

Modelling Thermal Responses of Metabolic Traits

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Abstract

Temperature is the one of the most important factors in determining metabolic rate and the optimisation of metabolic rate will tend to increase the fitness of any organism. Climate change is already having an enormous impact on all life on Earth and ectotherms can be particularly sensitive to environmental shifts in temperature. Many species are resorting to latitudinal range shifts, as well as behavioural and evolutionary adaptation, but extinction is often unavoidable. Improving our ability to model thermal responses is vital if we are to predict, understand, and hopefully prevent, future extinctions. This study compared the relative ability of three models, the cubic polynomial, Briere, and Schoolfield, each being fitted to 1367 datasets within the BioTraits meta-dataset. It was concluded that the Briere model was the best based on these data, affirmed by consistently better AIC_c scores.

I. Introduction

1 **I. Introduction**

2 Temperature is a fundamental parameter

3 in almost all biological processes and its

4 importance in metabolic biology is well doc-

5 umented (Brown et al. 2004, Montoya et al.

6 2012, Dell et al. 2011, DeLong et al. 2017).

7 Metabolic processes are often catalysed by

8 enzymes which depend on kinetic, and ulti-

9 mately heat, energy to function. As temper-

10 ature decreases, molecules have less and less

kinetic energy, decreasing the acquisition 11

rate of substrate by enzyme units, and thus 12

metabolic rates decrease accordingly. When 13

temperature increases, metabolic rates in- 14

crease steadily until the thermal optimum 15

is reached (DeLong et al. 2017, Dell et al. 16

2011); that is, the temperature at which 17

optimal metabolic rate occurs (T_{pk} or T_{opt}). 18

Beyond this, increasing temperature will 19

start to hinder the metabolic rate until the 20

21 lethal limit is reached. The mechanisms be-
 22 hind this sharp decline in metabolic rate
 23 are disputed but could potentially be due
 24 to protein degradation (Johnson & Lewin
 25 1946, Dell et al. 2011). These metabolic re-
 26 sponses to temperature exhibit a remarkably
 27 similar pattern graphically, often referred
 28 to as Thermal Performance Curves (TPCs),
 29 across a plethora of metabolic processes and
 30 taxa. This makes the study of TPCs a use-
 31 ful tool of comparison for all life on Earth as
 32 all living things depend on metabolism for
 33 their energy. Furthermore, an increased un-
 34 derstanding of how species respond to tem-
 35 perature is imperative in a rapidly warming
 36 world. If we can find some plasticity in
 37 a species' temperature tolerance then per-
 38 haps it will have a better chance of avoiding
 39 the mass extinction that is sweeping our
 40 planet, although some recent findings sug-
 41 gest the scope for adaptation may be limited
 42 (Tüzün & Stoks 2018). Of course, there are

other ways a species may adapt, for exam- 43
 ple through latitudinal range shifting (Haase 44
 et al. 2019) or evolution. However, the latter 45
 seems unlikely in the time-frame available, 46
 although some studies have suggested it is 47
 possible (DeLong et al. 2018), and the for- 48
 mer is only possible for mobile species with 49
 suitable habitats to move to. The database 50
 used in this study, BioTraits, was provided 51
 by my supervisor, Dr. Samraat Pawar, and 52
 is an extension of the database used by Dell 53
 et al. (2011) and Kontopoulos et al. (2018). 54
 It is one of the most extensive metabolic 55
 datasets ever amalgamated and provided 56
 an excellent way to compare three different 57
 models for TPCs. 58

i. Models 59

Three models were used in this study to 60
 compare their ability to fit to each dataset 61
 within BioTraits. Firstly, there is the phe- 62

63 nomenological cubic polynomial:

$$B = B_0 + B_1T + B_2T^2 + B_3T^3 \quad (1)$$

64 Where B is the responding trait value, T is
65 the temperature, B_0 is the y -intercept of the
66 curve, and B_1 , B_2 , and B_3 are coefficients
67 acting on T .

68 Secondly, the Briere model (Briere et al.
69 1999) was used as an alternative phenomono-
70 logical model:

$$B = B_0T(T - T_0)\sqrt{T_m - T} \quad (2)$$

71 Where T_0 and T_m are the minimum and
72 maximum thermal tolerances respectively
73 for the trait - B - and B_0 is a normalisation
74 constant. Whilst this model is technically
75 phenomenological, it can still provide useful
76 biological insight once fitted as it gives an
77 estimate of the thermal tolerance of a par-
78 ticular trait for a particular organism; as
79 it has x -intercepts (T), and its parameters
80 meet the definition of ‘ecological parameters’
81 coined by (Lamb et al. 1984). However, it

falls short of the definition of a mechanis- 82
tic model as the model provides no insight 83
into the underlying biological mechanisms 84
at work. 85

Finally, the third model used in this 86
study was a simplified version of the Sharpe- 87
Schoolfield (Schoolfield et al. 1981) model 88
to provide a mechanistic comparison as it is 89
derived from thermodynamic and enzyme 90
kinetic theory. The full model is given by: 91

$$B = \frac{B_0 e^{\frac{-E}{k}(\frac{1}{T} - \frac{1}{283.15})}}{1 + e^{\frac{E_l}{k}(\frac{1}{T_l} - \frac{1}{T})} + e^{\frac{E_h}{k}(\frac{1}{T_h} - \frac{1}{T})}} \quad (3)$$

Where k is the Boltzmann constant ($8.617 \times$ 92
 10^{-5} eV K^{-1} , Boltzmann (1872)), T is the 93
temperature in Kelvin, B_0 is the trait value 94
at a reference temperature - 283.15 K in 95
this study - E_l is the low-temperature de- 96
activation energy (eV) of the enzyme and 97
controls the behaviour of the curve at very 98
low temperatures and T_l is the tempera- 99
ture at which 50% of enzyme units have 100
been low-temperature deactivated. E_h is 101
the high-temperature deactivation energy of 102

the enzyme and controls the behaviour of the curve beyond T_{pk} , and T_h is the temperature at which 50% of enzyme units have been high-temperature deactivated. E is the activation energy which controls the behaviour of the curve in the enzyme's 'normal operating range', that is, before T_{pk} but not at low temperatures. This model attempts to capture the non-linear rate acceleration (E_l) at low temperatures, the linear rate increase, E , up to T_{pk} , and rapid non-linear deceleration, E_h , at high temperatures as the temperature approaches the lethal limit of an enzyme. This makes the Briere model an interesting comparison as this too attempts to capture the two non-linear and linear aspects of a typical TPC. There are also two simplified versions of the Schoolfield model, each with either the E_h or E_l element removed from the denominator. The removal of E_l and associated terms is sometimes intuitive as low-temperature deactivation of

enzymes is weak and there is often insufficient data at low temperatures, as is the case with BioTraits. For these reasons, this simplified version was used for this study and is given by:

$$B = \frac{B_0 e^{\frac{-E}{k}(\frac{1}{T} - \frac{1}{283.15})}}{1 + e^{\frac{E_h}{k}(\frac{1}{T_h} - \frac{1}{T})}} \quad (4)$$

Simplifying the model also allows for more datasets to be explained as the minimum number of datapoints required for the six-parameter model would be larger than for the four-parameter simplified version. Other studies, such as Alber & Schaffner (1993), have demonstrated that the simplified Schoolfield models often fit better. The reference temperature for this study was set at 283.15 K as it has proved a reliable reference in other studies of this nature (Dell et al. 2011) and seems more appropriate for the many cold-adapted species represented in BioTraits, despite Schoolfield et al. (1981) using a reference of 298.15 K. Assuming there has been no low- or high-temperature

deactivation B_0 can be used as an approximation of the trait value at this reference temperature. This study will be based on comparisons of these models, in the vein of Johnson & Omland (2004), as opposed to traditional null-hypothesis testing, as they demonstrated the power of model comparison with large sets of observational data.

predominantly used in this study to 'wrangle' BioTraits, that is, to prepare a filtered version for model fitting. It was then used post-fitting to visualise the models with optimised parameters, in particular making use of the package ggplot2 (3.1.0) (Wickham 2016) to create aesthetic and intuitive plots.

II. Methods

Python

i. Computing Languages

R

R (3.2.3) is a popular programming language within the biological sciences and is particularly powerful for statistical analysis, data manipulation, and data visualisation (R Core Team 2015). RStudio (1.1.419) was used as an interactive development environment (IDE) for R in this study as it permits the display of graphics and code within the same pane which is useful for visualisation (RStudio Team 2016). R was

Python (3.5.2) is an immensely popular, object-oriented programming language first developed by van Rossum (1991). It can lend its popularity, in part, to its extensive collection of packages, as well as its intuitive syntax, making it one of the most versatile programming languages. In particular, the package lmfit (Newville 2014) has robust functions for optimising model parameters using non-linear least-squares techniques which were used to fit the models in this study. Furthermore, Python is relatively fast, computationally speaking, at

189 looping, which gives it further advantage
 190 over R for fitting models to large datasets
 191 like BioTraits. Visual Studio Code (Mi-
 192 crosoft 2019) was used as an IDE for Python
 193 and IPython (P’erez & Granger 2007) was
 194 used as a command shell for testing code as
 195 it has particularly good debugging features.

196 **Bash**

197 Bash (4.3.48) is an open-source, UNIX shell
 198 and command language originally created by
 199 Fox (1989). It is the default shell language
 200 used for most Linux distributions such as
 201 Ubuntu 16.04 which was the operating sys-
 202 tem used in this study. A Bash script was
 203 used to run each of the scripts used in this
 204 project as it is a versatile language that can
 205 easily call and run other languages.

206 **ii. Data**

207 BioTraits consists of 2165 unique thermal
 208 responses of metabolic processes from 1010
 209 publications. Predominantly, respiration,

growth, and photosynthetic rate are the 210
 metabolic process being measured against 211
 temperature. BioTraits includes species 212
 from many Phyla with diverse life histo- 213
 ries, but a majority of representatives are 214
 terrestrial species, Arthropods being par- 215
 ticularly well-represented. As this dataset 216
 contains 155 columns and 25826 rows, it 217
 was first refined to a handful of columns 218
 relevant to this study to improve compu- 219
 tational speed, namely the trait value and 220
 temperature. Rows with missing values for 221
 these columns were removed and any dataset 222
 with less than six unique datapoints was also 223
 removed, as this is the minimum required 224
 to estimate four parameter models like the 225
 cubic polynomial and simplified Schoolfield. 226

227 **iii. Parameter estimation**

Using R, starting parameters for every 228
 dataset in BioTraits were calculated. For 229
 the cubic polynomial model, starting values 230
 of 1 were used for all four parameters. For 231

Briere, estimates for T_0 and T_m were made for E was taken as the gradient of this line,
 using the minimum and maximum observed with the Eh estimate being twice this value.
 temperatures respectively. For Schoolfield, a If regression failed, default estimates of 0.65
 reference temperature of 283.15 K was used for E and 1.3 for Eh were used as recom-
 and the parameter estimations were car- mended defaults from the literature, and E
 ried out following the method of Schoolfield was given bounds of 0 to 3, while Eh was
 et al. (1981). B_0 was estimated as the bound between 0 and 6 to maintain theo-
 recorded trait value nearest to this tem- retical accuracy (Montoya et al. 2012, Dell
 perature, as defined by the equation. The et al. 2011, Allen et al. 2006). Th was esti-
 peak metabolic rate B_{max} was then calcu- mated by calculating the nearest recorded
 lated (T_{pk} being the corresponding tempera- temperature to $\frac{B_{max}}{2}$ as Th is the temper-
 ture at this trait value) and the dataset was ature at which half the enzyme units have
 split either side of this value. If B_{max} oc- been made inactive, so this provides a good
 curred at the highest recorded temperature estimate, with a lower bound of T_{pk} applied
 (i.e. the rate had not started descending (Kontopoulos et al. 2018). For datasets with
 yet) the dataset was not split and the sub- no datapoints after B_{max} , Th was given a
 sequent regression was carried out on the starting value equal to T_{max} .
 whole dataset. The trait values were plot-
 ted on an Arrhenius plot ($\log(trait)$ against
 $\frac{1}{kT}$) with k being the Boltzmann constant.
 Linear regression was carried out on the
 left-hand (below T_{pk}) data and the estimate

iv. Model comparison

The Akaike Information Criterion (AIC)
 (Akaike 1974) was used to compare model
 fits within each dataset and is given by the
 formula, assuming the model is univariate,

is linear in its parameters, and has normally-distributed residuals:

$$AIC = 2k - 2\ln(\hat{L}) \quad (5)$$

Where \hat{L} is the maximum likelihood estimation of the model and k is the number of parameters in the model. AIC rewards the relative goodness of fit between models applied to the same data but penalises number of parameters used as this can sometimes lead to overfitting. This method was chosen over similar methods such as the Bayesian Information Criterion (BIC , Schwarz (1978)) as it can be modified for small datasets and has higher success rates in picking the best model if the 'true' model is not present in a sample of models (Vrieze 2012). However, despite the existing penalty, AIC can still be prone to favouring models with more parameters if the sample size is small (benchmark of at least 40 datapoints per model parameter suggested by Johnson & Omland (2004)) and BioTraits

includes several datasets that would be susceptible to this. This can be circumvented by using AIC_c (Hurvich & Tsai 1989), an extension of AIC with the addition of a further parameter penalty given by:

$$AIC_c = AIC + \frac{2k^2 + 2k}{n - k - 1} \quad (6)$$

Where n is the sample size and k is the number of parameters as before. It should be noted that as $n \rightarrow \infty$, $\frac{2k^2 + 2k}{n - k - 1} \rightarrow 0$ and thus $AIC_c \rightarrow AIC$, making it suitable for large samples too. As some of the datasets used in this analysis had only the minimum permissible number of datapoints to fit a four-parameter model, AIC_c was used to compare models instead of AIC .

In addition to AIC_c , adjusted R^2 , or \bar{R}^2 , was used as an alternative comparison tool. Generally attributed to Wright (1921), R^2 is purely a measure of goodness of fit and is given by:

$$R^2 = 1 - \frac{SS_{res}}{SS_{tot}} \quad (7)$$

Where SS_{res} is the sum of the squared residuals between the model and observed data and SS_{tot} is the total sum of squares. A score of 1 is a 'perfect' fit and a negative score is considered a worse fit than a straight, horizontal line through the observed mean. R^2 is very susceptible to overfitting as the addition of a new parameter will always improve the score and this can lead to poor parameter estimation which is particularly bad for model comparison (Johnson & Om-land 2004). Fortunately, this can be accounted for to an extent with adjusted R^2 , \bar{R}^2 :

$$\bar{R}^2 = 1 - (1 - R^2) \frac{n - 1}{n - k - 1} \quad (8)$$

Where n is the number of datapoints and k is the number of parameters. Unlike R^2 , it only improves its score if an additional parameter improves the model more than would be expected by chance, making it less susceptible to overfitting and a better comparative tool for this study.

III. Results

Of the 2165 datasets in BioTraits, 1367 remained after the vetting criteria was implemented. Starting parameters were then calculated for each dataset in R with varying success. These parameters were then optimised in Python with no failed convergences (Table 1). For Briere, the interquar-

Table 1: *Median parameter estimates after optimisation*

	Cubic	Briere	Schoolfield
B_0	2340	0.0001434	1.271
B_1	-23.62	-	-
B_2	0.08193	-	-
B_3	-9.163e-05	-	-
T_0	-	277	-
T_m	-	318.5	-
E	-	-	0.6356
E_h	-	-	1.558
T_h	-	-	303.2

tile ranges of B_0 , T_0 and T_m were fairly narrow, as were the converged parameters for Schoolfield, although these were subject to

far stricter bounds. The datasets and associated optimised parameters were returned to R for visualisation and comparative statistical analysis. An example plot is shown in Figure 1 and summarised AIC_c and \bar{R}^2 , values are shown in Table 2. Mean \bar{R}^2 indicated that the cubic model had the best average fit, with Schoolfield second and Briere last. Interestingly, Table 2 shows that despite this, if the best \bar{R}^2 counts are compared, that is, the number of times each model had the highest R^2 value rather than the average, cubic is still the best, but Briere replaces Schoolfield as second best. With regards to AIC_c , Briere is considered the best model in the highest number of datasets, with cubic second best and Schoolfield worst.

IV. Discussion

For Schoolfield, the regression carried out on the Arrhenius-transformed data was unreliable at providing 'reasonable' starting

Table 2: *Summary of comparative statistics for the three models. The first and second rows are counts and the third row is the median adjusted R^2*

	Cubic	Briere	Schoolfield
Best AIC_c	372	549	203
Best adjusted R^2	708	336	306
Average adjusted R^2	0.8507	0.6655	0.7343

estimates and defaults were often used as a result, as this provided better fits in those cases. 'Reasonable' estimates were defined as within or near the range established by Dell et al. (2011); for E this was 0.2 – 1.2. This may have been because a lot of the datasets used were quite small, and, after being split about B_{max} , could lead to unrepresentative regression on the Arrhenius plots, leading to poor estimates for E and E_h . However, the apparent weakness in starting parameter estimation seemed to be mitigated by the use of literature defaults as the average optimised parameters were similar to other studies (Dell et al. 2011, Kon-

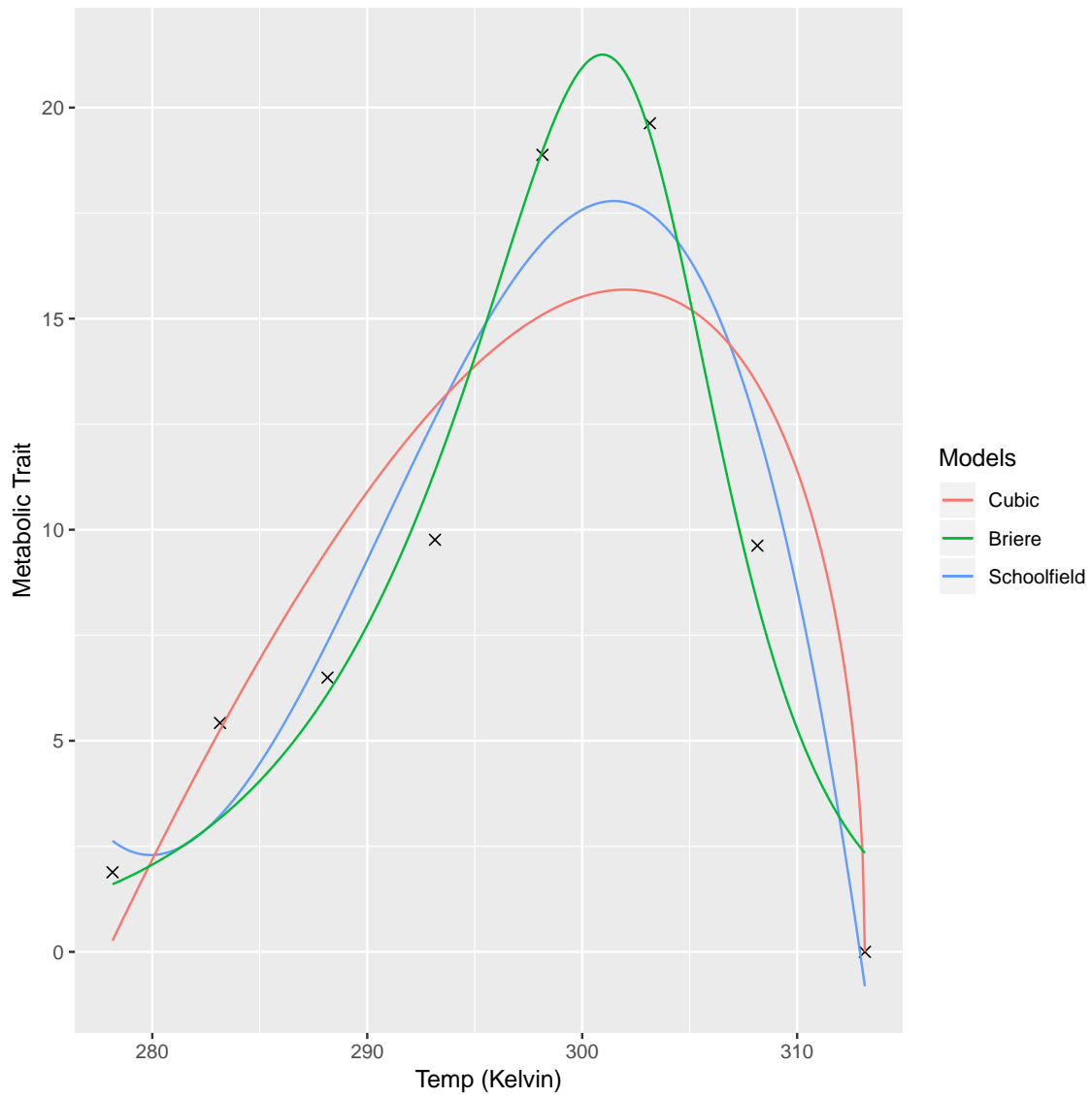


Figure 1: An example plot of the three models fitted to data used by (Li et al. 2009)

topoulos et al. 2018). Average optimised pa-
 rameters for T_0 and T_m in the Briere model
 were slightly lower and higher respectively
 than the estimates of (Briere et al. 1999)
 ($T_0 = 283.15K$, $T_m = 310.15K$). This may
 be due to the larger and more taxonomically
 diverse nature of BioTraits as Briere’s study
 was concerned exclusively with Arthropods.

Comparing the models produced some
 conflicting but altogether more interesting
 results. Cubic was the best fit according to
 the average \bar{R}^2 but this is not particularly
 surprising when one considers the freedom
 with which its parameters can be varied
 without theoretical and biological bounds.
 Of the other two models, Schoolfield had the
 higher average \bar{R}^2 but Briere had a higher
 \bar{R}^2 in more datasets when count was consid-
 ered, implying Briere had a greater range of
 fit consistency. As the Briere model is based
 predominantly on the thermal responses of
 Arthropod metabolism, perhaps for these

species it fit well and for other Phyla it
 was less competent. Conversely, comparison
 through AIC_c showed Briere to be the best
 model in the highest number of datasets,
 with cubic second and Schoolfield third. The
 discrepancy between the two comparative
 measures is intriguing and may be due to
 the additional parameter penalty that AIC_c
 incorporates which would favour the Briere
 model over the other two as it requires one
 fewer parameter. Moving forward, further
 research in this field would benefit by com-
 paring more models, such as those proposed
 by DeLong et al. (2017), and more data will
 undoubtedly improve reliability. Whilst Bio-
 Traits is one of the most extensive datasets
 of its kind, it still shows clear bias towards
 certain Phyla, life histories, and climates,
 which will hinder any extrapolations that
 can be drawn from models it fits well to.

In conclusion, the cubic model did fit
 well on average and was well-supported by

428 \bar{R}^2 , but the application of a purely phe- 450
 429 nomenological linear model to the proven 451
 430 non-linear response of metabolism to tem- 452
 431 peratures would limit any useful biological 453
 432 interpretation and prediction going forward. 454

433 The Schoolfield model has the advantage of
 434 parameters based on biological mechanisms
 435 but this study showed it was significantly
 436 less able to explain the data, particularly
 437 when AIC_c was considered. Briere, how-
 438 ever, proved to be the best model, at least
 439 for BioTraits. It also has a distinct advan-
 440 tage over the cubic model in that, despite
 441 its lack of mechanistic-underpinning, useful
 442 biological inferences can be made when it
 443 is fit well.

Levins (1966) said that the unavoidable 455
 paradox of model fitting in biology is that 456
 one must sacrifice either generality, preci- 457
 sion, or realism in the formulation of bio- 458
 logical models. Whilst many may dismiss 459
 the cubic model for its lack of mechanis- 460
 tic property, is sacrificing realism in using 461
 a phenomenological model like the cubic 462
 worse than sacrificing precision by using a 463
 mechanistic model like Schoolfield? I would 464
 argue it is not. 465

444 Ultimately, the 'best' model depends on
 445 the question you are asking. If the un-
 446 derstanding of mechanism is your goal,
 447 Schoolfield is still the best, as it is the only
 448 one of these three that is based on such
 449 theory. If you are interested in thermal tol-

Mention affect of bad B0 for schoolfield 466
 Akaike weights and tings Growth rate tends 467
 to have a non-linear relationship for arthro- 468
 pods (Briere et al. 1999) which may make 469
 the cubic model less useful for interpreta- 470
 tion. The Briere curve intercepts the tem- 471

472 perature axis at high and low temperatures,
 473 allowing estimation of an upper and lower
 474 trait threshold. AICc a much better com-
 475 parison tool than \bar{R}^2 (Johnson & Omland
 476 2004) The cubic polynomial may have no
 477 biological underpinning but is sacrificing re-
 478 alism much different to sacrificing precision
 479 (Levins 1966)?

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