipette or pour off RNAlater from cartridge into a new 2ml tube, spin down cells at 12,000xg for 5min, pour off RNAlater, place tube on ice.

### Step 6.

Cut filter off of cartridge using razor blade (autoclaved, ethanol-flamed), and place on sterile/autoclaved piece of foil.

### Step 7.

Using sterile, ethanol-flamed razor blade and forceps cut filter into 6-10 pieces and place into prepared 15ml tube.

### Step 8.

Add 250ul Lysis buffer to 2ml tube with spun down cells, resuspend by vortexing, add this to 15ml tube.

### Step 9.

Using adaptor, vortex 15ml tube for 7-10min, at medium-high speed.

### Step 10.

Add 100 ul Homogenate additive and place on ice for 10min.

### Step 11.

Centrifuge tube at 4000 x g for 2min at 4°C. 10. Remove lysate and place in 2ml tube. Repeat centrifugation if necessary to remove all lysate.

**Step 12.**

Add 1 part acid:phenol chloroform to tube (equal to amount of lysate removed) and centrifuge at 10,000xg for 5min. Follow procedure for using phenol, use fume hood, double glove, respirator etc.

**Step 13.**

Remove top aqueous layer and place in fresh 2ml tube. Be sure not to touch the bottom layer!

**Step 14.**

Heat the elution solution to 95°C in heat block (use a small aliquot in a 1.5mL tube).

**Step 15.**

Add 1.25x 100% ethanol. Mix well, and add to filter cartridge in a collection tube provided. Centrifuge at 10,000xg for 15sec, discard the flow through. Filter cartridge can only hold 700ul, so might need to do a few spins.

**Step 16.**

Add 700ul Wash Solution 1, spin at 10,000xg for 15sec, discard flow through.

**Step 17.**

Add 500ul Wash Solution 2/3, spin at 10,000xg for 15sec, discard flow through.

**Step 18.**

Repeat Step 16.

**Step 19.**

Put the filter cartridge back into the tube, centrifuge for 1.5 min at 10,000xg to remove residual fluid from the filter.

**Step 20.**

Transfer the filter cartridge into a fresh collection tube, apply 50 µl pre-heated (95 °C) Elution solution to the center of the filter, spin for 30 sec at 10,000 X g; repeat with another 50 µl pre-heated (95°C) Elution Solution.20. Freeze at -80°C long term. Short term (24-48hrs), RNA can be stored at -20°C.

**Step 21.**

DNase Treatment: adapted from Ambion Turbo-DNAase kit. Set heat block to 37°C. Transfer 100ul of sample from mirVana collection tube to 1.5 ml centrifuge tube.

**Step 22.**

Add 10ul 10x Buffer to 100ul RNA.

**Step 23.**

Add 1ul Turbo DNase, incubate @ 37°C for 20min.

**Step 24.**

Add another 1ul of Turbo DNase, incubate for 20min more.

**Step 25.**

Add 10ul Inactivation Reagent and vortex on and off for 5 min.

**Step 26.**

Spin tubes @ 10,000xg for 1.5min.

**Step 27.**

Pipet RNA into new 0.5ml tube, be sure NOT to touch inactivation reagent.

**Step 28.**

Freeze at -80°C long term. Short term (24-48hrs), RNA can be stored at -20°C.