

# **Research Forefronts: Estimation of serial intervals using pathogen genomic data**

with an application to  
COVID-19

Sometimes, our data  
might not be sufficient to  
fully reconstruct the  
transmission tree

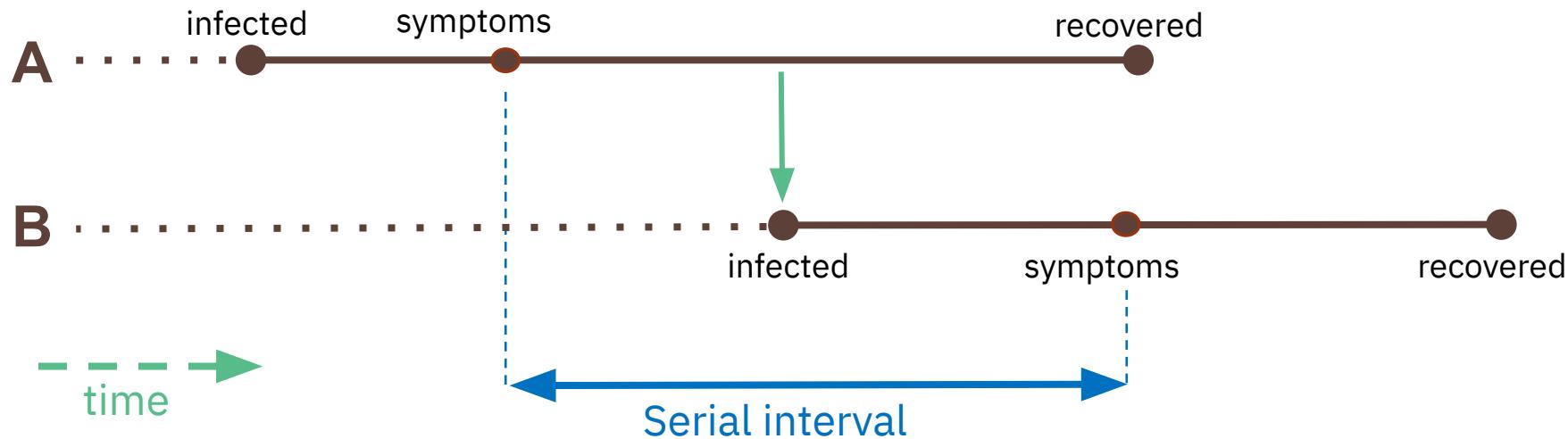
But that doesn't mean  
there's nothing we can  
learn...

We developed a method to  
estimate **serial intervals** using  
genomic data.

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# What is the serial interval?

Definition of the serial interval = length of time between successive cases in a chain of transmission  
= length of time between symptom onset in an infector and infectee



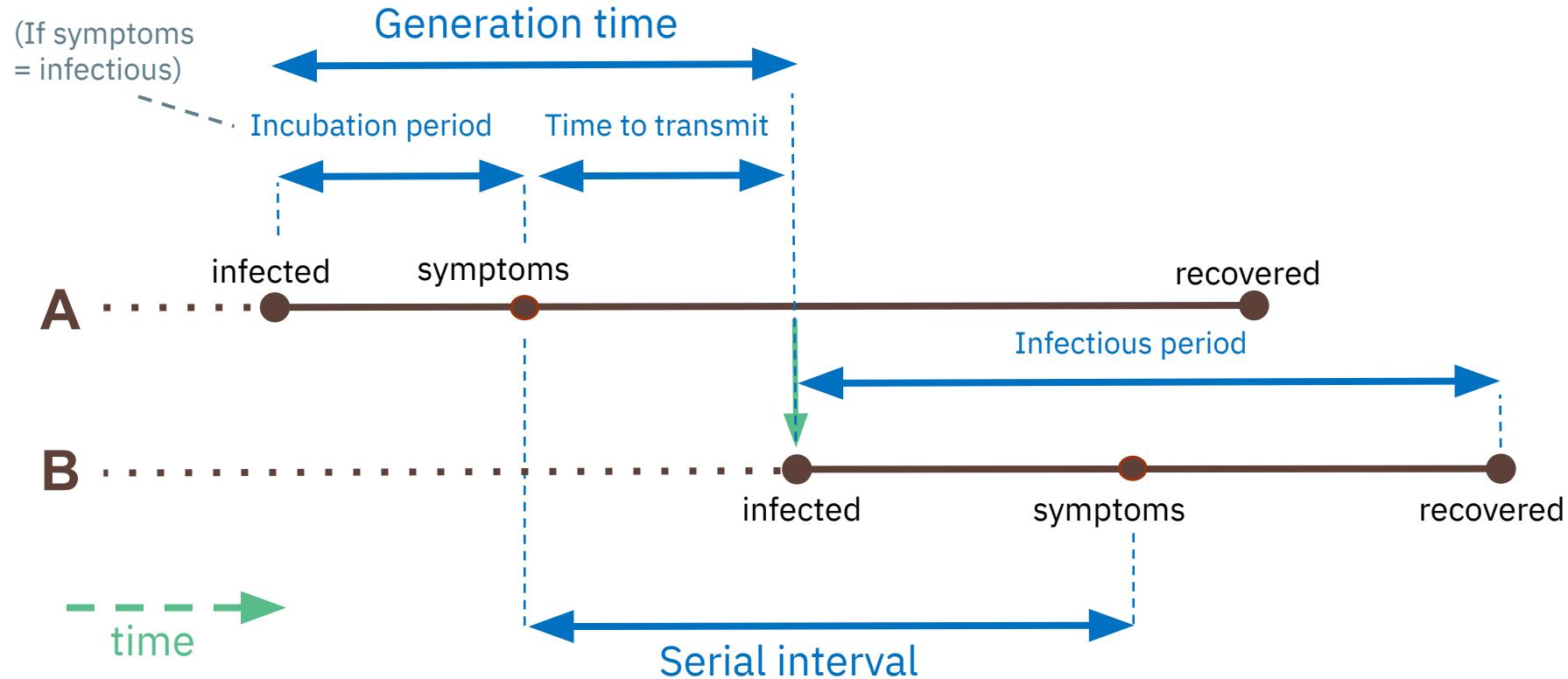
# Why is the serial interval important?

- ❖ Tells us about the speed of transmission...
- ❖ ...this informs surveillance efforts.
- ❖ Used to calculate quantities like  $R_0$ ,  $R_t$  ...
- ❖  $R_0 \approx 1 + rS$  ← Epidemic exponential growth rate x serial interval \*
- ❖ ...and hence in understanding herd immunity thresholds and more.

$R_0$  = the average number of cases caused by a single infected individual, in a wholly susceptible population  
 $R_t$  = the average number of cases caused by a single infected individual, at a specific time  $t$

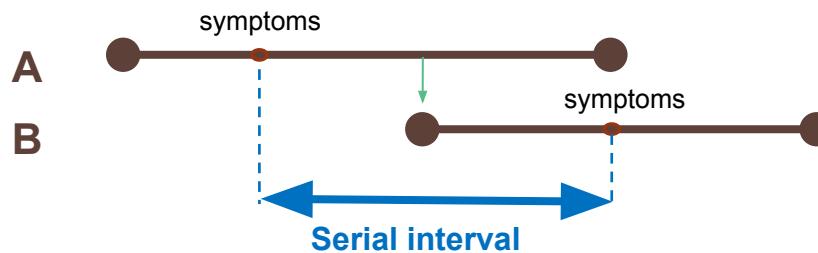
\* How generation intervals shape the relationship between growth rates and reproductive numbers, Wallinga and Lipsitch (2006) *Proceedings of the Royal Society B* 4

# It's also related to other disease intervals

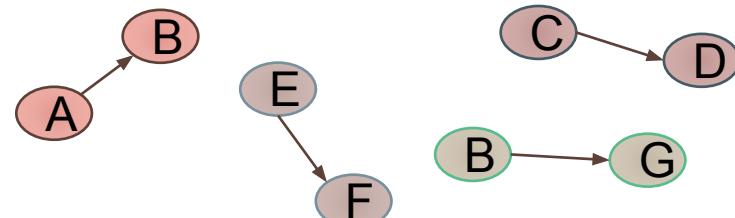


# Most existing methods for serial interval estimation assume direct observation of transmission pairs (infectors & infectees)

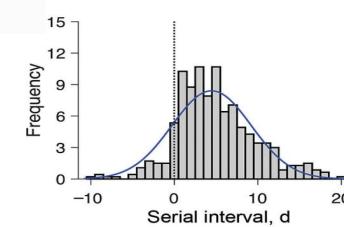
1. **Contact trace** pairs of cases which are assumed to represent direct transmission



3. **Parametric estimation** of the serial interval given this observed data



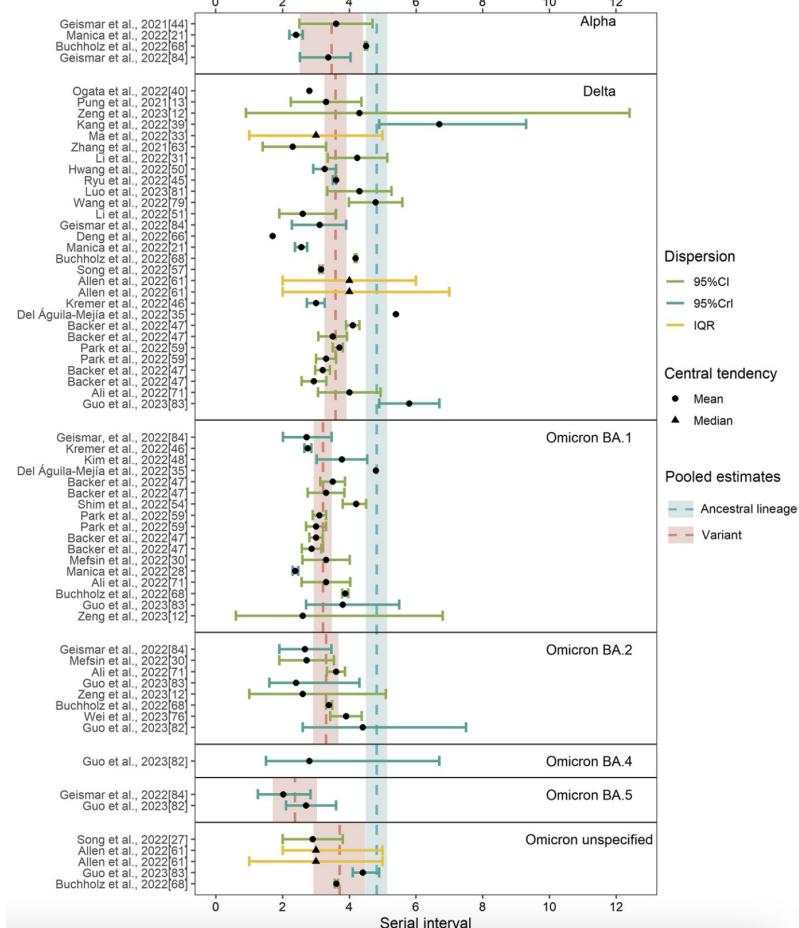
2. This provides **direct observations** of the serial interval



Serial Interval of COVID-19 among Publicly Reported Confirmed Cases  
Du et al. (2020) *Emerging Infectious Diseases*

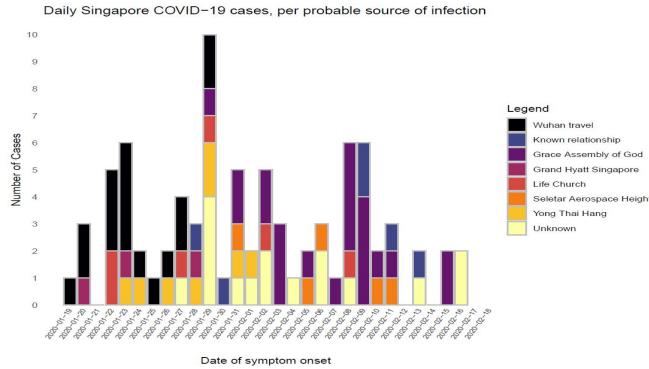
# Systematic Review and Meta-analysis of Serial Intervals for COVID-19

Xu et al (2023) estimate a mean serial interval from ancestral lineage SARS-CoV-2 of **4.82 days (95% CI 4.5 - 5.14)**



All included studies (98) use contact data, the majority of which assume direct contact-traced pairs and many of which were household pairs.

# How we got thinking about serial intervals...

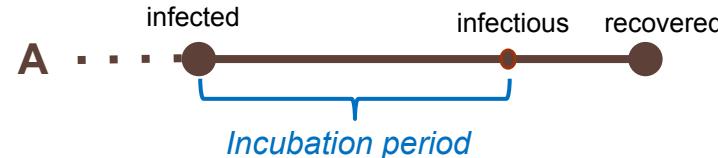


Evidence for transmission of COVID-19 prior to symptom onset. Tindale, Stockdale et al (2020) eLife

*“About 40% to 80% of the novel coronavirus transmission occurs two to four days before an infected person has symptoms”*

By collating contact data from outbreaks in Singapore and Tianjin, we estimated the amount of **pre-symptomatic transmission** of COVID-19.

This requires estimation of both the serial interval and incubation period.



We developed a new approach for estimating incubation periods whilst taking into account that observed pairs may not represent direct transmission

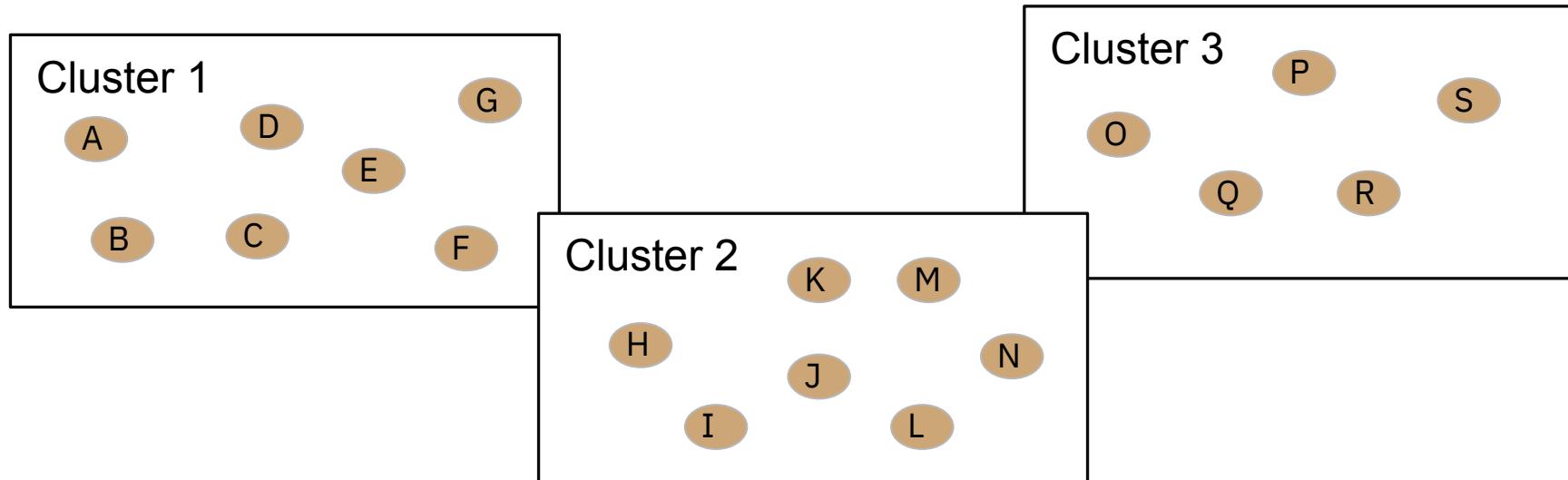
# Contact tracing approaches require detailed personal data, and are often limited to household studies

This motivated a new genomic approach:

- Use pathogen whole genome sequence data as a proxy for contact data
- Use in broader clusters than e.g. households
- Incorporate possibility of missing cases
- Fast and cluster-specific estimates: track the serial interval through time and under different settings or variants

# Estimating serial intervals with genomic data

Suppose we have a set of case clusters (perhaps genomic clusters, or clusters associated with e.g. schools, hospitals) from an outbreak of infectious disease. We wish to **estimate the serial interval in each cluster...**



We know each case's symptom onset time and pathogen sequence, but we don't know who infected whom.

# Whole genome sequences as a proxy for contact data

**The main idea:** As with the other methods in this course, differences in the sequences tell us how closely related people's infections are and therefore who might have infected whom.

Case A: ATC**G**TATCAGTCAG

Case B: ATC**A**GTATCAGTCAG

**However**, since we want to work with broad clusters where we don't necessarily sample a large proportion of cases, we need to consider

1. There may be **large** uncertainty in who infected whom
2. Inferred pairs (infector & infectee) might not represent direct transmission

# Take uncertainty in who infected whom into account, by sampling feasible transmission networks

## 1. Identify all plausible transmission pairs:

Pairs  $(i,j)$  in the same cluster, with closely related sequences & realistic timing, where infector  $i$  showed symptoms first.

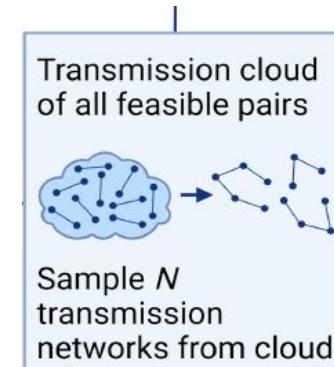
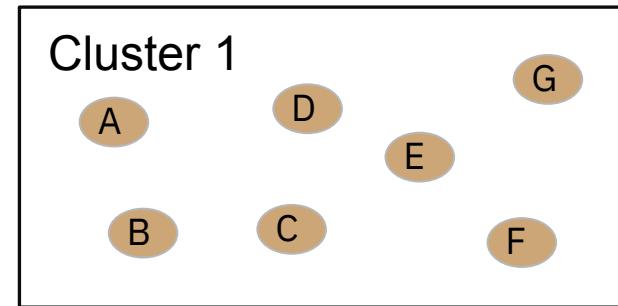
1. Difference in symptom onset date  $\leq T$
2. Pairwise genomic distance  $\leq G$

Case i: ATCGGTATCAG

Case j: ATCAGTATCAG

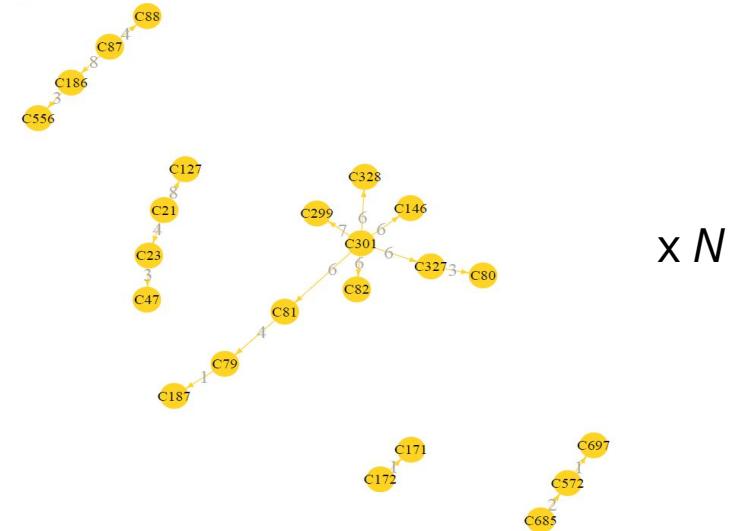
## 2. Sample plausible transmission networks:

Built from the plausible pairs, by sampling an infector for each infectee.



# Take uncertainty in who infected whom into account, by sampling feasible transmission networks

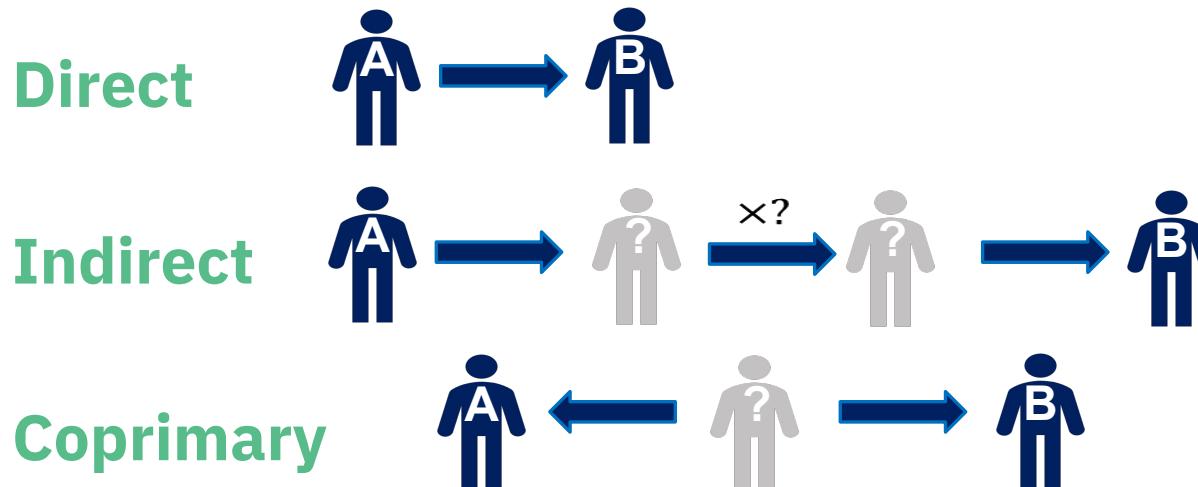
Incorporate uncertainty by sampling a set of networks:



We estimate the serial interval in each network independently – and then **average across networks**

# Estimate the serial interval distribution, taking indirect transmission into account

To take under-sampling into account, we consider that, for every infector-infectee pair (A, B) in every sampled network, transmission may have been:



Inspired by:

Serial Intervals of  
Respiratory Infectious  
Diseases: A Systematic  
Review and Analysis  
Vink et al. (2014)

We fit a mixture model to incorporate this idea...

# Estimate the serial interval distribution: possible pathways

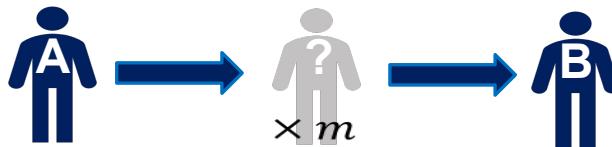
True serial interval distribution  $\sim \Gamma(\mu, \sigma)$

Direct



Observed time difference between A & B,  $T_{a,b} \sim \Gamma(\mu, \sigma)$

Indirect



$T_{a,b}$  = sum of  $m + 1$  serial intervals,  $m \sim \text{Geo}(\pi)$   
 $\sim \text{Compound Geometric Gamma}(\mu, \sigma, \pi)$

Proportion w of pairs

# Estimate the serial interval distribution: possible pathways

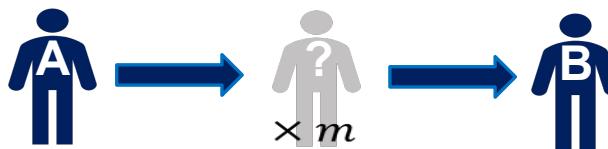
True serial interval distribution  $\sim \Gamma(\mu, \sigma)$

Direct



Observed time difference between A & B,  $T_{a,b} \sim \Gamma(\mu, \sigma)$

Indirect



Probability of sampling an infectee

$T_{a,b}$  = sum of  $m + 1$  serial intervals,  $m \sim \text{Geo}(\pi)$   
 $\sim \text{Compound Geometric Gamma}(\mu, \sigma, \pi)$

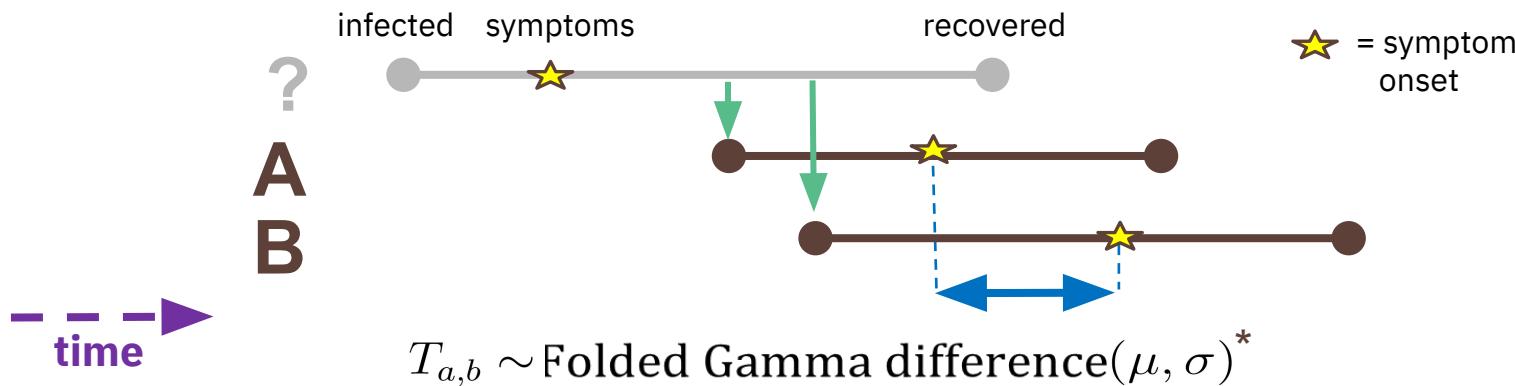
Proportion w of pairs

# Estimate the serial interval distribution: possible pathways

Coprimary



$T_{a,b}$  is the strictly non-negative difference of two  $\Gamma(\mu, \sigma)$  distributions



# Estimate the serial interval distribution: mixture model taking possible pathways into account

We combine these possible transmission pathways into a mixture model with log-likelihood:

$$l(\mu, \sigma, \pi, w | D) = \sum_{k=1}^n \log[w f_{CGG}(T_{a_k, b_k} | \mu, \sigma, \pi) + (1 - w) f_{FGD}(T_{a_k, b_k} | \mu, \sigma)]$$

Sum over all pairs in  
the network

Direct or indirect

Coprimary

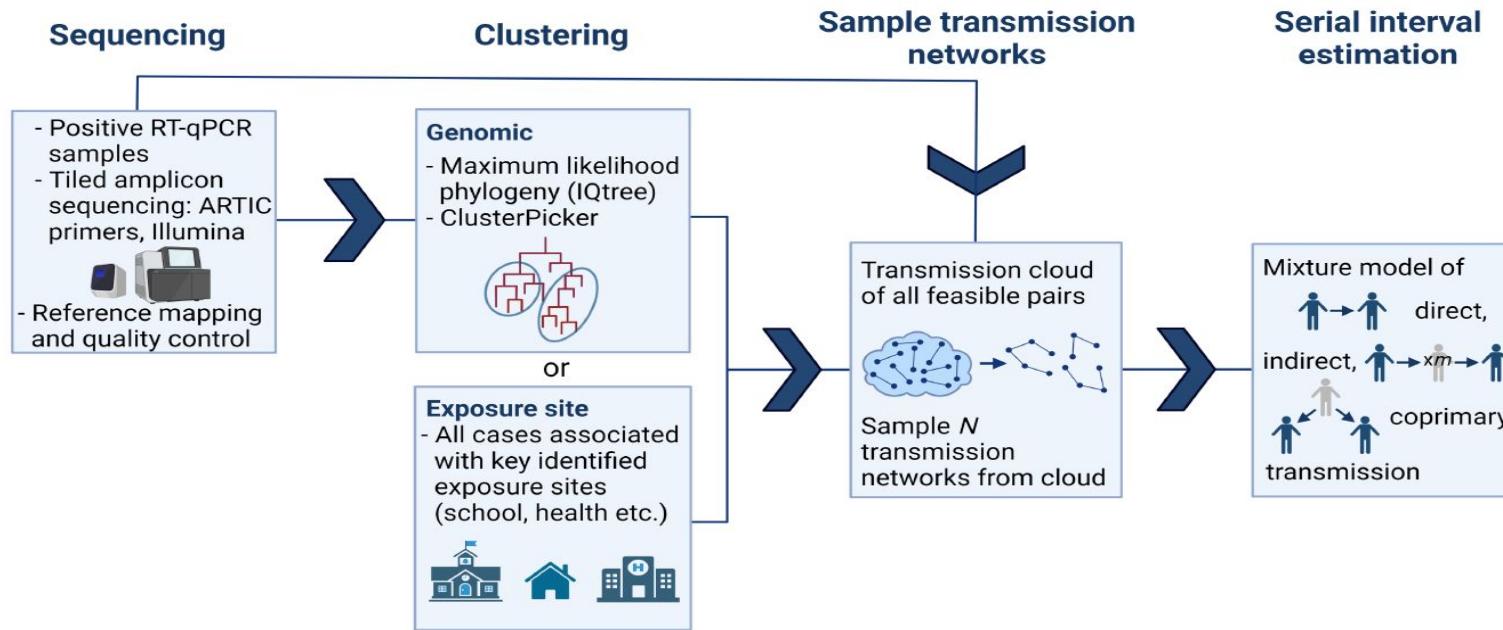
Instead of maximising this directly, we incorporate Beta distributed priors for  $w$  and  $\pi$ , and perform maximum a posteriori (MAP) estimation. We calculate the MAP for each sampled network, and then average over all networks

Our confidence intervals need to take into account uncertainty in each network, as well as uncertainty when combining across networks:

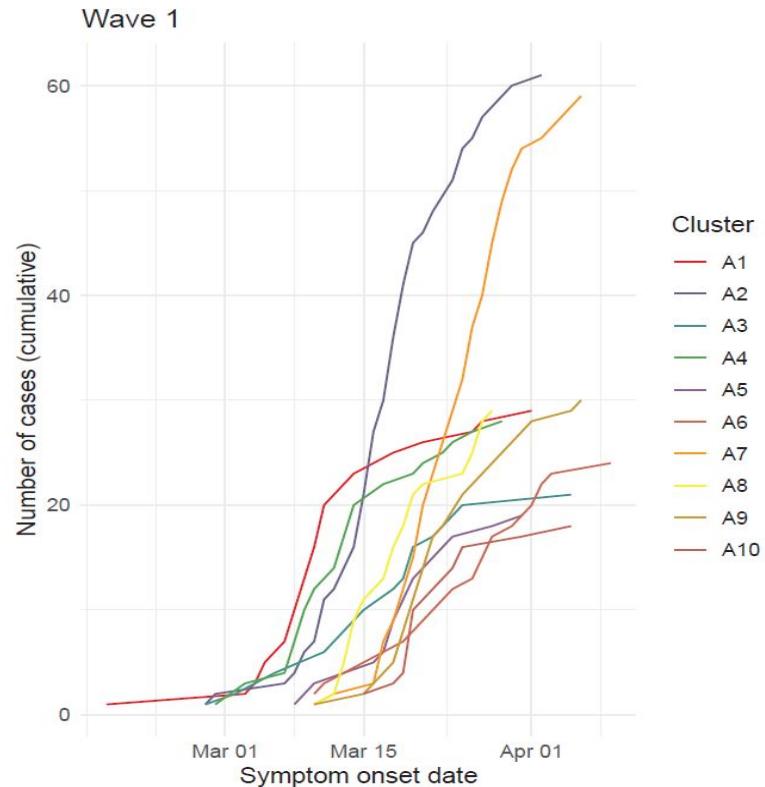
$$\hat{\text{Var}}(\hat{\mu}_c) = \mathbb{E}_{\tau} \left( \hat{s}\text{e}(\hat{\mu}_{c, \tau_k})^2 \right) + \text{Var}_{\tau} \left( \hat{\mu}_{c, \tau_k} \right).$$

For cluster  $c$  and each network  $\tau_k$

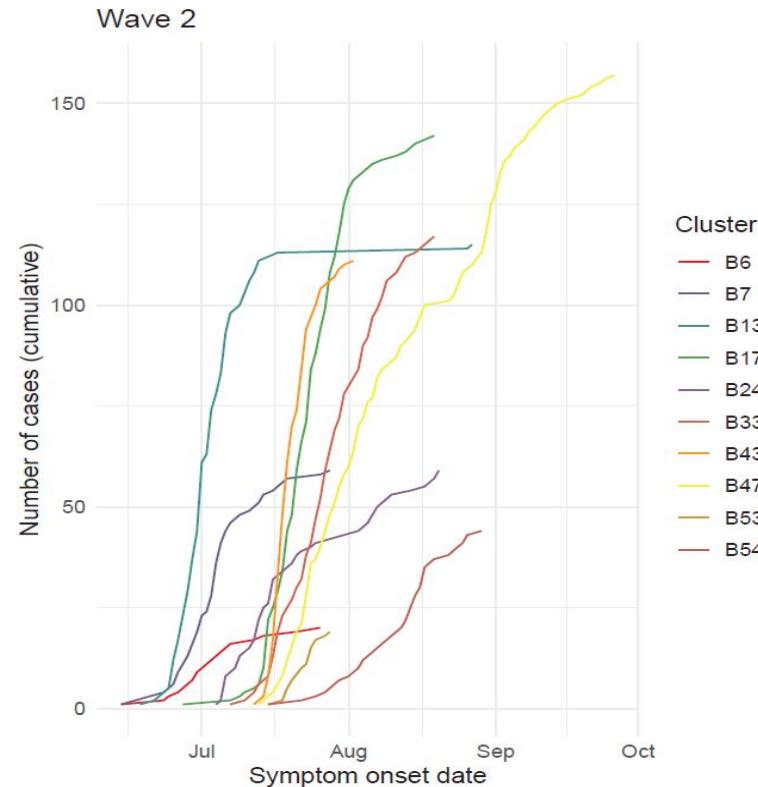
# A schematic view of the method



# Application: COVID-19 clusters in Victoria, Australia

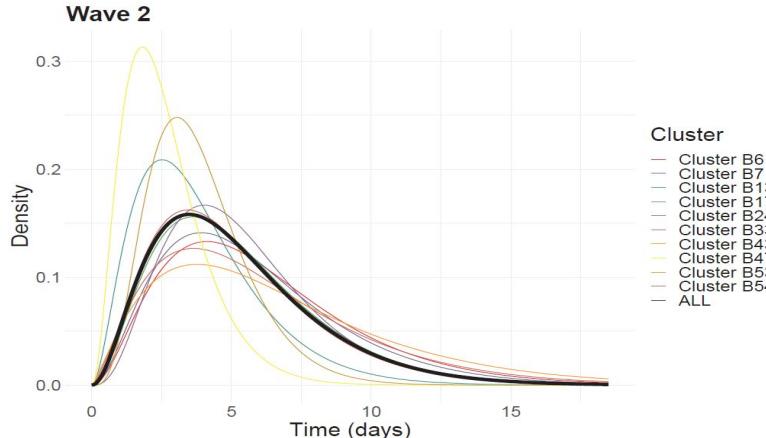
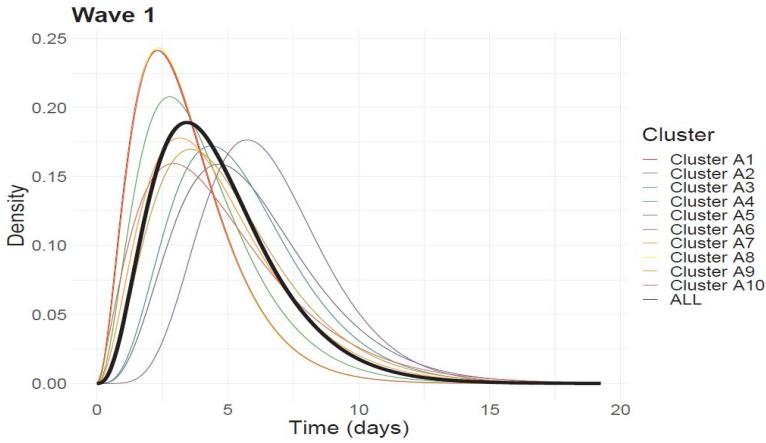
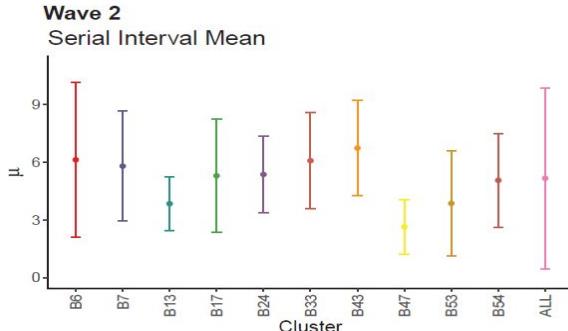
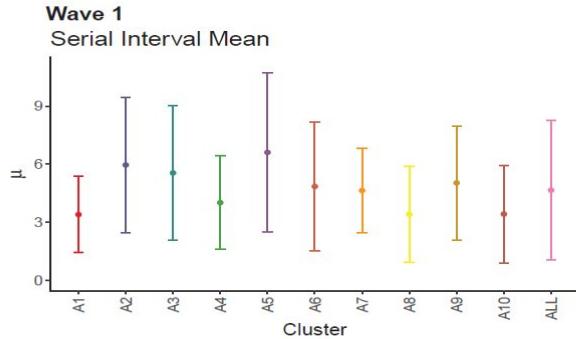


6 January–14 April 2020



1 June–28 October 2020

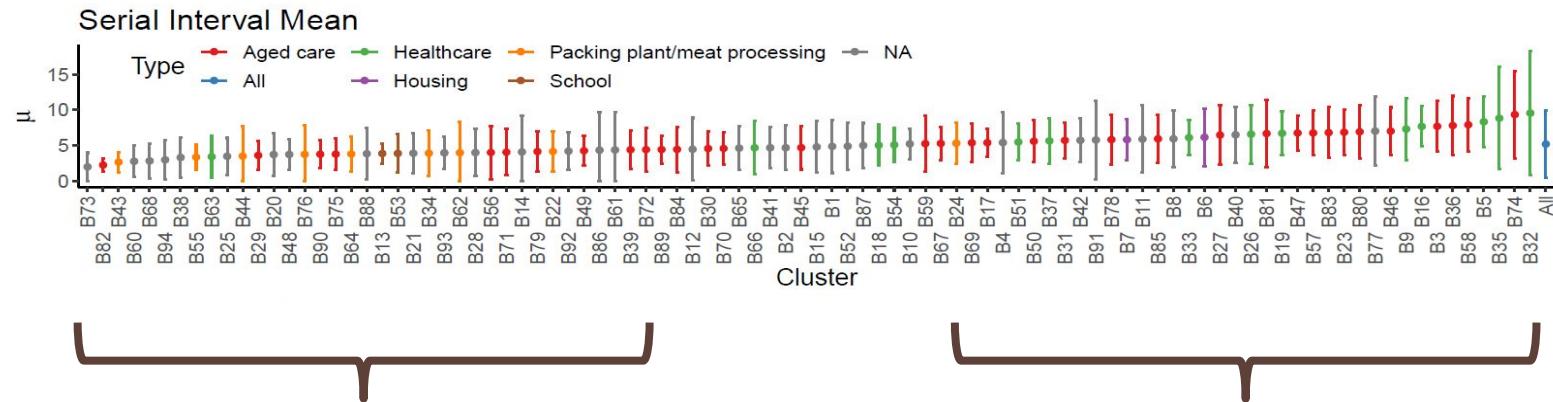
# Cluster-specific serial intervals: in line with published estimates, with some variation by cluster



Context: Early published estimates ~5 days

# Using a larger range of 2nd wave clusters, we can compare across different exposure settings

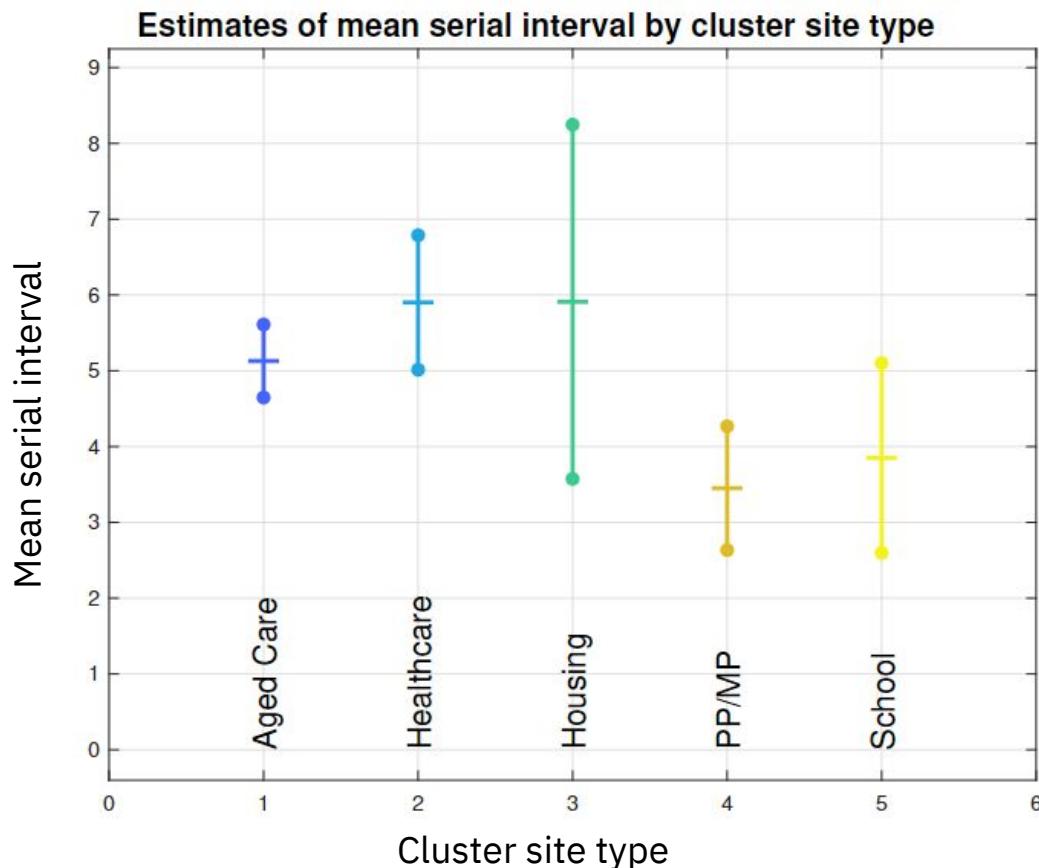
Estimates of the mean range from **2 to 9.5 days** (compared to standard estimates ~5 days)



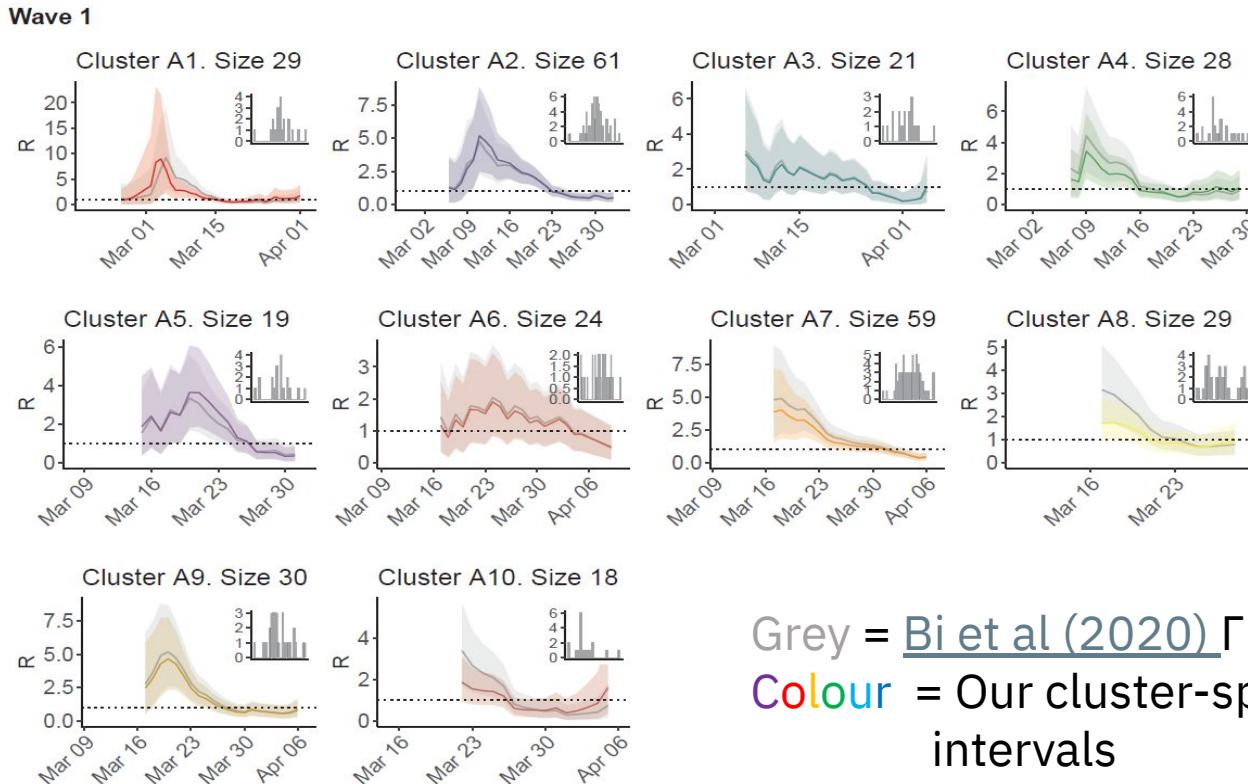
Among the shorter: packing  
plants, schools

Among the longer: aged-care,  
healthcare, housing

Using a larger range of 2nd wave clusters, we can compare across different exposure settings



# Estimates of $R_t$ are (sometimes) impacted by the underlying serial interval distribution

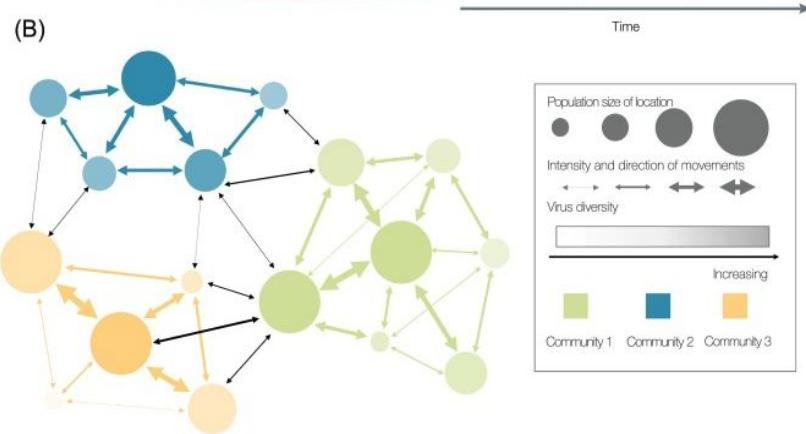
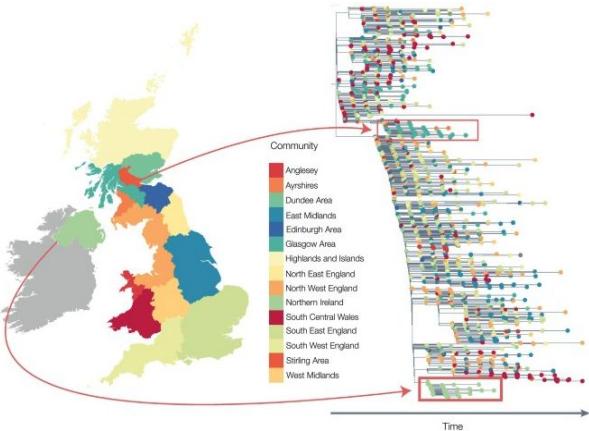


Grey = [Bi et al \(2020\)](#)  $\Gamma(\mu = 6.3, \sigma = 4.2)$   
Colour = Our cluster-specific serial intervals

# In conclusion

- It would have been difficult to do full transmission reconstruction (outbreaker, TransPhylo) for the Victoria data: **low diversity sequences, lots of missing cases (wave 2 particularly)**
- Even still, pathogen sequence data can help us learn about aspects of transmission. Here, we estimate serial intervals, without the need for contact studies
- Broad population sequencing makes it easier to compare serial intervals across time, space, setting, or Variants of Concern (VOC)

# Genomic epi at different scales



Population scale -  
phylogenetics,  
phyldynamics,  
phylogeography

Intermediate  
scale

Local scale -  
methods from  
this course



# Genomic epidemiology offers high resolution estimates of serial intervals for COVID-19

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[Link](#)



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Epidemics

journal homepage: [www.elsevier.com/locate/epidemics](http://www.elsevier.com/locate/epidemics)



A method to estimate the serial interval distribution under partially-sampled data

Kurnia Susvitasari <sup>\*</sup>, Paul Tupper, Jessica E. Stockdale, Caroline Colijn

*Department of Mathematics, Simon Fraser University, Canada*

[Link](#)





# **Methods comparison and opportunities**

# Methods comparison (these are abilities, not quality)

Method	Unsampled hosts	Phylogeny vs pairs	Multiple sequences per host	Simultaneous phylogeny and transmission	Bottleneck >1	Environmental organism	Incorporate epidemiological data (beyond times of collection, infectious period)
BEASTLIER	✗	phylogeny	✓	✓	✗	✗	✗
TransPhylo	✓	phylogeny	✗	✗	✗	✗	✗
Outbreaker 2	✓	pairs	✗	✗	✗	✗	✓(readily)
Phybreak	✗	phylogeny	✗	✓	✗	✗	✗
SCOTTI	✓ (limited)	phylogeny	✓	✓	✓	✓(limited)	✗
BREATH	✓	phylogeny	✗ (in progress)	✓	✗	✗	✗

# Some recent methods and studies

Ke and Vikalo,  
*Graph-Based Reconstruction and Analysis of Disease Transmission Networks Using Viral Genomic Data*, Journal of Computational Biology (2023)

Lindsey et al.  
*Characterising within-hospital SARS-CoV-2 transmission events using epidemiological and viral genomic data across two pandemic waves*, Nature Communications (2022).

Junhang Pan et al,  
*TransFlow: a Snakemake workflow for transmission analysis of Mycobacterium tuberculosis whole-genome sequencing data*, Bioinformatics (2023)

Van der Roest et al, *A Bayesian inference method to estimate transmission trees with multiple introductions; applied to SARS-CoV-2 in Dutch mink farms*, PLoS Comp Bio (2023)

Clustering + transmission reconstruction within clusters via graphs and host importance scores

•

Adapted *Outbreaker2* for hospital settings. Includes ward occupancy data

Pipeline from raw sequences to clustering to transmission reconstruction, combining various existing methods

Extension to Phybreak allowing for multiple pathogen introductions

# Areas that need more methods

Intermediate sampling: between 10-40%

Plasmids and bacteria together

Variable sampling:

- over time (in principle ok in TransPhylo implementation underway)
- across a dataset

Reinfections and coinfections

Environmental transmission

Incorporate more epidemiological data

Connect to forecasting

uncertainty  
environmental source  
variable sampling

host diversity  
unsampled host  
deep sequencing  
larger datasets  
intermediate sampling  
phylogeny

# The potential of genomics for infectious disease forecasting

Received: 1 March 2022

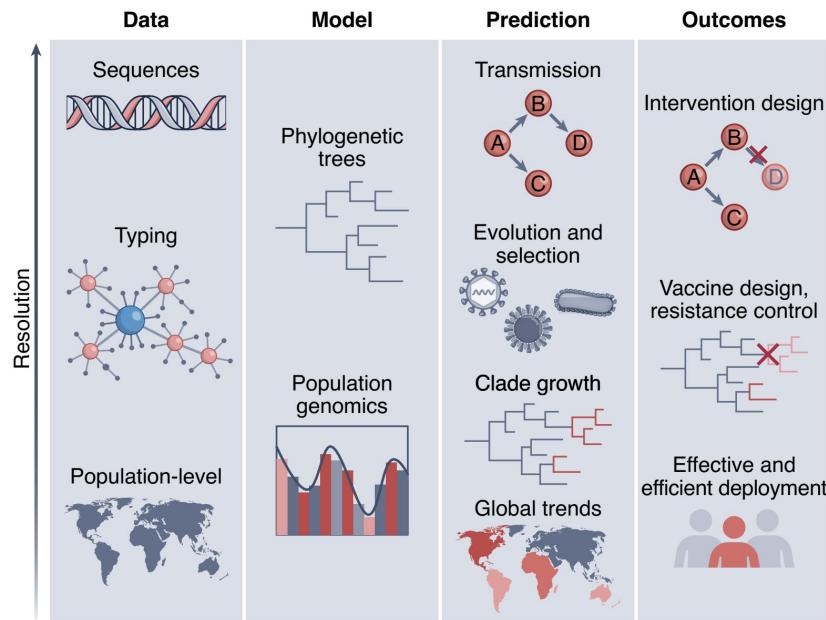
Jessica E. Stockdale , Pengyu Liu  and Caroline Colijn  

Accepted: 18 August 2022

Published online: 20 October 2022

 Check for updates

Genomic technologies have led to tremendous gains in understanding how pathogens function, evolve and interact. Pathogen diversity is measurable at high precision and resolution, in part because over t





# Questions and discussion

