

bggn213_HW_Class6

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Can you improve this analysis code?

```
library(bio3d)
s1 <- read.pdb("4AKE") # kinase with drug

## Note: Accessing on-line PDB file

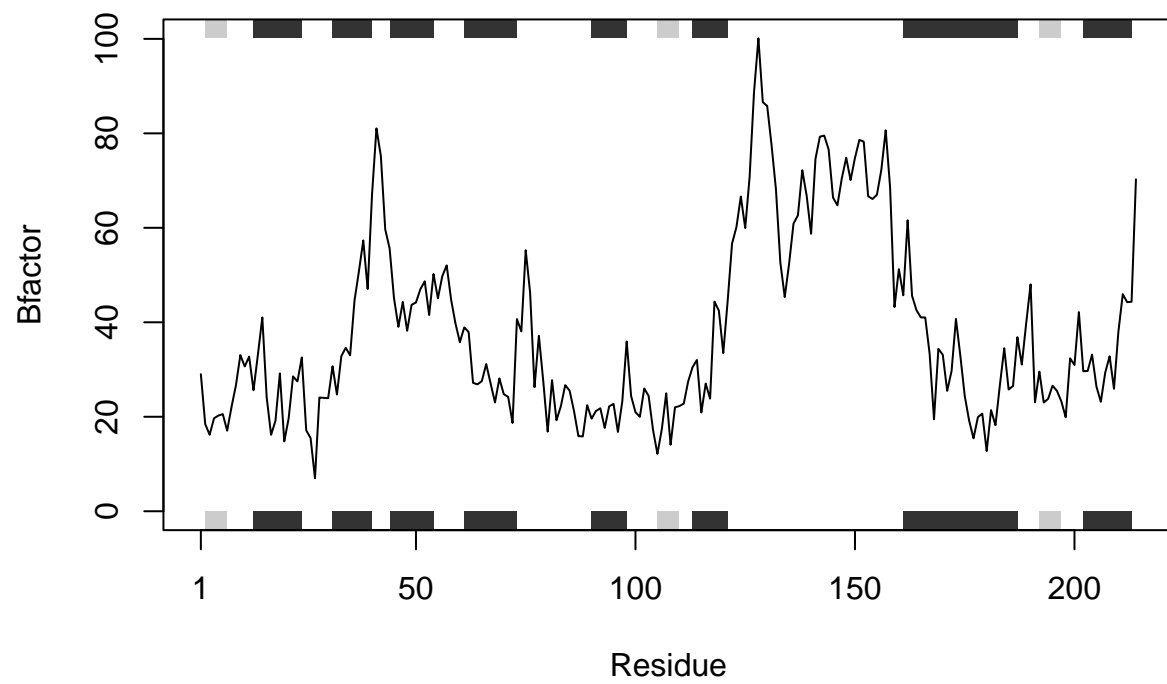
s2 <- read.pdb("1AKE") # kinase no drug

## Note: Accessing on-line PDB file
## PDB has ALT records, taking A only, rm.alt=TRUE

s3 <- read.pdb("1E4Y") # kinase with drug

## Note: Accessing on-line PDB file

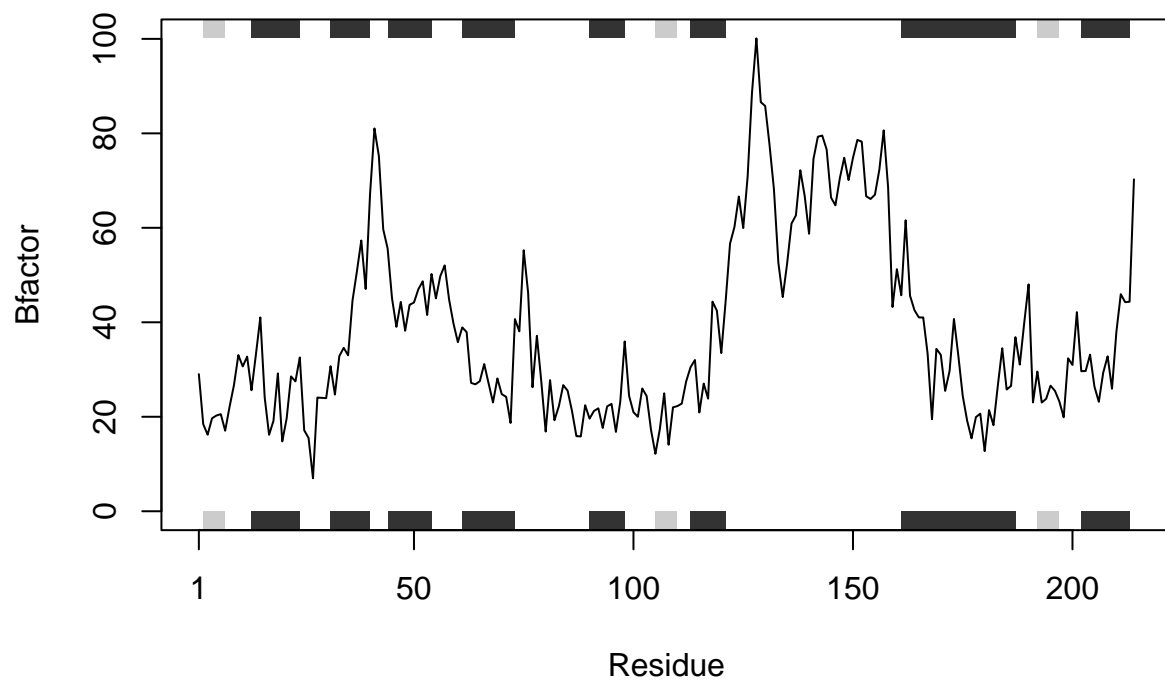
s1.chainA <- trim.pdb(s1, chain="A", elety="CA")
s2.chainA <- trim.pdb(s2, chain="A", elety="CA")
s3.chainA <- trim.pdb(s1, chain="A", elety="CA")
s1.b <- s1.chainA$atom$b
s2.b <- s2.chainA$atom$b
s3.b <- s3.chainA$atom$b
plotb3(s1.b, sse=s1.chainA, typ="l", ylab="Bfactor")
```



```
plotb3(s2.b, sse=s2.chainA, typ="l", ylab="Bfactor")
```



```
plotb3(s3.b, sse=s3.chainA, typ="l", ylab="Bfactor")
```



```
hc <- hclust( dist( rbind(s1.b, s2.b, s3.b) ) )
plot(hc)
```

Cluster Dendrogram



```
dist(rbind(s1.b, s2.b, s3.b))  
hclust (*, "complete")
```

A better plot to compare across the different proteins

`plotb3` function is useful for plotting per-residue numeric vectors for a given protein structure along with a schematic representation of major secondary structure elements.

`trim.pdb ()` can be used to produce a new smaller PDB object, containing a subset of atoms, from a given larger PDB object.

`atom` is a character matrix containing all atomic coordinate `ATOM` data, with a row per `ATOM` and a column per record type. See below for details of the record type naming convention (useful for accessing columns).

The B-factor describes the displacement of the atomic positions from an average (mean) value (mean-square displacement), which reflects the mobility or flexibility of various parts of the molecule.

- Input explanation
 - “`pdbname`” is a single element character vector containing the name of the PDB file to be read, or the four letter PDB identifier for online file access.
 - “`chain`” is a character vector of chain identifiers.
 - “`elety`” is a character vector of atom names.

```

library(bio3d)

Better_plotb3<- function(pdbname, chain, elety) {
  # assign different colors to each vector
  plot_colors <- c("red", "gray", "blue")

  # to read multiple PDB files and extract the "atom" & "b" information
  for (i in 1:length(pdbname)) {
    s1 <- read.pdb(pdbname[i])
    s1.chain_df <- trim.pdb(s1, chain = chain, elety = elety)
    s1.b<- s1.chain_df$atom$b

    # plot s1.b
    if (i == 1) {
      plotb3(s1.b, sse = s1.chain_df, typ = "l", ylab="B-factor",
             col = plot_colors[i])

      # adds additional lines to s1.b
    } else {
      lines(s1.b, col = plot_colors[i])
    }
  }

  # modify the legend
  legend("topright", title = "PDB Name", pdbname, fill = plot_colors,
        horiz=TRUE, cex = 0.5, inset = c(0.03, 0.06))
}

```

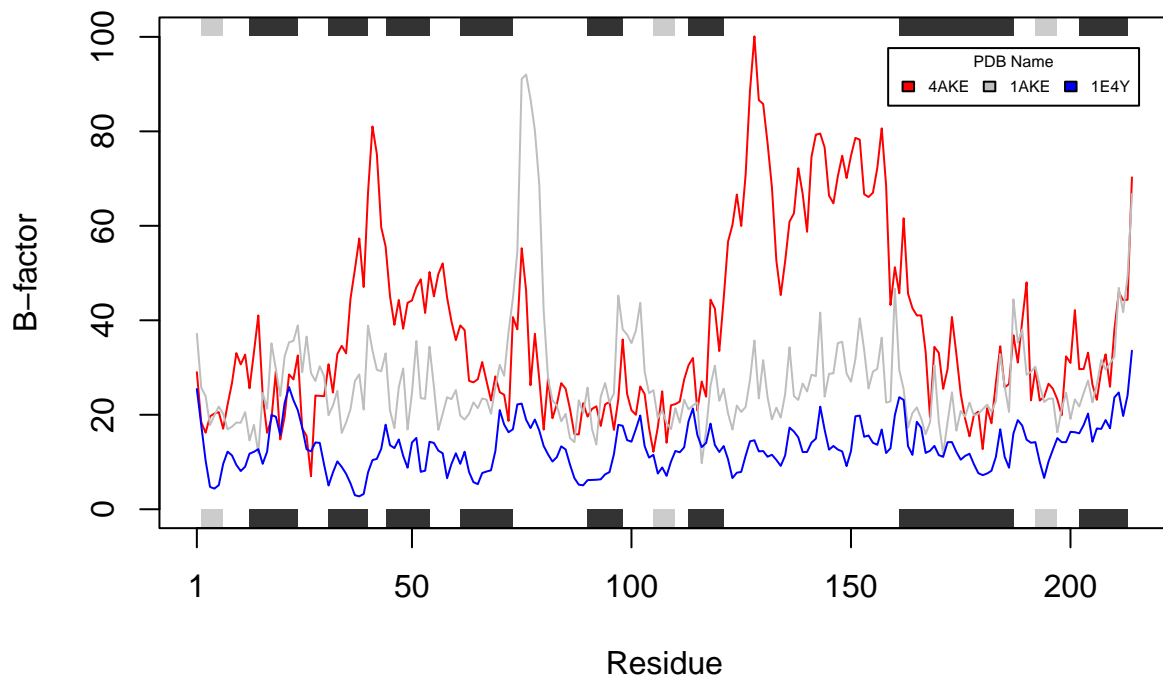
```

pdbname <- c("4AKE", "1AKE", "1E4Y")
chains <- "A"
elety <- "CA"

Better_plotb3(pdbname, chains, elety)

```

Note: Accessing on-line PDB file



```
## Note: Accessing on-line PDB file
## PDB has ALT records, taking A only, rm.alt=TRUE
## Note: Accessing on-line PDB file
```

To remove the secondary structure from the plot

```
library(bio3d)

Better_plotb3_2<- function(pdbname, chain, elety) {
  # assign different colors to each vector
  plot_colors <- c("red", "gray", "blue")

  # to read multiple PDB files and extract the "atom" & "b" information
  for (i in 1:length(pdbname)) {
    s1 <- read.pdb(pdbname[i])
    s1.chain_df <- trim.pdb(s1, chain = chain, elety = elety)
    s1.b<- s1.chain_df$atom$b

    # plot s1.b
    if (i == 1) {
      plotb3(s1.b, typ = "l", ylab="B-factor", col = plot_colors[i])
    }
  }
}
```

```

# adds additional lines to s1.b
} else {
  lines(s1.b, col = plot_colors[i])
}
}

# modify the legend
legend("topright", title = "PDB Name", pdbname, fill = plot_colors,
      horiz=TRUE, cex = 0.5, inset = c(0.03, 0.06))
}

```

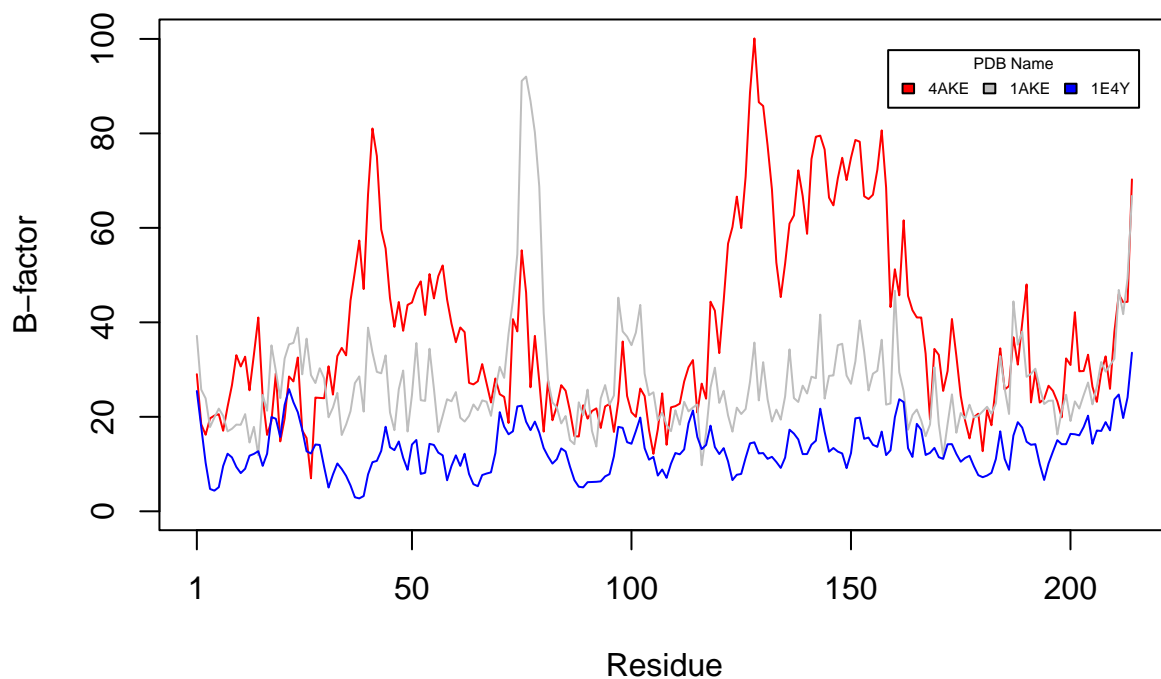
```

pdbname <- c("4AKE", "1AKE", "1E4Y")
chains <- "A"
elety <- "CA"

Better_plotb3_2(pdbname, chains, elety)

```

Note: Accessing on-line PDB file



```

## Note: Accessing on-line PDB file
## PDB has ALT records, taking A only, rm.alt=TRUE
## Note: Accessing on-line PDB file

```