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TotalSeq™-A Antibodies Protocol with 10x Single Cell 3' Reagent Kit v2 [↗](#)Sam Li<sup>1</sup><sup>1</sup>BioLegend

1 Works for me

[dx.doi.org/10.17504/protocols.io.8vyhw7w](https://doi.org/10.17504/protocols.io.8vyhw7w)

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## ABSTRACT

Buyer is solely responsible for determining whether Buyer has all intellectual property rights that are necessary for Buyer's intended uses of the BioLegend TotalSeq™ products. For example, for any technology platform Buyer uses with TotalSeq™, it is Buyer's sole responsibility to determine whether it has all necessary third party intellectual property rights to use that platform and TotalSeq™ with that platform.

The following protocol combines highly multiplexed protein marker detection with unbiased transcriptome profiling for thousands of single cells. Epitope detection is enabled by BioLegend's TotalSeq™ antibody-oligo conjugates. TotalSeq™ reagents are compatible with Cellular Indexing of Transcriptomes and Epitopes by sequencing (CITE-seq), RNA Expression And Protein sequencing assay (REAP-seq), and similar workflows.

Please read the entire protocol before starting the experiments.

## EXTERNAL LINK

<https://www.biolegend.com/en-us/protocols/totalseq-a-antibodies-protocol-with-10x-single-cell-3-reagent-kit-v2>

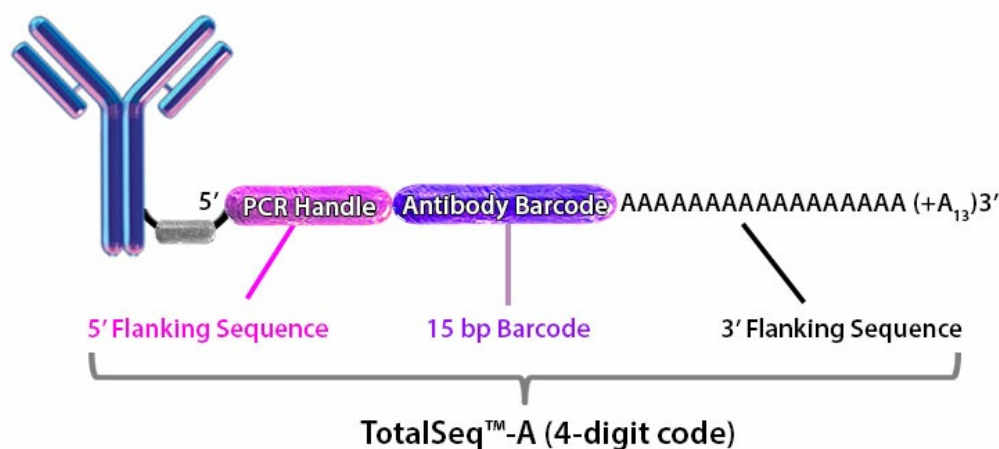
## GUIDELINES

**Notes:**

Oligonucleotide sequences:

TotalSeq™ antibodies. Each clone is barcoded with a unique oligonucleotide sequence. These contain standard small TruSeq RNA read 2 sequences and can be amplified using Illumina's TruSeq Small RNA primer sets (RPIx – primers, see example RPI1 below)

CCTTGGCACCCGAGAATTCCAACAAGACCTTGAGBAAAAAAAAAAAAAAAAAAAAAAAAAAAAA\*<sub>13</sub>A.



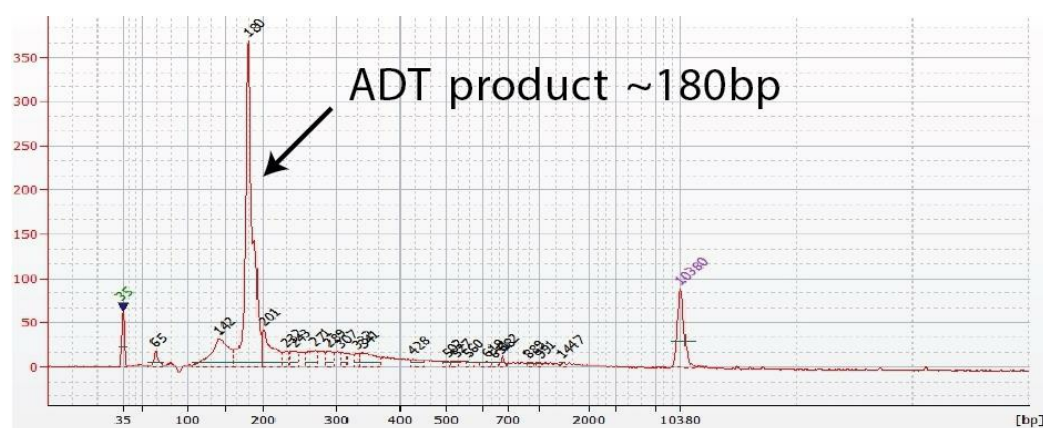
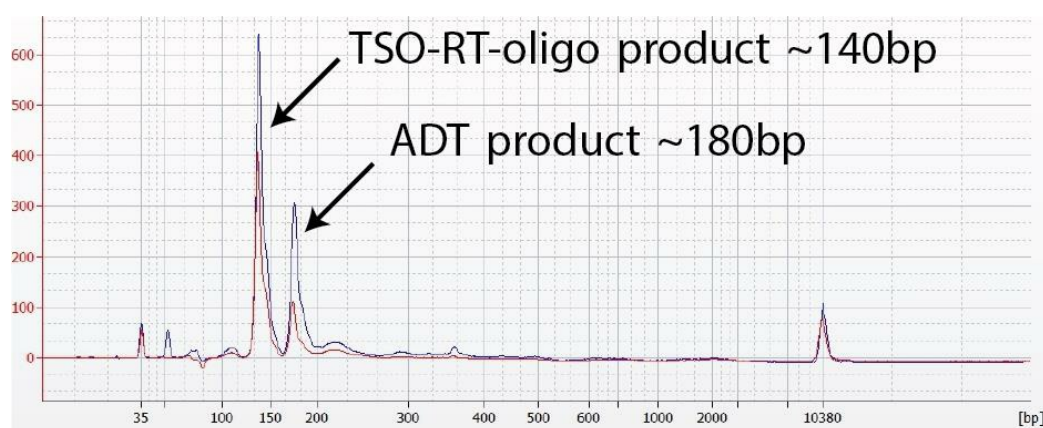
Please visit <https://www.biolegend.com/totalseq> for detailed information:

Oligos required for ADT library amplification:

- Drop-seq P5-SMART-PCR hybrid primer (for Drop-seq only)  
5'AATGATACGGCGACCACCGAGATCTACACGCTGTCCGCGGAAGCAGTGGTATCAACGCAGAGT\*A\*C
- 10x Genomics SI-PCR primer (for 10x Single Cell) 5'AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGC\*T\*C
- ADT cDNA PCR additive primer 5'CCTTGGCACCCGAGAATT\*C\*C
- Illumina Small RNA RPI1 primer (for ADT amplification; i7 index 1, Oligonucleotide sequences, Illumina)  
5'CAAGCAGAAGACGGCATACGAGATCGTGATGTGACTGGAGTTCCTTGGCACCCGAGAATTC\*C\*A

\* indicates a phosphorothioate bond

B indicates C or G or T; not A nucleotide



## MATERIALS TEXT

- TotalSeq™ antibody-oligo conjugates
- Biotinylated antibody and oligo [barcoded](#) streptavidin
- Human TruStain FcX™ (Fc Receptor Blocking Solution) (Cat# 422301/422302)
- 8-strip PCR tubes, emulsion safe (TempAssure PCR 8-strips, USA Scientific, Cat# 1402-4700)
- Nuclease-Free Pipette Tips (e.g. ThermoFischer Scientific AM12650, AM12660 or equivalent)
- Bioanalyzer chips and reagents (DNA High Sensitivity and small RNA kit, Agilent Cat# 5067-4626 and 5067-1548)
- Quantabio sparQ PureMag Beads (Quantabio, Cat# 951960) or SPRIselect reagent (Beckman Coulter, Cat# B23317)
- E-Gel™ EX Agarose Gels, 4% (ThermoFischer Scientific Cat# G401004)
- DNA LoBind Tubes (Eppendorf, Cat# 022431021)
- PCR Thermocycler (Bio-Rad, T100™ Thermal Cycler)
- TruStain fcX™ (anti-mouse CD16/32) Antibody (Cat#101319/101320)
- TruStain FcX™ PLUS (anti-CD16/32, recommended for mouse cells. BioLegend Cat# 156603)
- KAPA HiFi HotStart ReadyMix (2X) (Kapa Biosystems, Cat# KK2601)
- Specific primers, see “Notes” at the end of the protocol (primers not available from BioLegend, order from preferred supplier)
- Magnetic tube rack (e.g. ThermoFischer)
- Qubit (ThermoFischer Scientific, Cat# Q33226)
- Fuchs-Rosenthal Counting Chamber (Hemocytometer, VWR, Cat# 15170-230)
- Phosphate Buffered Saline (PBS) (BioLegend, Cat# 420501 or equivalent)
- Cell Staining Buffer (BioLegend, Cat# 420201)
- 80% Ethanol
- Flowmi™ Cell Strainer (Bel-Art, H-B Instrument, Cat# H13680-0040)

### I) Cell staining for Drop-seq or 10x Genomics platforms

- 1 Carefully count all cells to ensure accurate quantitation.
  - Make note of cell viability (>95%) and also include dead cells in the total cell count.
  - If high cell death is observed, live cell enrichment (e.g. by Flow Cytometry) is recommended.
- 2 Resuspend 1-2 million cells in 100 µl Cell Staining Buffer.
- 3 Add 5 µl of Human TruStain FcX™ Fc Blocking reagent.
- 4 Incubate for 5 – 10 minutes at 4°C.
- 5 While cells are incubating in Fc Block, prepare antibody-pool using 1 µg (or titrated amounts) of each TotalSeq™ and/or hashtag or biotinylated antibody.
- 6 To maximize performance, centrifuge the antibody pool at 14,000xg at 2 - 8°C for 10 minutes before adding to the cells.
- 7 Carefully pipette out the liquid, avoiding the bottom of the tube, and add the TotalSeq™ antibody cocktail to the cell suspension.
- 8 Incubate for 30 minutes at 4°C.

- 9 Wash cells 2 times with 1 mL PBS, spin 5 minutes 350g at 4°C.  
\*Please note that it has been observed in some cases that various factors, including cell/sample type, tube manufacturer, rotor type, wash buffer, etc., may result in an excessive number of cells coating the side of the tube. Please ensure that staining and washing conditions are appropriate for your sample type.
- 10 If using biotinylated antibodies, incubate with the appropriate oligo [barcoded](#) streptavidin at the recommended amount specified in the product technical datasheet for 20 minutes.
- 11 Wash cells 2 times with 1 mL PBS, spin 5 minutes 350g at 4°C.
- 12 Resuspend cells in PBS at appropriate concentration for downstream application. (e.g. for 10x Genomics 500 cells/μl; for Drop-seq 200 cells/μl).
- 13 Filter cells through 40 μm strainers.
- 14 Verify cell concentration by counting on hemocytometer after filtration.

II) Run Drop-seq (Macosko et al., 2015) or 10x Genomics single cell 3' v2 assay as described until before cDNA amplification.

15 **At cDNA amplification step:**

Add "additive" primers (0.2 μM) to cDNA PCR to increase yield of Antibody Derived Tag (ADT, cDNA derived from the TotalSeq™ antibodies) products, 1 μl (for 10x Genomics) or 0.4 μl (for Drop-seq). See notes at the end of the protocol for further details. Subtract the total volume of additive primer from the water added to the PCR reaction.

Please see example table below of 10X cDNA amplification reaction table (section 2.2, Chromium Single Cell 3' v2 Reagent Kit, Quick Reference Card (v2 Chemistry))

cDNA Amplification Mix	1X (μL)
Nuclease-Free Water	7
Amplification Master Mix	50
cDNA Additive	5
cDNA Primer Mix	2
ADT Additive Primer (0.2 μM stock)	1
<b>Total</b>	<b>65</b>

See notes in the Guidelines section or on our [website](#) for further details on primer sequences.

III) ADT and mRNA library preparation

- 16 **A) After cDNA amplification: Separate ADT-derived cDNAs (180bp) and mRNA-derived cDNAs (>300bp).**
  - 16.1 Perform sparQ or SPRI selection to separate mRNA-derived and ADT cDNAs. Follow manufacturer's recommendations.

- 16.2 Do not discard supernatant from 0.6X sparQ or SPRI. This contains the ADTs.
- Add 0.6X sparQ or SPRI (60 µl, based on 100 µl sample volume) to cDNA reaction as described in 10x Genomics or Drop-seq protocol.
  - Incubate 5 minutes and place on magnet.
  - Supernatant contains ADTs.
  - Beads contain full length mRNA-derived cDNAs.

17 **B) mRNA-derived cDNA >300bp (beads fraction).**

Proceed with standard 10x Genomics or Drop-seq protocol for cDNA sequencing library preparation.

18 **C) ADTs 180bp (supernatant fraction).**

18.1 1) Purify ADTs using two 2X sparQ or SPRI purifications per manufacturer protocol

- Add 1.4X sparQ or SPRI (140 µl, based on 100 µl sample volume) to supernatant to obtain a final sparQ or SPRI volume of 2X sparQ or SPRI.
- Transfer entire volume into a low-bind 1.5mL tube.
- Incubate 10 minutes at room temperature.
- Place tube on magnet and wait 2 minutes until solution is clear.
- Carefully remove and discard the supernatant.
- Add 400 µl 80% Ethanol to the tube without disturbing the pellet and stand for 30 seconds (only one Ethanol wash).
- Carefully remove and discard the ethanol wash.
- Centrifuge tube briefly and return it to magnet.
- Remove and discard any remaining ethanol.
- Resuspend beads in 50 µl water.
- Perform another round of 2X sparQ or SPRI purification by adding 100 µl sparQ or SPRI reagent directly onto resuspended beads.
- Mix by pipetting, and incubate 10 minutes at room temperature.
- Place tube on magnet and wait 2 minutes until solution is clear.
- Carefully remove and discard the supernatant.
- Add 200 µl 80% Ethanol to the tube without disturbing the pellet and stand for 30 seconds (1st Ethanol wash).
- Carefully remove and discard the ethanol wash.
- Add 200 µl 80% Ethanol to the tube without disturbing the pellet and stand for 30 seconds (2nd Ethanol wash).
- Carefully remove and discard the ethanol wash.
- Centrifuge tube briefly and return it to magnet.
- Remove and discard any remaining ethanol and allow the beads to air dry for 2 minutes (do not over dry beads).
- Resuspend beads in 45 µl water.
- Pipette mix vigorously and incubate at room temperature for 5 minutes.
- Place tube on magnet and transfer clear supernatant into PCR tube.

## 18.2 2) Amplify ADT sequencing library

Prepare 100µL PCR reaction with purified ADTs. For primer sequences please see "Notes" at the end of the protocol:

- 45 µl purified ADT fraction
- 50 µl KAPA HiFi HotStart ReadyMix (2X).
- 2.5 µl Truseq Small RNA RPLx primer (containing i7 index) 10 µM.
- 2.5 µl P5 oligo at 10 µM depending on application:
  - For Drop-seq use P5-SMART-PCR hybrid oligo.
  - For 10X use SI PCR oligo.

Cycling conditions:

95°C	3 min	
<b>95°C</b>	<b>20 sec</b>	
<b>60°C</b>	<b>30 sec</b>	<b>6-10 cycles</b>
<b>72°C</b>	<b>20 sec</b>	
72°C	5 min	

## 18.3 3) Purify PCR product using 1.6X sparQ or SPRI purification by adding 160 µl sparQ or SPRI reagent.

- Incubate 5 minutes at room temperature.
- Place tube on magnet and wait 1 minute until solution is clear.
- Carefully remove and discard the supernatant.
- Add 200 µl 80% Ethanol to the tube without disturbing the pellet and stand for 30 seconds (first Ethanol wash).
- Carefully remove and discard the ethanol wash.
- Add 200 µl 80% Ethanol to the tube without disturbing the pellet and stand for 30 seconds (second Ethanol wash).
- Carefully remove and discard the ethanol wash.
- Centrifuge tube briefly and return it to magnet.
- Remove and discard any remaining ethanol and allow the beads to air dry for 2 minutes.
- Resuspend beads in 20 µl water.
- Pipette mix vigorously and incubate at room temperature for 5 minutes.
- Place tube on magnet and transfer clear supernatant to PCR tube.

## 18.4 4) ADT libraries are now ready to be sequenced.

- Quantify libraries by standard methods (QuBit, BioAnalyzer, qPCR).
- ADT libraries will be around 180 bp (Figure 1).

#### IV) Sequencing CITE-seq libraries:

- 19 We estimate that an average of 100 molecules per ADT per cell is sufficient to achieve useful information. The number of reads required to obtain 100 molecules depends on the complexity of the sequencing library (e.g. duplication rate). ADT and mRNA cDNA sequencing libraries can be pooled at desired proportions. To obtain sufficient read coverage for both libraries we typically sequence ADT libraries in 5-10% of a lane and cDNA library fraction at 90% of a lane (HiSeq2500 Rapid Run Mode Flowcell).



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