

de.NBI and its Galaxy interface for RNA folding

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You can download the pdfs you will need today here

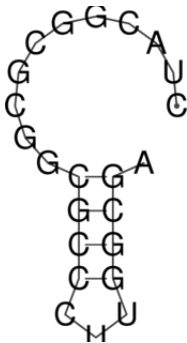
http://www.bioinf.uni-leipzig.de/~fall/RNA_folding_workshop-presentation.pdf

<http://www.bioinf.uni-leipzig.de/~fall/Exercises.pdf>

Goal: Use RNAfold to do a simple structure prediction.

- ▶ Upload the file `rna.fa` into your Galaxy session.
- ▶ Start RNAfold with standard parameters
- ▶ Look into the output

- ▶ CUACGGCGCGGGCGCCCUUGGCGA
- ▶((((...))). (-5.00)



Goal: Use RNAfold to do a structure prediction using the partition function

- ▶ Start RNAfold using --partfunc
- ▶ Look into the output

- ▶ CUACGGCGCGGGCGCCCUUGGCGA
- ▶ MFE:((((...))). (-5.00)
- ▶ PF:{, {{...|||...}}}. [-5.72]

The partition function is a rough measure for the well-definedness of the MFE structure. The third line shows a condensed representation of the pair probabilities of each nucleotide, similar to the dot-bracket notation, followed by the ensemble free energy ($-kT * \ln(Z)$) in kcal/mol

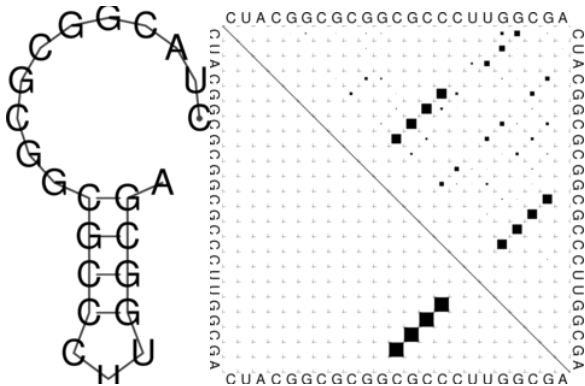
The partition function allows us to calculate the proportion of a certain structure in the ensemble.

Use RNAfold -p to get the ensemble free energy, which is related to the partition function via $\mathbf{F} = -\mathbf{RT} \cdot \ln(\mathbf{Q})$, for the unconstrained (F_u) and the constrained case (F_c), (use option -C), and evaluate $\mathbf{pc} = \exp((F_u - F_c)RT)$ to get the desired probability.

- ▶ CUACGGCGCGGGCGCCCUUGGCGA
- ▶ MFE:((((...))). (-5.00)
- ▶ PF:{, {{...|||...}}}. [-5.72]
- ▶ CS: { 0.00 d=4.66 }
- ▶ MEA:((...))((...))... { 2.90 MEA=14.79 }
- ▶ frequency of mfe structure in ensemble 0.311796; ensemble diversity 6.36

Pseudo bracket notation:

Here, the usual '(', ')', '.', represent bases with a strong preference (more than 2/3) to pair upstream (with a partner further 3'), pair down-stream, or not pair, respectively. '', '', and ',,' are just weaker version of the above and '|' represents a base that is mostly paired but has pairing partners both upstream and downstream. In this case open and closed brackets need not match up.

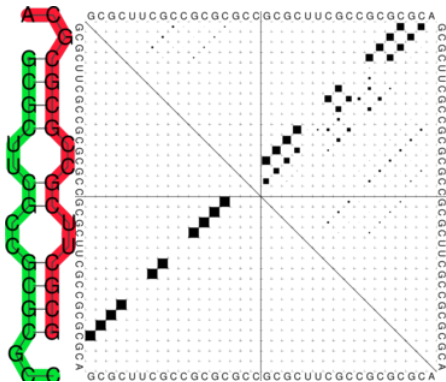


- Goal:** Use SHAPE-directed RNAfold to do a structure prediction
- ▶ Upload the file `rna.simple.shape` into your Galaxy session.
 - ▶ Start RNAfold using the shape file and `-shapeMethod=D`
 - ▶ Look into the output

Goal: Use RNAcofold to predict the cofolding of two sequences.

- ▶ Upload the file `cofold.txt` into your Galaxy session. (Look at it)
- ▶ Start RNAcofold using `cofold.txt` with the `--partfunc` option
- ▶ Look at the output.

- ▶ GCGCUUCGCCGCGCGCC&GCGCUUCGCCGCGCGCA
- ▶ (((((..((..(((...&))))..))..)))... (-17.70)
- ▶ (((((..(,(((,,&))))..),,)))... [-18.26]
- ▶ frequency of mfe structure in ensemble 0.401754 , delta G binding= -3.95



Cofold can use concentrations of molecules for duplex prediction, but this is slow for longer sequences.

Goal: Use RNAduplex to predict *only* intermolecular base pairs of two sequences.

- ▶ Upload the file duplex.txt into your Galaxy session. (Look at it)
- ▶ Start RNAduplex using duplex.txt with standard parameter
- ▶ Look at the output.

RNAduplex does not use concentrations and neglects intramolecular interactions, faster but less reliable, good prefilter.

Goal: Use RNAup to test the RNAduplex result.

- ▶ Start RNAup using duplex.txt with --include_both
- ▶ Look at the output and compare it with the RNAduplex result.

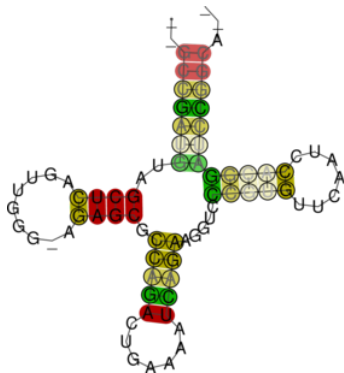
RNAup is also taking intramolecular interactions into account.

Goal: Use RNAalifold to predict the consensus structure

- ▶ Upload the clustal file `alifold.aln` into your Galaxy session.
- ▶ Edit the data type of `alifold.aln` to 'clustal'
- ▶ Use RNAalifold with the `alifold.aln` and `--partfunc` (Calculate partition function: 1)
- ▶ (Download the output) and look at it
- ▶ Bonus: Fold the sequences (`alifold.fa`) individually (RNAfold) and compare the results.

Goal: Use RNAalifold to predict *and visualize* the consensus structure

- ▶ Use RNAalifold with alifold.aln and --color and --aln
- ▶ (Download the output) and look at it



RNAalifold uses covariance information from sequence alignment to predict a consensus structure.

Goal: Use RNACode to predict coding sequences in a MAF alignment.

- ▶ Upload the file `oskar.27way.rnacode.maf` into your Galaxy session.
- ▶ (Change its data type to maf)
- ▶ Use RNACode with the maf file, `--cutoff 0.05`, `--best_region`, `--best_hit`, with GTF output
- ▶ (Download the output) and look at it

Goal: Use RNAz

- ▶ Upload the file `oskar.27way.rnaz.maf` into your Galaxy session.
- ▶ Use RNAz with the maf file
- ▶ (Download the output) and look at it