

Models for Count Data

FW8051 Statistics for Ecologists

Department of Fisheries, Wildlife and Conservation Biology



Learning Objectives

- Be able to fit regression models appropriate for count data in R and JAGS
 - Poisson regression models
 - Quasi-Poisson (R only)
 - Negative Binomial regression
- Be able to evaluate model fit
 - Residual plots
 - Goodness-of-fit tests
- Use deviances and AIC to compare models.
- Use an offset to model rates and densities, accounting for variable survey effort
- Be able to describe statistical models and their assumptions using equations and text and match parameters in these equations to estimates in computer output.

```
glmPdace<-glm(longnosedace~acreage+do2+maxdepth+no3+so4+temp,  
              data=longnosedace, family=poisson())  
summary(glmPdace)
```

Call:

```
glm(formula = longnosedace ~ acreage + do2 + maxdepth + no3 +  
     so4 + temp, family = poisson()), data = longnosedace)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-9.234	-4.086	-1.662	1.771	14.362

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)	
(Intercept)	-1.564e+00	2.818e-01	-5.551	2.83e-08	***
acreage	3.843e-05	2.079e-06	18.480	< 2e-16	***
do2	2.259e-01	2.126e-02	10.626	< 2e-16	***
maxdepth	1.155e-02	6.688e-04	17.270	< 2e-16	***
no3	1.813e-01	1.068e-02	16.974	< 2e-16	***
so4	-6.810e-03	3.622e-03	-1.880	0.0601	.
temp	7.854e-02	6.530e-03	12.028	< 2e-16	***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for poisson family taken to be 1)

Null deviance: 2766.9 on 67 degrees of freedom

Model

$$Dace_i \sim \text{Poisson}(\lambda_i)$$

$$\log(\lambda_i) = \beta_0 + \beta_1 Acreage_i + \beta_2 DO2_i + \beta_3 maxdepth_i + \\ \beta_4 NO3_i + \beta_5 SO4_i + \beta_6 temp_i$$

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$$\lambda_i = \exp(\beta_0 + \beta_1 Acreage_i + \beta_2 DO2_i + \beta_3 maxdepth_i + \beta_4 NO3_i + \\ \beta_5 SO4_i + \beta_6 temp_i)$$

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Interpretation

$$Dace_i \sim \text{Poisson}(\lambda_i) \quad (1)$$

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- $\exp(\beta_0)$ = expected mean when all predictors are equal to 0
- $\beta_2 = 0.226$ = expected change in the log mean [i.e., $\log(\lambda)$], per unit change in D02, while holding everything else constant.

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- $\exp(\beta_0)$ = expected mean when all predictors are equal to 0
- $\beta_2 = 0.226$ = expected change in the log mean [i.e., $\log(\lambda)$], per unit change in D02, while holding everything else constant.
- $\exp(\beta_2) = 1.25$ = we expect the mean to increase by a factor of 1.25 for every 1 unit change in D02 (and holding everything else constant)

Inference

Rely on asymptotic Normality for Maximum Likelihood Estimators

$$\hat{\beta} \sim N(\beta, I^{-1}(\theta))$$

```
# Store coefficients and their standard errors  
beta.hats <- coef(glmPdace)  
ses <- sqrt(diag(vcov(glmPdace)))  
round(cbind(beta.hats-1.96*ses, beta.hats+1.96*ses), 3)
```

```
##           [,1]    [,2]  
## (Intercept) -2.117 -1.012  
## acreage      0.000  0.000  
## do2          0.184  0.268  
## maxdepth     0.010  0.013  
## no3          0.160  0.202  
## so4         -0.014  0.000  
## temp         0.066  0.091
```

Profile confidence intervals

Inverts the likelihood ratio test to form confidence intervals
(see Section 10.10 of the book)

```
#' Or, can use profile likelihood intervals  
confint(glmPdata)
```

```
## Waiting for profiling to be done...
```

```
##              2.5 %          97.5 %  
## (Intercept) -2.117222e+00 -1.012594e+00  
## acreage      3.432623e-05  4.247872e-05  
## do2          1.841122e-01  2.674327e-01  
## maxdepth     1.023509e-02  1.285684e-02  
## no3          1.603516e-01  2.022292e-01  
## so4         -1.400207e-02  1.966821e-04  
## temp         6.576200e-02  9.136105e-02
```

Confidence Intervals for $\exp(\beta)$

1. Calculate a CI for β
2. Exponentiate the confidence limits

```
round(cbind(exp(beta.hats-1.96*ses), exp(beta.hats+1.96*ses)), 3)
```

```
##           [,1]  [,2]  
## (Intercept) 0.120 0.363  
## acreage     1.000 1.000  
## do2         1.202 1.307  
## maxdepth    1.010 1.013  
## no3         1.174 1.224  
## so4         0.986 1.000  
## temp        1.068 1.096
```


Residuals

Deviance residuals = $\text{sign}(y_i - \mu_i)\sqrt{d_i}$, where:

- d_i is the contribution of the i^{th} observation to the residual deviance (may be useful for spotting outliers/influential points)
- $\text{sign} = 1$ if $y_i > \mu_i$ and -1 if $y_i < \mu_i$.

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Often Deviance and Pearson residuals are similar.

Residual Plots

- Residuals versus fitted values
- Residuals versus predictors
- Residuals over time or space (to diagnose possible spatial/temporal correlation)

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See also `check_model` in the `performance` package.

Goodness-of-fit: Is the Poisson distribution appropriate?

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If the model is appropriate: “goodness-of-fit” statistics for real and simulated data should be similar

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- If the p-value is not small, then. . .

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- If the p-value is not small, then. . . we do not have enough evidence to suggest the model is not appropriate

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Loop:

- Simulate a random vector of β 's using $MVN(\hat{\beta}, \widehat{Cov}(\hat{\beta})^2)$
- Use these β 's to form λ_i 's = $\exp(\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i})$
- Simulate new data using these λ_i 's and `rpois()`.
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Lets do this!

Overdispersion, $Var(Y|X) > E(Y|X)$

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- Spatial, temporal, within-individual clustering (repeated measures)
- Response may be due to a mixture of random processes
 - Presence/absence determined by suitable habitat
 - Counts | suitable habitat may be Poisson
 - Leads to “zero-inflation” models

Testing for Overdispersion: Residual Deviance and Pearson's χ^2

Some compare:

$$\text{Residual deviance} \quad \text{or} \quad \sum_{i=1}^n \frac{(Y_i - E[Y_i|X_i])^2}{\text{Var}[Y_i|X_i]}$$

to a χ^2 distribution with $n - p$ degrees of freedom.

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So, best to test using (predictive) simulation techniques discussed earlier (e.g., using the Pearson χ^2 statistic).

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We may obtain reasonable estimates of β , but:

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Before 'correcting for overdispersion' , consider whether:

- You may have left out important predictors
- If you need to allow for non-linear relationships (residual plots).

How to deal with overdispersion

Relaxing the Poisson Assumption

Poisson regression:

- $\log(\lambda_i) = \log(\mu_i) = \beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}$
- $f(y_i) \sim \text{Poisson}(\lambda_i)$

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- $E[Y_i|X_i] = \lambda_i = e^{\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}}$
- $\text{Var}[Y_i|X_i] = \lambda_i = e^{\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}}$

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- $\text{Var}[Y_i|X_i] = \lambda_i = e^{\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}}$

What if $E[Y_i|X_i] = \exp(\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i})$ is appropriate, but the Poisson distribution is not?

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What if $E[Y_i|X_i] = \exp(\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i})$ is appropriate, but the Poisson distribution is not?

In particular, what if $\text{Var}[Y_i|X_i] > E[Y_i|X_i]$?

Relaxing the Poisson Assumption

Poisson regression:

- $\log(\lambda_i) = \log(\mu_i) = \beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}$
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Option 1: Bootstrap!

Option 2: Variance Inflation

Another option: add a scale parameter to inflate variances.

- $E[Y_i|X_i] = \mu = e^{\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}}$
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Estimate of overdispersion, $\hat{\phi}$ by either:

- $\hat{\phi} = \frac{\text{Residual deviance}}{(n-p)}$
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Zuur et al. recommend:

- If $\phi > 1.5$ should adjust for overdispersion
- If greater than 15 or 20, consider alternative methods (Negative Binomial, zero-inflation models, Poisson-Normal model)

Variance Inflation: Quasilikelihood

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- $\hat{\beta}$ will be unchanged, but SE will be larger by a factor of $\sqrt{\hat{\phi}}$.
- No longer “maximum likelihood”
- **quasiliquelihood** (more on this later)
- Modeling the first two moments of Y ($E[Y|X]$, $\text{Var}[Y|X]$)

Poisson Model with Overdispersion parameter

Fit to slug data:

- $\hat{\beta}$ does not change
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Lets do this!

Option 3: Use a different distribution than Poisson

$$Dace_i \sim \text{NegativeBinomial}(\mu_i, \theta)$$

$$\log(\mu_i) = \beta_0 + \beta_1 \text{Acreage}_i + \beta_2 \text{DO2}_i + \beta_3 \text{maxdepth}_i + \\ \beta_4 \text{NO3}_i + \beta_5 \text{SO4}_i + \beta_6 \text{temp}_i$$

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- $E[Y_i] = \mu_i$
- $\text{Var}[Y_i] = \mu_i + \mu_i^2 / \theta$

Poisson is a limiting case (when $\theta \rightarrow \infty$)

Negative Binomial Models in R

Can fit negative binomial models in R using the `glm.nb` function in the MASS library

```
glm.nb(y ~ x, data=)
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Lets do this and inspect goodness of fit!

Model comparisons

For large samples, the difference in deviances for nested models should be $\sim \chi^2$ with df = difference in number of parameters between the two models.

$$D_2 - D_1 \sim \chi^2_{df}$$

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- Can use `drop1(model, test="Chi")` (equivalent to a likelihood ratio test) or `Anova` in `car` package
- Can use forward, backwards, stepwise selection (with the same dangers/caveats related to overfitting); see `stepAIC` in `MASS` library (for backwards selection)

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- → smaller AIC is better

Can use to compare nested and non-nested models.

- Not always appropriate for certain types of models (problematic if you have latent variables, e.g., mixture models)

Negative Binomial in JAGS

JAGS: `dnegbin` specified in terms of parameters (p, θ)

Negative Binomial in JAGS

JAGS: **dnegbin** specified in terms of parameters (p, θ)

We will specify the model in terms of μ and θ , then solve for p :

```
log(mu[i]) <- alpha + beta*IRook[i]  
p[i] <- theta/(theta+mu[i])  
slugs[i] ~ dnegbin(p[i],theta)
```

Poisson-normal model

$$\begin{aligned} \log(\lambda_i) = \log(\mu_i) &= \beta_0 + \beta_1 X_{1,i} + \dots \beta_p X_{p,i} + \epsilon_i \\ \epsilon_i &\sim N(0, \sigma^2) \end{aligned}$$

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- $Var[Y_i|X_i] = \mu_i + (e^{\sigma^2} - 1)\mu_i^2$

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Count data, Y , are often collected:

- over varying lengths of time
- in sample units that have different areas

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We may be interested in modeling rates:

$$E[Y_i|X_i]/\text{Time}_i$$

Or densities:

$$E[Y_i|X_i]/\text{Area}_i$$

We may want to account for variable survey effort (varying times or areas)!

Offsets

Poisson and negative binomial models for rate data:

$$\log(E[Y_i|X_i]/\text{Time}_i) = \beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}$$

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$\log(\text{Time}_i)$ is called an **offset** and can be modeled using:

```
glm(y~x + offset(log(time)), data= , family =  
poisson())
```

An **offset** is an explanatory variable with a regression coefficient fixed at 1.

See **PoissonOffsetTemplate.R** and **PoissonOffset.R** (in the Generalized linear models folder) for an exercise fitting a Poisson model with an offset.

DIC

Martyn Plummer (creator of JAGS):

DIC [like AIC] is (an approximation to) a theoretical out-of-sample predictive error.

“The deviance information criterion (DIC) is widely used for Bayesian model comparison, despite the lack of a clear theoretical foundation. . . . valid only when the effective number of parameters in the model is much smaller than the number of independent observations. In disease mapping, a typical application of DIC, this assumption does not hold and DIC under-penalizes more complex models. Another deviance-based loss function, derived from the same decision-theoretic framework, is applied to mixture models, which have previously been considered an unsuitable application for DIC.”

Andrew Gelman: “I don’t really ever know what to make of DIC. On one hand, it seems sensible... On the other hand, I don’t really have any idea what I would do with DIC in any real example. In our book we included an example of DIC—people use it and we don’t have any great alternatives—but I had to be pretty careful that the example made sense. Unlike the usual setting where we use a method and that gives us insight into a problem, here we used our insight into the problem to make sure that in this particular case the method gave a reasonable answer.”

http://andrewgelman.com/2011/06/22/deviance_dic_ai/

Model comparisons

There are other potential options out there (e.g., WIC, cross-validation estimates of predictive error, etc)

Hooten, Mevin B, and N Thompson Hobbs. 2015. "A Guide to Bayesian Model Selection for Ecologists." *Ecological Monographs* 85 (1): 3–28.