JAGS Examples from Kery's book Introduction to WinBugs

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http://www.mbr/-pwrc.usgs.gov/software/kerybook/R/_WB/_code.txt Most of the code will run, with a few modifications, using JAGS. In particular, we have to add BUGSoutput when referencing any of the output from JAGS.

- In Kery's book: plot (out\$mean\$predicted, out\$mean\$residual)
- We need to use: plot (out\$BUGSoutput\$mean\$predicted, out\$BUGSoutput\$mean\$residual).

Trends in Wallcreeper Counts



- Proportion of sample quadrats where the bird is observed
- Assume deviations from a linear trend are normally distributed

Ch 8: Linear regression

Prior and likelihood is very similar to what we have already seen.

```
linreg<-function() {
# Priors
  alpha ~ dnorm(0,0.001)
  beta ~ dnorm(0,0.001)
  sigma ~ dunif(0, 100)

# Likelihood
  for (i in 1:n) {
     y[i] ~ dnorm(mu[i], tau)
     mu[i] <- alpha + beta*x[i]
}</pre>
```

Derived quantities

```
# Derived quantities
tau <- 1/ (sigma * sigma)
p.decline <- 1-step(beta) # Probability of decline</pre>
```

Step() = 1 if what is in the "()" is > 0 (and 0 otherwise). So, this calculates the probability that $\beta < 0$.

Residuals

```
for (i in 1:n) {
   residual[i] <- y[i]-mu[i]  # Residuals for observed data
   predicted[i] <- mu[i]  # Predicted values
   sq[i] <- pow(residual[i], 2)  # Squared residuals for observed of</pre>
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Note: we will get n.iter residuals for each data point! Why?

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Residual: $r[i] = y_i - (alpha + beta * x[i]).$

We generate n_{MCMC} values of alpha and beta, so n_{MCMC} values of r[i]!

- Generate new ("ideal") data: y.new[i] ~ dnorm(mu[i], tau)
 - Using the assumed model
 - Using parameters drawn from the posterior distribution (accounts for parameter uncertainty)

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- Calculate measure of "fit" (or lack of fit) for for the observed data and simulated data
 - fit <- sum(sq[]) sum of squared residuals for the observed data
 - fit.new <- sum(sq.new[]) (same for simulated data)

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- test = 1 if SS(simulated data) > SS(observed data), i.e., the model fits the observed data better than the simulated data.
- bpvalue <- mean(test) (how often does the model fit observed data better than the simulated data)
- Want bpvalue to be large (can't conclude there is a lack of fit).

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For each value of X = x (loop), he generates n_{MCMC} predictions.

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for(i in 1:length(x)) {
   predictions[i,] <- out$BUGSoutput$sims.list$alpha +
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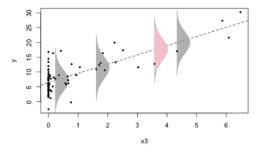
- Gives us n_{MCMC} draws of $E[Y|X=x]=\alpha+x\beta$. What does this tell us?
- This is the posterior distribution of the fitted line (tells us about the line and its uncertainty)

Prediction Intervals

What about prediction intervals? Intervals that are likely to contain a new observation, if collected?

Prediction Intervals

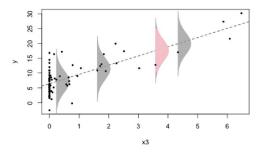
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Prediction Intervals

What about prediction intervals? Intervals that are likely to contain a new observation, if collected?



- Requires we also consider the variability about the line
- This variability is described by the errors $\sim N(0, \sigma^2)$.

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- random normal deviates
- add: rnorm(n_{MCMC} , mean = 0, sd = reference posterior for σ)
- gives us the posterior of $Y_{new}|X$ (rather than E[Y|X])
- Try to modify Kery's code to do this! Hint: to figure out n_{MCMC} , inspect the dimension of out\$BUGSoutput\$sims.list\$alpha

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Actually, we did do this when we generated y.new.

We could have just added y . new to our "parameters.to.save" vector and then summarized the quantiles of the jags output to get the prediction intervals.

Exercise 2: Prediction Intervals (frequentist)

For linear models fit in R, we can generate confidence and prediction intervals using the predict function, and more specifically, predict.lm

- Look up the help file for predict.lm and examples at bottom
- Calculate these intervals for the fitted linear model.
- Plot these intervals using the lines function along with Bayesian prediction intervals

Chapter 11: ANCOVA model (importance of priors and	
scaling)	

Relationship between Body Mass and Body Length



- Asp viper's from 3 populations (Pyrenees, Massif Central, Jura Mountains) in Switzerland
- Interested in population-specific differences in the body-mass relationship

Linear regression model

$$y_i = \alpha_0 + \beta_1 x_i + \epsilon_i$$

- y_i = body mass of individual i
- x_i = body length of snake i

How dow we capture "population-specific differences in the body-mass relationship"?

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How dow we capture "population-specific differences in the body-mass relationship"?

Include interactions!

Example CH 11.2

Effects parameterization:

$$y_i = \alpha_{pyr} + \beta_1 I_{MC,i} + \beta_2 I_{Jura,i} + \beta_3 x_i + \beta_4 x_i I_{MC,i} + \beta_5 x_i I_{Jur,i} + \epsilon_i$$

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Identify the parameters used in the simulation:

- Effects model (α and $\beta_1 \beta_5$)
- Means model (α and β) for each population.

Parameters

Effects parameterization

- $\alpha = -250$
- $\beta_1 = 150$
- $\beta_2 = 200$
- $\beta_3 = 6$
- β₄ -3
- β₅ -4

Means parameterization

- Pyrenees: $\alpha = -250$, $\beta = 6$
- Massif: $\alpha = -100$, $\beta = 3$
- Jura: $\alpha = -50$, $\beta = 2$

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```
curve(dnorm(x, 0,sqrt(1/0.001)), from=-250, to=250)
```

