# Supplemental Material: Base Diagnostics

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### Chapter 1

### Introduction

This is the supplemental material for experiments breaking down nondominated sorting into its two main components: phenotypic fitness sharing and nondominated front ranking. We evaluated these components, along with standard nondominated sorting, on the contradictory objectives diagnostic to measure their contribution on the overall effectiveness of nondominated sorting.

#### 1.1 About our supplemental material

This supplemental material is hosted on GitHub using GitHub pages. The source code and configuration files used to generate this supplemental material can be found in this GitHub repository. We compiled our data analyses and supplemental documentation into this nifty web-accessible book using bookdown.

Our supplemental material includes the following paper figures and statistics:

• Nondomintaed sorting breakdown (Section ??)

### 1.2 Contributing authors

- Jose Guadalupe Hernandez
- Alexander Lalejini
- Charles Ofria

### 1.3 Computer Setup

These analyses were conducted in the following computing environment:

#### print(version)

```
##
                   x86_64-pc-linux-gnu
## platform
## arch
                   x86_64
## os
                   linux-gnu
                   x86_64, linux-gnu
## system
## status
## major
## minor
                   3.1
                   2023
## year
## month
                   06
                   16
## day
```

```
## svn rev 84548
## language R
## version.string R version 4.3.1 (2023-06-16)
## nickname Beagle Scouts
```

#### 1.4 Experimental setup

```
Setting up required variables variables.
# libraries we are using
library(ggplot2)
library(cowplot)
library(dplyr)
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##
                   filter, lag
## The following objects are masked from 'package:base':
##
##
                   intersect, setdiff, setequal, union
library(PupillometryR)
## Loading required package: rlang
# data diractory for qh-pages
DATA_DIR = '/opt/ECJ-2023-Suite-Of-Diagnostic-Metrics-For-Characterizing-Selection-Schemes/DATA/CONTRAD
# data diractory for local testing
\# DATA_DIR = '-\('Desktop\)/Repositories\('ECJ-2023-Suite-Of-Diagnostic-Metrics-For-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Characterizing-Selection-Character
# graph variables
SHAPE = c(5,3,1)
cb_palette <- c('#88CCEE','#EE7733','#EE3377')</pre>
TSIZE = 26
p_theme <- theme(</pre>
     text = element_text(size = 18),
     plot.title = element_text( face = "bold", size = 20, hjust=0.5),
     panel.border = element_blank(),
     panel.grid.minor = element_blank(),
     legend.title=element_text(size=12),
     legend.text=element_text(size=12),
     axis.title = element_text(size=18),
     axis.text = element_text(size=18),
     legend.position="bottom",
     panel.background = element_rect(fill = "#f1f2f5",
                                                                                           colour = "white",
                                                                                           linewidth = 0.5, linetype = "solid")
)
# default variables
```

```
DIMENSIONALITY = 100

GENERATIONS = 50000

# selection scheme related stuff

ACRO = c('nds','nfr','pfs')

NAMES = c('Nondominated sorting (nds)','Nondominated front ranking (nfr)','Phenotypic fitness sharing (nfr)','Phenotypic
```

### Chapter 2

# Nondominated sorting breakdown

For these experiments we break down nondominated sorting into its two main components: phenotypic fitness sharing and nondominated front ranking. We evaluated these components, along with standard nondominated sorting, on the contradictory objectives diagnostic to measure their contribution on the overall effectiveness of nondominated sorting. Here we present the results for **activation gene coverage** and **satisfactory trait coverage** found by each selection scheme on the contradictory objectives diagnostic. 50 replicates are conducted for each scheme explored.

#### 2.1 Data setup

```
over_time_df <- read.csv(paste(DATA_DIR,'over-time.csv', sep = "", collapse = NULL), header = TRUE, str
over_time_df$scheme <- factor(over_time_df$scheme, levels = NAMES)
over_time_df$acro <- factor(over_time_df$acro, levels = ACRO)</pre>
```

### 2.2 Activation gene coverage over time

Activation gene coverage in a population over time. Data points on the graph is the average activation gene coverage across 50 replicates every 2000 generations. Shading comes from the best and worse coverage across 50 replicates.

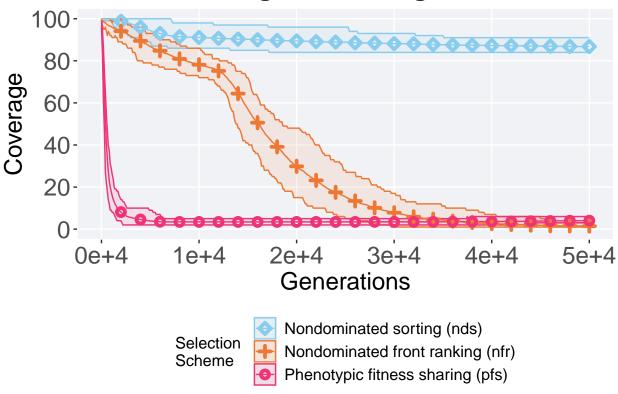
```
lines = over_time_df %>%
group_by(scheme, gen) %>%
dplyr::summarise(
   min = min(uni_str_pos),
   mean = mean(uni_str_pos),
   max = max(uni_str_pos)
)
```

```
## `summarise()` has grouped output by 'scheme'. You can override using the
## `.groups` argument.
```

```
over_time_plot = ggplot(lines, aes(x=gen, y=mean, group = scheme, fill = scheme, color = scheme, shape
  geom_ribbon(aes(ymin = min, ymax = max), alpha = 0.1) +
  geom_line(size = 0.5) +
  geom_point(data = filter(lines, gen %% 2000 == 0 & gen != 0), size = 1.5, stroke = 2.0, alpha = 1.0)
  scale_y_continuous(
   name="Coverage",
   limits=c(0, 100),
```

```
breaks=seq(0,100, 20),
   labels=c("0", "20", "40", "60", "80", "100")
  ) +
  scale_x_continuous(
   name="Generations",
   limits=c(0, 50000),
   breaks=c(0, 10000, 20000, 30000, 40000, 50000),
   labels=c("0e+4", "1e+4", "2e+4", "3e+4", "4e+4", "5e+4")
  ) +
  scale_shape_manual(values=SHAPE)+
  scale_colour_manual(values = cb_palette) +
  scale_fill_manual(values = cb_palette) +
  ggtitle('Activation gene coverage over time')+
 p_theme +
  guides (
    shape=guide_legend(ncol=1, title.position = "left", title = 'Selection \nScheme'),
    color=guide_legend(ncol=1, title.position = "left", title = 'Selection \nScheme'),
    fill=guide_legend(ncol=1, title.position = "left", title = 'Selection \nScheme')
over_time_plot
```

## Activation gene coverage over time



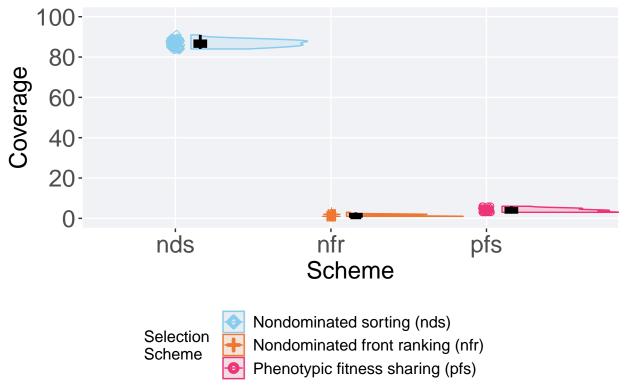
### 2.3 Final activation gene coverage

Activation gene coverage found in the final population at 50,000 generations.

```
plot = filter(over_time_df, gen == 50000) %>%
  ggplot(., aes(x = acro, y = uni_str_pos, color = acro, fill = acro, shape = acro)) +
  geom_flat_violin(position = position_nudge(x = .1, y = 0), scale = 'width', alpha = 0.2, width = 1.5)
  geom_boxplot(color = 'black', width = .07, outlier.shape = NA, alpha = 0.0, size = 1.0, position = po
  geom_point(position = position_jitter(width = 0.03, height = 0.02), size = 2.0, alpha = 1.0) +
  scale_y_continuous(
   name="Coverage",
   limits=c(0, 100),
    breaks=seq(0,100, 20),
   labels=c("0", "20", "40", "60", "80", "100")
  ) +
  scale_x_discrete(
    name="Scheme"
  scale_shape_manual(values=SHAPE)+
  scale_colour_manual(values = cb_palette, ) +
  scale_fill_manual(values = cb_palette) +
  ggtitle('Final activation gene coverage')+
  p_theme
plot_grid(
  plot +
    theme(legend.position="none"),
  legend,
 nrow=2,
  rel_heights = c(3,1)
```

```
## Warning: Using the `size` aesthetic with geom_polygon was deprecated in ggplot2 3.4.0.
## i Please use the `linewidth` aesthetic instead.
## This warning is displayed once every 8 hours.
## Call `lifecycle::last_lifecycle_warnings()` to see where this warning was
## generated.
```





#### 2.3.1 Stats

Summary statistics for the coverage found in the final population.

```
act_coverage = filter(over_time_df, gen == 50000)
act_coverage$acro = factor(act_coverage$acro, levels = c('nds','pfs','nfr'))
act_coverage %>%
group_by(acro) %>%
dplyr::summarise(
    count = n(),
    na_cnt = sum(is.na(uni_str_pos)),
    min = min(uni_str_pos, na.rm = TRUE),
    median = median(uni_str_pos, na.rm = TRUE),
    mean = mean(uni_str_pos, na.rm = TRUE),
    max = max(uni_str_pos, na.rm = TRUE),
    IQR = IQR(uni_str_pos, na.rm = TRUE)
)
```

```
## # A tibble: 3 x 8
                        min median mean
    acro count na_cnt
    <fct> <int> <int> <dbl> <dbl> <int> <dbl>
## 1 nds
             50
                    0
                         84
                                87 86.8
                                           91 2.75
             50
                    0
## 2 pfs
                          3
                                 4 4.04
                                            6 2
## 3 nfr
             50
                    0
                                 1 1.44
                          1
                                            3 1
```

Kruskal–Wallis test illustrates evidence of statistical differences.

```
kruskal.test(uni_str_pos ~ acro, data = act_coverage)
##
   Kruskal-Wallis rank sum test
##
## data: uni_str_pos by acro
## Kruskal-Wallis chi-squared = 133.91, df = 2, p-value < 2.2e-16
Results for post-hoc Wilcoxon rank-sum test with a Bonferroni correction.
pairwise.wilcox.test(x = act_coverage$uni_str_pos, g = act_coverage$acro, p.adjust.method = "bonferroni
                     paired = FALSE, conf.int = FALSE, alternative = '1')
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
##
## data: act_coverage$uni_str_pos and act_coverage$acro
##
##
       nds
              pfs
## pfs <2e-16 -
## nfr <2e-16 <2e-16
##
## P value adjustment method: bonferroni
```

#### 2.4 Satisfactory trait coverage over time

Satisfactory trait coverage in a population over time. Data points on the graph is the average activation gene coverage across 50 replicates every 2000 generations. Shading comes from the best and worse coverage across 50 replicates.

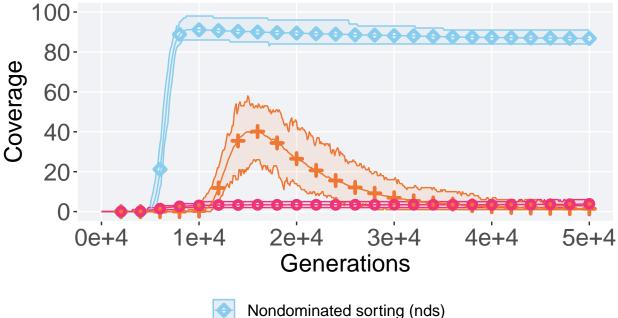
```
lines = over_time_df %>%
group_by(scheme, gen) %>%
dplyr::summarise(
   min = min(pop_uni_obj),
   mean = mean(pop_uni_obj),
   max = max(pop_uni_obj)
)
```

## `summarise()` has grouped output by 'scheme'. You can override using the
## `.groups` argument.

```
over_time_plot = ggplot(lines, aes(x=gen, y=mean, group = scheme, fill = scheme, color = scheme, shape
  geom_ribbon(aes(ymin = min, ymax = max), alpha = 0.1) +
  geom_line(size = 0.5) +
  geom_point(data = filter(lines, gen %% 2000 == 0 & gen != 0), size = 1.5, stroke = 2.0, alpha = 1.0)
  scale_y_continuous(
   name="Coverage",
   limits=c(0, 100),
   breaks=seq(0,100, 20),
   labels=c("0", "20", "40", "60", "80", "100")
) +
  scale_x_continuous(
   name="Generations",
   limits=c(0, 50000),
   breaks=c(0, 10000, 20000, 30000, 40000, 50000),
   labels=c("0e+4", "1e+4", "2e+4", "3e+4", "4e+4", "5e+4")
```

```
) +
scale_shape_manual(values=SHAPE)+
scale_colour_manual(values = cb_palette) +
scale_fill_manual(values = cb_palette) +
ggtitle('Satisfactory trait coverage over time')+
p_theme + theme(legend.title=element_blank(),legend.text=element_text(size=12)) +
guides(
    shape=guide_legend(ncol=1, title.position = "bottom"),
    color=guide_legend(ncol=1, title.position = "bottom"),
    fill=guide_legend(ncol=1, title.position = "bottom")
)
over_time_plot
```

## Satisfactory trait coverage over time



Nondominated sorting (nds)

Nondominated front ranking (nfr)

Phenotypic fitness sharing (pfs)

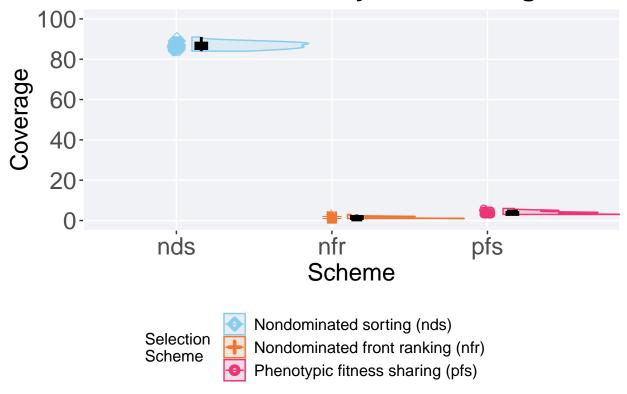
### 2.5 Final satisfactory trait coverage

Satisfactory trait coverage found in the final population at 50,000 generations.

```
plot = filter(over_time_df, gen == 50000) %>%
    ggplot(., aes(x = acro, y = pop_uni_obj, color = acro, fill = acro, shape = acro)) +
    geom_flat_violin(position = position_nudge(x = .1, y = 0), scale = 'width', alpha = 0.2, width = 1.5)
    geom_boxplot(color = 'black', width = .07, outlier.shape = NA, alpha = 0.0, size = 1.0, position = po
    geom_point(position = position_jitter(width = 0.03, height = 0.02), size = 2.0, alpha = 1.0) +
    scale_y_continuous(
        name="Coverage",
```

```
limits=c(0, 100),
    breaks=seq(0,100, 20),
    labels=c("0", "20", "40", "60", "80", "100")
  scale_x_discrete(
    name="Scheme"
  scale_shape_manual(values=SHAPE)+
  scale_colour_manual(values = cb_palette, ) +
  scale_fill_manual(values = cb_palette) +
  ggtitle('Final satisfactory trait coverage')+
  p_theme
plot_grid(
  plot +
    theme(legend.position="none"),
  legend,
  nrow=2,
  rel_heights = c(3,1)
```

## Final satisfactory trait coverage



#### 2.5.1 Stats

Summary statistics for the coverage found in the final population.

```
sat_coverage = filter(over_time_df, gen == 50000)
sat_coverage$acro = factor(sat_coverage$acro, levels = c('nds','pfs','nfr'))
sat_coverage %>%
  group_by(acro) %>%
 dplyr::summarise(
   count = n(),
   na_cnt = sum(is.na(pop_uni_obj)),
   min = min(pop_uni_obj, na.rm = TRUE),
   median = median(pop_uni_obj, na.rm = TRUE),
   mean = mean(pop_uni_obj, na.rm = TRUE),
   max = max(pop_uni_obj, na.rm = TRUE),
   IQR = IQR(pop_uni_obj, na.rm = TRUE)
## # A tibble: 3 x 8
                                                    IQR
    acro count na_cnt
                          min median mean
                                              max
     <fct> <int> <int> <dbl> <dbl> <int> <dbl>
                                               91 2.75
## 1 nds
              50
                      0
                           84
                                  87 86.8
## 2 pfs
              50
                      0
                            3
                                   4 3.86
                                               6 1
                      0
                                                3 1
## 3 nfr
              50
                            1
                                   1 1.4
Kruskal-Wallis test illustrates evidence of statistical differences.
kruskal.test(pop_uni_obj ~ acro, data = sat_coverage)
##
##
  Kruskal-Wallis rank sum test
##
## data: pop_uni_obj by acro
## Kruskal-Wallis chi-squared = 134.12, df = 2, p-value < 2.2e-16
Results for post-hoc Wilcoxon rank-sum test with a Bonferroni correction.
pairwise.wilcox.test(x = sat_coverage$pop_uni_obj, g = sat_coverage$acro, p.adjust.method = "bonferroni
                     paired = FALSE, conf.int = FALSE, alternative = '1')
##
  Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: sat_coverage$pop_uni_obj and sat_coverage$acro
##
       nds
              pfs
## pfs <2e-16 -
## nfr <2e-16 <2e-16
##
## P value adjustment method: bonferroni
```