

Models for Count Data

FW8051 Statistics for Ecologists

Department of Fisheries, Wildlife and Conservation Biology



```
glmPdace<-glm(longnosedace~acreage+do2+maxdepth+no3+so4+temp,
               data=longnosedace, family=poisson())
summary(glmPdace)
```

```
Call:
glm(formula = longnosedace ~ acreage + do2 + maxdepth + no3 +
    so4 + temp, family = poisson(), data = longnosedace)
```

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-1.564e+00	2.818e-01	-5.551	2.83e-08 ***
acreage	3.843e-05	2.079e-06	18.480	< 2e-16 ***
do2	2.259e-01	2.126e-02	10.626	< 2e-16 ***
maxdepth	1.155e-02	6.688e-04	17.270	< 2e-16 ***
no3	1.813e-01	1.068e-02	16.974	< 2e-16 ***
so4	-6.810e-03	3.622e-03	-1.880	0.0601 .
temp	7.854e-02	6.530e-03	12.028	< 2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for poisson family taken to be 1)

Null deviance: 2766.9 on 67 degrees of freedom
 Residual deviance: 1590.0 on 61 degrees of freedom
 AIC: 1936.9

Number of Fisher Scoring iterations: 5

- Be able to fit regression models appropriate for count data in R and JAGS
 - Poisson regression models
 - Quasi-Poisson (R only)
 - Negative Binomial regression
- Be able to evaluate model fit
 - Residual plots
 - Goodness-of-fit tests
- Use deviances and AIC to compare models.
- Use an offset to model rates and densities, accounting for variable survey effort
- Be able to describe statistical models and their assumptions using equations and text and match parameters in these equations to estimates in computer output.

Model

$$Dace_i \sim \text{Poisson}(\lambda_i)$$

$$\log(\lambda_i) = \beta_0 + \beta_1 \text{Acreage}_i + \beta_2 \text{DO2}_i + \beta_3 \text{maxdepth}_i +$$

$$\beta_4 \text{NO3}_i + \beta_5 \text{SO4}_i + \beta_6 \text{temp}_i$$

$$\lambda_i = \exp(\beta_0 + \beta_1 \text{Acreage}_i + \beta_2 \text{DO2}_i + \beta_3 \text{maxdepth}_i + \beta_4 \text{NO3}_i +$$

$$\beta_5 \text{SO4}_i + \beta_6 \text{temp}_i)$$

$$\lambda_i = e^{(\beta_0)} e^{(\beta_1 \text{Acreage}_i)} e^{(\beta_2 \text{DO2}_i)} e^{(\beta_3 \text{maxdepth}_i)} e^{(\beta_4 \text{NO3}_i)} e^{(\beta_5 \text{SO4}_i)} e^{(\beta_6 \text{temp}_i)}$$

Interpretation

$$Dace_i \sim \text{Poisson}(\lambda_i) \quad (1)$$

$$\log(\lambda_i) = \beta_0 + \beta_1 \text{Acreage}_i + \beta_2 \text{DO2}_i + \beta_3 \text{maxdepth}_i + \beta_4 \text{NO3}_i + \beta_5 \text{SO4}_i + \beta_6 \text{temp}_i \quad (2)$$

$$\lambda_i = e^{(\beta_0)} e^{(\beta_1 \text{Acreage}_i)} e^{(\beta_2 \text{DO2}_i)} e^{(\beta_3 \text{maxdepth}_i)} e^{(\beta_4 \text{NO3}_i)} e^{(\beta_5 \text{SO4}_i)} e^{(\beta_6 \text{temp}_i)}$$

- β_0 = expected log mean when all predictors are equal to 0
- $\exp(\beta_0)$ = expected mean when all predictors are equal to 0
- $\beta_2 = 0.226$ = expected change in the log mean [i.e., $\log(\lambda)$], per unit change in D02, while holding everything else constant.
- $\exp(\beta_2) = 1.25$ = we expect the mean to increase by a factor of 1.25 for every 1 unit change in D02 (and holding everything else constant)

Profile confidence intervals

Inverts the likelihood ratio test to form confidence intervals (see Section 10.10 of the book)

```
#' Or, can use profile likelihood intervals
confint(glmPdace)
```

```
## Waiting for profiling to be done...
```

```
##              2.5 %          97.5 %
## (Intercept) -2.117222e+00 -1.012594e+00
## acreage      3.432623e-05  4.247872e-05
## do2          1.841122e-01  2.674327e-01
## maxdepth     1.023509e-02  1.285684e-02
## no3          1.603516e-01  2.022292e-01
## so4          -1.400207e-02  1.966821e-04
## temp         6.576200e-02  9.136105e-02
```

Inference

Rely on asymptotic Normality for Maximum Likelihood Estimators

$$\hat{\beta} \sim N(\beta, I^{-1}(\theta))$$

```
# Store coefficients and their standard errors
beta.hats <- coef(glmPdace)
ses <- sqrt(diag(vcov(glmPdace)))
round(cbind(beta.hats*1.96*ses, beta.hats*1.96*ses), 3)
```

```
##              [,1]      [,2]
## (Intercept) -2.117 -1.012
## acreage      0.000  0.000
## do2          0.184  0.268
## maxdepth     0.010  0.013
## no3          0.160  0.202
## so4         -0.014  0.000
## temp         0.066  0.091
```

Confidence Intervals for $\exp(\beta)$

1. Calculate a CI for β
2. Exponentiate the confidence limits

```
round(cbind(exp(beta.hats*1.96*ses), exp(beta.hats*1.96*ses)), 3)
```

```
##              [,1]      [,2]
## (Intercept)  0.120  0.363
## acreage      1.000  1.000
## do2          1.202  1.307
## maxdepth     1.010  1.013
## no3          1.174  1.224
## so4          0.986  1.000
## temp         1.068  1.096
```

Residuals

Deviance residuals = $\text{sign}(y_i - \mu_i) \sqrt{d_i}$, where:

- d_i is the contribution of the i^{th} observation to the residual deviance (may be useful for spotting outliers/influential points)
- $\text{sign} = 1$ if $y_i > \mu_i$ and -1 if $y_i < \mu_i$.

Pearson residuals = $\frac{Y_i - E[Y_i|X_i]}{\sqrt{\text{Var}[Y_i|X_i]}} = \frac{Y_i - \lambda_i}{\sqrt{\lambda_i}}$ for Poisson

- If the Poisson model is appropriate, these residuals should have constant variance

Often Deviance and Pearson residuals are similar.

Residual Plots

- Residuals versus fitted values
- Residuals versus predictors
- Residuals over time or space (to diagnose possible spatial/temporal correlation)

See `residualPlots` in `car` library for useful plots.

See also `check_model` in the `performance` package.

Goodness-of-fit: Is the Poisson distribution appropriate?

How can we assess overall model fit?

Think "Bayesian p-value"...

- Account for uncertainty in $\hat{\beta}$ by drawing values of β from $N(\hat{\beta}, \widehat{\text{Cov}}(\hat{\beta})^2)$
- Use the values of β above to estimate λ (`lambda.hat`)
- Simulate new data (e.g., using `rpois(n, lambda.hat)`)
- Calculate a measure of fit for both real and simulated data
- Rinse, repeat.

If the model is appropriate: "goodness-of-fit" statistics for real and simulated data should be similar

Goodness of fit test

H_0 : the data were generated by the assumed model

H_A : the data were not generated by the assumed model

- The GOF statistics for the simulated data tell us what we might expect to see if the Null hypothesis is true.
- P-value = proportion of time the GOF statistic for **our observed data** is as or more extreme than GOF statistic for the simulated data
- If p-value is small, then we have evidence that the model is not appropriate.
- If the p-value is not small, then...we do not have enough evidence to suggest the model is not appropriate

What should we use as a measure of fit?

One option (see Kery Ch 13), Pearson χ^2 statistic:

$$\chi^2_{n-p} = \sum_{i=1}^n \frac{(Y_i - \hat{E}[Y_i|X_i])^2}{\widehat{\text{Var}}[Y_i|X_i]}$$

Loop:

- Simulate a random vector of β 's using $MVN(\hat{\beta}, \widehat{\text{Cov}}(\hat{\beta})^2)$
- Use these β 's to form λ_i 's = $\exp(\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i})$
- Simulate new data using these λ_i 's and `rpois()`.
- Calculate χ^2_{n-p} for real and simulated data (plugging in $\hat{E}[Y_i|X_i] = \widehat{\text{Var}}[Y_i|X_i] = \hat{\lambda}_i$).

p-value = proportion of times $\chi^2_{sim} \geq \chi^2_{real}$ (want large values)

Lets do this!

Overdispersion, $\text{Var}(Y|X) > E(Y|X)$

Reasons data may be overdispersed:

- Omitted variables
- Explanatory and response variables may be measured with error
- Model may be mis-specified (relationship between $\log(\mu)$ and x may be non-linear)
- Outliers
- Spatial, temporal, within-individual clustering (repeated measures)
- Response may be due to a mixture of random processes
 - Presence/absence determined by suitable habitat
 - Counts | suitable habitat may be Poisson
 - Leads to "zero-inflation" models

Testing for Overdispersion: Residual Deviance and Pearson's χ^2

Some compare:

$$\text{Residual deviance} \quad \text{or} \quad \sum_{i=1}^n \frac{(Y_i - E[Y_i|X_i])^2}{\widehat{\text{Var}}[Y_i|X_i]}$$

to a χ^2 distribution with $n - p$ degrees of freedom.

Large values can be caused by:

- Mis-specified model (missing predictors)
- Mis-specified distribution
- Outliers
- **Small numbers of observations for any set of unique covariate values**

So, best to test using (predictive) simulation techniques discussed earlier (e.g., using the Pearson χ^2 statistic).

Consequences of Overdispersion

We may obtain reasonable estimates of β , but:

- SE may be too small
- We may select overly complex models

Before 'correcting for overdispersion', consider whether:

- You may have left out important predictors
- If you need to allow for non-linear relationships (residual plots).

Relaxing the Poisson Assumption

Poisson regression:

- $\log(\lambda_i) = \log(\mu_i) = \beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}$
- $f(y_i) \sim \text{Poisson}(\lambda_i)$
- $E[Y_i|X_i] = \lambda_i = e^{\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}}$
- $\text{Var}[Y_i|X_i] = \lambda_i = e^{\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}}$

What if $E[Y_i|X_i] = \exp(\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i})$ is appropriate, but the Poisson distribution is not?

In particular, what if $\text{Var}[Y_i|X_i] > E[Y_i|X_i]$?

Option 1: Bootstrap!

Option 2: Variance Inflation

Another option: add a scale parameter to inflate variances.

- $E[Y_i|X_i] = \mu = e^{\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}}$
- $\text{Var}[Y_i|X_i] = \phi \mu = \phi e^{\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}}$

Estimate of overdispersion, $\hat{\phi}$ by either:

- $\hat{\phi} = \frac{\text{Residual deviance}}{(n-p)}$
- $\hat{\phi} = \frac{1}{n-p} \sum_{i=1}^n \frac{(Y_i - E[Y_i|X_i])^2}{\text{Var}[Y_i|X_i]}$ (what R does, probably better)

Zuur et al. recommend:

- If $\phi > 1.5$ should adjust for overdispersion
- If greater than 15 or 20, consider alternative methods (Negative Binomial, zero-inflation models, Poisson-Normal model)

Variance Inflation: Quasiliquelihood

In R, can use `glm` with `family = quasipoisson()`

Will estimate an inflation factor using:

$$\hat{\phi} = \frac{1}{n-p} \sum_{i=1}^n \frac{(Y_i - E[Y_i|X_i])^2}{\text{Var}[Y_i|X_i]}$$

- $\hat{\beta}$ will be unchanged, but SE will be larger by a factor of $\sqrt{\hat{\phi}}$.
- No longer "maximum likelihood"
- **quasiliquelihood** (more on this later)
- Modeling the first two moments of Y ($E[Y|X]$, $\text{Var}[Y|X]$)

Poisson Model with Overdispersion parameter

Fit to slug data:

- $\hat{\beta}$ does not change
- SE's inflated by $\sqrt{\hat{\phi}}$

Lets do this!

Option 3: Use a different distribution than Poisson

$$D_{ace_i} \sim \text{NegativeBinomial}(\mu_i, \theta)$$

$$\log(\mu_i) = \beta_0 + \beta_1 \text{Acreage}_i + \beta_2 \text{DO2}_i + \beta_3 \text{maxdepth}_i +$$

$$\beta_4 \text{NO3}_i + \beta_5 \text{SO4}_i + \beta_6 \text{temp}_i$$

$$\mu_i = \exp(\beta_0 + \beta_1 \text{Acreage}_i + \beta_2 \text{DO2}_i + \beta_3 \text{maxdepth}_i + \beta_4 \text{NO3}_i +$$

$$\beta_5 \text{SO4}_i + \beta_6 \text{temp}_i)$$

- $E[Y_i] = \mu_i$
- $\text{Var}[Y_i] = \mu_i + \mu_i^2 / \theta$

Poisson is a limiting case (when $\theta \rightarrow \infty$)

Negative Binomial Models in R

Can fit negative binomial models in R using the `glm.nb` function in the `MASS` library

```
glm.nb(y ~ x, data=)
```

Lets do this and inspect goodness of fit!

Model comparisons

For large samples, the difference in deviances for nested models should be $\sim \chi^2$ with df = difference in number of parameters between the two models.

$$D_2 - D_1 \sim \chi_{df}^2$$

- Can use `drop1(model, test="Chi")` (equivalent to a likelihood ratio test) or `Anova` in `car` package
- Can use forward, backwards, stepwise selection (with the same dangers/caveats related to overfitting); see `stepAIC` in `MASS` library (for backwards selection)

AIC

$\text{AIC} = -2 \times \log L(\hat{\theta}|y) + 2 \times \text{number of parameters}$

- measure of "fit" with "penalty" for model complexity
- the larger log-likelihood, the smaller AIC
- for similar likelihoods, AIC will be smaller for simpler models
- \rightarrow smaller AIC is better

Can use to compare nested and non-nested models.

- Not always appropriate for certain types of models (problematic if you have latent variables, e.g., mixture models)

Negative Binomial in JAGS

JAGS: **dnegbin** specified in terms of parameters (p, θ)

We will specify the model in terms of μ and θ , then solve for p :

```
log(mu[i]) <- alpha + beta*IBook[i]
p[i] <- theta/(theta+mu[i])
slugs[i] ~ dnegbin(p[i], theta)
```

Poisson-normal model

$$\log(\lambda_i) = \log(\mu_i) = \beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i} + \epsilon_i$$

$$\epsilon_i \sim N(0, \sigma^2)$$

- $E[Y_i|X_i, \epsilon_i = 0] = \mu_i = \exp^{\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}} \neq E[Y_i|X_i]$
- $E[Y_i|X_i] = e^{\beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i} + \frac{1}{2}\sigma^2}$
- $\text{Var}[Y_i|X_i] = \mu_i + (e^{\sigma^2} - 1)\mu_i^2$

Offsets

Count data, Y , are often collected:

- over varying lengths of time
- in sample units that have different areas

We may be interested in modeling rates:

$$E[Y_i|X_i]/\text{Time}_i$$

Or densities:

$$E[Y_i|X_i]/\text{Area}_i$$

We may want to account for variable survey effort (varying times or areas)!

Offsets

Poisson and negative binomial models for rate data:

$$\log(E[Y_i|X_i]/\text{Time}_i) = \beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}$$

$$\Rightarrow \log(E[Y_i|X_i]) - \log(\text{Time}_i) = \beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}$$

$$\Rightarrow \log(E[Y_i|X_i]) = \log(\text{Time}_i) + \beta_0 + \beta_1 X_{1,i} + \dots + \beta_p X_{p,i}$$

$\log(\text{Time}_i)$ is called an **offset** and can be modeled using:

```
glm(y~x + offset(log(time)), data= , family = poisson())
```

An **offset** is an explanatory variable with a regression coefficient fixed at 1.

See [PoissonOffsetTemplate.R](#) and [PoissonOffset.R](#) (in the Generalized linear models folder) for an exercise fitting a Poisson model with an offset.

Martyn Plummer (creator of JAGS):

DIC [like AIC] is (an approximation to) a theoretical out-of-sample predictive error.

"The deviance information criterion (DIC) is widely used for Bayesian model comparison, despite the lack of a clear theoretical foundation...valid only when the effective number of parameters in the model is much smaller than the number of independent observations. In disease mapping, a typical application of DIC, this assumption does not hold and DIC under-penalizes more complex models. Another deviance-based loss function, derived from the same decision-theoretic framework, is applied to mixture models, which have previously been considered an unsuitable application for DIC."

Andrew Gelman: "I don't really ever know what to make of DIC. On one hand, it seems sensible... On the other hand, I don't really have any idea what I would do with DIC in any real example. In our book we included an example of DIC—people use it and we don't have any great alternatives—but I had to be pretty careful that the example made sense. Unlike the usual setting where we use a method and that gives us insight into a problem, here we used our insight into the problem to make sure that in this particular case the method gave a reasonable answer."

http://andrewgelman.com/2011/06/22/deviance_dic_ai/

Model comparisons

There are other potential options out there (e.g., WIC, cross-validation estimates of predictive error, etc)

Hooten, Mevin B, and N Thompson Hobbs. 2015. "A Guide to Bayesian Model Selection for Ecologists." Ecological Monographs 85 (1): 3–28.