

Module 4.1 Exercise

Jelena H. Pantel

2023-12-05 12:18:29.636737

Exercise 1. Fitting data to a model: linear regression

A. Overview I indicated that even a linear regression is actually proposing a model - that a response variable y is driven by a driving variable x , and that this relationship is linear. For example, I might hypothesize that newborn baby weight (kg) is driven by the length of gestation (weeks). As a researcher, I collect data to test this hypothesis and see how well a linear model does describe the relationship, and I estimate the *parameters* in the linear model.

The data looks like this:

gestation	weight
30	1.6
32	1.7
34	2.5
36	2.8
38	3.2
40	3.5

The linear model looks like this:

$$y = m(x) + b$$

$$weight = m(gestationperiod) + b$$

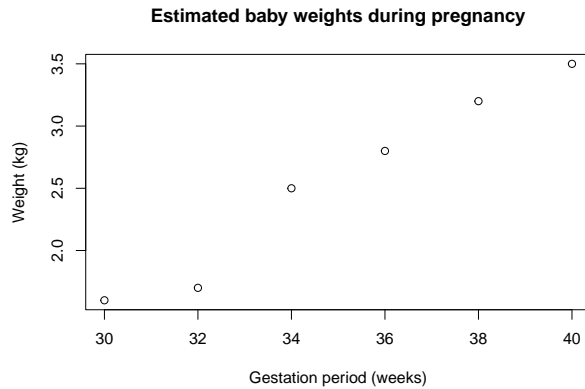
This model has variables y and x , and parameters m (the slope) and b (the y -intercept). Our goal is to **fit the data to the model**, and estimate the model parameters m and b that are most consistent with the data. You will learn how to do this in an Introduction to Statistics with R class - here I am using it to show you how to fit data to a model in the most basic case. This knowledge will come in handy later when we have more complex data and models. Please open a new R script and complete the following exercises (in some instances I provide the full code and you just need to type it into your R script. In other instances you will generate the code during class). Save your code periodically!

Step 1: Enter the data into R To enter the data into R, just paste this code in:

```
baby <- matrix(data = cbind(c(30, 32, 34, 36, 38, 40), c(1.6,
  1.7, 2.5, 2.8, 3.2, 3.5)), nrow = 6, ncol = 2, dimnames = list(NULL,
  c("gestation", "weight")))
baby <- as.data.frame(baby)
```

Step 2: Create a plot with gestation period on the x-axis and weight on the y-axis Please create a scatterplot with gestation period on the x-axis and weight on the y-axis. Add in informative x and y-axis labels. Mine is below.

```
plot(baby, main = "Estimated baby weights during pregnancy", xlab = "Gestation period (weeks)", ylab =
```



Step 3: Use least-squares regression to fit data to linear model, estimate the most likely parameter values given the data Paste the code I provide here, to fit the data to the linear model and estimate the values for b (the y-intercept) and m (the slope of the linear relationship).

```
# Fit linear model
model = lm(weight ~ gestation, data = baby)
# Examine model
summary(model)

##
## Call:
## lm(formula = weight ~ gestation, data = baby)
##
## Residuals:
##      1      2      3      4      5      6
## 0.07143 -0.23714  0.15429  0.04571  0.03714 -0.07143
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) -4.60000    0.64335   -7.15 0.002024 **
## gestation    0.20429    0.01829   11.17 0.000366 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.1531 on 4 degrees of freedom
## Multiple R-squared:  0.9689, Adjusted R-squared:  0.9611
## F-statistic: 124.7 on 1 and 4 DF, p-value: 0.0003661
```

Based on this, what are the most likely estimates for the slope and y-intercept?

```
##
```

```
## Call:
## lm(formula = weight ~ gestation, data = baby)
##
## Coefficients:
## (Intercept)    gestation
##      -4.6000      0.2043
```

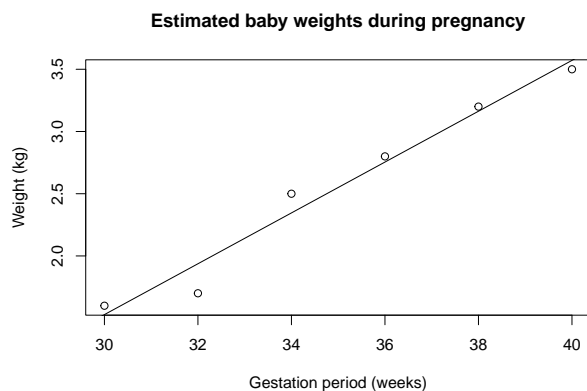
And what is the value of the residual sum of squares (RSS, sum of squared deviations from observed to model-predicted data)?

```
deviance(model)
```

```
## [1] 0.09371429
```

We can place the model-estimated line onto the scatterplot of the observed data to see the model-predicted values of weight for each gestation age *according to our model*. We use the command `abline(model)`:

```
plot(baby, main = "Estimated baby weights during pregnancy",
     xlab = "Gestation period (weeks)", ylab = "Weight (kg)")
abline(model)
```



Step 4: Repeat: use least-squares regression to fit data to linear model, estimate the most likely parameter values given the data To make sure this is clear, let's look at the sum of squared residuals for some data that is poor fit to a linear model. Enter the following data into R, and run a linear regression using the `lm` command, then calculate the *residual sum of squares (RSS)*:

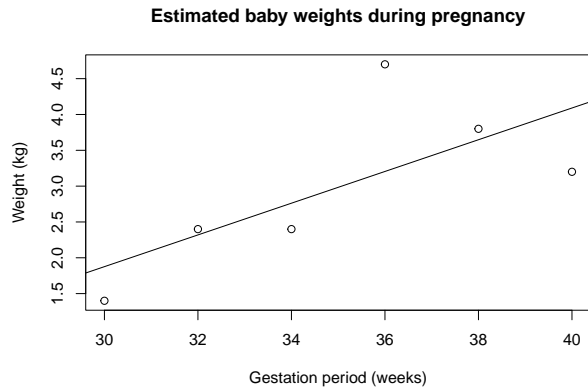
```
baby2 <- matrix(data=cbind(c(30,32,34,36,38,40),c(1.4,2.4,2.4,4.7,3.8,3.2)),nrow=6,ncol=2,dimnames=list(
baby2 <- as.data.frame(baby2)
model2 = lm(weight ~ gestation,data=baby2)
```

```
knitr::kable(baby2,format="markdown",padding=-1L,align="c")
```

gestation	weight
30	1.4
32	2.4

gestation	weight
34	2.4
36	4.7
38	3.8
40	3.2

```
plot(baby2, main = "Estimated baby weights during pregnancy", xlab = "Gestation period (weeks)", ylab = "Weight (kg)", abline(model2))
```



I got a value of sum of squared residuals equal to 3.4161905, indicating that the best-fit line (obtained by minimizing the RSS) is not as good a fit to this data as the dataset above.

I hope it makes sense now how we fit data to our hypothesized model and estimate the parameters of the model most likely to have produced our observed data.

Exercise 2. Fitting data to a model: nonlinear least squares (nls)

A. Overview We can use the same perspective - fitting data to a model by minimizing the RSS - to study non-linear models. This presentation is very straight-forward, but I will be transparent that non-linear least squares isn't always easy (I chose data that was relatively easy to fit). You would take more coursework to really be ready to apply this to more nonlinear models (though it certainly is done, usually for single-species growth or feeding curves).

Here, we will look at how organisms absorb and utilize nutrients - a nutrient-response curve (perhaps useful for studying phytoplankton nutrient uptake). In this instance, we still have a response variable y and a causal variable x , but the relationship between x and y is nonlinear. It has been observed that many nutrient-response curves reach an asymptote, where organisms become saturated, and no longer increase their absorption no matter how many more nutrients are available. The shape of one kind of useful saturation curve is described by a *Michaelis-Menten* model.

The general form of this is:

$$Y = \frac{aX}{b + X}$$

In our phytoplankton example, let's say that Y is the cell length (in μm) and X is the phosphorus concentration (in $\mu g/l$). The model parameters are a , the upper asymptote (for

$$X \rightarrow \infty$$

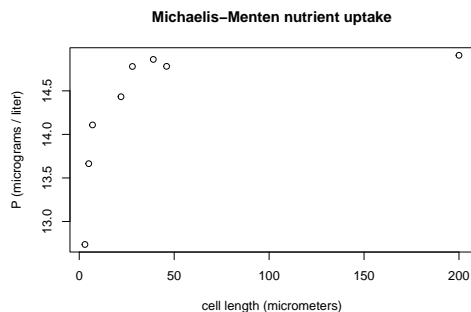
), and b , the the X value giving a response equal to $a/2$.

The data looks like this:

```
set.seed(1234)
X <- c(3, 5, 7, 22, 28, 39, 46, 200)
a <- 15; b <- 0.5
Ye <- as.numeric( SSmicmen(X, a, b) )
res <- rnorm(8, 0, 0.1)
Y <- Ye + res
knitr::kable(as.data.frame(cbind(Y,X)),format="markdown",padding=-2L)
```

Y	X
12.73644	3
13.66411	5
14.10844	7
14.43210	22
14.77975	28
14.86073	39
14.78124	46
14.90793	200

```
plot(X,Y, main = "Michaelis-Menten nutrient uptake", xlab = "cell length (micrometers)", ylab = "P (mic
```



Step 2: Use nonlinear least-squares to fit data to model, estimate the most likely parameter values given the data Non-linear least squares is included in base R and can be fit to data using the `nls` command. Use `?nls` to check the syntax needed to get this command to work: You'll need to specify the model formula, the data, and often starting values - these are beginning options for estimates for the nls estimator. `nls` has a difficult job, which is guessing the combination of parameters that are the best fit for the data - give it a hand by making reasonable suggestions for beginning values of model parameters that need to be estimated.

Note that `r` is also pre-supplied with *some* model formulas, including a Michaelis-Menten curve, that pre-chooses starting values for you. If you're lucky enough to be fitting a model already included in R's formulas, the job will be easier. You can find the self-starting models included with R by checking `?selfStart`, these include: `SSasymp`, `SSasympOff`, `SSasympOrig`, `SSbiexp`, `SSfol`, `SSfpl`, `SSgompertz`, `SSlogis`, `SSmicmen`, `SSweibull`.

To fit the data to the Michaelis-Menten model using the self-start formula `SSmicmen`:

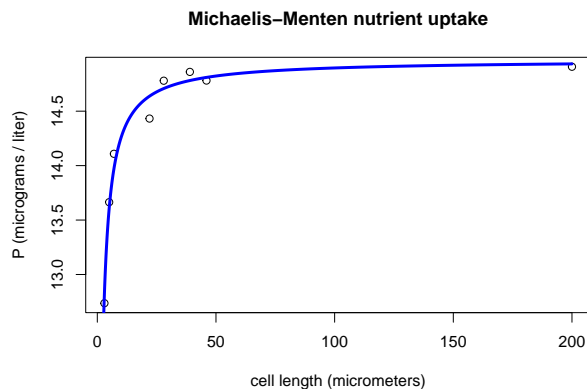
```
model.nls <- nls(Y ~ SSmicmen(X, a, b))
summary(model.nls)
```

```
##
## Formula: Y ~ SSmicmen(X, a, b)
##
## Parameters:
##   Estimate Std. Error t value Pr(>|t|)
## a 14.97166    0.06057  247.17 2.96e-13 ***
## b  0.50207    0.03345   15.01 5.51e-06 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.1196 on 6 degrees of freedom
##
## Number of iterations to convergence: 0
## Achieved convergence tolerance: 3.056e-06
```

Question: What are the most likely estimates for the parameters in the model, a and b

Let's plot the data and fitted curve:

```
newX <- as.data.frame(seq(0, 200, by = 0.1))
colnames(newX) <- "X"
pred <- predict(model.nls, newX)
plot(X, Y, main = "Michaelis-Menten nutrient uptake", xlab = "cell length (micrometers)",
     ylab = "P (micrograms / liter)")
lines(newX$X, pred, lwd = 3, col = "blue")
```



Step 3: Use nls to analyze data for 2 competing *Daphnia* over 9 weeks in a mesocosm I set up an experiment in 2020, where I placed *Daphnia pulex* and *Daphnia magna* in a 300-liter container, added water, nutrients, and phytoplankton. I let the *Daphnia* grow for 8 weeks, then fit the data to a Lotka-Volterra competition model, to estimate growth rates and interaction coefficients. I used nls for this, so let's take a look at the data and the fit of that curve.

For this, I will use a modified, discrete-time version of the Lotka-Volterra competition equation:

$$\begin{aligned} n_{1,t+1} &= n_{1,t} + n_{1,t}(L_1 e^{(-A_1 n_{1,t} - B_1 n_{2,t})}) \\ n_{2,t+1} &= n_{2,t} + n_{2,t}(L_2 e^{(-A_2 n_{2,t} - B_2 n_{1,t})}) \end{aligned}$$

Here, the model parameters are:

- L_1 and L_2 - the population growth rate
- A_1 and A_2 - the rate of intraspecific competition (competition with individuals of the same species)
- B_1 and B_2 - the rate of interspecific competition (competition with individuals of the other species)

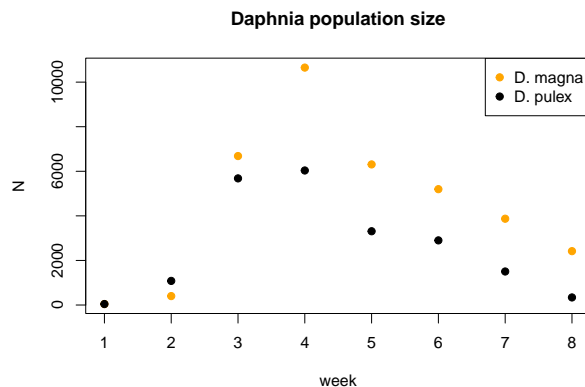
Please make a scatterplot of the data for the 2 species, with time (week) on the x-axis and points for the two species on the y-axis

I include my code here in case you just want to use mine:

```
dp <- matrix(data = cbind(1:8, c(44, 1082, 5681, 6036, 3310,
  2900, 1504, 340), c(44, 401, 6684, 10652, 6308, 5198, 3872,
  2417)), nrow = 8, ncol = 3, dimnames = list(NULL, c("week",
  "dp", "dm")))
dp <- as.data.frame(dp)
knitr::kable(dp, format = "markdown", padding = -1L, align = "c")
```

week	dp	dm
1	44	44
2	1082	401
3	5681	6684
4	6036	10652
5	3310	6308
6	2900	5198
7	1504	3872
8	340	2417

```
plot(dp$week, dp$dm, pch = 19, col = "orange", main = "Daphnia population size",
  xlab = "week", ylab = "N")
points(dp$week, dp$dp, pch = 19, col = "black")
legend("topright", legend = c("D. magna", "D. pulex"), pch = 19,
  col = c("orange", "black"))
```



Here is code to get the nls model to fit, and estimate parameters in the Lotka-Volterra competition equation.

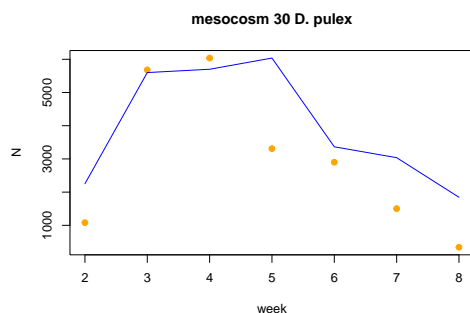
```
sub <- read.csv("https://raw.githubusercontent.com/jhpantel/ude-ecomod/main/data/sub.csv",
  header = T)
dp_mod <- nls(dp_Nt1 ~ dp_Nt + dp_Nt * ((L * exp(-A * dp_Nt -
  B * dm_Nt)))^days), data = sub, start = c(L = 1.5, A = 1e-04,
  B = 1e-04), na.action = na.omit)
```

The code seen here estimates the value of L_1 , A_1 , and B_1 - all parameters for the D. pulex growth equation. **What are the model-estimated values for those parameters, given the data?** Hint: use `summary(dp_mod)`

```
summary(dp_mod)
```

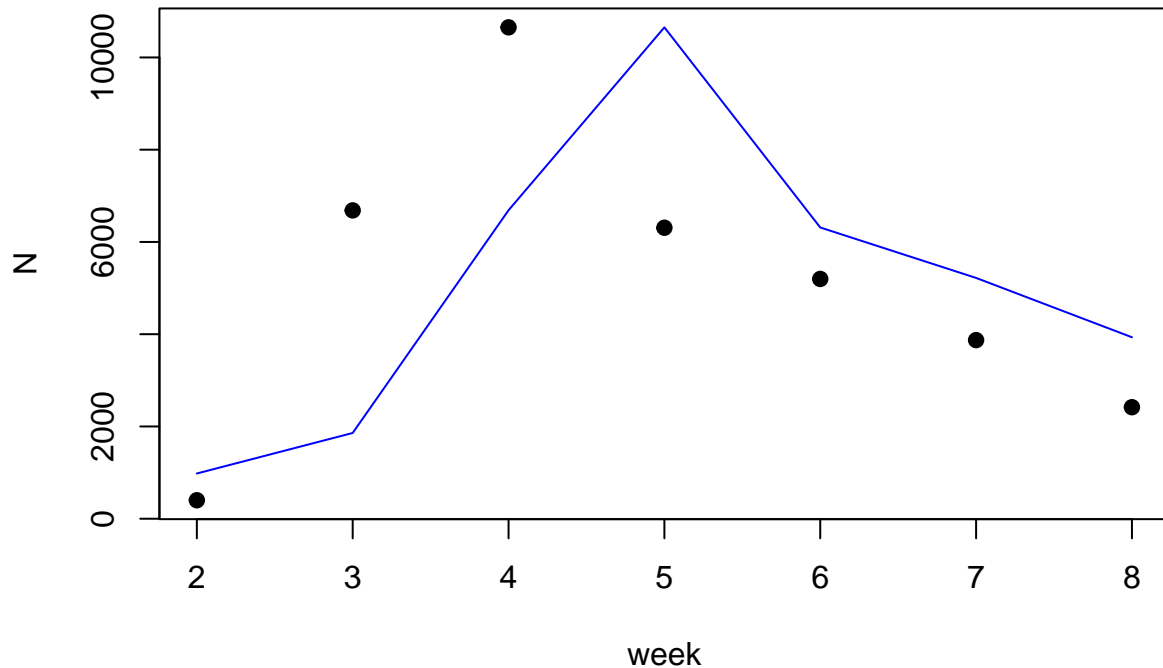
```
##
## Formula: dp_Nt1 ~ dp_Nt + dp_Nt * ((L * exp(-A * dp_Nt - B * dm_Nt)))^days)
##
## Parameters:
##      Estimate Std. Error t value Pr(>|t|)
## L 1.397e+00  5.095e-02  27.422  <2e-16 ***
## A 8.251e-05  4.250e-05   1.942   0.061 .
## B 1.022e-04  8.364e-05   1.222   0.231
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 2428 on 32 degrees of freedom
##
## Number of iterations to convergence: 13
## Achieved convergence tolerance: 8.292e-06
```

```
sub$yhat <- predict(dp_mod)
plot(sub$week[sub$meso == 30], sub$dp_Nt1[sub$meso == 30], main = "mesocosm 30 D. pulex",
  xlab = "week", ylab = "N", pch = 19, col = "orange")
lines(sub$week[sub$meso == 30], sub$yhat[sub$meso == 30], col = "blue")
```



Finally, please edit the code to fit the model for the D. magna data. Give the most likely parameter estimates, and a plot of the data and model-fit line.

mesocosm 30 D. magna



```
##
## Formula: dm_Nt1 ~ dm_Nt + dm_Nt * ((L * exp(-A * dm_Nt - B * dp_Nt))^days)
##
## Parameters:
##      Estimate Std. Error t value Pr(>|t|)
## L  1.302e+00  7.753e-02  16.790  <2e-16 ***
## A  2.022e-04  8.778e-05   2.304   0.0279 *
## B -1.810e-06  1.969e-05  -0.092   0.9273
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 1839 on 32 degrees of freedom
##
## Number of iterations to convergence: 8
## Achieved convergence tolerance: 7.778e-06
```

Exercise 3. Fitting data to Lotka-Volterra competition model using `gauseR`

A. Overview We will use data from Gause's experiments with *Paramecium*, tracking competitive interactions between *P. aurelia* and *P. caudatum*.

```
# load competition data
data("gause_1934_science_f02_03")

# subset out data from species grown in mixture
```

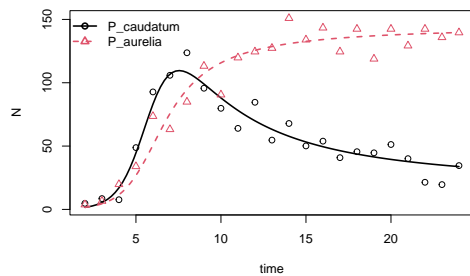
```

mixturedat <- gauge_1934_science_f02_03[gauge_1934_science_f02_03$Treatment ==
  "Mixture", ]

# extract time and species data
time <- mixturedat$Day
species <- data.frame(mixturedat$Volume_Species1, mixturedat$Volume_Species2)
colnames(species) <- c("P_caudatum", "P_aurelia")

# run wrapper
gause_out <- gaugeR::gause_wrapper(time = time, species = species)

```



What were the parameter estimates?

```
gause_out$parameter_intervals
```

##		lower_sd	mu	upper_sd
##	P_caudatum0	0.01033120	1.595268713	2.463299e+02
##	P_aurelia0	0.08812049	1.631618399	3.021066e+01
##	r1	0.34795149	1.259232133	4.557145e+00
##	r2	0.47571115	1.026156214	2.213521e+00
##	a11	-0.02061162	-0.005157869	-1.290709e-03
##	a12	-0.02361279	-0.008000167	-2.710508e-03
##	a21	-0.01100428	-0.001974871	-3.544183e-04
##	a22	-0.01459709	-0.006851130	-3.215572e-03