

# nf-core/taxprofiler: highly parallelised and flexible pipeline for metagenomic taxonomic classification and profiling

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## 1 Abstract

Metagenomic classification tackles the problem of characterising the taxonomic source of all DNA sequencing reads in a sample. A common approach to address the differences and biases between the many different taxonomic classification tools is to run metagenomic data through multiple classification tools and databases. This, however, is a very time-consuming task when performed manually - particularly when combined with the appropriate preprocessing of sequencing reads before the classification.

Here we present nf-core/taxprofiler, a highly parallelised taxonomic classification and processing pipeline that allows for automated and simultaneous classification and/or profiling of both short- and long-read metagenomic sequencing libraries against a large number of taxonomic classifiers and profilers as well as databases within a single pipeline run. Implemented in Nextflow and as part of the nf-core initiative, the pipeline benefits from high levels of scalability and portability, accommodating from

36 small to extremely large projects on a wide range of computing infrastructure, as well  
37 as best-practise software development and community support to ensure longevity  
38 and adaptability of the pipeline, keeping up with the field of metagenomics.

## 39 2 Introduction

40 Whole-genome, metagenomic sequencing offers strong benefits to the taxonomic clas-  
41 sification of DNA samples over targeted approaches (Eloe-Fadrosh et al. 2016; Florian  
42 P. Breitwieser, Lu, and Salzberg 2019). While metabarcoding approaches targeting the  
43 16S rRNA or other marker genes are widely used due to low cost and large, diverse  
44 reference databases (Yilmaz et al. 2014; Lynch and Neufeld 2015), metagenomic ap-  
45 proaches have been gaining popularity with the increasingly lower costs of, for exam-  
46 ple, shotgun sequencing. These metagenomic analyses have been shown to provide  
47 a similar resolution on microbial genomes during taxonomic classification (Hillmann  
48 et al. 2018), with the added benefit of having greater reusability potential of the data,  
49 via whole genome reconstruction and also functional classification of metagenomics  
50 (Sharpton 2014; Quince et al. 2017).

51 Taxonomic classifiers (sometimes referred to as taxonomic bidders) aim to identify  
52 the original ‘taxonomic source’ of a given DNA sequence (Ye et al. 2019; Meyer et al.  
53 2022; Govender and Eyre 2022). In metagenomics, this typically consists of compar-  
54 ing millions of DNA sequences against hundreds or thousands of reference genomes  
55 either via alignment or ‘k-mer matching’ (Sharpton 2014; Sun et al. 2021), with the  
56 most close match being considered the most likely original ‘source’ organism of that  
57 sequence. Taxonomic profilers additionally will also try to infer species abundance  
58 of the organism in the original sample, based on the sequence abundance (Nayfach  
59 and Pollard 2016). We will use classifiers and profilers interchangeably throughout  
60 the publication.

61 Due to the scale of the problem, taxonomic profiling remains an ‘unresolved prob-  
62 lem’ in bioinformatics. Having to identify the original source of many sequences out  
63 of many reference genomes, but in an *efficient* manner, is understandably a difficult  
64 problem. Therefore a plethora of tools have been developed to address this challenge,  
65 all with their own biases and specific contexts (Sczyrba et al. 2017; Meyer et al. 2022).  
66 Additionally, each tool often produces tool-specific output formats making it difficult  
67 to efficiently cross compare results. Thus, no established ‘gold standard’ method cur-  
68 rently exists.

69 One solution to addressing the problem of choice among the range of different tools  
70 is to run all of them in parallel, and cross compare the results. This can be useful both  
71 for benchmarking studies (e.g. Sczyrba et al. 2017; Meyer et al. 2022), but also to  
72 build consensus profiles whereby confidence of a particular taxonomic identification  
73 can be increased when it is detected by multiple tools (McIntyre et al. 2017; Ye et al.  
74 2019).

75 A second challenge in taxonomic classification is a question of databases. As with  
76 tools, there is no one set ‘gold standard’ database. Different questions and contexts

77 require different databases, such as when a researcher wants to search for both bacte-  
78 rial and viral species in samples, and as an extension of this, taxonomic classifiers  
79 may need different settings for each database. Furthermore, as genomic sequenc-  
80 ing becomes cheaper and more efficient, the number of publicly available reference  
81 genomes is rapidly increasing (Nasko et al. 2018). Consequently, the size of reference  
82 databases of taxonomic classifiers is also growing, often outpacing the computational  
83 capacity available to researchers. In fact, while this was one of the main motivations  
84 behind classifiers such as Kraken2 (Wood, Lu, and Langmead 2019), these algorithmic  
85 techniques are already becoming insufficient (Wright, Comeau, and Langille 2023).

86 Finally, with the decrease of costs, the possibility for larger and larger metagenomic  
87 sequencing datasets increases, leading to increasing sample sizes in studies, as ex-  
88 emplified by the doubling of the number of metagenomes on the European Bioin-  
89 formatic Institute’s MGnify database in two years (Mitchell et al. 2019). Altogether  
90 this highlights the need for methods to efficiently profile many samples using many  
91 tools. Manually setting up bioinformatic jobs for classification tasks for each database  
92 and settings against different tools on traditional academic computing infrastructure  
93 (e.g. high performance computing clusters or ‘HPC’ clusters) can be very tedious. Ad-  
94 ditionally, particularly for very large sample sets, there is increasing use of cloud plat-  
95 forms that have greater scalability than traditional HPCs. Being able to reliably and  
96 reproducibly execute taxonomic classification tasks across infrastructure with mini-  
97 mal intervention would therefore be a boon for the metagenomics field.

98 Here we present nf-core/taxprofiler, a pipeline designed to allow users to effi-  
99 ciently and simultaneously taxonomically classify and profile short- and long-read  
100 sequencing data against multiple tools and databases in a single pipeline run.  
101 nf-core/taxprofiler utilises Nextflow (Di Tommaso et al. 2017) to ensure efficiency,  
102 portability, and scalability, and has been developed within the nf-core initiative of  
103 Nextflow pipelines (Ewels et al. 2020) to ensure high quality coding practises and  
104 user accessibility, including detailed documentation and a graphical-user-interface  
105 (GUI) execution interface.

### 106 3 Implementation

107 nf-core/taxprofiler aims to facilitate three main steps of a typical whole-genome,  
108 metagenomic sequencing analysis workflow (Chiu and Miller 2019). Taking in short-  
109 (e.g. Illumina) or long-read (e.g. Nanopore) FASTQ or FASTA files, it can (1) perform  
110 a range of appropriate read preprocessing steps, (2) perform taxonomic classification  
111 and profiling against a range of different tools depending on user preferences, and  
112 finally (3) perform post-classification aggregation and standardisation of the resulting  
113 profiles with the possibility of visualisation to the outputs (Figure 1). All relevant  
114 preprocessing statistics are displayed in an interactive and dynamic MultiQC report  
115 (Ewels et al. 2020).

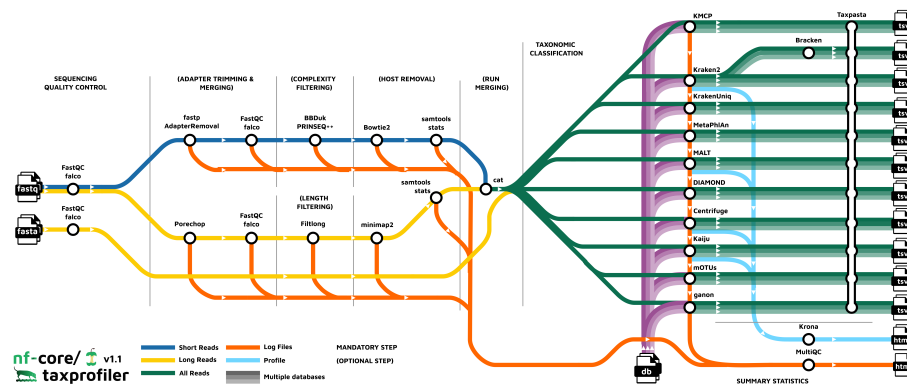


Figure 1: Visual overview of the nf-core/taxprofiler workflow. nf-core/taxprofiler can take in FASTQ (short or long reads) or FASTA files (long reads), that will optionally go through sequencing quality control (e.g. with FastQC), read preprocessing (e.g. removal of adapters), complexity filtering, host removal, and run merging before performing taxonomic classification and/or profiling with a user-selected range of tools and databases. Output from all classifiers and profilers are standardised into a common taxon table format, and when supported visualisations of the profiles are generated.

### 3.1 Input and Execution

The pipeline can be executed via typical Nextflow commands, or using the standard nf-core 'launch' GUI (<https://nf-co.re/taxprofiler/launch>), making the pipeline accessible for both computationally experienced as well as less experienced researchers. In addition to the general usage and parameter documentation of the pipeline (<https://nf-co.re/taxprofiler>). The GUI offers immediate assistance and guidance to users on what each parameter does, both in short- and long-form, with long-form parameter descriptions additionally describing which tool-specific parameters are being modified for each pipeline parameter (Figure 2). The GUI also includes controlled user input by providing strict drop-down lists and input validation prior execution of the pipeline to reduce the risk of typos and other mistakes (in contrast to the command-line interface (CLI) that only includes validation at pipeline run-time).

An example nf-core command line execution of the pipeline can be seen in Code Block 1, where two input files are supplied: one file specifying paths of FASTQ files of metagenomic samples and necessary metadata for preprocessing (such as sample ID and sequencing platform), and the second file specifying paths to the user-defined databases with per-database classification parameters. Various parameters are available to select different preprocessing steps, and provide additional configuration such as tool selection and value options. Note that even if a user supplies a given database in the database input sheet, the corresponding profiling tool must still be activated with the corresponding pipeline parameter (e.g. `--run_kraken2`). Per-classifier flags are also available for the optional saving of additional non-profile output files.

Preprocessing short-read QC options

Launch

--shortread\_qc\_minlength

15

?

Specify the minimum length of reads to be retained

Specifying a minimum read length filtering can speed up profiling by reducing the number of short unspecific reads that need to be match/aligned to the database.

Modifies tool parameter(s):

- removed from reads --length\_required
- AdapterRemoval: --min length

--perform\_shortread\_complexityfilter

☐ True
☒ False

?

Turns on nucleotide sequence complexity filtering

--shortread\_complexityfilter\_tool

bbduk

?

Specify which tool to use for complexity filtering

[ Select an option ]

bbduk

prinseqplusplus

fastp

--shortread\_complexityfilter\_entropy

?

Specify the minimum sequence entropy level for complexity filtering

--shortread\_complexityfilter\_bbduk\_windowsize

50

?

On this page

Nextflow command-line flags

> Input/output options

Preprocessing general QC options

**Preprocessing short-read QC options**

Preprocessing long-read QC options

Preprocessing host removal options

Preprocessing run merging options

Profiling options

Postprocessing and visualisation options

Show hidden params

Figure 2: Screenshot of the nf-core pipeline launch graphical user interface with nf-core/taxprofiler options displayed. The web browser-based interface provides guidance for how to configure each pipeline parameter by providing both short and long help descriptions to help guide users in which contexts to configure each parameter. Additional elements such as radio buttons, drop down menus, and background regular expressions check for validity of input. When pressing launch, a prepared configuration file and command is provided that can be copied and pasted by the user into the terminal

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**Listing 1** Example nf-core/taxprofiler command for running short-read quality control, removal of host DNA and executing the k-mer based Kraken2 and marker gene alignment MetaPhlAn3 tools.

---

```

$ nextflow run nf-core/taxprofiler \
  -r 1.1.0 \
  -profile singularity,<institute> \
  --input <samplesheet.csv> \
  --databases <database.csv> \
  --perform_shortread_qc \
  --shortread_qc_minlength 20 \
  --preprocessing_qc_tool falco \
  --run_host_removal --hostremoval_reference 'host_genome.fasta' \
  --run_kraken2 --kraken2_save_reads \
  --run_metaphlan3 \
  --run_krona \
  --run_profile_standardisation

```

---

All nf-core pipelines are strictly versioned (specified with the Nextflow `-r` flag), and to ensure reproducibility, each version of the pipeline has a fixed set of software used for each step of the pipeline. The fixed set of software are controlled through the use of the conda package manager or containers (e.g., Docker, Apptainer [previously known as Singularity]) from the stable Bioconda (Grüning et al. 2018) or BioContainers (Veiga Leprevost et al. 2017) repositories. This, coupled with the intrinsic Nextflow ability to execute on most infrastructure whether that is a local laptop (resource requirements permitting), traditional HPC, as well across common cloud providers also makes nf-core/taxprofiler a very portable pipeline that can be used in many contexts.

## 3.2 Preprocessing

Preprocessing steps in nf-core/taxprofiler are aimed at removing laboratory and sequencing artefacts that may influence taxonomic profiling, either for computing resource consumption or and/or false-positive or false-negative classification reasons. First sequencing quality control with FastQC (Andrews 2010) or Falco (Sena Brandine and Smith 2021) is carried out. Falco was included for reduced memory requirements, in particular for long read sequencing data. Artificial library adapter sequences added during sequencing reduce sequencing matching accuracy by reducing sequence specificity, and in some cases, may result in false-positive hits due to adapter sequence contamination in reference genomes (Schäffer et al. 2018; F. P. Breitwieser, Baker, and Salzberg 2018) <sup>1</sup>. Additionally, paired-end merging may provide longer sequences

---

<sup>1</sup>For an ‘infamous’ case of adapter sequences in a published eukaryotic genome, see the following blog posts

Graham Etherington: <https://web.archive.org/web/20201219022000/http://grahametherington.blogspot.com/2014/09/why-you-should-qc-your-reads-and-your.html?m=1> why-you-should-qc-your-reads-and-your.html Sixing Huang: <https://web.archive.org/web/20220904205331/https://dgg32.medium.com/carp->

158 that will allow for more specific classification when paired-end alignment is not sup-  
159 ported by a given classifier. For these tasks nf-core/taxprofiler can apply either fastp  
160 (Chen et al. 2018) or AdapterRemoval2 (Schubert, Lindgreen, and Orlando 2016) for  
161 short reads, and currently Porechop (Wick et al. 2017) for Oxford Nanopore long-read  
162 data. For both short and long reads, FastQC or Falco is run again to allow assessment  
163 on the performance of the adapter removal and/or pair-merging step.

164 Low complexity sequences, e.g. sequences containing long stretches of mono- or  
165 di-nucleotide repeats provide little specific genetic information that contribute to  
166 taxonomic identification, as they can align to many different reference genomes  
167 (Schmieder and Edwards 2011; Clarke et al. 2019). Including such reads during  
168 taxonomic profiling can increase run-time and memory usage for little gain, as  
169 during lowest-common-ancestor (LCA) classification steps they will be assigned to  
170 high-level taxonomic ranks (e.g. Kingdom). nf-core/taxprofiler performs removal of  
171 these reads through complexity filtering algorithms as provided by fastp, BBDuk  
172 (Bushnell 2022), or PRINSEQ++ (Cantu, Sadural, and Edwards 2019). Long read  
173 sequences often do not have such reads, as lengths are sufficient enough to capture  
174 greater sequence diversity - but it is sometimes desirable to only classify reads longer  
175 than a certain length - as these provide more precise taxonomic information (Dilthey  
176 et al. 2019; Portik, Brown, and Pierce-Ward 2022). Therefore, nf-core/taxprofiler can  
177 remove reads shorter than a user-defined length using Filtlong.

178 Removing host DNA is another common preprocessing step in metagenomic studies.  
179 This can help speed up run-time, particularly in microbiome studies, where detection  
180 of microbes are of interest. Furthermore, host-contamination of reference genomes in  
181 public databases is common (Longo, O'Neill, and O'Neill 2011; Kryukov and Imanishi  
182 2016; Florian P. Breitwieser et al. 2019) and therefore the removal of such sequences  
183 can also decrease the risk of false positive taxonomic assignment. To remove multiple  
184 hosts or other sequences, all reference genomes can be combined into a single FASTA  
185 reference file. Short read host removal can be carried out with Bowtie2 (Langmead  
186 and Salzberg 2012; Langmead et al. 2019) and minimap2 (Li 2018) for long reads, both  
187 in combination with SAMtools (Li et al. 2009; Danecek et al. 2021), where reads are  
188 aligned against the reference genome and the off-target (unaligned) reads are then  
189 converted back to FASTQ format for classification.

190 Finally, nf-core/taxprofiler can optionally perform run merging where libraries have  
191 been sequenced over multiple lanes to generate one profile per sample or library. The  
192 final set of reads used for profiling can be optionally saved for downstream re-use.  
193 Throughout all steps, relevant statistics and log files are generated and used both for  
194 the final pipeline run report as well as saved into the results directory of the pipeline  
195 run for further inspection where necessary.

### 3.3 Profiling

There are many types of metagenomic profiling techniques, from profiling against whole-genome references with alignment or k-mer based approaches, to methods involving alignment to species-specific marker-gene families (Quince et al. 2017; Ye et al. 2019). nf-core/taxprofiler aims to support and include all established classification or profiling tools as requested by the community. The choice of tools used in a pipeline run is up to the user, with a tool being executed when both the corresponding database and --run\_<tool> flag is provided. Specific classification settings for each tool and database are specified in the database CSV input sheet. Some tools also have pipeline level command-line flags for controlling certain aspects of output files.

As of version 1.1.0, the following classifiers and profilers are available: Kraken2 (Wood, Lu, and Langmead 2019), Bracken (Lu et al. 2017), KrakenUniq (F. P. Breitwieser, Baker, and Salzberg 2018), Centrifuge (Kim et al. 2016), MALT (Vågane et al. 2018), DIAMOND (Buchfink, Reuter, and Drost 2021), Kaiju (Menzel, Ng, and Krogh 2016), MetaPhlAn (Blanco-Míguez et al. 2023), mOTUs (Ruscheweyh et al. 2022), ganon (Piro et al. 2020), KMCP (Shen et al. 2023). ?@tbl-tool-summaries summarises the category and reference database type for each tool.

| Sequence Matching Type | Primary Algorithm | Reference Type | Method     | Tool    |
|------------------------|-------------------|----------------|------------|---------|
| Nucleotide             | k-mer based       | whole-genome   | classifier | Kraken2 |
| Amino Acid             | k-mer based       | whole-genome   | classifier | Kaiju   |
| Nucleotide             | k-mer based       | whole-genome   | profiler   | Bracken |

Nucleotide | k-mer based | whole-genome | profiler | KrakenUniq |  
Nucleotide | k-mer based | whole-genome | profiler | ganon |  
Nucleotide | k-mer based | whole-genome | profiler | KMCP |  
Nucleotide/Amino Acid | alignment based | whole-genome | classifier | MALT |  
Amino Acid | alignment based | whole-genome | classifier | DIAMOND |  
Nucleotide | alignment based | whole-genome | profiler | Centrifuge |  
Nucleotide | alignment based | marker-gene | profiler | MetaPhlAn |  
Nucleotide | alignment based | marker-gene | profiler | mOTUS |

: List of nf-core/taxprofiler supported taxonomic/classifiers profilers as of version 1.1 and their approximate method and supported input database types. Sequencing matching type refers to which ‘molecular alphabet’ is primarily used for matching between a query (read) and a reference (genome/gene). Primary algorithm refers to the algorithm type used for sequencing matching. Reference type refers to the typical sequence type used in database construction of the tool. Method refers to whether the tool performs just read classification (classifier) or additionally abundance estimation (profiler) {#tbl-tool-summaries}



229 By default, nf-core/taxprofiler produces the per-sample main taxonomic classification  
230 profile from a tool or a tool's report generation tool. The output is normally in the  
231 form of counts per reference sequencing, with additional statistics about the hits of  
232 a particular organism (estimated abundance, taxonomic level etc.). Users can also op-  
233 tionally request output of per-read classification output, and output such as classified  
234 and unclassified reads in FASTQ format, where supported.

235 The pipeline provides high efficiency, particularly during the metagenomic classifica-  
236 tion stage, through the inherent parallelisation provided by Nextflow. While metage-  
237 nomic classification is comparatively computationally intensive (in terms of mem-  
238 ory and execution time; due to a combination of sequencing depth and number of  
239 reference genomes), Nextflow automatically optimises the execution order of all the  
240 steps in pipeline, maximising the number parallel running of multiple profilers and/or  
241 databases at any given time point, as far as the available computational resources al-  
242 low. For local machines such as laptops or desktops, Nextflow will automatically  
243 detect all available computational resources but this is customisable using Nextflow  
244 configuration files. For HPC and cloud infrastructure, users typically have to define  
245 the computational infrastructural environment the pipeline is being executed on (CPU  
246 or memory limitations, queues, instance types, etc.). To facilitate the pipeline set-up,  
247 nf-core/taxprofiler supports pre-defined centralised generic and pipeline-specific in-  
248 stitutional Nextflow configurations as provided by nf-core/configs ([https://nf-co.re/](https://nf-co.re/configs)  
249 [configs](https://nf-co.re/configs); more than 90 institutions at the time of writing). However, users are still wel-  
250 come to supply their own custom configuration files, further refining computational  
251 limitations or execution specifications.

### 252 3.4 Post-profiling

253 In metagenomic studies, it is common practise to compare the profiles among many  
254 samples, and the results of multiple profiles are normally stored in 'taxon tables', i.e.  
255 counts per reference taxon (rows), for each sample (columns). When available, nf-  
256 core/taxprofiler supports the option to produce the 'native' taxon table of each classi-  
257 fication tool when multiple samples are run.

258 One of the challenges that researchers face when comparing multiple taxonomic clas-  
259 sifiers or profilers is the heterogenous output formats that are produced, that often  
260 require custom parsing and merging scripts for each tool to standardise. To facilitate  
261 more user-friendly cross-comparisons between tools, nf-core/taxprofiler utilises the  
262 TAXPASTA tool (Beber et al. 2023) to generate standardised profiles and generate  
263 multi-sample tables.

264 Summary statistics for the entire pipeline are visualised and displayed in a customis-  
265 able MultiQC report (Ewels et al. 2020). When supported, quality control of data and  
266 pipeline runs are shown for manual verification. Krona plots (Ondov, Bergman, and  
267 Phillippy 2011) can also optionally be generated for supported tools to help provide  
268 further visualisation of taxonomic profiles.

## 269 3.5 Output

270 To summarise, the main default output from nf-core/taxprofiler are both classifier  
271 ‘native’ and standardised single- and multi-sample taxonomic profiles with counts  
272 per-taxon and an interactive MultiQC run report with all run statistics, in addition to  
273 the raw log files themselves where available.

274 The MultiQC run report displays statistics and summary visualisations for all steps of  
275 the pipeline where possible, lists of versions for all tools of each step of the pipeline,  
276 and provides a dynamically-constructed text for the recommended ‘methods’ text for  
277 reporting how the pipeline was executed (including relevant citations) that users can  
278 use in their own publications.

279 Optional outputs can include other types of profiles (e.g. per read classification) and  
280 in other formats as produced by the tools themselves, as well as raw reads from pre-  
281 processing steps and output visualisations from Krona. Nextflow resource usage and  
282 trace reports are also by default produced for users to check pipeline performance.

## 283 4 Discussion

### 284 4.1 Comparison with other solutions

285 nf-core/taxprofiler has been specifically developed for the analysis of whole-genome,  
286 *metagenomic* sequencing data. While other types of taxonomic profiling data such  
287 as 16S amplicon sequencing are well established fields with a range of popular high-  
288 quality and best-practise tools pipelines (e.g. (Blanco-Míguez et al. 2023; Schloss et  
289 al. 2009)) and databases (DeSantis et al. 2006; Yilmaz et al. 2014), ‘gold standard’  
290 tools and databases for metagenomics remain much less established. Thus, the need  
291 for highly-multiplexed classification is more desirable for the newer metagenomics  
292 methods. Despite this, tools such as METAXA2 (Bengtsson-Palme et al. 2015) that  
293 use shotgun sequencing reads to recover 16S sequences from metagenomic samples.

294 A range of pipelines already exists for taxonomic profiling, however, each have  
295 their own particular purpose and capabilities. Here we compare the functionality  
296 of nf-core/taxprofiler against four other recently published or released pipelines,  
297 selected based on their similarity of purpose to nf-core/taxprofiler. We searched  
298 Google Scholar for open-source pipelines published or released in the last 5 years (at  
299 the time of writing, since 2018) that were designed primarily for metagenomic classi-  
300 fication screening, that supported at least 2 classifiers, had at least one preprocessing  
301 step and were not specifically targeted at read classification of specific domains of  
302 taxa (e.g. viruses or bacteriophages only). We also included an additional pipeline  
303 at the recommendations of the authors of the pipeline due to the functional overlap  
304 to nf-core/taxprofiler. We then evaluated the pipelines based on their publications  
305 and documentation for typical metagenomic profiling workflow steps, and a range  
306 of criteria related to expectations of modern bioinformatic workflows that can be  
307 summarised in the following four criteria: reproducibility, accessibility, scalability,  
308 and portability (Wratten, Wilm, and Göke 2021). After searching, we selected the

309 following pipelines for comparison with nf-core/taxprofiler: sunbeam [v4; Clarke et  
310 al. (2019)], Unipro UGENE [v48; Rose et al. (2019)], TAMA [github: 3a22c8f; Sim et  
311 al. (2020)], and StaG-mwc [0.7.0; Boulund et al. (2023)].

312 In terms of accessibility, all pipelines have documentation describing the installation  
313 steps, usage instructions, and output files. However, there are varying levels of de-  
314 tail and comprehensiveness. In particular, StaG-mwc and nf-core/taxprofiler have  
315 the most detailed descriptions of all possible output files for every supported mod-  
316 ule, whereas Unipro UGENE and sunbeam have very minimal to possibly unfinished  
317 output documentation. For execution options, most of the pipelines provide CLI ex-  
318 ecution, except for Unipro UGENE which offers only GUI-based pipeline set-up (de-  
319 spite a command-line execution of the GUI generated configuration). In particular, nf-  
320 core/taxprofiler is the only pipeline providing both CLI and GUI interfaces for pipeline  
321 run execution.

322 Criteria covering portability also overlap with accessibility, as it implies options for  
323 and ease of different users running on different types of computing infrastructure,  
324 whether that is on their own laptop, on an HPC cluster, or in the cloud. Unipro  
325 UGENE is the only pipeline that supports execution on all three major operating sys-  
326 tems (Linux, OSX, Windows), whereas StaG-mwc and nf-core/taxprofiler can be run  
327 on unix operating systems, and sunbeam and TAMA are only being supported on  
328 Linux. While all pipelines support 'local' machine execution (e.g. personal laptops or  
329 desktops), a large portion of academic users execute computationally intensive bioin-  
330 formatic tasks on HPC clusters. In these contexts, pipeline task submissions are nor-  
331 mally managed by job schedulers, thus integration with schedulers is an important  
332 criterion for running large multi-step and parallelised pipelines. The three pipelines  
333 leveraging workflow managers (Snakemake (Mölder et al. 2021) and Nextflow) sup-  
334 port integration with schedulers (StaG-mwc, sunbeam, and nf-core/taxprofiler) with  
335 nf-core/taxprofiler supporting the most by far ([>10 scheduling systems](#)) as natively  
336 offered by Nextflow. This allows the greatest possible choice for users in terms of  
337 which HPC infrastructure they can execute their pipeline on. As an extension of this,  
338 only nf-core/taxprofiler has explicit support for cloud computing (e.g. AWS, GCP, or  
339 Microsoft Azure), again maximising user choice and portability when it comes to run-  
340 ning the pipeline.

341 In terms of scalability, the aforementioned integration with schedulers and cloud com-  
342 puting support implicitly maximises efficiency and parallelisation of pipeline runs,  
343 providing good scalability for varying numbers of input files and steps in the pipeline.  
344 Again, the three workflow manager based pipelines provide scalability, whereas there  
345 is no mention neither Unipro UGENE nor TAMA in reference to parallel task execu-  
346 tion. Furthermore, all pipelines except TAMA, allowed per-process customisation of  
347 computational resources, something critical for maximising efficient scalability to en-  
348 sure only the necessary resources for a given step of a pipeline are requested.

349 In terms of reproducibility, all five pipelines are good at ensuring reproducibility in  
350 terms of pipeline and software versioning (allowing re-execution of pipeline runs us-  
351 ing the same software), with only tama not having stable versioned releases. However,  
352 installing software manually across different infrastructures can result in variability

353 in the execution of each software <sup>2</sup> (Di Tommaso et al. 2017). The current most popular  
354 solution to the problem of inconsistent software environments is to use container  
355 engines such as Docker or Apptainer to run container images which are isolated, deter-  
356 ministic computing environments which can be executed by any system providing a  
357 container runtime. Only Unipro UGENE does not document the use of a container sys-  
358 tem, with nf-core/taxprofiler offering the biggest choice for users courtesy of Nextflow  
359 (6 different engine systems at the time of writing).

360 Finally, we compared metagenomics related functionality between the pipelines. All  
361 pipelines support short-read FASTQ input, but only nf-core/taxprofiler explicitly re-  
362 ports long-read support, while the documentation in Unipro UGENE states that assem-  
363 bled contigs are possible input to some of the profilers. All pipelines support read pre-  
364 processing (adapter clipping, and merging). In terms of tools used for preprocessing,  
365 Trimmomatic (Bolger, Lohse, and Usadel 2014) is popular across the other pipelines  
366 but is not supported in nf-core/taxprofiler. Only sunbeam and nf-core/taxprofiler sup-  
367 port complexity filtering to remove low sequence diversity reads. In fact within sun-  
368 beam, the authors developed their own dedicated, performant complexity filtering  
369 tool Komplexity (Clarke et al. 2019). Most pipelines support some form of host re-  
370 moval (only TAMA did not support this), and it is likely possible with Unipro UGENE  
371 through user customisation of the workflow. In all cases, host removal consists of  
372 mapping processed reads with an aligner and using the off-target reads for down-  
373 stream profiling (as implemented in nf-core/taxprofiler), however StaG-mwc has an  
374 additional separate metagenomic host removal step with Kraken2. nf-core/taxprofiler  
375 supports by far the largest number of taxonomic classifiers and profilers at 11 as of  
376 v1.1.0 - providing the greatest choice to users - with StaG-mwc offering 7, and the  
377 remaining pipelines only 3. Only nf-core/taxprofiler and partly StaG-mwc explicitly  
378 support running each profiler with multiple databases. nf-core/taxprofiler is the only  
379 pipeline that supports running an arbitrary number of different metagenomic profiler  
380 databases each with their own settings - making it useful for tool parameter compar-  
381 ison, testing different databases, or reducing the size of each database (e.g. per domain)  
382 to make it more flexibility for running on smaller computational infrastructure. StaG-  
383 mwc allows multiple references for their short-read alignment steps rather than the  
384 metagenomic profilers. For output, nf-core/taxprofiler, StaG-mwc, and sunbeam (via  
385 an extension) support a singular run report for summarising all preprocessing step.  
386 Only nf-core/taxprofiler and TAMA produce standardised output for all taxonomic  
387 profilers (via TAXPASTA). However Unipro UGENE additionally offers a 'consensus'  
388 profile using WEVOTE (Metwally et al. 2016).

389 To summarise, many of the pipelines reviewed here offer similar functionality, with  
390 particularly StaG-mwc having a strong overlap with nf-core/taxprofiler. Thus, users  
391 in most cases will be able to select the pipeline depending on which framework they  
392 feel most comfortable with. However the advantages of nf-core/taxprofiler mainly  
393 come from the offering of the greatest choice of tools, the benefits provided by  
394 Nextflow whereby it provides the greatest number of computational infrastructure

<sup>2</sup>As demonstrated in this blogpost from Paweł Przytuła: <https://web.archive.org/web/20230320223436/https://appsilon.com/reproducible-research-when-your-results-cant-be-reproduced/> (Accessed 2023-08-25)

types the pipeline can be executed on, and container systems can be used to ensure reproducibility, and the support of the nf-core community due to the centralised pool of ‘plug-and-play’ modules to make it easier to update the pipeline over time to add new tool.

The functionality offered by other pipelines not currently supported by nf-core/taxprofiler include sequencing saturation estimation (StaG-mwc), taxonomy-free composition comparison (StaG-mwc), functional profiling (StaG-mwc), *de novo* assembly (sunbeam), and reference mapping (StaG-mwc, sunbeam). We do not plan to support *de novo* assembly or functional profiling in nf-core/taxprofiler as we feel this better served by other existing dedicated pipelines (e.g. Uritskiy, DiRuggiero, and Taylor 2018; Krakau et al. 2022).

We note there exists a range of other pipelines that also include some form of taxonomic classification. However often these pipelines have been developed with a different main purpose (e.g. Assembly and binning for nf-core/mag (Krakau et al. 2022), MetaWRAP (Uritskiy, DiRuggiero, and Taylor 2018), SqueezeMeta (Tamames and Puente-Sánchez 2018), or MEDUSA (Morais et al. 2022); Metagenomic read alignment with CCMetaGen (Marcelino et al. 2020) and Wochenende (Rosenboom et al. 2022)).

## 4.2 Development roadmap

An important advantage of nf-core/taxprofiler is that it is being developed within the nf-core community (<https://nf-co.re>), that provides strong long-term support for the continued community-based development and maintenance of its pipelines. In this framework, we will continue to add additional preprocessing, metagenomic classification, and profiling tools as they become established and as requested by the metagenomics community, for example, we feel that the inclusion of steps such as sequencing saturation estimation as already being performed by StaG-mwc would be beneficial to the nf-core/taxprofiler workflow (possibly with dedicated tools such as Nonpareil (Rodriguez-R et al. 2018)), and/or more performant complexity filtering tools such as Komplexity as offered by sunbeam. This also applies to extend support to other sequencing platforms; nf-core/taxprofiler already supports Nanopore long-read data, however the use of long-read PacBio data for metagenomic data is growing in interest (Portik, Brown, and Pierce-Ward 2022). We are therefore considering adding dedicated preprocessing steps for this type of sequencing data.

A remaining major challenge for metagenomics researchers (and not supported in the same workflow by any of the compared pipelines above) is the construction of databases for each profiling tool. Given there still are no curated, high-quality ‘gold standard’ databases in metagenomics, and while nf-core/taxprofiler allows the profiling against multiple databases and settings in parallel, currently the pipeline still requires users to construct these manually and to supply to the pipeline. While we feel this is currently a reasonable investment as such databases can be repeatedly reused, we are exploring the possibility to add an additional complementary workflow in the pipeline to allow automated database construction of all classification tools,

437 given a set of FASTA reference files.

438 Finally, once an overall taxonomic profile is generated, researchers often wish to val-  
439 idate hits through more sensitive and accurate methods such as with read-mapping  
440 alignment. While read alignment is supported by other pipelines such as StaG-mwc,  
441 this happens in-parallel to the taxonomic profiling and requires prior expectation of  
442 which reference genomes to map against. Instead, nf-core/taxprofiler could be easily  
443 extended to have a validation step similar to that of the ancient DNA metagenomic  
444 pipeline aMeta (Pochon et al. 2022) where, utilising Nextflow’s execution parallelism,  
445 the input sequences could be aligned back to the reference genomes of only those  
446 species with hits from the taxonomic classification with dedicated accurate short- or  
447 long-read aligners. In addition to the more precise classification, post-classification  
448 read-alignment could also be particularly useful for researchers in palaeogenomics  
449 who wish to use tools other than KrakenUniq for initial classification (as in aMeta),  
450 where alignment information can be used to authenticate ancient DNA within their  
451 samples but also in clinical metagenomics to identify potential pathogens at much  
452 finer resolution (e.g. down to strain level).

453 Another motivation for developing nf-core/taxprofiler, despite the large number  
454 of existing metagenomics pipelines is that by establishing a taxonomic profiling  
455 pipeline within the nf-core ecosystem, it is possible to begin building both standalone  
456 but also an integrated suite of powerful interconnected pipelines for the major  
457 stages of metagenomic workflows. Existing microbial- and metagenomics- related  
458 pipelines within the nf-core initiative include nf-core/ampliseq, nf-core/mag, and  
459 nf-core/funcscan. We expect over time the ability to link inputs and outputs of each  
460 workflow to develop comprehensive metagenomic analyses, while still maintaining  
461 powerful standalone pipelines, providing maximal user choice.

## 462 **5 Conclusion**

463 nf-core/taxprofiler is an accessible, efficient, and scalable pipeline for metagenomic  
464 taxonomic classification and profiling that can be executed on anywhere from laptops  
465 to the cloud. Offering, to our knowledge, the largest number of taxonomic profilers  
466 across similar pipelines, it provides flexibility for users not just on choice of profiling  
467 tool but also with databases and database settings, with any number being able to be  
468 supplied to the pipeline in a single run. We hope that through detailed documentation  
469 and a range of execution options, nf-core/taxprofiler will make reproducible and high-  
470 throughput metagenomics more accessible for a wide range of disciplines.

## 471 **6 Data Availability**

472 All data used in this publication

## 473 7 Code Availability

474 nf-core/taxprofiler source code is available on GitHub at <https://github.com/nf-core/taxprofiler>, and each release is archived on Zenodo (latest version DOI: [10.5281/zenodo.7728364](https://doi.org/10.5281/zenodo.7728364))

477 The version of the pipeline described in this paper is version (1.1.0) (release specific  
478 Zenodo archive DOI: [10.5281/zenodo.8358147](https://doi.org/10.5281/zenodo.8358147))

## 479 8 Supplementary Data

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