





# Effect of Water, Sanitation, Handwashing, and Nutrition Interventions on Enteropathogens in Children 14 Months Old: A Cluster-Randomized Controlled Trial in Rural Bangladesh

Jessica A. Grembi, <sup>1,0</sup> Audrie Lin, <sup>2</sup> Md. Abdul Karim, <sup>3</sup> Md. Ohedul Islam, <sup>3</sup> Rana Miah, <sup>3</sup> Benjamin F. Arnold, <sup>4</sup> Elizabeth T. Rogawski McQuade, <sup>5</sup> Shahjahan Ali, <sup>3,0</sup> Md. Ziaur Rahman, <sup>3</sup> Zahir Hussain, <sup>3</sup> Abul K. Shoab, <sup>3</sup> Syeda L. Famida, <sup>3</sup> Md. Saheen Hossen, <sup>3</sup> Palash Mutsuddi, <sup>3</sup> Mahbubur Rahman, <sup>3,0</sup> Leanne Unicomb, <sup>3</sup> Rashidul Haque, <sup>3</sup> Mami Taniuchi, <sup>5</sup> Jie Liu, <sup>5</sup> James A. Platts-Mills, <sup>5</sup> Susan P. Holmes, <sup>6</sup> Christine P. Stewart, <sup>7,0</sup> Jade Benjamin-Chung, <sup>2</sup> John M. Colford Jr, <sup>2</sup> Eric R. Houpt, <sup>5</sup> and Stephen P. Luby. <sup>1,0</sup>

<sup>1</sup>Division of Infectious Diseases and Geographic Medicine, Stanford University, Stanford, California, USA, <sup>2</sup>Division of Epidemiology and Biostatistics, School of Public Health, University of California, Berkeley, Berkeley, California, USA, <sup>3</sup>Infectious Diseases Division, International Centre for Diarrhoeal Disease Research, Bangladesh, Dhaka, Bangladesh, <sup>4</sup>Francis I. Proctor Foundation, University of California, San Francisco, San Francisco, California, USA, <sup>5</sup>Division of Infectious Diseases and International Health, University of Virginia, Charlottesville, Virginia, USA, <sup>6</sup>Department of Statistics, Stanford University, Stanford, California, USA, and <sup>7</sup>Institute for Global Nutrition, University of California, Davis, Davis, California, USA

*Background.* We evaluated the impact of low-cost water, sanitation, and handwashing (WSH) and child nutrition interventions on enteropathogen carriage in the WASH Benefits cluster-randomized controlled trial in rural Bangladesh.

*Methods.* We analyzed 1411 routine fecal samples from children  $14 \pm 2$  months old in the WSH (n = 369), nutrition counseling plus lipid-based nutrient supplement (n = 353), nutrition plus WSH (n = 360), and control (n = 329) arms for 34 enteropathogens using quantitative polymerase chain reaction. Outcomes included the number of co-occurring pathogens; cumulative quantity of 4 stunting-associated pathogens; and prevalence and quantity of individual pathogens. Masked analysis was by intention-to-treat.

**Results.** Three hundred twenty-six (99.1%) control children had 1 or more enteropathogens detected (mean,  $3.8 \pm 1.8$ ). Children receiving WSH interventions had lower prevalence and quantity of individual viruses than controls (prevalence difference for noro-virus: -11% [95% confidence interval {CI}, -5% to -17%]; sapovirus: -9% [95% CI, -3% to -15%]; and adenovirus 40/41: -9% [95% CI, -2% to -15%]). There was no difference in bacteria, parasites, or cumulative quantity of stunting-associated pathogens between controls and any intervention arm.

**Conclusions.** WSH interventions were associated with fewer enteric viruses in children aged 14 months. Different strategies are needed to reduce enteric bacteria and parasites at this critical young age.

Keywords. enteric pathogens; water, sanitation, and handwashing; WSH; nutrition; Bangladesh; child health.

Diarrheal disease remains the fifth leading cause of child mortality, with the highest burdens in low- and middle-income countries [1]. Children who survive often have persistent deficits in physical growth and cognitive development [2–4]. Asymptomatic polymicrobial pathogen carriage is common in areas with high diarrheal disease burden [5, 6]. In some cases, subclinical pathogen carriage had stronger negative dose-dependent associations with child growth than pathogen-associated diarrhea [5–8]. Mucosal damage induced by specific

formance of oral vaccines, again irrespective of diarrheal symptoms [9, 10].

Interventions targeting household dripking water can

enteropathogens has also been correlated with decreased per-

Interventions targeting household drinking water, sanitation, and handwashing (WSH) practices aim to reduce fecal-oral transmission of enteropathogens in areas without municipal water and sewerage. Household WSH trials have reported varied success in reducing diarrhea and/or specific enteropathogens [11-14]. WSH interventions combined with child-specific nutrition interventions could be synergistic in improving growth, especially when dietary diversity and caloric intake are limited. Nutritional interventions might support improved gut mucosal immune function and a more resilient commensal gut microbiota to provide colonization resistance and pathogen clearance after infection [15, 16]. However, there is potential for nutritional interventions supplying iron to provide a growth advantage to iron-scavenging pathogens over beneficial commensals that promote intestinal barrier function [17]. Meta-analyses of

#### The Journal of Infectious Diseases® 2023;227:434–47

© The Author(s) 2020. Published by Oxford University Press for the Infectious Diseases Society of America. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited. https://doi.org/10.1093/infdis/jiaa549

Received 26 May 2020; editorial decision 21 August 2020; accepted 25 August 2020; published online August 29, 2020.

Correspondence: Jessica A. Grembi, PhD, Division of Infectious Diseases and Geographic Medicine, Stanford University, 300 Pasteur Dr, L-134, Stanford, CA 94305 (jgrembi@stanford.edu).

multiple-micronutrient fortification of complementary foods, covering mostly low-income countries across Asia, Africa, and the Americas over the past decade, report no impact or increased risk of diarrhea [18–20]. None of these studies measured enteropathogens.

Direct measurement facilitates objective assessment of pathogen carriage [21]. Most studies have been limited by the measurement of a small number of enteropathogens. Only the Sanitation, Hygiene, Infant Nutrition Efficacy (SHINE) trial in Zimbabwe has evaluated the impact of WSH, child nutrition, and combined child nutrition and WSH (N+WSH) interventions on a comprehensive suite of enteropathogens. Compared with controls, no intervention reduced enteropathogen prevalence or quantity in children 6–12 months old, indicating interventions might have been insufficient to disrupt pathogen transmission [13].

The WASH Benefits Bangladesh cluster-randomized controlled trial reported 31%-38% relative reductions in child diarrhea from WSH, nutrition, and N+WSH interventions; and 0.13-0.25 higher length-for-age z scores at 22 months of age for children receiving nutrition and N+WSH interventions compared with controls [22]. A sister trial in Kenya found comparable growth improvements for children receiving nutrition and N+WSH interventions but no reduction in diarrhea for any group, despite similar interventions and sample size, results consistent with the SHINE trial [23]. In Bangladesh, health promoters had more frequent contact with study households and intervention uptake was higher than the other trials [23]. An evaluation of 6 enteric parasites in children  $30 \pm 2$  months old from the Bangladesh trial reported lower prevalence of Giardia and hookworm in WSH and N+WSH arms than controls [24, 25]. In Kenya, children from the WSH and N+WSH arms had lower Ascaris prevalence than controls [26]. These findings suggest that WSH interventions can reduce parasites in young children; however, the leading etiologies of diarrheal disease for children <5 years old are bacterial and viral [27, 28].

This study assessed the impact of WSH, nutrition, and N+WSH interventions on 34 bacterial, viral, and parasitic enteropathogens in a subset of WASH Benefits Bangladesh children  $14 \pm 2$  months of age. We evaluated the impact of interventions on prevalence and quantity of individual pathogens, as well as composite measures: the number of unique enteropathogens detected, in total and stratified by bacteria, viruses, and parasites; and a novel measure of the quantity of the 4 most prominent stunting-associated pathogens from the Etiology, Risk Factors, and Interactions of Enteric Infections and Malnutrition and the Consequences for Child Health and Development Project (MAL-ED) (ie, enteroaggregative *Escherichia coli* [EAEC], *Shigella*/enteroinvasive *E. coli* [EIEC], *Campylobacter* spp, and *Giardia* spp) [5].

#### **MATERIALS AND METHODS**

## **Study Design and Population**

The WASH Benefits Bangladesh study measured the impact of WSH and nutrition interventions on child growth, development, and parasitic infection (ClinicalTrials.gov, NCC01590095) [22, 24, 25, 29, 30]. Intervention arms included drinking water treatment, sanitation, handwashing, combined WSH, nutrition, and combined N+WSH. This analysis includes children enrolled in the environmental enteric dysfunction substudy, which was a subsample of clusters from the WSH, nutrition, N+WSH, and control arms (in a 1:1:1:1 ratio) [31, 32]. Interventions were delivered while mothers were pregnant, so children were exposed to interventions from birth; a detailed description of interventions has been provided previously [22]. In brief, the WSH intervention consisted of chlorine tablets and a safe drinking water storage vessel; a dual-pit latrine with a water seal, child potties, and hoes for feces disposal; and handwashing stations (including detergent soap with dispensers) near the latrine and kitchen. The nutrition intervention consisted of age-appropriate infant feeding recommendations plus lipid-based nutrient supplements twice daily from age 6 months to 24 months. The N+WSH intervention combined the WSH and nutrition packages. Behavior change messaging was delivered 6 times per month to intervention households, which resulted in high adherence [22]. Rotavirus vaccination had not been implemented in Bangladesh at the time of the study.

Although enteropathogen prevalence tends to increase with age over the first 2 years of life, the importance of early pathogen carriage on later health outcomes (eg, stunting and cognitive deficits [5, 33]) motivated us to evaluate children at a younger age  $(14 \pm 2 \text{ months})$  than the parasite studies  $(30 \pm 2 \text{ months})$  [31, 32]. Written informed consent was obtained from parents of all children. The trial was approved by human subjects committees at the International Centre for Diarrhoeal Disease Research, Bangladesh (icddr,b; PR-11063), the University of California, Berkeley (2011-09-3652), and Stanford University (25863).

## Fecal Sample Collection, Total Nucleic Acid Extraction, and Pathogen Quantification

Primary caregivers collected child fecal samples, which were placed on cold chain 155 minutes (interquartile range, [IQR], 80–529) after defecation, transported on dry ice to the laboratory, and stored at –80°C. DNA and RNA were extracted using the QIAamp Fast DNA Stool Mini kit (Qiagen, Venlo, The Netherlands) and a modified protocol, which included spikeins of 2 extrinsic controls to monitor extraction and amplification efficiency [34]. Enteropathogens (Supplementary Table 1) were measured via quantitative polymerase chain reaction (PCR) using a TaqMan array card (validation, conditions, and quality controls measures reported previously [28, 34]) at icddr,b. Quantification cycle value was used as an inverse

measure of pathogen quantity, with 1 unit corresponding to a doubling of pathogen quantity and the analytical limit of detection at quantification cycle 35 [27]. Pathogen quantities were normalized based on per-sample extraction/amplification efficiency. For nondetects, quantity was set to half of the detection limit (1.8495 log<sub>10</sub> copies/g of stool).

#### **Statistical Analysis**

Analysis was by intention-to-treat. Analysis plans describing pathogen outcomes were prespecified (see https://osf.io/ ky275/). This study did not target collection of diarrheal stools. The subset of stools from children with reported diarrheal symptoms in the previous 7 days was small and not representative of all diarrhea episodes, which prohibited an analysis of diarrheal stools. Therefore, our analysis focuses on outcomes of enteropathogen carriage in children, including both symptomatic and asymptomatic pathogen detection. We also conducted 3 post hoc analyses that (1) excluded samples from children with reported diarrheal symptoms, (2) evaluated seasonal effect modification of intervention efficacy, and (3) adjusted for only child age and season of sample collection. Investigators were masked to group assignment until primary analysis was complete. Statistical analyses were performed in R software (version 3.5.2) [35].

Composite outcomes included the number of pathogens in total and by type (bacteria, virus, parasite) and an aggregate metric for the quantity of 4 stunting-associated organisms (EAEC, *Shigella*/EIEC, *Campylobacter* spp, *Giardia* spp) that were independently associated with linear growth deficits in a longitudinal analysis of the MAL-ED study where the strength of association was robust to the presence of other enteropathogens [5]. We used g-computation [13] to estimate the absolute difference in the outcome between study arms (using Poisson regression for the number of pathogens and linear regression for the composite stunting-associated pathogen quantity). We obtained confidence intervals (CIs) with a nonparametric bootstrap (B = 1000) that resampled clusters with replacement.

For the 18 pathogens detected in >5% of samples, we estimated prevalence differences and ratios between study arms using generalized linear models with robust standard errors [22, 31]. We also estimated differences in  $\log_{10}$  quantity using the g-computation estimator with a 2-component model, including logistic and log-linear regression steps to account for the sparse, semicontinuous quantity data [5, 13]. In addition to 95% CIs, we report P values corrected for multiple comparisons using the Benjamini–Hochberg procedure [36] within each treatment contrast and model type. For quantity analyses, we employed a double bootstrap (B = 1000 for outer, and BB = 25 for inner bootstrap) to estimate P values on g-computation results before applying the Benjamini–Hochberg correction.

We prespecified a fully adjusted analysis to improve precision of estimates and control for potential residual confounding

from baseline and sample-specific covariates (Supplementary Table 3). Covariates for adjusted analyses were prescreened with a likelihood ratio test, and those with P < .1 in bivariate analysis with the outcome were retained for the fully adjusted model. A final analysis accounted for differential missingness of the outcome due to incomplete sample collection using data-adaptive, targeted maximum likelihood estimation, and inverse probability of censoring weighting [37].

#### **RESULTS**

Of 1532 eligible children, 1411 fecal samples with valid qPCR results were obtained from children aged 14 ± 2 months enrolled in the WSH (n = 369), nutrition (n = 353), N+WSH (n = 360), and control (n = 329) arms of the WASH Benefits Bangladesh trial (110 did not provide a fecal sample; 21 did not provide sufficient sample). Diarrheal symptoms were reported in the past 7 days for 214 (14.0%) children, and the proportion of children who did not provide a fecal sample was not different between those with (0.06 [95% CI, .03-.10]) or without (0.07 [95% CI, .06-.09]) reported diarrheal symptoms. Household enrollment characteristics were balanced across study arms and were similar to the main trial (Table 1 and Supplementary Table 2). Fewer children from the control arm provided fecal samples, and there was imbalance across study arms for 2 prognostic covariates related to sample collection: season (monsoon vs dry; Supplementary Figure 1) and child age (Welch's t test between control and each intervention arm: season P < .006, age P < .0001; Table 1). Several bacterial enteropathogens were more prevalent during the monsoon season across all arms (Supplementary Figure 2). Imbalance by arm and strong relationship between enteropathogen carriage and both monsoon season and age led us to rely on adjusted analyses for primary inference, according to our analysis plan.

Only 18 organisms were detected in >5% of stool samples (Supplementary Figure 3). The most prevalent bacteria were EAEC (76%), Campylobacter spp (47%), atypical enteropathogenic E. coli (41%), and enterotoxigenic E. coli (40%; 20% each for heat-labile and heat-stable toxin, 5% of samples had both). The most prevalent viruses were norovirus (17%; 15% for genogroup GII, 3% for GI, 1% of samples had both), sapovirus (12%), and adenovirus 40/41 (8%). Rotavirus was present in 2.2% of all samples, and 4.0% of samples where caregivers reported diarrhea in the past 7 days. The parasites detected at >5% prevalence were Giardia (14%), Cryptosporidium (12%), and Enterocytozoon bieneusi (10%).

## Impact of Interventions on Composite Pathogen Load

Enteropathogen carriage was high: 326 (99.1%) control children had at least 1 pathogen detected, with an average of 3.8 (standard deviation [SD], 1.8) co-occurring pathogens: 2.8 (SD, 1.4) bacteria, 0.6 (SD, 0.6) viruses, and 0.4 (SD, 0.7) parasites. Children in the WSH group had 0.50 (95% CI, .07–.90) fewer total

Table 1. Household Enrollment and Child/Sample Characteristics by Intervention Arm

Characteristic	Control $(n = 499)$	WSH (n = 446)	Nutrition $(n = 435)$	N+WSH (n = 447)
Household enrollment characteristics				
Maternal				
Age, y	23 (5)	24 (5)	24 (5)	24 (5)
Height, cm	151 (5)	150 (5)	150 (6)	150 (5)
Years of education	7 (3)	6 (3)	6 (4)	6 (3)
Paternal				
Years of education	5 (4)	5 (4)	5 (4)	5 (4)
Works in agriculture	104 (23%)	128 (29%)	148 (34%)	127 (28%)
Household				
No. of people	5 (2)	5 (2)	5 (2)	5 (2)
No. of children <18 y	2 (1)	2 (1)	2 (1)	2 (1)
Has electricity	269 (60%)	278 (62%)	269 (62%)	272 (61%)
Has a cement floor	75 (17%)	55 (12%)	50 (11%)	53 (12%)
Acres of agricultural land owned	0.18 (0.25)	0.17 (0.26)	0.17 (0.30)	0.13 (0.18)
Drinking water				
Shallow tubewell is primary water source	329 (73%)	337 (76%)	309 (71%)	318 (71%)
Has stored water at home	230 (51%)	199 (45%)	209 (48%)	229 (51%)
Reported treating water yesterday	1 (0%)	0 (0%)	0 (0%)	1 (0%)
Minutes to primary drinking water source	1 (2)	1 (6)	1 (2)	1 (2)
Sanitation				
Daily defecating in the open				
Adult men	19 (4%)	29 (7%)	38 (9%)	38 (9%)
Adult women	12 (3%)	16 (4%)	23 (5%)	21 (5%)
Children aged 8 to <15 y	9 (5%)	17 (8%)	13 (8%)	22 (11%)
Children aged 3 to <8 y	65 (30%)	89 (37%)	90 (40%)	92 (37%)
Children aged 0 to <3 y <sup>a</sup>	71 (72%)	73 (75%)	68 (80%)	79 (88%)
Latrine				
Owned <sup>b</sup>	271 (60%)	244 (55%)	234 (54%)	230 (51%)
Concrete slab	426 (97%)	400 (93%)	382 (93%)	399 (94%)
Functional water seal	157 (38%)	95 (26%)	114 (32%)	111 (31%)
Visible stool on slab or floor	197 (45%)	225 (54%)	210 (52%)	222 (53%)
Owned a child potty	37 (8%)	20 (4%)	27 (6%)	21 (5%)
Human feces observed				
In the house	25 (6%)	36 (8%)	41 (9%)	36 (8%)
In the child's play area	5 (1%)	4 (1%)	7 (2%)	6 (1%)
Handwashing location				
Within 6 steps of latrine				
Has water	84 (21%)	51 (13%)	38 (10%)	54 (13%)
Has soap	45 (11%)	32 (8%)	23 (6%)	27 (7%)
Within 6 steps of kitchen				
Has water	48 (12%)	40 (10%)	43 (11%)	42 (10%)
Has soap	18 (4%)	11 (3%)	19 (5%)	14 (3%)
lutrition				
Household is food secure <sup>c</sup>	331 (74%)	298 (67%)	308 (71%)	317 (71%)
Child/sample characteristics		()		
Female child	225 (50%)	238 (53%)	228 (52%)	214 (47%)
Child stool sample collected	377 (84%)	394 (88%)	379 (87%)	380 (84%)
Child diarrhea reported at stool collection visit	67 (18%)	43 (11%)	61 (16%)	43 (11%)
Child age at stool collection, d	455 (66)	415 (57)	424 (54)	417 (57)
Sample collected during monsoon season <sup>d</sup>	155 (41%)	283 (72%)	244 (64%)	269 (71%)
Time until sample placed on cold chain, mine, median (IQR)	160 (76–767)	159 (79–495)	148 (80–398)	158 (90–525

Values represent either the mean (standard deviation) or No. (%) unless otherwise indicated.

 $Abbreviations: IQR, interquartile\ range; N+WSH,\ nutrition\ plus\ water,\ sanitation,\ and\ handwashing; WSH,\ water,\ sanitation,\ and\ handwashing; WSH,\ water,\ sanitation,\ and\ handwashing; water,\ sanitation,\ sanitat$ 

<sup>&</sup>lt;sup>a</sup>Open defecation does not include diaper disposal of feces.

<sup>&</sup>lt;sup>b</sup>Households that do not own a latrine typically share a latrine with extended family members who live in the same compound.

 $<sup>^{\</sup>rm c}\!$  Assessed by the Household Food Insecurity Access Scale.

<sup>&</sup>lt;sup>d</sup>Monsoon season is May-October.

eTime between when caregiver reported collecting child's fecal sample and field staff received sample and placed it on cold chain; values were not normally distributed.

pathogens and 0.28 (95% CI, .09–.48) fewer total viruses compared with control children (Table 2). Children who received nutritional interventions had fewer total viruses than controls (0.17 [95% CI, .05–.38] for nutrition; 0.21 [95% CI, .06–.37] for N+WSH). There was no difference in total pathogens—nor when broken down by bacteria, viruses, or parasites—between the combined N+WSH intervention and either the WSH- or nutrition-only arm. Virus results were robust across unadjusted and adjusted models (Supplementary Figure 4A).

In control children, the stunting-associated pathogen composite score (including EAEC, *Shigella*/EIEC, *Campylobacter* spp, and *Giardia* spp) was associated with concurrent, but not future, length-for-age *z* score (Supplementary Figure 5). There was no difference in the total load of the stunting-associated pathogens between children in any of the intervention arms compared with controls, nor between N+WSH children compared with children receiving single WSH or nutrition interventions (Supplementary Figure 4*B*).

#### Impact of Interventions on Individual Pathogen Prevalence and Quantity

Compared with controls, children receiving WSH interventions had significantly lower prevalence of norovirus, sapovirus, and adenovirus 40/41 (Figure 1 for prevalence ratios, Table 3 for prevalence differences). This corresponded to a mean 0.45 (95% CI, .21–.70)  $\log_{10}$  fewer norovirus, 0.38 (95% CI, .07–.74)  $\log_{10}$  fewer sapovirus, and 0.47 (95% CI, .10–.84)  $\log_{10}$  fewer adenovirus 40/41 when evaluated quantitatively (Figure 2 and Supplementary Table 4). These findings were consistent with unadjusted estimates. Children receiving the nutrition-only intervention had an absolute 8%–10% lower prevalence of EAEC and sapovirus than controls (Table 3), with similar results obtained for pathogen quantity (0.62 [95% CI, .09–1.11]  $\log_{10}$  fewer EAEC; 0.33 [95% CI, .03–.72]  $\log_{10}$  fewer sapovirus; Figure 2 and Supplementary Table 4), but neither was significant after multiple comparisons correction.

There was no difference in the prevalence or quantity of individual pathogens when comparing N+WSH children to those from the single nutrition arm, but there was a 6% (95% CI, 1%–12%) higher prevalence and 0.28 (95% CI, .05–.50) higher  $\log_{10}$  quantity of norovirus compared with children receiving the WSH-only intervention, although neither was significant after multiple comparisons correction (Figures 1 and 2, Table 3, and Supplementary Table 4).

Notably, there were no differences in protozoa between study arms. Results across all outcomes were comparable when adjusting for missing outcomes (children who did not provide fecal samples), when adjusting only for child age and season of sample collection, or when evaluating only samples from children with no reported diarrheal symptoms within the past 7 days (Supplementary Tables 5 and 6 and Supplementary Figure 4). Seasonal effect modification of intervention efficacy was not observed; however, point estimates suggested slightly

greater efficacy during the dry season for some pathogens (Supplementary Figures 6–8).

#### **DISCUSSION**

Children in rural Bangladesh receiving low-cost WSH, nutrition, or N+WSH interventions [38, 39] had similar prevalence and quantity of bacterial and parasitic enteropathogens in their stool at 14 months old compared with controls. Importantly, the 4 bacterial/protozoan enteropathogens most commonly associated with stunting [5] were not impacted by the interventions, both when evaluated individually or in total. This is consistent with lack of effect of the WSH interventions on child growth in the main trial, and a modest effect of nutrition interventions that was equivalent to other nutrition-only trials [22, 40]. However, the WSH intervention was associated with fewer enteric viruses, as evidenced by lower total viruses and lower prevalence and/or log10 quantity of norovirus, sapovirus, and adenovirus 40/41. Viruses are among the top diarrhea-causing pathogens in this age group [27, 28], and the combined reduction in viruses (~30%) is consistent with the diarrhea reductions reported for children whose households received WSH interventions [22], providing strong evidence for causality. Despite similar pathogen carriage profiles, children receiving nutrition interventions had lower diarrhea prevalence than controls [22], suggesting the nutrition intervention might be preventing illness when children are exposed to enteropathogens. Some parasite results were discordant with those from the main trial when children were older, which is discussed below.

Our findings parallel those of the SHINE trial in rural Zimbabwe. Both trials found no impact of nutrition interventions on enteropathogens [13]. The SHINE trial found that WSH interventions reduced *neither* diarrhea nor enteropathogens for children 6–12 months old [13, 41], whereas our study found *both* lower diarrhea *and* lower viral pathogens for children 14 months old receiving WSH interventions compared with controls. Less frequent intervention promotion could have led to lower adherence in the SHINE trial [23].

Our nutritional intervention was not associated with increased diarrhea or enteropathogens. Importantly, our supplement was consumed twice daily (4.5 mg iron per dose) to increase host absorption and reduce bacterial iron scavenging, compared with micronutrient powders that supply 1 daily 12.5-mg iron dose [17]. Furthermore, children receiving the nutrition intervention had less transferrin receptor [32], one viral mechanism for cellular invasion [42], possibly explaining the lower total viruses for this group. Although nonsignificant after multiple hypothesis correction, norovirus prevalence and quantity were higher in the N+WSH arm than the WSH arm, suggesting a possible interaction between the nutrition and WSH interventions. A potential explanation is that the nutrition intervention increased specific populations of gut bacteria, resulting in improved norovirus survivability and infectivity [43].

Table 2. Differences in the Number of Co-occurring Pathogens Between Intervention Arms in Fecal Samples Collected From Rural Bangladeshi Children Aged 14 Months Enrolled in the WASH Benefits Randomized Controlled Trial Environmental Enteric Dysfunction Substudy

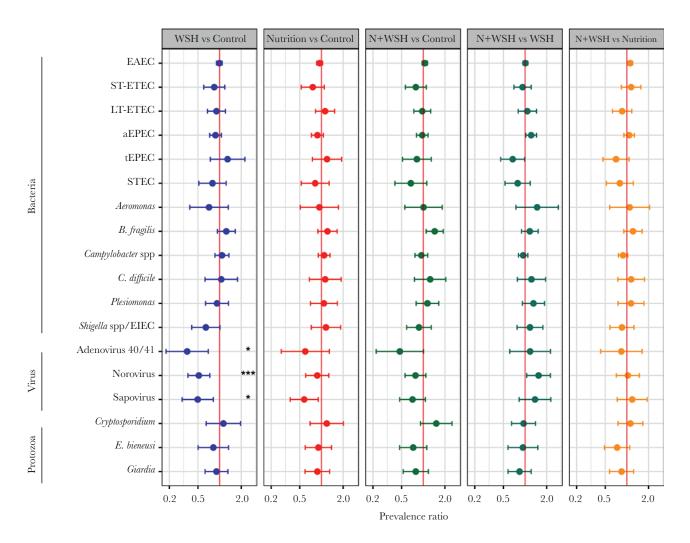
			No. of Co-oo	No. of Co-occurring Pathogens				
		Total <sup>a</sup>	Mons	Monsoon Season	Ī	Dry Season	Pathogen Diffe	Pathogen Difference <sup>b</sup> (95% CI)
Pathogen Group, Study Arm	No.	Mean (SD)	No.	Mean (SD)	No.	Mean (SD)	Intervention vs Control	N+WSH vs Intervention
All pathogens								
Control	299	3.8 (1.80)	116	3.8 (1.69)	183	3.9 (1.87)	AN	NA
WSH	351	3.8 (1.72)	266	4.1 (1.66)	84	2.9 (1.61)	-0.50 (90 to07)	0.17 (05 to .52)
Nutrition	344	4.0 (1.84)	233	4.3 (1.80)	111	3.4 (1.80)	-0.19 (68 to .18)	-0.02 (38 to .27)
N+WSH	339	4.1 (1.89)	251	4.3 (1.91)	88	3.4 (1.67)	-0.16 (65 to .17)	NA
Bacteria								
Control	323	2.80 (1.38)	121	2.8 (1.34)	202	2.8 (1.41)	ZA	NA
WSH	363	3.0 (1.53)	273	3.3 (1.49)	68	2.2 (1.33)	-0.18 (50 to .10)	0.08 (11 to .35)
Nutrition	348	3.1 (1.57)	236	3.4 (1.60)	112	2.4 (1.31)	-0.07 (42 to .15)	0.02 (25 to .32)
N+WSH	352	3.1 (1.56)	257	3.4 (1.60)	92	2.5 (1.26)	-0.04 (35 to .22)	NA
Viruses								
Control	312	0.60 (0.63)	123	0.60 (0.70)	189	0.60 (0.58)	NA	NA
WSH	358	0.30 (0.56)	271	0.30 (0.56)	98	0.40 (0.55)	-0.28 (48 to09)	0.15 (02 to .24)
Nutrition	349	0.40 (0.64)	237	0.40 (0.59)	112	0.50 (0.73)	-0.17 (38 to05)	-0.02 (16 to .10)
N+WSH	349	0.40 (0.63)	258	0.40 (0.60)	91	0.50 (0.69)	-0.21 (37 to06)	NA
Parasites								
Control	315	0.40 (0.65)	118	0.30 (0.58)	197	0.40 (0.68)	٩Z	NA
WSH	358	0.40 (0.65)	270	0.50 (0.67)	87	0.30 (0.55)	-0.01 (17 to .09)	0.01 (06 to .17)
Nutrition	347	0.50 (0.78)	234	0.50 (0.80)	113	0.50 (0.70)	0.03 (06 to .28)	-0.01 (21 to .09)
N+WSH	346	0.50 (0.70)	254	0.50 (0.70)	92	0.40 (0.71)	0.02 (09 to .20)	NA

Bold font indicates significance.

Abbreviations: Cl, confidence interval; NA, not applicable; N+WSH, nutrition plus water, sanitation, and handwashing; WSH, water, sanitation, and handwashing.

<sup>a</sup>Includes both monsoon (May–October) and dry (November–April) seasons.

bdistred for prespecified covariates using generalized linear models and g-computation estimator: household food insecurity, child age, child sex, child birth order, season of sample between defecation and sample placed on cold chain, mother's education level, number of children <18 years of age in the household, number of individuals living in the compound, distance in minutes to the primary water source, household floor and wall materials, household mother's age, mother's education level, number of children <18 years of age in the household, number of individuals living in the compound, distance in minutes to the primary water source, household floor and wall materials, household



**Figure 1.** Impact of interventions on prevalence ratio of individual pathogens in 14-month-old children from rural Bangladesh. Point estimates and 95% confidence intervals were determined with a generalized linear model adjusting for covariates associated with each pathogen outcome (likelihood ratio test P < .1 in bivariate analysis): household food insecurity, child age, child sex, child birth order, season of sample collection, time between defecation and sample placed on cold chain, mother's age, mother's height, mother's education level, number of children aged <18 years in the household, number of individuals living in the compound, distance in minutes to the primary water source, household floor and wall materials, and household assets. Pathogens significant after correction for false discovery rate are annotated: \*P < .05, \*\*\*P < .05. Abbreviations: aEPEC, atypical enteropathogenic *Escherichia coli*; EAEC, enteroaggregative *Escherichia coli*; EIEC, enteroinvasive *Escherichia coli*; LT-ETEC, enterotoxigenic *Escherichia coli* with heat-labile toxin; N+WSH, nutrition plus water, sanitation, and handwashing; STEC, Shiga toxin—producing *Escherichia coli*; ST-ETEC, enterotoxigenic *Escherichia coli* with heat-stable toxin; tEPEC, typical enteropathogenic *Escherichia coli*; WSH, water, sanitation, and handwashing.

This is not at odds with the growth outcomes from the main trial as viruses are not the top pathogen predictors of stunting.

An important limitation of this study was imbalance in child age and season between study arms at the time of fecal sample collection. Political unrest associated with the Bangladesh 2014 general election disproportionately impacted our ability to collect samples in control villages, where community members were not familiar with the project due to lack of visible interventions or routine visits by health promoters. Imbalance in age between arms is important because pathogen prevalence increases over the first 2 years of life [5]; control children from our substudy were already walking at the time of sample collection (study children began walking at 13.0 [IQR, 11.9–14.0] months [29]), potentially increasing their environmental exposure to pathogens. The observed seasonal variability of enteropathogens

is consistent with previous data from Bangladesh and seasonal diarrhea patterns from the WASH Benefits trial [2, 22], which could have biased our findings toward the null. To address these limitations, we relied on adjusted analyses that controlled for these covariates. We found no effect modification of intervention effectiveness by season, but estimates were imprecise due to low power to detect interaction.

Another limitation was the use of a single assessment to measure the impact of interventions on enteropathogens. Incidence of enteric viruses peaks at the age we evaluated [27]; however, the median age at initial detection of *Giardia* in surveillance stools in Bangladesh was 18 months [44]. Persistent *Giardia* and helminth infections are common, increasing the proportion of infected children at older ages; thus, cumulative differences would increase with a successful intervention as

Downloaded from https://academic.oup.com/jid/article/227/3/434/5899213 by guest on 31 August 2024

Table 3. Prevalence Difference Between Intervention Arms for Individual Pathogens

			Pathogen	en Prevalence				revalence Differen	Prevalence Difference Between Arms <sup>a</sup>	
		Total <sup>b</sup>	Mon	Monsoon Season	Dr	Dry Season	Intervention vs Control	Control	N+WSH vs Intervention	vention
Pathogen and Arm	o N	Mean (SD)	o N	Mean (SD)	O	Mean (SD)	PD (95% CI)	PValue <sup>c</sup>	PD (95% CI)	PValue⁵
EAEC										
Control	328	0.76 (0.43)	123	0.8 (0.4)	205	0.73 (0.44)	NA	AN	NA	NA
WSH	364	0.79 (0.41)	274	0.83 (0.38)	68	0.69 (0.47)	-0.01 (08 to .06)	.84	0.01 (05 to .06)	.83
Nutrition	320	0.72 (0.45)	237	0.75 (0.43)	113	0.65 (0.48)	-0.1 (17 to02)	.10	0.08 (.01–.14)	.29
N+WSH	354	0.8 (0.4)	259	0.81 (0.39)	92	0.76 (0.43)	0 (08 to .08)	1.00	NA	NA
STETEC										
Control	328	0.18 (0.38)	123	0.24 (0.43)	205	0.14 (0.34)	NA	A A	AA	A A
WSH	364	0.22 (0.42)	274	0.27 (0.45)	68	0.07 (0.25)	-0.02 (09 to .04)	.67	-0.01 (07 to .05)	18.
Nutrition	350	0.18 (0.38)	237	0.24 (0.43)	113	0.06 (0.24)	-0.05 (11 to .02)	.65	0.03 (03 to .09)	.65
N+WSH	354	0.21 (0.41)	259	0.27 (0.44)	92	0.04 (0.2)	-0.04 (1 to .03)	.54	AN	ΥZ
LFETEC										
Control	326	0.19 (0.39)	123	0.17 (0.38)	203	0.2 (0.4)	NA	Ϋ́	AN	ΥZ
WSH	364	0.19 (0.39)	274	0.22 (0.42)	68	0.08 (0.27)	-0.04 (1 to .01)	.42	0.02 (04 to .07)	18.
Nutrition	350	0.23 (0.42)	237	0.26 (0.44)	113	0.18 (0.38)	0.02 (04 to .09)	.65	-0.03 (1 to .04)	.65
N+WSH	354	0.21 (0.41)	259	0.23 (0.42)	92	0.16 (0.37)	0 (06 to .05)	1.00	AN	₹Z
aEPEC										
Control	327	0.46 (0.50)	122	0.38 (0.49)	205	0.51 (0.50)	NA	A A	AN	٨Z
WSH	364	0.36 (0.48)	274	0.34 (0.47)	68	0.43 (0.5)	-0.05 (13 to .02)	.47	0.08 (.01–.15)	.26
Nutrition	350	0.4 (0.49)	237	0.4 (0.49)	113	0.41 (0.49)	-0.06 (14 to .02)	.65	0.04 (03 to .11)	.65
N+WSH	353	0.44 (0.5)	258	0.41 (0.49)	92	0.51 (0.5)	0 (09 to .08)	1.00	ΝΑ	Ϋ́Z
tEPEC										
Control	328	0.08 (0.28)	123	0.11 (0.32)	205	0.06 (0.24)	NA	AN	AN	Ϋ́Z
WSH	364	0.15 (0.36)	274	0.17 (0.38)	68	0.08 (0.27)	0.03 (02 to .08)	.49	-0.05 (09 to 0)	.35
Nutrition	350	0.13 (0.34)	237	0.19 (0.39)	113	0.03 (0.16)	0.02 (03 to .07)	.65	-0.04 (09 to .01)	.58
N+WSH	353	0.1 (0.3)	258	0.14 (0.34)	92	0.01 (0.1)	-0.01 (05 to .03)	.79	AN	ΥZ
STEC										
Control	327	0.11 (0.31)	122	0.18 (0.39)	205	0.07 (0.25)	NA	A A	NA	AN
WSH	364	0.13 (0.34)	274	0.14 (0.35)	68	0.09 (0.29)	-0.02 (07 to .03)	.67	-0.03 (07 to .02)	99.
Nutrition	350	0.12 (0.32)	237	0.14 (0.35)	113	0.07 (0.26)	-0.03 (08 to .03)	.65	-0.02 (07 to .02)	.65
N+WSH	354	0.1 (0.3)	259	0.11 (0.31)	92	0.08 (0.28)	-0.04 (1 to .01)	.46	NA	ΥN
Aeromonas										
Control	327	0.06 (0.25)	123	0.03 (0.18)	204	0.08 (0.28)	NA	A A	ΝΑ	Ϋ́Z
WSH	364	0.05 (0.22)	274	0.07 (0.25)	68	(0) 0	-0.02 (06 to .01)	.49	0.02 (02 to .06)	99.
Nutrition	320	0.06 (0.24)	237	0.08 (0.27)	113	0.03 (0.16)	-0.01 (04 to .03)	.75	0.01 (03 to .05)	.77
N+WSH	354	0.07 (0.25)	259	0.09 (0.29)	92	0.01 (0.1)	0 (04 to .04)	1.00	NA	ΥZ
Bacteroides fragilis										
Control	328	0.19 (0.39)	123	0.14 (0.35)	205	0.21 (0.41)	NA	N A	AA	₹ Z
WSH	364	0.23 (0.42)	274	0.24 (0.43)	68	0.2 (0.4)	0.04 (01 to .1)	.42	0.04 (03 to .11)	99.

Table 3. Continued

			Pathogen	en Prevalence			ā.	revalence Differen	Prevalence Difference Between Arms <sup>a</sup>	
		Total <sup>b</sup>	Mons	Monsoon Season	Dr	Dry Season	Intervention vs Control	ontrol	N+WSH vs Intervention	vention
Pathogen and Arm	O Z	Mean (SD)	No.	Mean (SD)	No.	Mean (SD)	PD (95% CI)	PValue <sup>c</sup>	PD (95% CI)	PValue⁰
Nutrition	350	0.23 (0.42)	237	0.23 (0.42)	113	0.21 (0.41)	0.04 (02 to .1)	.65	0.05 (02 to .12)	.65
N+WSH	354	0.28 (0.45)	259	0.29 (0.45)	92	0.25 (0.44)	0.08 (.0215)	.18	NA	NA A
Campylobacter spp										
Control	328	0.41 (0.49)	123	0.39 (0.49)	205	0.43 (0.5)	NA	ΥN	NA	AN A
WSH	364	0.49 (0.5)	274	0.53 (0.5)	88	0.36 (0.48)	0.04 (06 to .13)	.67	-0.03 (1 to .04)	99.
Nutrition	350	0.52 (0.5)	237	0.56 (0.5)	113	0.42 (0.5)	0.04 (04 to .12)	.65	-0.07 (14 to .01)	.58
N+WSH	354	0.47 (0.5)	259	0.52 (0.5)	92	0.34 (0.48)	-0.03 (11 to .06)	.79	AN	A Z
Clostridioides difficile										
Control	328	0.08 (0.28)	123	0.08 (0.27)	205	0.08 (0.28)	Ϋ́	Ϋ́	AN	A Z
WSH	363	0.09 (0.28)	273	0.1 (0.29)	88	0.07 (0.25)	0 (04 to .05)	8.	0.02 (02 to .06)	99.
Nutrition	348	0.09 (0.29)	236	0.08 (0.28)	112	0.12 (0.32)	0.01 (03 to .06)	.71	0.02 (03 to .06)	99.
N+WSH	353	0.11 (0.31)	258	0.1 (0.3)	92	0.14 (0.35)	0.02 (03 to .07)	.67	AA	AN
Plesiomonas										
Control	328	0.14 (0.34)	123	0.1 (0.3)	205	0.16 (0.37)	NA	ΥN	NA	NA
WSH	363	0.15 (0.35)	273	0.18 (0.39)	88	0.03 (0.18)	-0.01 (06 to .04)	69.	0.04 (01 to .1)	.65
Nutrition	348	0.16 (0.36)	236	0.19 (0.39)	112	0.08 (0.27)	0.01 (05 to .07)	.75	0.02 (05 to .09)	99.
N+WSH	353	0.18 (0.39)	258	0.2 (0.4)	92	0.15 (0.36)	0.02 (04 to .08)	.75	NA	A A
Shigella spp/EIEC										
Control	328	0.13 (0.33)	123	0.15 (0.36)	205	0.11 (0.31)	AN	ΥN	NA	NA
WSH	364	0.13 (0.34)	274	0.16 (0.37)	88	0.03 (0.18)	-0.04 (1 to .01)	.42	0.02 (04 to .08)	69.
Nutrition	350	0.17 (0.37)	237	0.19 (0.4)	113	0.12 (0.32)	0.02 (04 to .09)	.65	-0.02 (09 to .04)	99.
N+WSH	354	0.15 (0.36)	259	0.19 (0.39)	92	0.05 (0.22)	-0.01 (07 to .04)	.79	NA	NA
Adenovirus 40/41										
Control	328	0.1 (0.31)	123	0.22 (0.42)	205	0.03 (0.18)	NA	٩Z	NA	NA A
WSH	364	0.06 (0.23)	274	0.07 (0.25)	88	0.03 (0.18)	-0.09 (15 to02)	80.	0.01 (03 to .05)	.81
Nutrition	350	0.08 (0.28)	237	0.08 (0.28)	113	0.08 (0.27)	-0.05 (12 to .03)	.65	-0.01 (06 to .04)	.68
N+WSH	354	0.07 (0.26)	259	0.07 (0.25)	92	0.08 (0.28)	-0.06 (14 to .01)	.46	NA	NA A
Norovirus										
Control	329	0.23 (0.42)	124	0.21 (0.41)	205	0.24 (0.43)	NA	Y V	NA	A A
WSH	369	0.11 (0.32)	274	0.13 (0.33)	94	0.07 (0.26)	0.11 (17 to05)	.004	0.06 (.01–.12)	.26
Nutrition	353	0.18 (0.38)	237	0.14 (0.35)	116	0.25 (0.43)	-0.03 (1 to .05)	.65	0.01 (06 to .07)	88.
N+WSH	360	0.18 (0.38)	260	0.18 (0.38)	100	0.18 (0.39)	-0.05 (12 to .02)	.46	NA	NA
Sapovirus										
Control	313	0.19 (0.4)	124	0.15 (0.35)	189	0.23 (0.42)	NA	Ϋ́	NA	NA
WSH	364	0.09 (0.28)	272	0.08 (0.27)	91	0.1 (0.3)	-0.09 (15 to03)	.03	0.03 (02 to .08)	99.
Nutrition	352	0.1 (0.3)	237	0.09 (0.29)	115	0.12 (0.33)	-0.08 (14 to02)	.10	0.02 (03 to .07)	99.
N+WSH	355	0.12 (0.32)	259	0.1 (0.3)	96	0.17 (0.37)	-0.05 (12 to .01)	.46	NA	₹N V

Table 3. Continued

			Pathogen	en Prevalence			1	Prevalence Difference Between Arms <sup>a</sup>	ce Between Arms <sup>a</sup>	
		Total <sup>b</sup>	Mons	Monsoon Season	Dry	Dry Season	Intervention vs Control	Control	N+WSH vs Intervention	vention
Pathogen and Arm	o N	Mean (SD)	o N	Mean (SD)	O	Mean (SD)	PD (95% CI)	PValue <sup>c</sup>	PD (95% CI)	$P$ Value $^{\circ}$
Cryptosporidium										
Control	328	0.08 (0.27)	123	0.09 (0.29)	205	0.07 (0.26)	AA	Ϋ́	NA	₹ Z
WSH	364	0.15 (0.35)	274	0.17 (0.38)	68	0.07 (0.25)	0.02 (03 to .07)	.67	-0.01 (06 to .05)	.83
Nutrition	350	0.12 (0.33)	237	0.14 (0.35)	113	0.09 (0.29)	0.02 (03 to .07)	.65	0.01 (04 to .07)	.68
N+WSH	354	0.14 (0.35)	259	0.14 (0.35)	92	0.14 (0.35)	0.04 (01 to .1)	.46	NA	₹Z
Enterocytozoon bieneusi										
Control	327	0.11 (0.31)	123	0.09 (0.29)	204	0.12 (0.32)	AN	ΑN	NA	₹Z
WSH	364	0.1 (0.3)	274	0.11 (0.31)	68	0.08 (0.27)	-0.02 (07 to .03)	.67	-0.01 (05 to .04)	.83
Nutrition	350	0.12 (0.32)	237	0.14 (0.34)	113	0.08 (0.27)	-0.01 (06 to .04)	.70	-0.03 (08 to .01)	.58
N+WSH	354	0.09 (0.29)	259	0.1 (0.3)	92	0.06 (0.24)	-0.03 (07 to .01)	.47	AN	٩Z
Giardia										
Control	328	0.14 (0.34)	123	0.07 (0.26)	205	0.18 (0.38)	AN	₹ V	NA	∢ Z
WSH	364	0.15 (0.35)	274	0.15 (0.36)	68	0.12 (0.33)	-0.01 (07 to .04)	69.	-0.02 (07 to .03)	99.
Nutrition	350	0.14 (0.34)	237	0.12 (0.33)	113	0.17 (0.38)	-0.02 (07 to .04)	.65	-0.02 (07 to .03)	.65
N+WSH	354	0.13 (0.33)	259	0.13 (0.34)	92	0.12 (0.32)	-0.03 (08 to .02)	.53	NA	ΑN

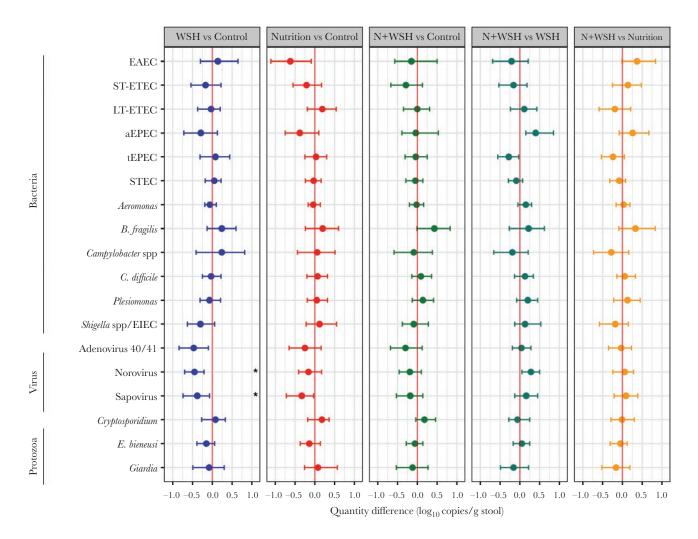
Fecal samples (n = 1411) were collected from rural Bangladeshi children aged 14 months enrolled in the WASH Benefits randomized controlled trial environmental enteric dysfunction substudy. Bold font indicates significance after correcting for false discovery rate using the Benjamini-Hochberg procedure.

Abbreviations: aEPEC, atypical enteropathogenic Escherichia coli, Cl, confidence interval; EAEC, enteroaggregative Escherichia coli; EIEC, enteroinvasive Escherichia coli; UTETEC, enterotoxigenic Escherichia coli with heat-stable toxin; tEPEC, typical enteropathogenic N+WSH, nutrition plus water, sanitation, and handwashing; PD, prevalence difference; SD, standard deviation; STEC, Shiga toxin-producing Escherichia coli; STETEC, enterotoxigenic Escherichia coli with heat-stable toxin; tEPEC, typical enteropathogenic Escherichia coli; WSH, water, sanitation, and handwashing

Adjusted for prespecified covariates with a likelihood ratio test P < .1 in bivariate analysis with the outcome: household food insecurity; child age, sex, and birth order; season of sample collection; time until sample placed on cold chain, mother's age, height and education level; number of children <18 years of age in the household; number of individuals living in the compound; distance in minutes to the primary water source; household floor and wall materials; household assets.

<sup>b</sup>Includes both monsoon (May-October) and dry (November-April) seasons.

°P values shown are adjusted for false discovery rate using the Benjamini–Hochberg procedure.



**Figure 2.** Impact of interventions on quantity (log<sub>10</sub> copies/gram stool) of individual pathogens in 14-month-old rural Bangladeshi children. Point estimates and 95% confidence intervals determined with a parametric g-formula including both logistic and log-linear regression steps with generalized linear models adjusting for covariates associated with each pathogen outcome (likelihood ratio test P < .1 in bivariate analysis): household food insecurity, child age, child sex, child birth order, season of sample collection, time between defecation and sample placed on cold chain, mother's age, mother's height, mother's education level, number of children aged <18 years in the household, number of individuals living in the compound, distance in minutes to the primary water source, household floor and wall materials, and household assets. Pathogens significant after correction for false discovery rate are annotated: \*P < .05. Abbreviations: aEPEC, atypical enteropathogenic *Escherichia coli*; EIEC, enteroinvasive *Escherichia coli*; LT-ETEC, enterotoxigenic *Escherichia coli* with heat-labile toxin; N+WSH, nutrition plus water, sanitation, and handwashing; STEC, Shiga toxin—producing *Escherichia coli*; ST-ETEC, enterotoxigenic *Escherichia coli* with heat-stable toxin; tEPEC, typical enteropathogenic *Escherichia coli*; WSH, water, sanitation, and handwashing.

children age [44]. In contrast to our observations in 14-monthold children, those who received WSH and N+WSH interventions had a reduced prevalence of *Giardia* and hookworm at age 30 months [24, 25]. These former analyses had greater statistical power from a 50% larger sample size and geographic block–matched analysis. Thus, we cautiously interpret our parasite results: The young age at which we measured was not ideal to evaluate protozoa/helminths, and smaller sample size and covariate imbalances weakened our ability to detect differences.

Notably, our strongest findings were for enteric viruses, the pathogens least influenced by season and child age in this study. Estimated differences in virus quantities were small in comparison to fecal loads (difference of approximately 0.5 of 5–7 log<sub>10</sub> copies/g stool). However, viruses are excreted at orders

of magnitude higher per gram of feces and have low infectious doses (10–1000 viral particles) [45]. Thus, interventions that disrupt transmission of all types of enteropathogens might see greater effects in enteric viruses compared with bacteria/parasites with similar infectious doses and even greater effects than those with higher infectious doses. The prevalence of norovirus, sapovirus, and adenovirus 40/41 were each approximately 10% lower in WSH children than controls, and these pathogens account for 25% of all diarrhea cases for children <24 months old in Bangladesh [27, 28]. This suggests that WSH interventions can be clinically relevant for diarrheal disease. Our adjusted and unadjusted analyses for viruses led to the same scientific inference, and our data are consistent with the reductions in diarrhea reported in the main trial for the WSH intervention, which

gives us high confidence that WSH interventions reduced enteric viruses in 14-month-old children.

Rotavirus prevalence in our study was similar to previous cross-sectional studies where most children do not have diarrhea [46]. However, the prevalence in children with diarrhea was low for Bangladesh prior to implementation of rotavirus vaccination [27], likely because our sample collection period (February–November) did not include peak rotavirus season (December–January) [47].

There is no evidence for differences in efficacy of WSH interventions against bacteria compared with viruses. Handwashing with soap had similar removal efficacy for both, chlorine inactivation is comparable, and no difference in decay rates for bacteria and viruses have been observed on foods or household inanimate objects [48–50]. The evidence that WSH interventions impacted viruses is stronger than for bacteria given the influence of seasonality on some bacterial pathogens. Thus, we are less confident that our results indicate definitively that WSH and nutrition interventions did not impact bacterial pathogens.

WSH interventions were associated with lower enteric viruses in children 14 months old compared with controls, which is a modest impact on overall enteropathogens. These viruses account for a quarter of the diarrheal episodes for children <2 years old in Bangladesh, indicating a potentially clinically meaningful impact on childhood diarrhea. Our results suggest that neither low-cost household-level WSH nor nutrition interventions are sufficient to disrupt nonviral enteropathogen carriage at this critical early age. This is consistent with the lack of effect of WSH interventions on growth as the primary stuntingassociated pathogens are thought to be bacterial and protozoan. Future interventions should be designed with consideration for transmission pathways and environmental or zoonotic reservoirs of bacterial and parasitic enteropathogens to maximize efficacy for improving child health beyond diarrheal disease to include ponderal growth and cognitive development.

#### **Supplementary Data**

Supplementary materials are available at *The Journal of Infectious Diseases online*. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

## Notes

Author contributions. J. A. G. led the analysis and drafted the manuscript with input from coauthors. M. R., L. U., S. P. L., C. P. S., J. M. C., B. F. A., J. B.-C., and A. L. helped develop interventions and oversaw trial implementation. S. A., S. L. F., M. S. H., P. M., Z. H., M. Z. R., A. K. S., A. L., and J. A. G. were responsible for field sample and data collection/management. M. A. K., R. M., M. O. I., J. L., M. T., R. H., and J. A. G. contributed to pathogen data acquisition. J. A. G., E. T. R. M., B. F. A.,

A. L., J. B.-C., J. A. P.-M., S. P. H., J. L., and S. P. L. contributed to data analysis and interpretation. S. P. L., J. A. G., and E. R. H. conceived of the study.

Acknowledgments. We gratefully acknowledge the study participants and their families. We thank the intervention delivery, sample collection, and laboratory teams at icddr,b for their tremendous dedication in conducting high-quality fieldwork, including during periods of civil unrest.

**Data sharing.** The WASH Benefits trial protocol was previously published [30], and the statistical analysis plan, de-identified data, and analysis files for this study are published online at: https://osf.io/ky275/.

**Disclaimer.** The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Financial support. This work was supported by the Bill & Melinda Gates Foundation (award numbers OPPGD759 and OPP1161946 for project funds, and OPP1179069 to J. A. P.-M.); the National Science Foundation (graduate research fellowship to J. A. G.); Stanford University (Stanford Interdisciplinary Graduate Fellowship to J. A. G.); and the National Institute of Allergy and Infectious Diseases, National Institutes of Health (grant number K01AI130326 to E. T. R. M.).

**Potential conflicts of interest.** All authors: No reported conflicts of interest.

All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

#### References

- 1. Troeger C, Blacker BF, Khalil IA, et al. Estimates of the global, regional, and national morbidity, mortality, and aetiologies of diarrhoea in 195 countries: a systematic analysis for the Global Burden of Disease Study 2016. Lancet Infect Dis 2018; 18:1211–28.
- Black RE, Brown KH, Becker S, Alim AR, Huq I. Longitudinal studies of infectious diseases and physical growth of children in rural Bangladesh. II. Incidence of diarrhea and association with known pathogens. Am J Epidemiol 1982; 115:315–24.
- Berkman DS, Lescano AG, Gilman RH, Lopez SL, Black MM. Effects of stunting, diarrhoeal disease, and parasitic infection during infancy on cognition in late childhood: a follow-up study. Lancet 2002; 359:564–71.
- 4. Tarleton JL, Haque R, Mondal D, Shu J, Farr BM, Petri WA. Cognitive effects of diarrhea, malnutrition, and *Entamoeba histolytica* infection on school age children in Dhaka, Bangladesh. Am J Trop Med Hyg **2018**; 74:475–81.
- Rogawski ET, Liu J, Platts-Mills JA, et al; MAL-ED Network Investigators. Use of quantitative molecular diagnostic methods to investigate the effect of enteropathogen

- infections on linear growth in children in low-resource settings: longitudinal analysis of results from the MAL-ED cohort study. Lancet Glob Health **2018**; 6:e1319–28.
- Platts-Mills JA, Taniuchi M, Uddin MJ, et al. Association between enteropathogens and malnutrition in children aged 6-23 mo in Bangladesh: a case-control study. Am J Clin Nutr 2017; 105:1132–8.
- Lee G, Pan W, Peñataro Yori P, et al. Symptomatic and asymptomatic *Campylobacter* infections associated with reduced growth in Peruvian children. PLoS Negl Trop Dis 2013; 7:e2036.
- 8. Amour C, Gratz J, Mduma E, et al; Etiology, Risk Factors, and Interactions of Enteric Infections and Malnutrition and the Consequences for Child Health and Development Project (MAL-ED) Network Investigators. Epidemiology and impact of *Campylobacter* infection in children in 8 low-resource settings: results from the MAL-ED study. Clin Infect Dis **2016**; 63:1171–9.
- 9. Taniuchi M, Platts-Mills JA, Begum S, et al. Impact of enterovirus and other enteric pathogens on oral polio and rotavirus vaccine performance in Bangladeshi infants. Vaccine **2016**; 34:3068–75.
- Parker EP, Kampmann B, Kang G, Grassly NC. Influence of enteric infections on response to oral poliovirus vaccine: a systematic review and meta-analysis. J Infect Dis 2014; 210:853–64.
- 11. Luby SP, Agboatwalla M, Painter J, et al. Combining drinking water treatment and hand washing for diarrhoea prevention, a cluster randomised controlled trial. Trop Med Int Heal **2006**; 11:479–89.
- 12. Zambrano LD, Priest JW, Ivan E, et al. Use of serologic responses against enteropathogens to assess the impact of a point-of-use water filter: a randomized controlled trial in Western Province, Rwanda. Am J Trop Med Hyg 2017; 97:876–87.
- 13. Rogawski McQuade ET, Platts-Mills JA, Gratz J, et al. Impact of water quality, sanitation, handwashing, and nutritional interventions on enteric infections in rural Zimbabwe: the Sanitation Hygiene Infant Nutrition Efficacy (SHINE) trial. J Infect Dis 2019; 221:1379–86.
- 14. Patil SR, Arnold BF, Salvatore AL, et al. The effect of India's total sanitation campaign on defecation behaviors and child health in rural Madhya Pradesh: a cluster randomized controlled trial. PLoS Med 2015; 11:e1001709.
- 15. Stecher B, Hardt WD. Mechanisms controlling pathogen colonization of the gut. Curr Opin Microbiol **2011**; 14:82–91.
- 16. Endt K, Stecher B, Chaffron S, et al. The microbiota mediates pathogen clearance from the gut lumen after non-typhoidal *Salmonella* diarrhea. PLoS Pathog **2010**; 6:e1001097.
- 17. Paganini D, Zimmermann MB. The effects of iron fortification and supplementation on the gut microbiome and

- diarrhea in infants and children: a review. Am J Clin Nutr **2017**; 106:1688–93.
- 18. Dewey KG, Yang Z, Boy E. Systematic review and metaanalysis of home fortification of complementary foods. Matern Child Nutr **2009**; 5:283–321.
- Das JK, Salam RA, Kumar R, Bhutta ZA. Micronutrient fortification of food and its impact on woman and child health: a systematic review. Syst Rev 2013; 2:67.
- 20. Gera T, Pena-Rosas JP, Boy-Mena E, Sachdev HS. Lipid based nutrient supplements (LNS) for treatment of children (6 months to 59 months) with moderate acute malnutrition (MAM): a systematic review. PLoS One 2017; 12:e0182096.
- 21. Brown J, Cumming O. Stool-based pathogen detection offers advantages as an outcome measure for water, sanitation, and hygiene trials. Am J Trop Med Hyg 2020; 102:260–1.
- 22. Luby SP, Rahman M, Arnold BF, et al. Effects of water quality, sanitation, handwashing, and nutritional interventions on diarrhoea and child growth in rural Bangladesh: a cluster randomised controlled trial. Lancet Glob Heal 2018; 6:30490–4.
- 23. Pickering AJ, Null C, Winch PJ, et al. The WASH Benefits and SHINE trials: interpretation of WASH intervention effects on linear growth and diarrhoea. Lancet Glob Health **2019**; 7:e1139–46.
- 24. Ercumen A, Benjamin-Chung J, Arnold BF, et al. Effects of water, sanitation, handwashing and nutritional interventions on soil-transmitted helminth infections in young children: a cluster-randomized controlled trial in rural Bangladesh. PLoS Negl Trop Dis 2019; 13:e0007323.
- 25. Lin A, Ercumen A, Benjamin-Chung J, et al. Effects of water, sanitation, handwashing, and nutritional interventions on child enteric protozoan infections in rural Bangladesh: a cluster-randomized controlled trial. Clin Infect Dis 2018; 67:1515–22.
- 26. Pickering AJ, Njenga SM, Steinbaum L, et al. Effects of single and integrated water, sanitation, handwashing, and nutrition interventions on child soil-transmitted helminth and *Giardia* infections: a cluster-randomized controlled trial in rural Kenya. PLoS Med 2019; 16:e1002841.
- 27. Platts-Mills JA, Liu J, Rogawski ET, et al. Use of quantitative molecular diagnostic methods to assess the aetiology, burden, and clinical characteristics of diarrhoea in children in low-resource settings: a reanalysis of the MAL-ED cohort study. Lancet Glob Heal 2018; 6:e1309–18.
- 28. Liu J, Platts-Mills JA, Juma J, et al. Use of quantitative molecular diagnostic methods to identify causes of diarrhoea in children: a reanalysis of the GEMS case-control study. Lancet **2016**; 388:1291–301.
- 29. Tofail F, Fernald LC, Das KK, et al. Effects of water quality, sanitation, handwashing, and nutritional interventions on

- child development in rural Bangladesh: a cluster randomised controlled trial. Lancet Glob Heal **2018**; 4642:30490–4.
- 30. Arnold BF, Null C, Luby SP, et al. Cluster-randomised controlled trials of individual and combined water, sanitation, hygiene and nutritional interventions in rural Bangladesh and Kenya: the WASH Benefits study design and rationale. BMJ Open **2013**; 3:e003476.
- 31. Lin A, Ali S, Arnold BF, et al. Effects of water, sanitation, handwashing, and nutritional interventions on environmental enteric dysfunction in young children: a cluster-randomized, controlled trial in rural Bangladesh. Clin Infect Dis 2020; 70:738–47.
- 32. Stewart CP, Dewey KG, Lin A, et al. Effects of lipid-based nutrient supplements and infant and young child feeding counseling with or without improved water, sanitation, and hygiene (WASH) on anemia and micronutrient status: results from 2 cluster-randomized trials in Kenya and Bangladesh. Am J Clin Nutr 2018; 109:148–64.
- 33. Murray-Kolb LE, Acosta AM, De Burga RR, et al. Early childhood cognitive development is affected by interactions among illness, diet, enteropathogens and the home environment: findings from the MAL-ED birth cohort study. BMJ Glob Health 2018; 3:752.
- 34. Liu J, Gratz J, Amour C, et al. Optimization of quantitative PCR methods for enteropathogen detection. PLoS One **2016**; 11:1–11.
- 35. R Core Team. R: a language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing, 2018.
- 36. Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. J R Stat Soc Ser B **1995**; 57:289–300.
- 37. Gruber S, van der Laan MJ. tmle: an R package for targeted maximum likelihood estimation. J Stat Softw **2012**; 51:1–35.
- 38. Sultana F, Unicomb LE, Nizame FA, et al. Acceptability and feasibility of sharing a soapy water system for handwashing in a low-income urban community in Dhaka, Bangladesh: a qualitative study. Am J Trop Med Hyg **2018**; 99:502–12.
- Hussain F, Clasen T, Akter S, et al. Advantages and limitations for users of double pit pour-flush latrines: a qualitative study in rural Bangladesh. BMC Public Health 2017; 17:515.

- 40. Das JK, Salam RA, Hadi YB, et al. Preventive lipid-based nutrient supplements given with complementary foods to infants and young children 6 to 23 months of age for health, nutrition, and developmental outcomes. Cochrane Database Syst Rev 2019; 5:CD012611.
- 41. Humphrey JH, Mbuya MNN, Ntozini R, et al; Sanitation Hygiene Infant Nutrition Efficacy (SHINE) Trial Team. Independent and combined effects of improved water, sanitation, and hygiene, and improved complementary feeding, on child stunting and anaemia in rural Zimbabwe: a clusterrandomised trial. Lancet Glob Health 2019; 7:e132–47.
- 42. Wessling-Resnick M. Crossing the iron gate: why and how transferrin receptors mediate viral entry. Annu Rev Nutr **2018**; 38:431–58.
- 43. Jones MK, Watanabe M, Zhu S, et al. Enteric bacteria promote human and mouse norovirus infection of B cells. Science **2014**; 346:755–9.
- 44. Rogawski ET, Bartelt LA, Platts-Mills JA, et al; MAL-ED Network Investigators. Determinants and impact of *Giardia* infection in the first 2 years of life in the MAL-ED birth cohort. J Pediatric Infect Dis Soc **2017**; 6:153–60.
- 45. Julian TR. Environmental transmission of diarrheal pathogens in low and middle income countries. Environ Sci Process Impacts **2016**; 18:944–55.
- 46. Platts-Mills JA, Babji S, Bodhidatta L, et al. Pathogenspecific burdens of community diarrhoea in developing countries: a multisite birth cohort study (MAL-ED). Lancet Glob Heal **2015**; 3:564–75.
- 47. Das SK, Begum D, Ahmed S, et al. Geographical diversity in seasonality of major diarrhoeal pathogens in Bangladesh observed between 2010 and 2012. Epidemiol Infect **2014**; 142:2530–41.
- 48. Kramer A, Schwebke I, Kampf G. How long do nosocomial pathogens persist on inanimate surfaces? A systematic review. BMC Infect Dis **2006**; 6:130.
- Edmonds SL, McCormack RR, Zhou SS, Macinga DR, Fricker CM. Hand hygiene regimens for the reduction of risk in food service environments. J Food Prot 2012; 75:1303–9.
- 50. Centers for Disease Control and Prevention. Effect of chlorination on inactivating selected pathogens. https:// www.cdc.gov/safewater/effectiveness-on-pathogens.html. Accessed 18 August 2019.