Bioinformatic approaches to regulatory genomics and epigenomics

376-1347-00L - 2022 | week 05

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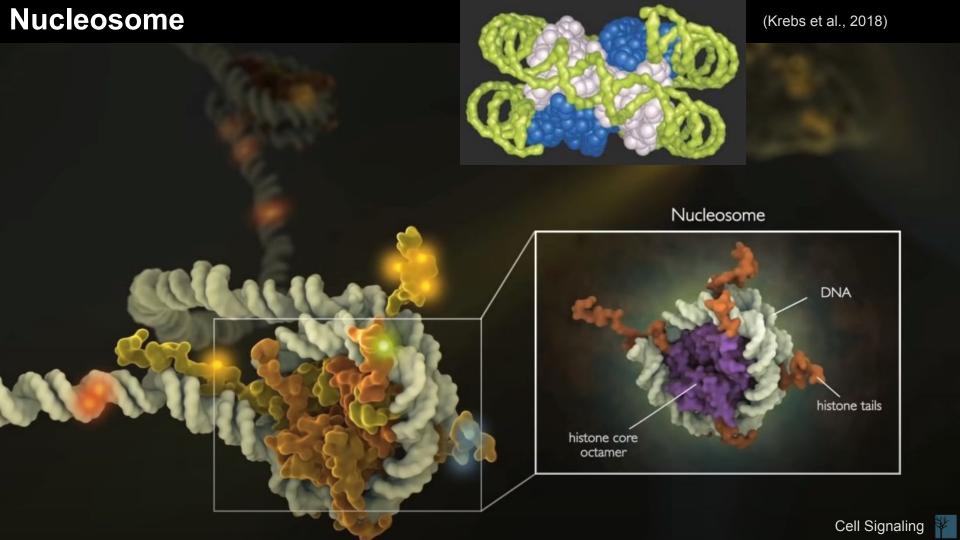


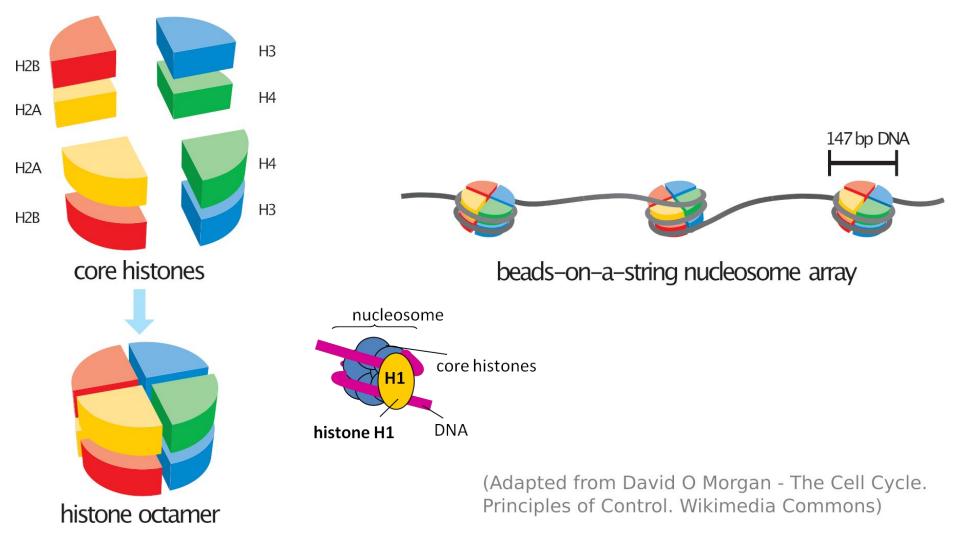
Plan

• The 'histone code' & functional elements

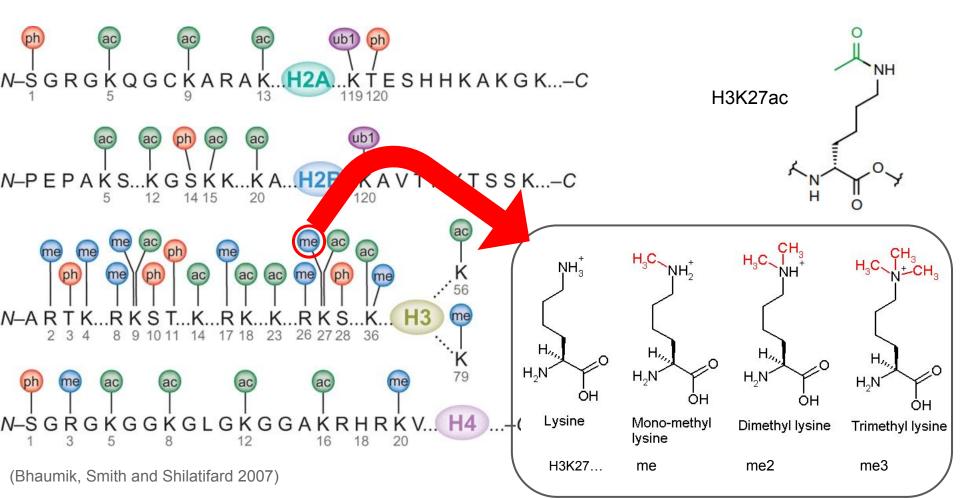
- Debriefing on the assignment
- Exploring overlaps

p300 & enhancers

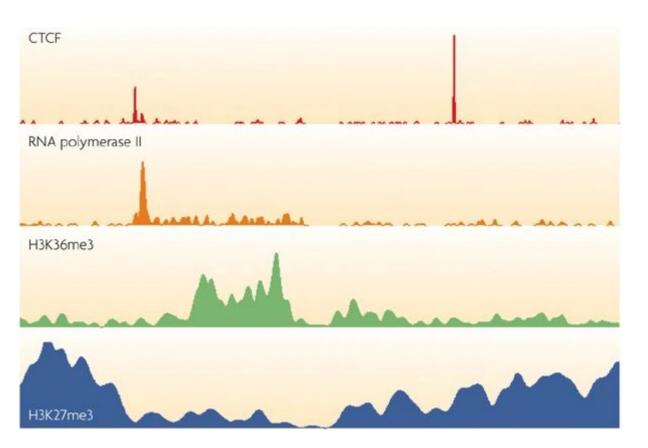




Many residues on the histone tails can be post-translationally modified

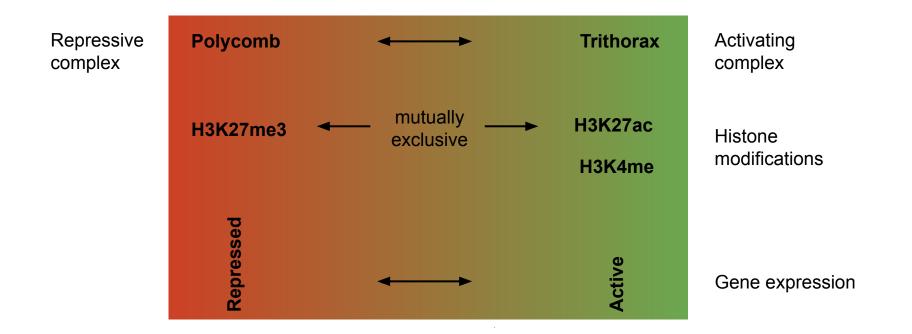


Some histone modifications appear to be very localized, e.g. happening on a specific nucleosome, while most are much more broadly distributed



The strategy of calling 'peaks' must therefore be adapted (e.g. "broad" option of most peak-callers)

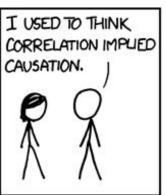
There is a very strong association of certain histone marks and activation or repression

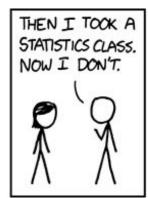


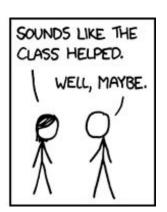
But which comes first?

Causality or correlation?

Are histone modifications responsible for activation/repression, or are they merely associated side-effects?

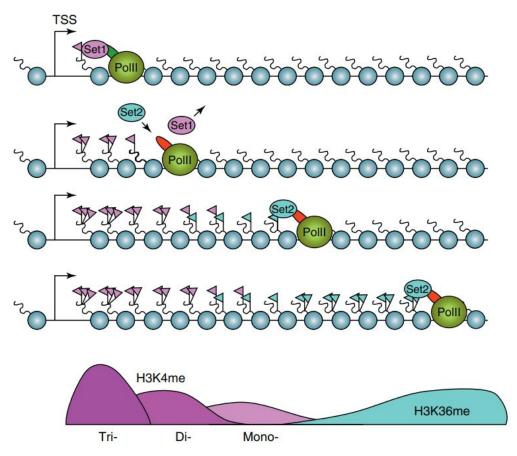




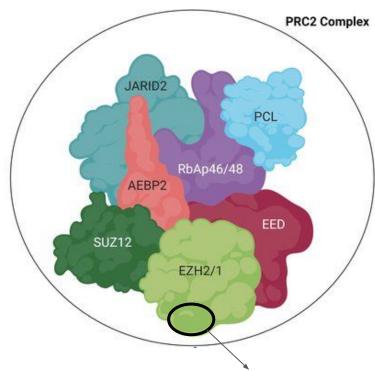


(https://xkcd.com/552)

Transcription-mediated histone modification



The example of H3K27me3, chiefly deposited by the polycomb repressive complex (PRC2)



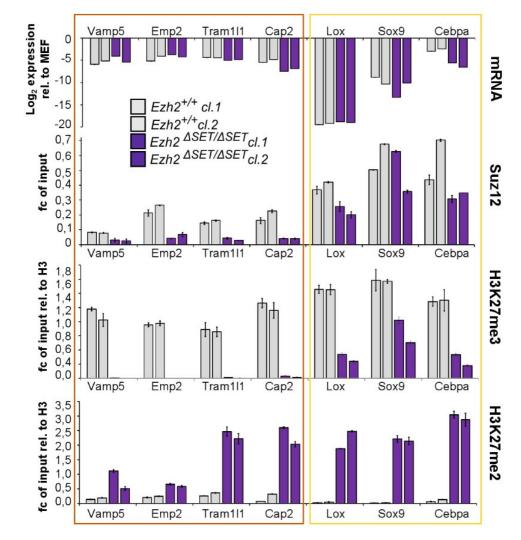
EHZ2's SET domain catalyzes the addition of a 3rd methyl group to H3K27, i.e. H3K27me2 → H3K27me3

Abolishing the enzymatic activity of *Ezh2*, the gene responsible for depositing H3K27me3, abolishes (most of) the mark but does not prevent the repression of the target genes, nor cellular reprogramming

(Fragola et al., PLoS Genetics 2013)

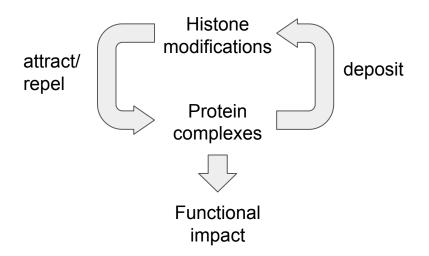
Similarly, the loss of H3K4me3 appears to have no effect on nascent transcription

(Murray et al., bioRxiv 2019)



Causality or correlation?

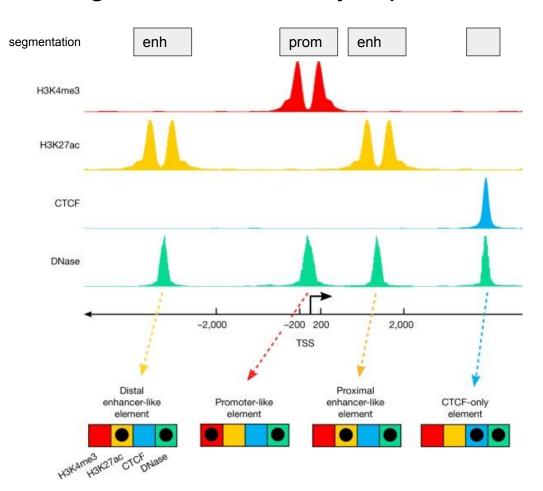
The jury is still out, but the current most likely view is somewhere in the middle



Whether they're causative or not, they can serve as **proxies** for function.

This means that profiling a few histone modifications gives an overview of the epigenomic landscape of a cellular state which would otherwise require profiling all the potentially-relevant factors/complexes

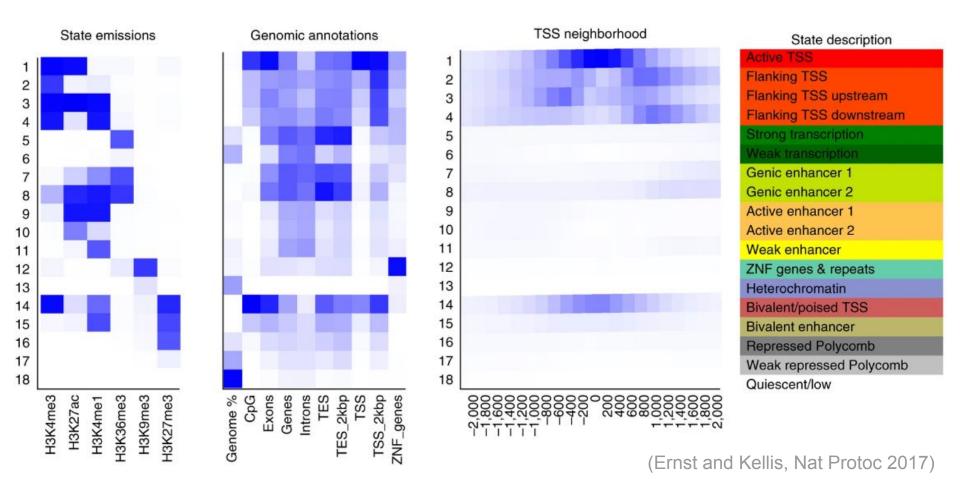
A signature-based encyclopedia of DNA elements



ENCODE's "signature strategy":

- Different types of functional genetic elements are associated with different chemical signatures
- We can identify functional elements by identifying these signatures genome-wide

So how many kinds of functional elements/states are there?



Some stuff is pretty clear:

- Transcription start site (TSS):
 - H3K4me3 is almost always associated with a
 - Active TSS are marked by H3K27ac
 - So-called "poised" (or bivalent) TSS are instead marked by both H3K4me3 and H3K27me3

Enhancers:

- Most enhancers have H3K4me1
- Active enhancers are marked by H3K27ac
- So-called "poised" (or bivalent) enhancers are marked by H3K4me1 and H3K27me3
- Repressed elements are marked by H3K27me3
- Heterochromatin is marked by H3K9me3
- Insulators: CTCF+cohesin

Assignment debriefing

& practical

Inconsistent seqlevels

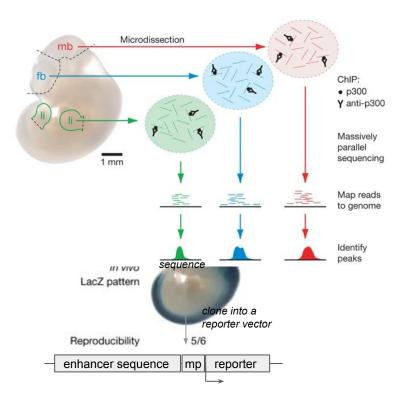
```
overlap_H3K27ac <- overlapsAny(peaks_p300, peaks_H3K27ac)

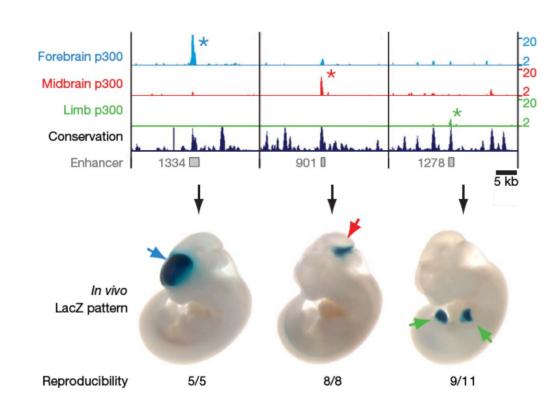
## Warning in .Seqinfo.mergexy(x, y): The 2 combined objects have no sequence levels in common. (Use
## suppressWarnings() to suppress this warning.)

table(overlap_H3K27ac)

## overlap_H3K27ac
## FALSE
## 6394</pre>
```

p300 and validation of enhancer activity





Assignment

 Using the peaks you downloaded last week, identify bivalent domains (H3K27me3 + H3K4me3) in mouse embryonic stem cells (mESC)

- What happens to those regions upon differentiation?
 - Choose a differentiated cell type (e.g. hepatocytes, neural progenitor, or smooth muscle cells)
 - Download the H3K27me3 and H3K4me3 peaks from this cell type
 - How many of the mESC bivalent domains are, in this differentiated cell type, overlapping either mark or their combination?