

7: Graphics

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```
doFigs <- FALSE
```

```
fig7.1 <- function(){  
  ## ---- brain-body ----  
  plot(Brainwt ~ Bodywt, xlim=c(0, 300),  
        ylim=c(0,1500), data=primates,  
        xlab="Body weight (kg)",  
        ylab="Brain weight (g)")  
  # Specify xlim to allow room for the labels  
  with(primates,  
        text(Brainwt ~ Bodywt,  
              labels=rownames(primates), pos=4))  
  # pos: pos=1 (below), 2 (left), 3 (above)  
}
```

```
supp7.1 <- function(){  
  if(!require(RColorBrewer))stop("RColorBrewer must be installed")  
  ## ---- col-plots ----  
  opt <- options(width=75)  
  colpal <- rev(list(  
    "Default palette" = palette()[1:8], cm.colors = cm.colors(12),  
    terrain.colors = terrain.colors(12), heat.colors = heat.colors(12),  
    blueRamp = colorRampPalette(c(blues9, "white"))(12),  
    "Brewer-Set1" = brewer.pal(8, "Set1"),  
    "Brewer-Dark2" = brewer.pal(8, "Dark2")))  
  palnam <- names(colpal)  
  plot(1, 1, xlim=c(0.5,12.5), ylim=c(0,length(palnam)+0.5), type="n",  
        axes=FALSE, xlab="", ylab="")  
  for(i in 1:length(palnam)){  
    len <- length(colpal[[i]])  
    points(1:len, rep(i,len), pch=15, col=colpal[[i]], cex=5.5)  
    legend(1, i+0.025, palnam[i], adj=0, box.col="white", bg="white",  
          x.intersp=0, y.intersp=0, yjust=0)
```

```

}
options(opt)
}

```

```

fig7.2 <- function(){
## ---- alpha-ex ----
## Sample from the 15992 rows
dfsamp <- cps1[sample(nrow(cps1), 3000), ]
plot(re78 ~ re75, data=dfsamp, pch=20, cex=0.5,
     col="black")
mtext(side=3, line=0.5, "A: 100% opacity", adj=0)
plot(re78 ~ re75, data=dfsamp, pch=20, cex=0.5,
     col=adjustcolor("black", alpha=0.4))
mtext(side=3, line=0.5, "B: 40% opacity", adj=0)
blueRamp <- colorRampPalette(c("white", blues9))
with(dfsamp, smoothScatter(re75~re74,
                          colramp=blueRamp))
mtext(side=3, line=0.5, "C: Color density plot",
     adj=0)
}

```

```

fig7.3 <- function(){
## ---- aspect ----
plot((1:30)*0.92, sin((1:30)*0.92),
     xlab="", ylab="")
}

```

```

supp7.2 <- function(){
## ---- Animals-body-brain ----
## Supplementary figure 7.2
library(MASS)
oldpar <- par(pch=16, pty="s", mfrow=c(2,2))
with(Animals, {      # bracket several R statements
  plot(body, brain)
  plot(sqrt(body), sqrt(brain))
  plot(body^0.1, brain^0.1)
  plot(log(body), log(brain))
})                    # close both sets of brackets
par(oldpar)          # Restore former settings
}

```

```

fig7.4 <- function(){
  ## ---- poss-hist ----
  ftotlen <- subset(possum, sex=="f")[, "totlngth"]
  ## Left panel: breaks at 72.5, 77.5,..
  hist(ftotlen, breaks = 72.5 + (0:5)*5, freq=FALSE,
       xlab="Total length", ylim=c(0,0.11),
       main="A: Breaks at 72.5, 77.5,...")
  ## Now superimpose a density curve, as in Fig. 7.3
  lines(density(ftotlen))
  ##
  ## Panel B: breaks at 75, 80, ...
  hist(ftotlen, breaks = 75 + (0:5)*5, freq=FALSE,
       xlab="Total length", ylim=c(0,0.11),
       main="B: Breaks at 75, 80, ...")
}

```

```

supp7.3 <- function(){
  ## ---- fposs-density ----
  ## Supplementary figure 7.3
  with(subset(possum, sex=="f"),
       plot(density(totlngth), type="l"))
}

```

```

fig7.5 <- function(){
  ## ---- boxplot ----
  ## Code
  with(subset(possum, sex=="f"),
       {boxplot(totlngth, horizontal=TRUE)
        rug(totlngth)} )
}

```

```

fig7.6 <- function(){
  ## ---- possum-qqn ----
  ftotlen <- subset(possum, sex == "f")[, "totlngth"]
  ylabel <- expression(bold("Data"))
  xy <- qqnorm(ftotlen, xlab="", ylab=ylabel)
  usr <- par()$usr
  fillcol <- adjustcolor("gray60", alpha=0.2)
  rect(usr[1], usr[3],
      usr[2], usr[4], col=fillcol)
  for(i in 1:7) qqnorm(rnorm(43), xlab="",
                      ylab="Simulated",
                      main="Q-Q: Simulated")
}

```

```
# rnorm() generates random samples from a normal
# distribution with, by default, mean 0 and SD=1.
}
```

```
fig7.7 <- function(){
## ---- plot-expr ----
yl <- expression("Area = " * pi * r~~2)
plot(1:5, pi*(1:5)^2, xlab="Radius (r)", ylab=yl)
}
```

```
fig7.8 <- function(){
## ---- cline-gph ----
## On the command line: Create and print object
xyplot(Brainwt ~ Bodywt, data=primates)
}
```

```
fig7.9 <- function(){
## ---- rowSwim ----
xyplot(ht ~ wt | sport, groups=sex, data=ais,
       par.settings=simpleTheme(pch=c(4,1)),
       scales=list(tck=0.5),
       auto.key=list(space="right"),
       subset=sport%in%c("Row", "Swim"))
}
```

```
fig7.10 <- function(){
## ---- grog-simple ----
## Simple version of plot
grogplot <- xyplot(
  Beer+Spirit+Wine ~ Year | Country,
  data=grog, outer=FALSE,
  auto.key=list(space="right"))
## ---- grog-update ----
## Update trellis object, then print
ylab <- "Amount consumed (per person)"
parset <- simpleTheme(pch=c(1,3,4))
finalplot <- update(grogplot, ylim=c(0,5.5),
  xlab="", ylab=ylab,
  par.settings=parset)
print(finalplot)
}
```

```

fig7.11 <- function(){
  ## ---- strip-grob ----
  library(grid)
  plotnam <- "Stripplot of cuckoo data"
  stripplot(species ~ length, xlab="", data=cuckoos,
    legend=list(top=list(fun=textGrob,
      args=list(label=plotnam,
        x=0))))
}

```

```

fig7.12 <- function(){
  ## ---- jobs-basic ----
  ## 1. Create a basic version of the graphics object
  jobsB.xyplot <-
    xyplot(Ontario+Quebec+BC+Alberta+Prairies+Atlantic ~ Date,
      data=jobs, type="b", layout=c(3,2), outer=TRUE,
      ylab="Number of jobs",
      scales=list(y=list(relation="sliced", log=TRUE)))
  ## ---- jobs-enhance ----
  ## 2. Code for the enhancements to jobsB.xyplot
  ylabpos <- exp(pretty(log(unlist(jobs[, -7])), 100))
  ylabels <- paste0(round(ylabpos), "\n(", log(ylabpos), ")")
  ## Create a date object 'startofmonth'; use instead of 'Date'
  atdates <- seq(from=95, by=0.5, length=5)
  datelabs <- format(seq(from=as.Date("1Jan1995", format="%d%b%Y"),
    by="6 month", length=5), "%b%y")
  update(jobsB.xyplot, xlab="", between=list(x=0.5, y=0.5),
    scales=list(x=list(at=atdates, labels=datelabs),
      y=list(at=ylabpos, labels=ylabels), tck=0.6) )
}

```

```

fig7.13 <- function(){
  ## ---- strip-bw ----
  stripplot(species ~ length, data=cuckoos,
    xlab="Cuckoo egg length (mm)")
  bwplot(species ~ length, data=cuckoos,
    xlab="Cuckoo egg length (mm)")
}

```

```

fig7.14 <- function(){
  ## ---- lattice-density ----
  ## Code
  colset <- c("gray", "black")
}

```

```
densityplot(~ earconch | sex, groups=Pop,
            data=possum,
            par.settings=simpleTheme(col=colset),
            auto.key=list(space="right"))
}
```

```
fig7.15 <- function(){
  ## ---- gph-from ----
  gph <- xyplot(Brainwt ~ Bodywt, data=primates,
               xlim=c(0,300))
  ## ---- my-panel ----
  my.panel <- function(x,y){
    panel.xyplot(x,y)
    panel.text(x,y, labels=rownames(primates), pos=4)
  }
  update(gph, panel=my.panel)
}
```

```
fig7.16 <- function(){
  print(c("The Figure 7.16 screen display was obtained using the playwith GUI.",
        "See text for details."), quote=FALSE)
}
```

```
fig7.17 <- function(){
  ## ---- qplot-smooth ----
  ## Default loess smooth, with SE bands added.
  quickplot(Year, mdbRain, data=bomregions2015,
            geom=c("point", "smooth"), xlab="",
            ylab="Av. rainfall , M-D basin")
}
```

```
fig7.18 <- function(){
  ## ---- overlay-dens-simple ----
  ## Overlay with boxplots and density contours
  ggplot2::quickplot(wt, ht, data=DAAG::ais,
                    geom=c("boxplot", "point", "density2d"),
                    facets = . ~ sex)
}
```

```

supp7.4 <- function(){
  ## ---- twenty5080 ----
  ## Supplementary figure 4
  quickplot(Year, mdbRain, data=bomregions2015) +
    geom_quantile(formula = y ~ ns(x,5),
                  quantiles=c(0.2,0.5,0.8) )
}

```

```

fig7.19 <- function(){
  ## ---- aisRS ----
  ## Extract from ais data for rowers and swimmers
  aisRS <- subset(DAAG::ais, sport %in% c("Row","Swim"))
  aisRS$sport <- droplevels(aisRS$sport)
  ggplot2::ggplot(aisRS) +
    ggplot2::geom_point(aes(wt, ht,
                           color=sex,
                           shape=sport),
                       size=2)
}

```

```

figset7 <- function(){
  if(!require(DAAG, quietly=TRUE))stop('DAAG must be installed')
  if(!require(MASS, quietly=TRUE))stop('MASS must be installed')
  if(!require(latticeExtra, quietly=TRUE))stop('latticeExtra must be installed')
  if(!require(ggplot2, quietly=TRUE))stop('ggplot2 must be installed')
  if(!require(splines, quietly=TRUE))stop('splines must be installed')
  if(!require(quantreg, quietly=TRUE))stop('quantreg must be installed')
  if(!require(RColorBrewer, quietly=TRUE))stop('RColorBrewer must be installed')
}

```

```
figset7()
```

Attaching package: 'MASS'

The following object is masked from 'package:DAAG':
hills

Attaching package: 'ggplot2'

The following object is masked from 'package:latticeExtra':
layer

Attaching package: 'SparseM'

The following object is masked from 'package:base':
backsolve

```
fig7.1()
```

```
Error in fig7.1(): could not find function "fig7.1"
```

Figure 1: Plot of brain weight against body weight, for selected primates.

```
supp7.1()
```

```
Error in supp7.1(): could not find function "supp7.1"
```

Supplementary Figure 1: Color palettes.

```
fig7.2()
```

```
Error in fig7.2(): could not find function "fig7.2"
```

Figure 2: In Panel A, points are plotted with the 100% opacity, i.e., no transparency. In Panel B, `alpha=0.4`, i.e., 40% opacity. Panel C uses the function `smoothScatter()` to show a smoothed color density representation of the data.

```
par(fig=c(0,0.265, 0,1))
```

```
fig7.3()
```

```
Error in fig7.3(): could not find function "fig7.3"
```

```
mtext(side=3, line=0.5, "A: 1:1 aspect ratio", adj=0)
```

```
Error in mtext(side = 3, line = 0.5, "A: 1:1 aspect ratio", adj = 0):  
plot.new has not been called yet
```

```
par(fig=c(0.265, 1, 0,1), new=TRUE)
```

```
Warning in par(fig = c(0.265, 1, 0, 1), new = TRUE): calling par(new=TRUE)  
with no plot
```

```
fig7.3()
```

```
Error in fig7.3(): could not find function "fig7.3"
```

```
mtext(side=3, line=0.5, "B: 1:3.5 aspect ratio", adj=0)
```

```
Error in mtext(side = 3, line = 0.5, "B: 1:3.5 aspect ratio", adj = 0):  
plot.new has not been called yet
```

Figure 3: Both figures show the same data, but with very different aspect ratios.


```
supp7.2()
```

```
Error in supp7.2(): could not find function "supp7.2"
```

Supplementary Figure 2: The figure shows a 2 by 2 layout of plots.

```
fig7.4()
```

Figure 4: The two panels show the same data, but with a different choice of breakpoints.

```
supp7.3()
```

```
Error in supp7.3(): could not find function "supp7.3"
```

Supplementary Figure 3: Density plot for lengths of female possums.

```
fig7.5()
```

Figure 5: Distribution of lengths of female possums. The bars (together making up a 'rug') show actual data values.

```
fig7.6()
```

```
Error in fig7.6(): could not find function "fig7.6"
```

Figure 6: Normal probability plots. The top left panel shows the 43 lengths of female possums. Other panels are for independent normal random samples of size 43.

```
fig7.7()
```

Figure 7: A mathematical expression is included as part of the y -axis label..

```
fig7.20 <- function(){
## ---- rgl-demo ----
## The Rcmdr and rgl packages must be installed
if(!requireNamespace('rgl', quietly=TRUE))stop("Package 'rgl' must be available.")
if(!requireNamespace('car', quietly=TRUE))stop("Package 'car' must be available.")
if(!require(DAAG, quietly=TRUE))stop("Package 'DAAG' must be available.")
rgl::open3d()                # Precedes the call to par3d()
rgl::par3d(cex=0.75)         # Optional
                             # Other params: see help(par3d)
with(nihills, car::scatter3d(x=log(dist), y=log(climb),
                             z=log(time),
                             grid=FALSE,
                             surface=FALSE,
                             point.col="black",
                             axis.scales=FALSE))
}
```

```
supp7.5 <- function(){
if(!require(googleVis))stop("Package 'googleVis' must be available.")
if(!exists("WorldBank"))stop("Requires dataset 'Worldbank' to be available.")
M <- gvisMotionChart(WorldBank, idvar="country",
                     timevar="year",
                     xvar="life.expectancy",
                     yvar="fertility.rate",
                     colorvar="region", sizevar="population",
                     options=list(width=700, height=600))
## Now display the motion chart
```

```
fig7.8()
```

```
Error in fig7.8(): could not find function "fig7.8"
```

Figure 8: Use of lattice function `xyplot()` to give a graph.

```
fig7.9()
```

```
Error in fig7.9(): could not find function "fig7.9"
```

Figure 9: Height (`ht`) versus Weight (`wt`), for rowers (`Row`) and swimmers (`Swim`). Different plotting symbols are used to distinguish males from females.

```
plot(M)  
}
```

```
fig7.20()
```

```
supp7.4()
```

```
Error in supp7.4(): could not find function "supp7.4"
```

Supplementary Figure 4: Annual rainfall, from 1901 to 2012, for the Murray-Darling basin region of Australia. Curves that estimate the 20%, 50% and 80% quantiles have been fitted.

```
fig7.10()
```

```
Error in fig7.10(): could not find function "fig7.10"
```

Figure 10: Australian and New Zealand apparent per person annual consumption (in liters) of the pure alcohol content of liquor products, for 1998 to 2006.

```
fig7.11()
```

```
Error in fig7.11(): could not find function "fig7.11"
```

Figure 11: The argument `legend` has been used to add text, supplied as a 'grob'.

```
fig7.12()
```

```
Error in fig7.12(): could not find function "fig7.12"
```

Figure 12: Jobs growth in Canadian provinces, between January 1995 and December 1996.

Figure 13: A stripplot and a dotplot, side by side for comparison.

```
fig7.14()
```

```
Error in fig7.14(): could not find function "fig7.14"
```

Figure 14: Lattice style density plot comparing possum earconch measurements, separately for males and females, between Victorian and other populations. Observe that the scatter of data values is shown along the horizontal axis.

```
fig7.15()
```

```
Error in fig7.15(): could not find function "fig7.15"
```

Figure 15: Here, labels have been added to the points. This can be done by updating a graph that has the points, by use of a panel function that both plots points and adds labels, or by adding a new layer to a graph that shows the points.

```
fig7.17()
```

```
Error in fig7.17(): could not find function "fig7.17"
```

Figure 17: Annual rainfall, from 1901 to 2012, for the Murray-Darling basin region of Australia. The curve is fitted using the default loess smoother. The pointwise standard error bands assume that errors about the curve are independent; this is unlikely to be strictly true. To suppress these bands, specify `se=FALSE`.

```
fig7.18()
```

```
Error in fig7.18(): could not find function "fig7.18"
```

Figure 18: Height versus weight, by sex, for Australian athletes in the `ais` data set. Box-plots that show the distributions of heights, and two-dimensional density contours have been added.

```
fig7.19()
```

```
Error in fig7.19(): could not find function "fig7.19"
```

Figure 19: Use `color` for distinguishing `sexes`, `shapes` for `sports`.

Figure 20: Snapshot of 3-D dynamic display.