Digestion Notl

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Abstract

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Protocol

Step 1.

Add:

- nuclease-free water qsp 50 μ L > 47 μ L
- 5µL 10X Buffer O
- $1\mu L$ DNA ($1 \mu g/\mu L$)
- $-1 \mu L Notl 1 (10U/\mu L) *$
- Mix gently and spin down for a few seconds.

Step 2.

Incubate at 37°C for 1-16 hours. The digestion reaction may be scaled either up or down > It's possible to process extended incubation by adding 0.25U/μg of DNA in 50 μL of reaction volume.

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Step 3.

incubation at 80°C for 20 min.

Step 4.

- 1. Measure the volume of the DNA sample.
- $> 50 \mu L$
- 2. Add 1/10 volume of sodium acetate, pH 5.2, (final concentration of 0.3 M) These amounts assume that the DNA is in TE only; if DNA is in a solution containing salt, adjust salt accordingly to achieve the correct final concentration.
- $> 5 \mu L$
- 3. Mix well.
- 4. Add 2 to 2.5 volumes of cold 100% ethanol (calculated after salt addition).
- > 110 (2V)
- 5. Mix well.
- 6. Place on ice or at -20 degrees C for >20 minutes.

- 7. Spin a maximum speed in a microfuge 10-15 min.
- 8. Carefully decant supernatant.
- 9. Add 1 ml 70% ethanol. Mix. Spin briefly. Carefully decant supernatant.
- 10. Air dry or briefly vacuum dry pellet.
- 11. Resuspend pellet in the appropriate volume of TE or water.