# Code Documentation Chromosome 8

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### **Overall Schema:**

- Access database for list of genes and gene identifiers.
   -get genes (pymysql data access script)
- 2) Create gene objects and compile list of genes and identifiers (xml file) for front-end.
  - gene module
  - getGeneList.py
- 3) Front-end uses accession number to call functions/scripts for details page.
  - seq\_module
  - codon\_usage
- 4) Scripts access database using accession number to:
  - -get sequence
  - -get exons
- 5) Use sequence and exon list to calculate annotated sequence, coding sequence, translation, enzyme sites, and codon usage. (Codon usage for entire chromosome is precalculated and stored in flat file).
  - whole\_genome\_freq

## Instructions for front-end:

Front end should import three modules: gene module, seg module and codon usage.

In order to populate list of all genes from Chromosome 8, they will need to run: getGeneList

For the gene detail page, they will need to use these functions with the accession number:

annotateSeq to return sequence with exon boundaries marked

codingSeq to return coding sequence translate to return peptide translation

getEnzymes to return restriction enzyme information

getCodonusage to return codon usage statistics

Additionally, they will need file of codon usage for entire chromosome using: whole genome freq

# Module Descriptions:

# I. gene\_module

<u>Class</u> <u>Method</u>

Gene \_\_\_init\_\_ Create gene object

Gene.total Static method for total number of gene

objects created

Gene.number Assigns id for each gene object Gene.acc Accession number (primary id)

Gene genid Gene name/identifier

Gene.product Product name

Gene.location Chromosomal location

Gene.\_registry.append(self) Creates registry of gene objects

string method for printing attributes of

gene object

# II. seq\_module

This module contains the functions for returning various sequences and annotations.

#### **Function**

getSequence (acc) Returns genomic DNA sequence dictionary using accession

number (key=bp number, value=base)

annotateSeg(acc) Returns genomic sequence with exon boundaries indicated

with "exon/exon" annotations (string)

codingSeq(acc) Returns coding sequence for particular gene (string)

translate(acc) Uses coding sequence to translate into peptide sequence

(string)

\_\_str\_\_

enz cut(acc, seq, enzyme) Returns dictionary of enzyme cleavage sites for any of 6

common enzymes that may cut the genomic sequence, as well as optional custom cleavage sequence (optional enzyme parameter). Sequence is optional and used for testing coding

sequence.

getEnzyme(acc, enzyme) Returns list of enzymes which cut the sequence and includes

'good' or 'bad' indication to show whether it is useful ('good' does not cut in coding region, 'bad' enzymes do), number of cleavage sites and position of cleavage sites (enzyme parameter is optional).

#### III. codon\_usage

This module calculates codon usage information for indicated gene.

#### **Function**

codonFreq(acc, dna)

Analyses the frequency of every possible codon in coding

sequence of specified gene

codonPercent(acc, freq\_table)

Uses frequency information to calculate the codon

percentage (number of times codon used per 100bp)

codonRatio(acc, freq\_table) Uses frequency information to calculate the relative ratio of

codon usage for each amino acid in the peptide product

getCodonusage(acc)

Returns two dictionaries: synonymous codons {amino acid: codons} and usage dictionary {codon: usage ratio, percent(freq/100bp)} for indicated gene.

# **Other Programs**

getGenelist.py

Uses gene\_module to produce a list of all genes and their identifiers (accession number, gene name, protein product, chromosomal location). Stores list as .xml file.

whole\_genome\_freq.py

Uses gene\_module to get registry of gene objectes, seq\_module to obtain coding sequence for each gene and codon\_usage module to aggregate frequency/percent codon usage. Uses whole\_genome\_freq() function to calculate relative codon usage ratios across the chromosome. Returns 2 dictionaries. Stores this information in a .txt file.