Bacterial communities on classroom surfaces

2 Manuscript demo

- ₃ James F Meadow¹
- ⁴ Biology and the Built Environment Center, Institute of Ecology and Evolution, University of Oregon,
- 5 Eugene, OR USA, jfmeadow@gmail.com

7 Introduction

- ⁸ The data used here are a small subset (first 20,000 quality-filtered sequences) of those previously published
- 9 [1]. This demo illustrates a few basic multivariate analysis methods with a sample dataset. In the original
- manuscript, we investigated the sources of microbes on classroom surfaces, and whether those microbial
- communities reflect common human contact with indoor surfaces.

12 Methods

- This sequence dataset was processed using QIIME 1.8 [2] with a default MacQIIME installation (http://dx.default.com/
- 14 //www.wernerlab.org/software/macqiime). Scripts for processing raw data are in the ../QIIME/ folder. To
- pick OTUs in that folder, you will execute the pickTheseOTUs.sh script sitting in that folder. This script
- wants to run MacQIIME, so if you are not using MacQIIME, you'll need to alter the top line to reflect your
- 17 system.
- 18 For statistical analyses, we primiarily used the phyloseq package to handle QIIME output files, and vegan
- and labdsv for multivariate ecology stats [3–5]. All sequences were rarefied to an equal sampling depth (100
- 20 sequences per sample) prior to analysis. Beta-diversity was calculated using the Canberra taxonomic metric.
- 21 The Canberra metric is defined as:

$$d_{jk} = \frac{1}{NZ} \sum \frac{x_{ij} - x_{ik}}{x_{ij} + x_{ik}}$$

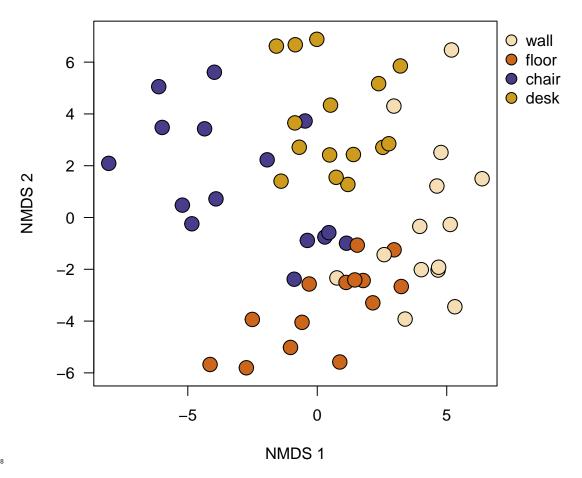
- where NZ is the number of non-zero entries. Reproducible documents were created with the knitr package
- 23 in R [6].

24 Results

- Out of a total 1.5923×10^4 sequences that passed quality filtering, we analyzed 5800 sequences in 58 samples
- distributed among 964 OTUs (97% sequence similarity). The most abundant OTU in the dataset was a
- 27 Cyanobacterium (3.14% of all sequences). The most abundant taxa are shown in Table 1.

Phylum	Family	Genus	Species	RelAbu
Cyanobacteria	Xenococcaceae	_	_	3.137931
Proteobacteria	Sphingomonadaceae	Sphingomonas	_	2.327586
Firmicutes	Staphylococcaceae	Staphylococcus	epidermidis	2.155172
Actinobacteria	Corynebacteriaceae	Corynebacterium	_	2.017241
Firmicutes	Alicyclobacillaceae	Alicyclobacillus	_	1.965517
	Cyanobacteria Proteobacteria Firmicutes Actinobacteria	Cyanobacteria Xenococcaceae Proteobacteria Sphingomonadaceae Firmicutes Staphylococcaceae Actinobacteria Corynebacteriaceae	Cyanobacteria Xenococcaceae – Proteobacteria Sphingomonadaceae Sphingomonas Firmicutes Staphylococcaceae Staphylococcus Actinobacteria Corynebacteriaceae Corynebacterium	CyanobacteriaXenococcaceaeProteobacteriaSphingomonadaceaeSphingomonas-FirmicutesStaphylococcaceaeStaphylococcusepidermidisActinobacteriaCorynebacteriaceaeCorynebacterium-

Table 1: Most abundant taxa across all surfaces.



Df ${\bf SumsOfSqs}$ ${\bf Mean Sqs}$ F.ModelR2Pr(>F)map\$SurfaceType 3 2.1962510.73208361.8481440.09311420.001Residuals 21.3903850.3961182NA0.9068858NA54 Total 57 23.586635NANA1.0000000NA

Table 2: Surface type explains a significant amount of variation among communities.

We found that surface type explained a significant amount of community variation (p = 0.001; from

- ³⁰ PERMANOVA on Canberra distances).
- Next, we tested for a quasi-distance-decay relationship. This is the sort of pattern we see in just about every
- ecosystem with most forms of life. We even found this to be a stong predictor in the dust sampled from
- the entire building [7]. So we can use the x and y coordinates as a map of samples, and then calculate the
- Euclidean pairwise distance between all samples. Then that goes through a mantel test to determine if these
- 35 distance are correlated with the community distances.
- We did not find any significant coorelation between community similarity and spatial distance (p = 0.653;
- ₃₇ from Mantel test) when considering all samples together. Likewise, individual sample types tested alone
- showed no relationship with spatial distance (p > 0.1 for all four sample types).

39 Discussion

- 40 So it looks like the type of surface, potentially as a proxy for human contact, explains a significant amount of
- variation, in the microbial communities on those surfaces, but their proximity to each other around the room
- doesn't seem to matter at all.

References

- 1. Meadow JF, Altrichter AE, Kembel SW, Moriyama M, O'Connor TK, et al. (2014) Bacterial communities on classroom surfaces vary with human contact. Microbiome 2: 7.
- ⁴⁶ 2. Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD, et al. (2010) QIIME allows analysis of high-throughput community sequencing data. Nature Methods 7: 335–336.
- 3. McMurdie PJ, Holmes S (2013) phyloseq: an R package for reproducible interactive analysis and graphics of microbiome census data. PloS one 8: e61217. Available: http://dx.plos.org/10.1371/journal.pone.0061217.
- Oksanen J, Blanchet FG, Kindt R, Legendre P, O'Hara RB, et al. (2011) vegan: Community Ecology
 Package. Available: http://cran.r-project.org/package=vegan.
- 52 5. Roberts DW (2010) labdsv: Ordination and Multivariate Analysis for Ecology. Available: http://cran.
 53 r-project.org/package=labdsv.
- 6. Xie Y (2013) knitr: A Comprehensive Tool for Reproducible Research in R. In: Stodden V, Leisch F,
 Peng RD, editors. Implementing reproducible computational research. Chapman; Hall/CRC. Available: http://www.crcpress.com/product/isbn/9781466561595.
- 7. Kembel SW, Meadow JF, O'Connor TK, Mhuireach G, Northcutt D, et al. (2014) Architectural design drives
 the biogeography of indoor bacterial communities. PLOS ONE 9: e87093. doi:10.1371/journal.pone.0087093.