

May 16,
2019

Working

CODEX Oligo-labeled Antibody Conjugation [↗](#)Yury Goltsev¹, Nikolay Samusik¹, Julia Kennedy-Darling¹, Salil Bhate¹, Matthew Hale¹, Gustavo Vazquez¹, Sarah Black¹, Garry Nolan¹¹Stanford University[dx.doi.org/10.17504/protocols.io.2qngdve](https://doi.org/10.17504/protocols.io.2qngdve) Gustavo Vazquez
Stanford University 

ABSTRACT

CODEX is a technology that uses oligo labeled antibodies, specialized fluorescent probes, and a companion instrument along-side a standard fluorescence microscope to create single-cell resolution fluorescence data across a multitude of parameters within spatial context in a single tissue. CODEX was developed by Yury Goltsev and Nikolay Samusik in the laboratory of Garry Nolan at Stanford. The technology is being commercialized by [Akoya Biosciences](#).

EXTERNAL LINK

[https://www.cell.com/cell/pdf/S0092-8674\(18\)30904-8.pdf](https://www.cell.com/cell/pdf/S0092-8674(18)30904-8.pdf)

GUIDELINES

Purified antibody stock can not be stored with any carrier protein or glycerol. Request the vendor to provide purified antibody in a BSA-free form.

A pre-purification process must be performed if a BSA-free antibody stock cannot be found.

Measure concentration of purified antibody with nanodrop before conjugation procedure. Concentrations on the tube label are not always accurate.

MATERIALS TEXT

PRODUCT	PROVIDER	CATALOG NUMBER
50KDa MWCO filter	Millipore	UFC505096
Antibody Reduction Solution 1	Akoya	RGT2004EA
Antibody Reduction Solution 2	Akoya	RGT2004EA
Antibody Conjugation Solution	Akoya	RGT2004EA
Filter Blocking Solution	Akoya	RGT2004EA
CODEX Antibody Tags	Akoya	RGT2004EA
Antibody Purification Solution	Akoya	RGT2004EA
Antibody Storage Solution	Akoya	RGT2004EA

Antibody Disulfide Reduction Reaction

- 1 Block nonspecific antibody binding to MWCO filter by adding 500ul Filter Blocking Solution to the top of each column and spinning down at 12,000g for 2 minutes.
- 2 Remove all liquid from the top of each column and discard flow-through. Exercise caution and do not scrape and damage filter with aspirator tip.

Measure concentration of stock purified antibody by nanodrop with pre-set IgG settings. Calculate the volume needed for 50ug of antibody

- 3 based on the nanodrop measurement.
- 4 Add 50ug of antibody to the top of each column and spin down at 12,000g for 8 minutes. The resulting solution should be 50ug of antibody concentrated into approximately 25ul.
- 5 Discard flow-through.
- 6 Prepare **Antibody Reduction Master Mix** by combining **Antibody Reduction Solution 1** with **Antibody Reduction Solution 2**.
20ul of **Antibody Reduction Solution 1** is needed for every 3 antibodies to conjugate.
20ul of **Antibody Reduction Solution 1** is mixed with 825ul of **Antibody Reduction Solution 2**.
- 7 Add 260ul **Antibody Reduction Master Mix** to the top of each column. Gently pipet up and down to mix reagent with antibody.
- 8 Incubate at RT for 30 minutes.
- 9 After 30 minutes spin down the columns at 12,000g for 8 minutes.
- 10 Discard flow-through.

Antibody Conjugation Reaction

- 11 Add 450ul **Antibody Conjugation Solution** to the top of each column and spin down at 12,000g for 8 minutes. During this spin, prepare the antibody tags.
- 12 The antibody tag preparation is time sensitive and must be done immediately prior to use. The antibody tag aliquots are to be used once for every 50ug antibody.
- 13 Pipet 220ul **Antibody Conjugation Solution** into each antibody tag aliquot. Ensure that the aliquot is dissolved with gentle pipetting.
- 14 After the spin in step 11 is completed, discard flow-through and add the dissolved antibody tag solutions from step 13 into each corresponding filter. Pipet up and down gently to mix the reagents.
- 15 Close the column lids and incubate the conjugation reaction at RT for 2 hours.
- 16 After 2 hours, spin down the columns at 12,000g for 8 minutes.
- 17 Discard flow-through.

- 18 Add 450ul **Antibody Purification Solution** to each column and spin down at 12,000g for 8 minutes.
Discard flow-through.
- 19 Add another 450ul **Antibody Purification Solution** to each column and spin down at 12,000g for 8 minutes.
Discard flow-through.
- 20 Add another 450ul **Antibody Purification Solution** to each column and spin down at 12,000g for 8 minutes.
Discard flow-through.
- 21 Add 100ul **Antibody Storage Solution** to the top of each column. Gently pipet up and down 10 or more times and wash the sides of the filters in the column.
- 22 Invert the filters into a fresh collection tube. Spin down at 3000g for 2 minutes. KEEP THE COLLECTED SOLUTION.
- 23 50ug antibody should be dissolved in 100ul **Antibody Storage Solution**. If the conjugation scale is larger than 50ug, add more **Antibody Storage Solution** according to this ratio.
- 24 Pipet the conjugated antibody solutions into sterile screw-top tubes and store at 4C.



This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited