

TaqMan[™] Gene Expression Assays —single-tube assays

USER GUIDE

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For descriptions of symbols on product labels or product documents, go to thermofisher.com/symbols-definition.

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R	17 December 2019	Removed troubleshooting and replaced with link to FAQs; updated multiplex reaction Application Note and url; updated endogenous control reference to Pub. No. COL33019 0619.
Q	15 May 2018	<ul style="list-style-type: none">Updated thermal cycling conditions for TaqMan™ Fast Advanced Master Mix.Added option of cDNA preamplification.Add option of C_{rt} algorithm for troubleshooting.Corrected troubleshooting for inhibitors in the real-time PCR reaction.
P	22 November 2017	<ul style="list-style-type: none">Added new instruments, Master Mixes, and other products applicable for the workflows.Removed content that is described in other resources; added references as appropriate.Streamlined and clarified content for ease of use and reading.Updated for general style, formatting, and branding.
N	November 2010	Baseline for this revision history.

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Product information

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IMPORTANT! Before using this product, read and understand the information in the “Safety” appendix in this document.

Product description

TaqMan™ Gene Expression Assays are a comprehensive collection of predesigned, preformulated primer and probe sets to perform quantitative gene expression studies on a variety of species. For a current list of available species and assays, use the Assay Search Tool at thermofisher.com/taqmangeneexpression.

- **TaqMan™ Gene Expression Assays**
 - A general collection of assays that target protein-coding transcripts from a variety of species and for specific diseases, pathways, or biological processes.
 - TaqMan™ Non-coding RNA Assays that target long non-coding RNA (ncRNA) in human, mouse, and rat species. These assays are designed for ncRNAs that are > 60 nt in length.
- **Endogenous control assays** (see page 18 for more information).

Note: Custom TaqMan™ Gene Expression Assays can also be designed. To design a custom assay, go to www.thermofisher.com/cadt. For more information on the design tool, see *Custom TaqMan™ Assays Design and Ordering Guide* (Pub. No. 4367671).

This document provides guidance for preparing cDNA templates (see page 12) and protocols for performing real-time PCR using a variety of compatible instruments and Master Mixes (see page 13).

For detailed information about TaqMan™ Gene Expression Assays, see page 18.

Contents and storage

Item	Storage
TaqMan™ Gene Expression Assay (single-tube format)	–25°C to –15°C ^[1]

^[1] Shipped at ambient temperature. See thermofisher.com/ambientshipping.

Go to thermofisher.com/taqmanfiles, then enter your order number to download the following files.

- Assay information files (AIFs)
- User Instruction Documents (Protocols, User Guides, and Quick Reference Cards)
- Certificates of Analysis
- Safety Data Sheets

For detailed information about the shipment and assay information files (AIF), see *Understanding Your Shipment* (Pub. No. MAN0017153).

TaqMan™ Gene Expression Assay formulations

To find and order predesigned, preformulated primer and probe sets in a variety of species, go to thermofisher.com/taqmangeneexpression, then use the Assay Search Tool.

Table 1 Standard formulations

Product	Dye	Size	Number of 20-µL reactions	Cat. No.	Concentration
TaqMan™ Gene Expression Assays	FAM™	Extra Small	75	4453320 ^[1] or 4448892	20X
		Small	250	4331182 ^[1]	
		Small	360	4351372	
		Medium	750	4351370	
		Large	2900	4351368	60X
	VIC™	Small	360	4448489	20X
		Medium	750	4448490	
		Large	2900	4448491	60X
TaqMan™ Gene Expression Assays, Primer-Limited (PL)	VIC™	Small	360	4448484	20X
		Medium	750	4448485	
		Large	2900	4448486	60X
TaqMan™ Non-coding RNA Assay	FAM™	Small	360	4426961	20X
		Medium	750	4426962	
		Large	2900	4426963	60X

^[1] This product is inventoried.

**Custom TaqMan™
Gene Expression
Assay
formulations**

Use the Custom TaqMan™ Assay Design Tool (www.thermofisher.com/cadt) to enter and submit sequences for new assay design. The tool also supports submission files created using FASTA file format. For details, see the *Custom TaqMan™ Assays Design and Ordering Guide* (Pub. No. 4367671).

For a comparison of Custom and Custom Plus Assay products go to thermofisher.com/customtaqmangex.

Table 2 Custom formulations

Product	Dye	Size	Number of 20 µL reactions	Cat. No.	Concentration
Custom Plus TaqMan™ RNA Assay	FAM™	Small	360	4441114	20X
		Medium	750	4441117	
		Large	2900	4441118	60X
	VIC™	Small	360	4448514	20X
		Medium	750	4448515	
		Large	2,900	4448516	60X
Custom PlusTaqMan™ RNA Assay, Primer-Limited (PL)	VIC™	Small	360	4448511	20X
		Medium	750	4448512	
		Large	2900	4448513	60X
Custom TaqMan™ Gene Expression Assays	FAM™	Small	360	4331348	20X
		Medium	750	4332078	
		Large	2900	4332079	60X
	VIC™	Small	360	4448508	20X
		Medium	750	4448509	
		Large	2900	4448510	60X
Custom TaqMan™ Gene Expression Assay, Primer-Limited (PL)	VIC™	Small	360	4448487	20X
		Medium	750	4448488	
		Large	2900	4448492	60X

Assay primer and
probe
concentrations

Assay type	Concentration					
	Forward primer		Reverse primer		Probe	
	1X	20X	1X	20X	1X	20X
TaqMan™ Gene Expression Assays (FAM™ or VIC™)	900 nM	18 µM	900 nM	18 µM	250 nM	5 µM
TaqMan™ Gene Expression Assays (VIC™, Primer Limited) ^[1]	150 nM	3 µM	150 nM	3 µM		

^[1] Recommended for multiplexing. For more information, see “Guidelines for duplex reactions using TaqMan™ Gene Expression Assays” on page 22.

Required materials not supplied

Unless otherwise indicated, all materials are available through **thermofisher.com**.
MLS: Fisher Scientific (**fisherscientific.com**) or other major laboratory supplier.

Table 3 Recommended products for isolation of RNA

Item	Source
Kits for RNA isolation	thermofisher.com/ rnaisolation

Table 4 Recommended products for preparation of cDNA

Item	Source
cDNA kit or cDNA Master Mix, one of the following:	
SuperScript™ IV VIL0™ Master Mix	11756050
SuperScript™ IV VIL0™ Master Mix with ezDNase™ Enzyme	11766050
High-Capacity cDNA Reverse Transcription Kit	4368813

Table 5 PCR Master Mixes

Item	Source
(Recommended) TaqMan™ Fast Advanced Master Mix	4444556
TaqMan™ Gene Expression Master Mix	4369016
TaqMan™ Universal Master Mix II, with UNG	4440038
TaqMan™ Universal Master Mix II, no UNG	4440047
TaqMan™ Fast Universal PCR Master Mix, no AmpErase™ UNG	4352042

Table 6 Other materials and equipment required for the workflow

Item	Source
Real-time PCR instrument, one of the following:	
QuantStudio™ 6 Pro and 7 Pro Real-Time PCR Systems	Contact your local sales office
QuantStudio™ 3 or 5 Real-Time PCR System	
QuantStudio™ 6 / QuantStudio™ 7 Flex Real-Time PCR System	
QuantStudio™ 12K Flex Real-Time PCR System	
StepOne™ or StepOnePlus™ Real-Time PCR System	
ViiA™ 7 Real-Time PCR System	
7500/7500 Fast Real-Time PCR System	

Item	Source
Software	
<i>(Optional)</i> Relative Quantification application	Available on the Connect
<i>(Optional)</i> Standard Curve application	
<i>(Optional)</i> ExpressionSuite™ Software	Available at thermofisher.com/expressionsuite
Equipment	
Thermal cycler, one of the following (or equivalent): <ul style="list-style-type: none"> • Veriti™ Thermal Cycler • SimpliAmp™ Thermal Cycler • ProFlex™ PCR System 	Contact your local sales office
Centrifuge, with adapter for 96-well or 384-well plates	MLS
Microcentrifuge	MLS
Vortex mixer	MLS
<i>(Optional)</i> Eppendorf™ MixMate™ (shaker)	Fisher Scientific™ 21-379-00
Pipettes	MLS
Tubes, plates, and other consumables	
Tubes, plates, and film	thermofisher.com/plastics
Aerosol-resistant barrier pipette tips	MLS
Disposable gloves	MLS
Reagents	
Nuclease-free Water	AM9930
RNase Inhibitor	N8080119
RNaseOUT™ Recombinant Ribonuclease Inhibitor	10777019
TURBO DNA-free™ KitDNase	AM1907
TE, pH 8.0	AM9849
<i>(Optional)</i> TaqMan™ PreAmp Master Mix	4391128
<i>(Optional)</i> TaqMan™ PreAmp Master Mix Kit	4384267

Workflow

Start with cDNA templates prepared from RNA samples (page 12)



Prepare the PCR Reaction Mix (page 14)



Set up and run the real-time PCR instrument (page 15)



Analyze the results (page 16)

Guidelines for preparation of cDNA

Guidelines for isolation of high-quality RNA

- See Table 3 on page 9 for recommended RNA isolation kits.
- *(Optional)* Use DNase to ensure minimal genomic DNA contamination of the RNA.

Guidelines for preparing cDNA templates

- See Table 4 on page 9 for recommended cDNA synthesis kits.
- Use the same reverse transcription procedure for all samples.
- For optimal reverse transcription, input RNA should be:
 - Free of inhibitors of reverse transcription (RT) and PCR
 - Dissolved in PCR-compatible buffer
 - Free of RNase activity

Note: We recommend using RNase Inhibitor (Cat. No. N8080119) or RNaseOUT™ Recombinant Ribonuclease Inhibitor (Cat. No. 10777019).

- Nondegraded total RNA (not applicable for double-stranded templates)

IMPORTANT! Degradation of the RNA may reduce the yield of cDNA for some gene targets.

- For the input RNA amount, follow the recommendations provided by the cDNA kit.
- Small amounts of cDNA can be pre-amplified.
Use TaqMan™ PreAmp Master Mix (Cat. No. 4391128) or TaqMan™ PreAmp Master Mix Kit (Cat. No. 4384267).
- Calculate the number of required reactions. Scale reaction components based on the single-reaction volumes, then include 10% overage, unless otherwise indicated.
- If using strip tubes to prepare cDNA templates, change to a new cap after each step or incubation.
- See your instrument user guide for detailed instructions about using plates, tubes, or strip tubes to prepare cDNA templates.

3

Perform real-time PCR

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Procedural guidelines for performing real-time PCR

- Follow best-practices when preparing or performing PCR (see page 22).
- Prepare the real-time PCR reactions in an area free of artificial templates and siRNA transfections. High-copy-number templates can easily contaminate the real-time PCR reactions.
- Configure run documents according to the instructions provided in the real-time PCR instrument user documents.
- Protect the assays from light and store as indicated until ready for use. Excessive exposure to light can negatively affect the fluorescent probes of the assays.
- Run technical replicates in triplicate to identify outliers.

Before you begin (60X assays)

Dilute 60X assays to 20X working stocks with TE, pH 8.0, then divide the solutions into smaller aliquots to minimize freeze-thaw cycles. The size of the aliquots depends upon the number of PCR reactions you typically run. An example dilution is shown in the following table.

1. Gently vortex the tube of 60X assay, then centrifuge briefly to spin down the contents and eliminate air bubbles.
2. In a 1.5-mL microcentrifuge tube, dilute sufficient amounts of 60X assay for the required number of reactions.

Component	Volume
TaqMan™ Gene Expression Assays (60X) or Custom TaqMan™ Gene Expression Assays (60X)	40 µL
TE, pH 8.0 (1X)	80 µL
Total aliquot volume	120 µL

3. Store aliquots at –20°C until use.

Prepare the PCR Reaction Mix

Thaw the cDNA samples on ice. Resuspend the cDNA samples by inverting the tube, then gently vortexing.

1. Mix the Master Mix thoroughly but gently.
2. Combine the PCR Reaction Mix and assays in an appropriately-sized microcentrifuge tube according to the following table.

Component	Volume for 1 reaction	
	Standard 96-well or 48-well Plates	384-well Plate or 96-well Fast Plate
Master Mix (2X) ^[1,2]	10 µL	5 µL
TaqMan™ Gene Expression Assay (20X) or Custom TaqMan™ Gene Expression Assay (20X)	1 µL	0.5 µL
Nuclease-free water ^[3]	7 µL	3.5 µL
Total PCR Reaction Mix volume	18 µL	9 µL

^[1] Recommended: TaqMan™ Fast Advanced Master Mix

^[2] [Optional] If you add AmpErase™ UNG (uracil-N-glycosylase), the final concentration must be 0.01U/ µL. Reduce the volume of water in the PCR reaction mix to compensate for additional volume from the UNG.

^[3] Adjust the volume of nuclease-free water for a larger volume of cDNA.

3. Vortex to mix the PCR Reaction Mix thoroughly, then centrifuge briefly to collect the contents at the bottom of the tube.
4. Transfer the appropriate volume of PCR Reaction Mix to each well of an optical reaction plate.
5. Add cDNA template (1 pg–100 ng in nuclease-free water), or nuclease-free water for NTC, to each well.
 - 1 µL for a 384-well plate or 96-well Fast Plate
 - 2 µL for a 96-well and 48-well Standard Plate

Note: Be sure to adjust the volume of nuclease-free water in the PCR reaction mix for a larger volume of cDNA.

IMPORTANT! For optimal results when using TaqMan™ Fast Universal PCR Master Mix, no AmpErase™ UNG, prepare the plate on ice. Run the plate within 2 hours of preparation, or store the plate at 2–8°C for up to 24 hours.

6. Seal the plate with a MicroAmp™ Optical Adhesive Film, then vortex briefly to mix the contents.
7. Centrifuge the plate briefly to collect the contents at the bottom of the wells.

Set up and run the real-time PCR instrument

See the appropriate instrument user guide for detailed instructions to program the thermal-cycling conditions or to run the plate.

Note: The instrument must be configured with the block appropriate for the plate type.

1. Select the cycling mode appropriate for the Master Mix.

IMPORTANT! The cycling mode depends on the Master Mix that is used in the reaction. The cycling mode does not depend on a Standard or a Fast plate format.

2. Set up the thermal protocol for your instrument.

See “Thermal protocols” on page 21 for the thermal protocols for other Master Mixes.

Table 7 TaqMan™ Fast Advanced Master Mix (StepOne™, StepOnePlus™, ViiA™ 7, and QuantStudio™ systems with fast cycling mode)

Step	Temperature	Time	Cycles
UNG incubation ^[1]	50°C	2 minutes	1
Enzyme activation	95°C	20 seconds ^[2]	1
Denature	95°C	1 second	40
Anneal / Extend	60°C	20 seconds	

^[1] Optional, for optimal UNG activity.

^[2] Enzyme activation can be up to 2 minutes. The time should not cause different results. See Enzyme activation time.

Table 8 TaqMan™ Fast Advanced Master Mix (7500 and 7500 Fast systems with fast cycling mode)

Step	Temperature	Time	Cycles
UNG incubation ^[1]	50°C	2 minutes	1
Enzyme activation	95°C	20 seconds ^[2]	1
Denature	95°C	3 seconds	40
Anneal / Extend	60°C	30 seconds	

^[1] Optional, for optimal UNG activity.

^[2] Enzyme activation can be up to 2 minutes. The time should not cause different results. See Enzyme activation time.

3. Set the reaction volume appropriate for the reaction plate.
 - **96-well Standard (0.2-mL) Plate:** 20 µL
 - **96-well Fast (0.1-mL) Plate and 384-well Plate:** 10 µL
4. Load the plate into the real-time PCR instrument.
5. Start the run.

Analyze the results

For detailed information about data analysis, see the appropriate documentation for your instrument. Use the absolute or relative quantification ($\Delta\Delta C_t$) methods to analyze results.

The general guidelines for analysis include:

- View the amplification plot; then, if needed:
 - Adjust the baseline and threshold values.
 - Remove outliers from the analysis.
- In the well table or results table, view the C_t values for each well and for each replicate group.

Perform additional analysis using any of the following software:

Software	Resource
Relative Quantification application	thermofisher.com/cloud
Standard Curve application	
ExpressionSuite™ Software ^[1]	thermofisher.com/expressionsuite

^[1] Can automatically define the baseline. Files from a QuantStudio™ 3 or 5 System are not compatible.

For more information about real-time PCR, see *Introduction to Gene Expression Getting Started Guide* (Pub. No. 4454239) or go to thermofisher.com/qpcr/education.



Troubleshooting and FAQs

Visit our online FAQ database for tips and tricks for conducting your experiment, troubleshooting information, and FAQs. The online FAQ database is frequently updated to ensure accurate and thorough content.

- For troubleshooting information and FAQs for this product: **thermofisher.com/taqmangesingletube FAQs**
- To browse the database and search using keywords: **thermofisher.com/faqs**



Supplemental information

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Endogenous controls

An endogenous control shows gene expression that is relatively constant and moderately abundant across tissues and cell types and treatment protocols. Normalization to endogenous control genes is currently the most accurate method to correct for potential biases that are caused by:

- Sample collection
- Variation in the amount of starting material
- Reverse transcription (RT) efficiency
- Nucleic acid (RNA/DNA) preparation and quality

No single control can act as a universal endogenous control for all experimental conditions, so we recommend verifying the chosen endogenous control or set of controls for the sample tissue, cell, or treatment. See *Using TaqMan™ Endogenous Control Assays to select an endogenous control for experimental studies* (Pub. No. COL33019 0619), available from **thermofisher.com**.

To select and order endogenous control assays, go to **thermofisher.com/taqmancontrols**.

TaqMan™ Gene Expression Assays chemistry overview

TaqMan™ MGB probes

TaqMan™ MGB probes contain:

- A reporter dye (for example, FAM™ dye) at the 5' end of the probe.
- A non-fluorescent quencher (NFQ) dye at the 3' end of the probe.
The NFQ dye does not fluoresce, which allows the real-time PCR system to measure the reporter dye contributions more accurately.
- A minor groove binder (MGB) at the 3' end of the probe that:
 - Increases the melting temperature (T_m) without increasing the probe length.
 - Allows for the design of shorter probes.

About the 5' nuclease assay

Note: The following figures are general representations of real-time PCR with TaqMan™ MGB probes and TaqMan™ Gene Expression Assays. The sequence regions are not necessarily drawn to scale.

The 5' nuclease assay process takes place during PCR amplification. It occurs in every cycle and does not interfere with the exponential accumulation of cDNA synthesis product.

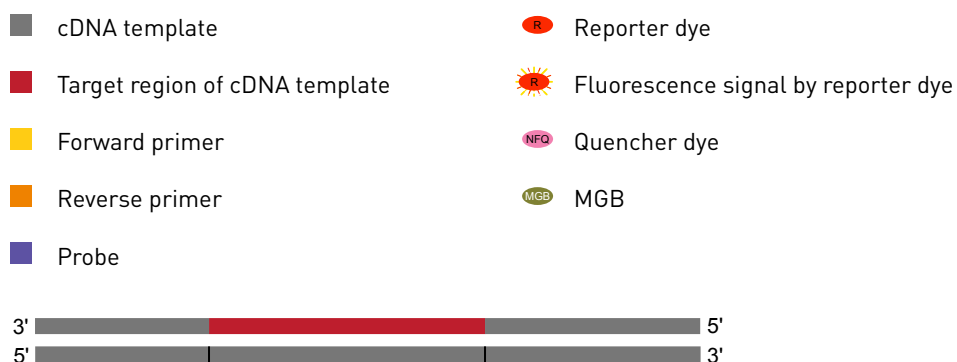


Figure 1 cDNA synthesis product

During the PCR, the forward and reverse primers anneal to complementary sequences along the denatured cDNA template strands (see Figure 2).

The TaqMan™ MGB probe anneals specifically to a complementary sequence between the forward and reverse primer sites (see Figure 2). When the probe is intact, the proximity of the reporter dye and quencher dye suppresses the reporter fluorescence, primarily by Förster-type energy transfer.

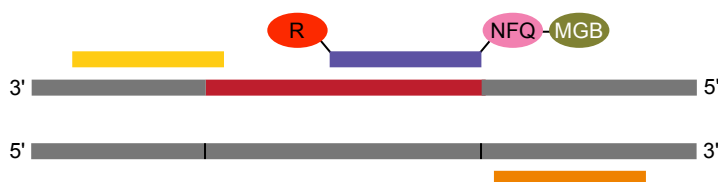


Figure 2 Annealing of probes and primers to cDNA strands

During polymerization, the DNA polymerase only cleaves probes that hybridize to the target sequence. Cleavage separates the reporter dye from the probe. The separation of the reporter dye from the quencher dye results in increased fluorescence by the reporter dye (see Figure 3).

This increase in fluorescence occurs only if the probe is complementary to the target sequence and if the target sequence is amplified during PCR. Because of these conditions, nonspecific amplification is not detected.

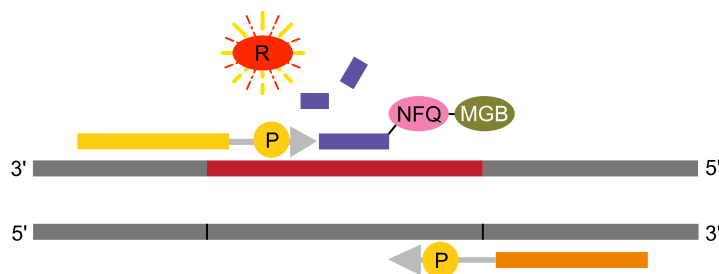


Figure 3 Initial polymerization and cleavage of reporter dye

Polymerization of the strand continues (see Figure 4), but because the 3' end of the probe is blocked, no extension of the probe occurs during PCR.

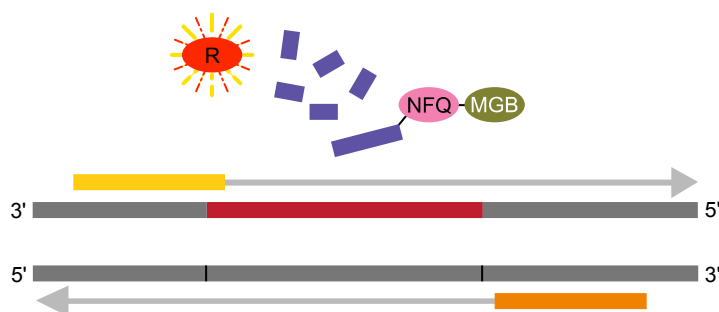


Figure 4 Completion of polymerization

Enzyme activation time

Using TaqMan™ Fast Advanced Master Mix, the enzyme activation step can range from 20 seconds to 2 minutes. A 20-second enzyme activation step is sufficient when the template is cDNA. A longer enzyme activation time will not affect the results.

The enzyme activation time for the default fast thermal cycling conditions on the instruments is 20 seconds. If a longer enzyme activation time is required, change the thermal cycling conditions before starting the run. A longer enzyme activation time can help to denature double-stranded genomic DNA when genomic DNA is used.

Algorithms for data analysis

Table 9 Algorithm recommendations for single-tube assays

Algorithm	Recommendation
Threshold (C_t)	Recommended.
Relative threshold (C_{rt})	<i>(Optional)</i> Use for troubleshooting abnormal or unexpected results.

The relative threshold algorithm is available in the Relative Quantification application on Connect (thermofisher.com/connect).

Thermal protocols

The thermal protocol settings depend on:

- The real-time PCR instrument
- Whether the Master Mix requires fast or standard cycling mode based on its chemistry
- Whether the Master Mix contains UNG

The thermal protocols in “Set up and run the real-time PCR instrument” on page 15 are optimized for the TaqMan™ Fast Advanced Master Mix.

The following tables provide thermal protocols for other Master Mixes that are compatible with TaqMan™ Gene Expression Assays.

IMPORTANT! The cycling mode depends on the Master Mix that is used in the reaction. The cycling mode does not depend on a Standard or a Fast plate format.

Table 10 TaqMan™ Gene Expression Master Mix or TaqMan™ Universal Master Mix II, with UNG (any compatible instrument)

Step	Temperature	Time (standard cycling mode)	Cycles
UNG incubation ^[1]	50°C	2 minutes	1
Enzyme activation	95°C	10 minutes	1
Denature	95°C	15 seconds	40
Anneal / Extend	60°C	1 minute	

^[1] For optimal UNG activity.

Table 11 TaqMan™ Universal Master Mix II, no UNG (any compatible instrument)

Step	Temperature	Time (standard cycling mode)	Cycles
Enzyme activation	95°C	10 minutes	1
Denature	95°C	15 seconds	40
Anneal / Extend	60°C	1 minute	

Table 12 TaqMan™ Fast Universal PCR Master Mix, no AmpErase™ UNG (StepOne™, StepOnePlus™, ViiA™ 7, or QuantStudio™ system)

Step	Temperature	Time (fast cycling mode)	Cycles
Enzyme activation	95°C	20 seconds	1
Denature	95°C	1 second	40
Anneal / Extend	60°C	20 seconds	

Table 13 TaqMan™ Fast Universal PCR Master Mix, no AmpErase™ UNG (7500 or 7500 Fast system)

Step	Temperature	Time (fast cycling mode)	Cycles
Enzyme activation	95°C	20 seconds	1
Denature	95°C	3 seconds	40
Anneal / Extend	60°C	30 seconds	

Guidelines for duplex reactions using TaqMan™ Gene Expression Assays

Duplex real-time PCR is the simultaneous amplification and measurement of two target sequences in one reaction. TaqMan™ Gene Expression Assays can be used in duplex real-time PCR when using a FAM™ dye-labeled assay in combination with a primer-limited, VIC™ dye-labeled assay. When setting up a duplex reaction:

- Validate that your duplex assay combinations provide similar results to your singleplex reactions.
- Consider the relative expression levels of each target.
- Perform serial dilutions of your sample in both singleplex and duplex reactions, and compare the results for relative expression.
- Select the higher-expressing target as the primer-limited, VIC™ dye-labeled assay.
- Use TaqMan™ Fast Advanced Master Mix, which has been optimized for duplexing reactions.

For more details on how to validate your duplex assay reactions and interpret the results, see *Factors Influencing Multiplex Real-Time PCR Application Note* (Pub. No. 136AP04-01) or go to thermofisher.com/taqmangeneexpressionduplex.

Best practices for PCR and RT-PCR experiments

Good laboratory practices for PCR and RT-PCR

- Wear clean gloves and a clean lab coat.
 - Do not wear the same gloves and lab coat that you have previously used when handling amplified products or preparing samples.

- Change gloves if you suspect that they are contaminated.
- Maintain separate areas and dedicated equipment and supplies for:
 - Sample preparation and reaction setup.
 - Amplification and analysis of products.
- Do not bring amplified products into the reaction setup area.
- Open and close all sample tubes carefully. Avoid splashing or spraying samples.
- Keep reactions and components capped as much as possible.
- Use a positive-displacement pipettor or aerosol-resistant barrier pipette tips.
- Clean lab benches and equipment periodically with 10% bleach solution or DNA decontamination solution.

Use UNG to prevent false-positive amplification

Carryover amplicons can result in false-positive amplification during PCR. Use a Master Mix that contains uracil-N-glycosylase (UNG; also known as uracil-DNA glycosylase (UDG)) to degrade many contaminating carryover amplicons.

UNG enzymatic activity occurs during an initial incubation at 50°C. UNG is partially inactivated during the 95°C incubation step for template denaturation and polymerase activation. Because UNG is not completely deactivated during the 95°C incubation, it is important to keep the annealing temperatures greater than 55°C and to refrigerate PCR products at 2°C to 8°C in order to prevent amplicon degradation.

To ensure the desired UNG activity:

- Use PCR components and thermal cycling conditions as specified.
UNG-containing Master Mixes incorporate the optimal concentration of UNG to prevent cross-contamination while not affecting real-time PCR performance.
- Do not attempt to use UNG-containing Master Mixes in subsequent amplification of dU-containing PCR products, such as in nested-PCR protocols. The UNG will degrade the dU-containing PCR products, preventing further amplification.

Although treatment with UNG can degrade or eliminate large numbers of carryover PCR products, use good laboratory practices to minimize cross-contamination from non-dU-containing PCR products or other samples.

Detect fluorescent contaminants

Fluorescent contaminants can generate false positive results. To help detect these contaminants, we recommend including a no-amplification control reaction that contains sample, but no master mix.

After PCR, if the absolute fluorescence of the no-amplification control is greater than the fluorescence of the no template control (NTC), fluorescent contaminants may be present in the sample or in the heat block of the real-time PCR instrument.



Safety



WARNING! GENERAL SAFETY. Using this product in a manner not specified in the user documentation may result in personal injury or damage to the instrument or device. Ensure that anyone using this product has received instructions in general safety practices for laboratories and the safety information provided in this document.

- Before using an instrument or device, read and understand the safety information provided in the user documentation provided by the manufacturer of the instrument or device.
 - Before handling chemicals, read and understand all applicable Safety Data Sheets (SDSs) and use appropriate personal protective equipment (gloves, gowns, eye protection, and so on). To obtain SDSs, see the “Documentation and Support” section in this document.
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Chemical safety



WARNING! GENERAL CHEMICAL HANDLING. To minimize hazards, ensure laboratory personnel read and practice the general safety guidelines for chemical usage, storage, and waste provided below. Consult the relevant SDS for specific precautions and instructions:

- Read and understand the Safety Data Sheets (SDSs) provided by the chemical manufacturer before you store, handle, or work with any chemicals or hazardous materials. To obtain SDSs, see the "Documentation and Support" section in this document.
- Minimize contact with chemicals. Wear appropriate personal protective equipment when handling chemicals (for example, safety glasses, gloves, or protective clothing).
- Minimize the inhalation of chemicals. Do not leave chemical containers open. Use only with sufficient ventilation (for example, fume hood).
- Check regularly for chemical leaks or spills. If a leak or spill occurs, follow the manufacturer cleanup procedures as recommended in the SDS.
- Handle chemical wastes in a fume hood.
- Ensure use of primary and secondary waste containers. (A primary waste container holds the immediate waste. A secondary container contains spills or leaks from the primary container. Both containers must be compatible with the waste material and meet federal, state, and local requirements for container storage.)
- After emptying a waste container, seal it with the cap provided.
- Characterize (by analysis if needed) the waste generated by the particular applications, reagents, and substrates used in your laboratory.
- Ensure that the waste is stored, transferred, transported, and disposed of according to all local, state/provincial, and/or national regulations.
- **IMPORTANT!** Radioactive or biohazardous materials may require special handling, and disposal limitations may apply.



WARNING! HAZARDOUS WASTE (from instruments). Waste produced by the instrument is potentially hazardous. Follow the guidelines noted in the preceding General Chemical Handling warning.



WARNING! 4L Reagent and Waste Bottle Safety. Four-liter reagent and waste bottles can crack and leak. Each 4-liter bottle should be secured in a low-density polyethylene safety container with the cover fastened and the handles locked in the upright position.

Biological hazard safety



WARNING! Potential Biohazard. Depending on the samples used on this instrument, the surface may be considered a biohazard. Use appropriate decontamination methods when working with biohazards.



WARNING! BIOHAZARD. Biological samples such as tissues, body fluids, infectious agents, and blood of humans and other animals have the potential to transmit infectious diseases. Conduct all work in properly equipped facilities with the appropriate safety equipment (for example, physical containment devices). Safety equipment can also include items for personal protection, such as gloves, coats, gowns, shoe covers, boots, respirators, face shields, safety glasses, or goggles. Individuals should be trained according to applicable regulatory and company/ institution requirements before working with potentially biohazardous materials. Follow all applicable local, state/provincial, and/or national regulations. The following references provide general guidelines when handling biological samples in laboratory environment.

- U.S. Department of Health and Human Services, *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, 5th Edition, HHS Publication No. (CDC) 21-1112, Revised December 2009; found at:
<https://www.cdc.gov/labs/pdf/CDC-BiosafetymicrobiologicalBiomedicalLaboratories-2009-P.pdf>
 - World Health Organization, *Laboratory Biosafety Manual*, 3rd Edition, WHO/CDS/CSR/LYO/2004.11; found at:
www.who.int/csr/resources/publications/biosafety/Biosafety7.pdf
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Documentation and support

Related documentation

Document	Pub. No.
<i>TaqMan™ Gene Expression Assays Quick Reference—single-tube assays</i>	4401212
<i>Introduction to Gene Expression Getting Started Guide</i>	4454239
<i>Understanding Your Shipment</i>	MAN0017153
<i>Custom TaqMan™ Assays Design and Ordering Guide</i>	4367671
<i>TaqMan™ Assay Multiplex PCR Optimization User Guide</i>	MAN0010189
<i>TaqMan™ PreAmp Master Mix User Guide</i>	4384557
<i>TaqMan™ PreAmp Master Mix Quick Reference</i>	4384556
QuantStudio™ 3 or 5 Real-Time PCR System	
<i>QuantStudio™ 3 and 5 Real-Time PCR Systems Installation, Use, and Maintenance Guide</i>	MAN0010407
<i>QuantStudio™ Design and Analysis Desktop Software User Guide</i>	MAN0010408
QuantStudio™ 6 / QuantStudio™ 7 Flex Real-Time PCR System	
<i>QuantStudio™ 6 and 7 Flex Real-Time PCR Systems Maintenance and Administration Guide</i>	4489821
<i>QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide</i>	4489822
QuantStudio™ 12K Flex Real-Time PCR System	
<i>QuantStudio™ 12K Flex Real-Time PCR System Maintenance and Administration Guide</i>	4470689
<i>QuantStudio™ 12K Flex Real-Time PCR System: Multi-Well Plates and Array Card Experiments User Guide</i>	4470050
StepOne™ or StepOnePlus™ Real-Time PCR System	
<i>StepOne™ and StepOnePlus™ Real-Time PCR Systems Installation, Networking and Maintenance User Guide</i>	4376782
<i>Applied Biosystems™ StepOne™ and StepOnePlus™ Real-Time PCR Systems Relative Standard Curve and Comparative C_t Experiments Getting Started Guide</i>	4376785
ViiA™ 7 Real-Time PCR System	
<i>Applied Biosystems™ ViiA™ 7 Real-Time PCR System User Guide: Calibration, Maintenance, Networking, and Security</i>	4442661
<i>Applied Biosystems™ ViiA™ 7 Real-Time PCR System Getting Started Guide</i>	4441434

Document	Pub. No.
7500/7500 Fast Real-Time PCR System	
<i>Applied Biosystems™ 7300/7500/7500 Fast Real-Time PCR System Installation and Maintenance Guide</i>	4347828
<i>Applied Biosystems™ 7500/7500 Fast Real-Time PCR System Getting Started Guide: Relative Standard Curve and Comparative C_t Experiments</i>	4387783
Data analysis	
<i>Real-Time PCR Systems Chemistry Guide: Applied Biosystems™ 7900HT Fast Real-Time PCR System and 7300/7500 Real-Time PCR Systems</i>	4348358
<i>Applied Biosystems™ 7900HT Fast Real-Time PCR System Relative Quantitation Using Comparative C_T Getting Started Guide</i>	4364016
<i>Applied Biosystems™ 7900HT Fast Real-Time PCR System Absolute Quantitation Using Standard Curve Getting Started Guide</i>	4364014
<i>Applied Biosystems™ 7300/7500/7500 Fast Real-Time PCR System Getting Started Guide: Absolute Quantitation using Standard Curve</i>	4347825
<i>Applied Biosystems™ 7300/7500/7500 Fast Real-Time PCR System Getting Started Guide: Relative Quantitation using Comparative C_t</i>	4347824
<i>Applied Biosystems™ StepOne™ and StepOnePlus™ Real-Time PCR Systems Relative Standard Curve and Comparative C_t Experiments Getting Started Guide</i>	4376785
<i>Applied Biosystems™ Relative Quantitation Analysis Module User Guide</i>	MAN0014820
<i>Applied Biosystems™ Standard Curve Analysis Module User Guide</i>	MAN0014819

Customer and technical support

Visit thermofisher.com/support for the latest service and support information.

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 - Product FAQs
 - Software, patches, and updates
 - Training for many applications and instruments
- Order and web support
- Product documentation
 - User guides, manuals, and protocols
 - Certificates of Analysis
 - Safety Data Sheets (SDSs; also known as MSDSs)

Note: For SDSs for reagents and chemicals from other manufacturers, contact the manufacturer.

Limited product warranty

Life Technologies Corporation and/or its affiliate(s) warrant their products as set forth in the Life Technologies' General Terms and Conditions of Sale at www.thermofisher.com/us/en/home/global/terms-and-conditions.html. If you have any questions, please contact Life Technologies at www.thermofisher.com/support.

