

1000genomes Selection Exercise

To get some exposure to different approaches for detecting recent positive selection from population genetic data, we will be using the 1000 Genomes data. We will present two approaches for detecting recent positive selection. The first is an approach based on differentiation between populations, that uses the PBS (population-branch statistic) to detect selection that occurred in the history of a particular population. The second is an approach based on haplotype patterns - using the iHS statistic. We'll suppose a particular gene is of interest (whose transcript start/stop is marked with vertical lines below)

We will interact with the 1000 Genomes data using **vcf** (variant call format) files. These files have one line per variant, and contain information about each individual's genotype as columns. These files are so large they are most often kept in a **gzip** compressed format, which you will recognize usually by seeing a filename that ends with **.vcf.gz**.

Before anything, we need to set up a unix-based analysis environment with the appropriate programs:

- **tabix**: For quickly retrieving specific lines from a **.vcf.gz** file. It creates an index of a **.vcf.gz** file and then uses the index to quickly find the lines a user requests.
- **bcftools**: For manipulating **vcf** files. Here we use it for some basic filtering.
- **plink2** / **plink-1.9**: Second-generation of the multi-faceted **plink** tool. It facilitates a number of common analyses in statistical genetics and population genetics. Here we use it to compute F_{ST} which we will need to compute the PBS statistic. We assume you will run it using the command **plink2**.
- **selscan**: For computing haplotype-based statistics that are sensitive to signatures of selective sweeps (EHH, iHS, XP-EHH, nSL, for example)
- **norm**: A program from the **selscan** package that will normalize iHS or XP-EHH values.

We have included an appendix, titled “Setting up your environment” that contains the appropriate instructions for installing these. For the moment, we assume you have gone through this, or are using an account on a server that has been pre-configured for you, and so we will jump right into some analysis. To be sure, check and see if the list of programs above can be found in a ‘bin/’ subdirectory of the main project directory here.

As a disclaimer, in what follows - we take an approach that builds intuition and take some shortcuts to make analyses feasible in a classroom setting. For publishable results, more rigor to assess the statistical significance of results is necessary than what we show here.

Preparing files that describe the 1000 Genomes individuals

To get started, we will want some basic information on the individuals in the 1000 Genomes data. We also want to simplify our analysis by just focusing on individuals from the CHB (Han Chinese from Beijing), YRI (Yoruba from Ibadan, Nigerai), and FIN (Finnish from Finland). Let's get started...

Downloading Individual-Level Data [SSS 2015 read-only, data already in data/]

We can download info on the 1000 Genomes phase III individuals with the following commands. The **integrated_call_samples_v3.20130502.ALL.panel** file we are obtaining contains the sample id and associated population and gender info for each individual.

```
cd ~/1000genomes_Selection_Exercise/data
# download 1000genomes_phase3 individual information
wget ftp://ftp.1000genomes.ebi.ac.uk/vol1/ftp/release/20130502/integrated_call_samples_v3.20130502.ALL.
```

[Click here](#) for a for explanation of the population codes.

Setting up .clst.txt and .iids files for plink and bcftools analyses

In order to extract individuals and define groups for computing selection statistics within we need to prepare a set of `.clst.txt` and `.iids` files. You will inspect each file after they are created to see the basic structure of these files (e.g. using the `head` command to see the start of the file and `tail` to see the end of the file).

The `.iids` files just contain a list of the individual ids we will include in our analysis.

The `.clst.txt` files define a “cluster” of individuals to consider as a single unit for analysis (e.g. a cluster might represent a population). Here we create a such file for each pair of populations we will be analyzing.

Here we use `awk` commands - `awk` is very useful unix-based tool for basic file manipulation.

```
cd ~/data
# use awk to produce a file that has the format of a .clst.txt file, with IID | IID | CLST for CHB, YRI
awk 'NR>1{print $1, $1, $2}' integrated_call_samples_v3.20130502.ALL.panel > 1kgv3_clst.txt

# Visually inspect the top and botom of the files.
head 1kgv3_clst.txt
tail 1kgv3_clst.txt
```

Downloading data for a specific 3Mb genomic region [SSS 2015 read-only, data already in data/]

Now, let's download a 3Mb part of the the 1000 Genomes Phase3 project data to use.

```
cd ~/data

# download a 3MB region (chr2:108013601-111013601) from the 1000genomes project and store as .vcf.gz fi
tabix -h ftp://ftp.1000genomes.ebi.ac.uk/vol1/ftp/release/20130502/ALL.chr2.phase3_shapeit2_mvncall_int
```

We will use our list of individual ids from the YRI, FIN, and CHB to extract just those individuals from the larger 1000 genomes data. We also want to filter out indels and multi-allelic sites.

```
# filter out indels (-V indels), multi-allelic sites (-m 2 -M 2). (Approx 30 sec - 1 min)
cd ~/data
../bin/bcftools view -v snps -m 2 -M 2 -O z 1000genomes_phase3_chr2_108013601_111013601.vcf.gz > snv_1000
```

Try running `ls -thrl` to verify that the output file `snv_1000genomes_chr2_108013601_111013601.vcf.gz` is smaller than the input.

Now we want to filter out sites that violate Hardy-Weinberg equilibrium. We first calculate genotype genotypes within each cluster.

```
tag=chr2_108013601_111013601
cd ~/data
../bin/plink-1.9 --vcf snv_1000genomes_$tag.vcf.gz \
    --within 1kgv3_clst.txt \
    --hardy midp\
    --keep-cluster-names FIN \
    --out tmp_fin
../bin/plink-1.9 --vcf snv_1000genomes_$tag.vcf.gz \
    --within 1kgv3_clst.txt \
    --hardy midp\
    --keep-cluster-names CHB \
    --out tmp_chb
../bin/plink-1.9 --vcf snv_1000genomes_$tag.vcf.gz \
    --within 1kgv3_clst.txt \
```

```

--hardy midp\
--keep-cluster-names YRI \
--out tmp_yri

# Pull out all of the IDs for SNPs that fail HWE (with a loose criterion) in at least one population
awk '$9<1e-5{print $2}' tmp_fin.hwe tmp_chb.hwe tmp_yri.hwe | sort | uniq > tmp.exclude.txt

# Filter out HW failures
../bin/bcftools view snv_1000genomes_${tag}.vcf.gz --exclude 'ID=@tmp.exclude.txt' -O z > snv_1000genomes.

```

We filtered out SNPs based on a criteria of $p < 10^{-5}$, where the p-values were calculated using an exact test of Hardy-Weinberg equilibrium. Note that a more rigorous approach is to conduct a χ^2 test for the joint set of genotype counts for each sub-population (in this case a 5 degree of freedom test).

Population-differentiation-based selection scan

One approach to detecting recent positive selection is to see if a local genomic region shows extensive differentiation between *two* populations (using a population differentiation statistic such as F_{ST} or a more formal inferential methods like BayesScan not shown here). Let's do this for the CHB vs FIN comparison for our focal region.

Computing pairwise F_{ST} s

```

# Fst between CHB and FIN
tag=chr2_108013601_111013601
cd ~/data
../bin/plink-1.9 --vcf snv_1000genomes_${tag}_HWEfiltered.vcf.gz \
--fst \
--within 1kgv3_clst.txt \
--out CHB_FIN_${tag}_HWEfiltered \
--keep-cluster-names FIN CHB

```

Use head to take a look at the structure of this file.

Visualizing pairwise F_{ST} values

Make sure your working directory in R is set to the data subdirectory. (Use the “Session” tab or `setwd()` command to change your working directory, e.g. `setwd("~/git/1000genomes_Selection_Exercise/data")`).

```

if(!require(dplyr)) install.packages("dplyr")
if(!require(ggplot2)) install.packages("ggplot2")
if(!require(reshape2)) {install.packages("stringi"); install.packages("reshape2")}
setwd("~/data")

# read data
fst_CHB_FIN_df <- read.table('CHB_FIN_chr2_108013601_111013601_HWEfiltered.fst', header = TRUE) %>%
  rename(FST_CF = FST) %>%
  filter(is.nan(FST_CF) == FALSE) %>%
  select(SNP, POS, FST_CF)

# Prepare the plot
fst_CHB_FIN_plot <- ggplot(fst_CHB_FIN_df, aes(x = POS, y = FST_CF)) + geom_point() + xlab('Position on

```

```
theme_bw() + ylab("") + geom_vline(xintercept = 109510927) + geom_vline(xintercept = 109605828)

# View the plot
fst_CHB_FIN_plot + ggsave(' ../results/fst_CHB_FIN_plot.png',width=7,height=4)
```

Questions

1. Suppose the 99.9th percentile of F_{ST} between CHB and FIN genome-wide is 0.5. Do you see any variants in this 3Mb region that are among the most differentiated in the genome?
2. Can you assess from this data whether selection took place just in CHB, FIN, or both?

PBS

A drawback of such an approach is we do not know which of the two populations (or if both) have experienced selection causing the allele frequencies to differentiate. Using *three* populations, we can compute a statistic that isolates a single populations. Specifically we compute an estimate of how much differentiation occurred on the branch leading to a particular population in a three-population tree consisting of a focal population, a sister population, and an outgroup population. These PBS statistics are based on F_{ST} , so we start by computing pairwise F_{ST} values.

```
# Fst between CHB and YRI
tag=chr2_108013601_111013601
cd ~/data
../bin/plink-1.9 --vcf snv_1000genomes_${tag}_HWEfiltered.vcf.gz \
  --fst \
  --within 1kgv3_clst.txt \
  --out CHB_YRI_chr2_108013601_111013601_HWEfiltered \
  --keep-cluster-names CHB YRI

# Fst between FIN and YRI
../bin/plink-1.9 --vcf snv_1000genomes_${tag}_HWEfiltered.vcf.gz \
  --fst \
  --within 1kgv3_clst.txt \
  --out FIN_YRI_chr2_108013601_111013601_HWEfiltered \
  --keep-cluster-names FIN YRI
```

Let's now compute the PBS statistic for the FIN and the CHB to see if we can determine which population the selection occurred in.

$$T = -\log(1 - F_{st})$$

$$PBS_{CHB} = \frac{T_{CY} + T_{CF} - T_{FY}}{2}$$

$$PBS_{FIN} = \frac{T_{FY} + T_{CF} - T_{CY}}{2}$$

```
setwd("~/data")
fst_CHB_YRI_df <- read.table('CHB_YRI_chr2_108013601_111013601_HWEfiltered.fst', header = TRUE) %>%
  rename(FST_CY = FST) %>%
  filter(is.nan(FST_CY) == FALSE) %>%
  select(SNP, POS, FST_CY)
```

```
fst_FIN_YRI_df <- read.table('FIN_YRI_chr2_108013601_111013601_HWEfiltered.fst', header = TRUE) %>%
  rename(FST_FY = FST) %>%
  filter(is.nan(FST_FY) == FALSE) %>%
  select(SNP, POS, FST_FY)

# join fst data frames and compute pbs
pbs_df <- fst_CHB_FIN_df %>%
  inner_join(fst_CHB_YRI_df, by = c("SNP", "POS")) %>%
  inner_join(fst_FIN_YRI_df, by = c("SNP", "POS")) %>%
  mutate(T_CY = -log(1 - FST_CY), T_CF = -log(1 - FST_CF), T_FY = -log(1 - FST_FY)) %>%
  mutate(PBS_CHB = (T_CY + T_CF - T_FY) / 2) %>%
  mutate(PBS_FIN = (T_CF + T_FY - T_CY) / 2) %>%
  select(POS, PBS_CHB, PBS_FIN)

# melt data for faceting in ggplot
melted_pbs_df <- melt(pbs_df, id = c("POS"))

# plot!
PBS_plot <- ggplot(melted_pbs_df, aes(x = POS, y = value)) + geom_point() + xlab('Position on chr2') +
  theme_bw() + ylab("") + facet_grid(variable ~ .) + geom_vline(xintercept = 109510927) + geom_vline(xintercept = 109510927)
PBS_plot + ggsave('../results/PBS_plot.png', width=7, height=4)
```

Questions

1. What is the major difference between the PBS values in this region for CHB vs Finland?
2. Draw the 3 population tree of (CHB, FIN, YRI) and mark which branch is extended the most in this region.
3. In which population would one conclude that there has been the most accelerated differentiation?
4. (Advanced) What are some possible interpretations for when the PBS statistic becomes negative?

iHS

Now's let turn towards looking at haplotype-based signatures of selection. We'll focus on the CHB for this analysis, so we'll begin by preparing a new data file with just the CHB individuals.

We will filter the file to only include variants with minor allele frequency >5%. In principle the rare variants increase our power, but for efficiency's sake we'll exclude them here, as most of the information for these statistics comes from common variants.

```
tag=chr2_108013601_111013601
cd ~/data
# write a file of just IIDs from CHB
awk '($3 == "CHB") {print $1}' 1kgv3_clst.txt > CHB_iids.txt

head CHB_iids.txt

# keep CHB individuals from the filtered vcf and filter out rare variants (at maf < 5%)
# this will be used for haplotype based selection analysis.
../bin/bcftools view -S CHB_iids.txt \
  -q '.05 minor' \
  -O z \
  snv_1000genomes_${tag}_HWEfiltered.vcf.gz > CHB_${tag}_HWEfiltered.vcf.gz
```

Haplotype-based methods use recombination rates between markers to calibrate the extent of haplotype homozygosity observed. To that end, we need to set up a list of all the markers in the format of a plink .bim file, and interpolate a genetic map at each of those positions. Here are the steps:

```
tag=chr2_108013601_111013601
cd ~/data
# write bim file from vcf (-H suppress header)
../bin/bcftools view CHB_${tag}_HWEfiltered.vcf.gz \
    -H | awk '{print $1, $3, "0", $2, $4, $5}' > CHB_${tag}_HWEfiltered.bim

head CHB_${tag}_HWEfiltered.bim

# get genetic positions and interpolate genetic positions for variants not present in the genetic_map
../bin/plink-1.9 --bim CHB_${tag}_HWEfiltered.bim \
    --cm-map genetic_map_chr2_combined_b37.txt 2 \
    --make-just-bim \
    --out ${tag}_HWEfiltered_gm

# format as map file
awk '{print $1, $2, $3, $4}' ${tag}_HWEfiltered_gm.bim > ${tag}_HWEfiltered_gm.map

# Inspect the map file - the columns are <CHR> <POS> <Position in centiMorgans> <Position in physical space>
head ${tag}_HWEfiltered_gm.map
```

Now we can finally play! This selscan command will compute iHS values for us, and the norm command normalizes them to have mean 0 and standard deviation 1 (which is useful for assessing outlier values).

This command will take about 5 minutes.

```
tag=chr2_108013601_111013601
cd ~/data
# compute iHS! This will take about 5 minutes
../bin/selscan --ihs \
    --vcf CHB_${tag}_HWEfiltered.vcf.gz \
    --map ${tag}_HWEfiltered_gm.map \
    --out CHB_${tag}_HWEfiltered
```

Normally we would normalize the iHS – but because we are only using a 3Mb region the normalization makes little sense. Here's what the command would like if we actually ran it.

```
# norm --ihs --bins 20 --files CHB_${tag}_HWEfiltered.ihs.out
```

Make plot of iHS scores

Let's make a plot of the iHS values in this region.

```
setwd("~/data")

# read iHS data
ihs_df <- read.table("CHB_chr2_108013601_111013601_HWEfiltered.ihs.out", header = FALSE)
colnames(ihs_df) <- c("SNP", "POS", "FREQ", "ihh1", "ihh0", "unstd_iHS")

IHS_plot <- ggplot(ihs_df, aes(x = POS, y = unstd_iHS)) + geom_point() + xlab('Position on chr2') + theme_minimal()

IHS_plot + ggsave("../results/IHS_plot.png")
```

Questions

1. Are there any provocative outliers among the iHS scores?
2. Suppose you are unconvinced that there are interesting patterns of iHS in this region - is it possible that the PBS signal in this region is real? Put another way - does positive selection in the past necessarily leave a signature in the iHS score of a population observed today?

Make a plot of allele frequencies

It also possible to make a plot representing allele frequencies in the region:

```
setwd("~/data")
freq_plot<-ggplot(ihs_df, aes(x = POS, y = FREQ)) + geom_point() + xlab('Position on chr2') + theme_bw()

freq_plot + ggsave("../results/freq_plot.png")
```

Or a histogram of the frequencies (an allele frequency spectrum):

```
setwd("~/data")

hist(ihs_df$FREQ,xlab="Non-reference frequency", ylab="# of SNPs",main="Allele frequency spectra",breaks=30)

png("allele_frequency_spectra.png", width=5, height=3, units="in", res=300)
hist(ihs_df$FREQ,xlab="Non-reference frequency", ylab="# of SNPs",main="Allele frequency spectra",breaks=30)
dev.off()
```

Questions

1. Non-reference alleles are typically derived (i.e. novel alleles relative to the ancestral allele) - so for simplicity sake let's assume they are. Does the allele frequency distribution seen here seem consistent with selection? Why or why not?

Additional Regions

The 1000 Genomes selection browser (<http://hsb.upf.edu/>) allows one to quickly browse various selections statistics that have been computed for major populations from the project.

Take a look at a few random regions of the genome of your choice. Compare what you see for the iHS and Fst tracks for the region surrounding the lactase gene LCT (near the gene there are variants affecting the persistence of lactase expression), SLC24A5 (a gene affecting skin pigmentation), and for KITLG (another gene affecting skin pigmentation). These are all genes that have been argued to show signatures of recent positive selection in humans, presumably as adaptations to novel environments (specifically high latitudes with low sunlight in the case of the skin pigmentation, and to dairy culture in the case of lactase)

Being cautious about selective scans

In a classic paper from 1979 titled “The Spandrels of San Marco and the Panglossian Paradigm: A critique of the Adaptationist Programme”, Stephen J Gould and Richard Lewontin, wrote eloquently about how easy it is to fall into the intellectual trap of taking an adaptationist worldview and explaining all features of organismal biology as due to the direct outcome of natural selection. Some features may be indirectly selected or due to chance events and genetic drift. While population genetic approaches provide more rigor, the tests are still ultimately statistical, and there is often a non-negligible chance of confusing patterns due simply due to genetic drift with those of selection. With that in mind, we should be critical (as with GWAS) about constructing just-so stories about a plausible gene without doing additional functional follow-up or gathering

other lines of evidence to support a claim about recent positive selection acting on a locus. Some useful perspectives on selective sweeps are Teshima et al (2006, Genome Research) and Fu and Akey (2013, Annual Review of Genomics and Human Genetics)

References

- **tabix** : (<http://www.htslib.org/doc/tabix.html>)
- **bcftools** : (<https://samtools.github.io/bcftools/bcftools.html>)
- **plink1.9** : Chang CC, Chow CC, Tellier LCAM, Vattikuti S, Purcell SM, Lee JJ (2015) Second-generation PLINK: rising to the challenge of larger and richer datasets. GigaScience, 4. (<https://www.cog-genomics.org/plink2>)
- **selscan** : ZA Szpiech and RD Hernandez (2014) selscan: an efficient multi-threaded program to calculate EHH-based scans for positive selection. Molecular Biology and Evolution, 31: 2824-2827. (<https://github.com/szpiech/selscan>)
- **iHS**: Voigt et al (2006) A Map of Recent Positive Selection in the Human Genome. PloS Genetics 4(3): e72.
- **PBS**: Yi et al (2010) Sequencing of 50 Human Exomes Reveals Adaptation to High Altitude. Science 329:75-78.

Appendix: Setting up your Environment

Downloading and Building Methods

This section can be skipped for now as the software tools are already built on the cluster. This can be used if you'd like to use these methods later. For now, scroll down to *Downloading Data*.

Download using conda

Conda is an excellent way to install programs for an analysis in a way that you can easily share your environment. This way someone else can replicate your analysis (just like we did here!). To install everything necessary to run this analysis run the following commands in the terminal.

```
git clone https://github.com/NovembreLab/1000genomes_Selection_Exercise.git
cd 1000genomes_Selection_Exercise
conda env create -f selection_exercise_environment.yml
source activate selex
```

Download from source

```
mkdir src
cd src/

# download tabix
git clone https://github.com/samtools/tabix

# download htslib & bcftools
wget http://sourceforge.net/projects/samtools/files/samtools/1.2/htslib-1.2.1.tar.bz2
wget http://sourceforge.net/projects/samtools/files/samtools/1.2/bcftools-1.2.tar.bz2

# clone selscan
git clone -b devel https://github.com/szpiech/selscan
```



```
# download plink (double check that the correct OS is specified)
# For linux:
wget https://www.cog-genomics.org/static/bin/plink150602/plink_linux_x86_64_dev.zip
# For mac:
wget https://www.cog-genomics.org/static/bin/plink150615/plink_mac.zip
```

Build

```
# make a bin directory under main directory if not there already
mkdir ../bin/

# tabix
cd tabix/
make
mv tabix ../../bin/

# bcftools
cd ../
tar xvjf htlib-1.2.1.tar.bz2
tar xvjf bcftools-1.2.tar.bz2
cd bcftools-1.2/
make
mv bcftools ../../bin/

# plink
cd ../
mkdir plink
unzip plink_linux_x86_64_dev.zip -d plink
mv plink/plink ../bin

# selscan
cd selscan/

# open Makefile make sure the correct OS is specified
cd src/
make
mv norm ../../bin/
mv selscan ../../bin/

# lets go home
cd ../../..
ls bin/

mv bin/plink bin/plink-1.9
# you should see binaries for tabix, bcftools, plink, selscan, and norm

# Add the current directory (bin) to the export path using this command
export PATH=$(pwd):$PATH
```