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# Abstract

# Introduction

Temperature is an important abiotic variable with increasing relevance as warming global temperatures alter the diversity, structure, and functioning of Earth’s ecosystems (Walther et al. 2002). Temperature’s effect on ecosystems manifest through complex direct and indirect pathways such as shifting species ranges [][] and subsequent changes to local and regional communities [], species adaptations (Gibert and DeLong 2017), and through effects on individual metabolic rates (Gillooly et al. 2001, Brown et al. 2004). While a growing body of theoretical and empirical study has enhanced our knowledge of temperature-mediated changes to ecosystems (O’Connor et al. 2009), general patterns are uncertain and empirical studies often idiosyncratic (Nelson et al. 2017a, Zhang et al. 2017), especially at higher levels of organizations such as communities and food webs (Walther et al. 2002, Woodward et al. 2010). Yet, identifying general effects of temperature is a necessary step towards understanding how future warming may alter the functioning of ecosystems and the services they provide.

Temperature has the potential to alter the provision and maintenance of ecosystem services by modifying the interactions among species (Woodward et al. 2010, Brose et al. 2012) that underpin ecosystem functions (de Ruiter et al. 1995, Thompson et al. 2012). The signature of temperature on the stability and dynamics of food webs is present across global climate gradients (Baiser et al. 2019) where its direct and indirect effects alter the magnitudes and relative distributions among species and trophic levels (May 1972, McCann et al. 1998, Barnes et al. 2018). Direct changes through species richness effects (Gibert 2019). Indirect through the distribution of consumer and prey species and it effects on the acquisition and allocation of resources among species (Zhang et al. 2017) –macroecology of food web dynamics and stability are controlled by the .

Temperature can directly alter the distribution of species interactions by asymmetric responses in organismal traits among species [e.g., attack rate, handling time, growth rates, etc.; Dell et al. (2014)].

Metabolic rates

acquisition of resources by consumer Generally, temperature is predicted to have disproportionately higher effects on consumers relative to resources (Allen et al. 2005, Vasseur and McCann 2005, **oconnor2011?**)

Here, we measured the patterning and distribution of organic matter fluxes within invertebrate food webs across a natural stream temperature gradient (~5 - 28C). Previous research in these streams has shown a strong positive effect of temperature on primary production both among streams (Padfield et al. 2017, **demars2011?**) and within streams seasonally (O’Gorman et al. 2012, Hood et al. 2018). Consumers rely largely on autochthonous resource (O’Gorman et al. 2012, **nelson2019?**) and therefore the dynamics of primary production have a strong control on consumer energy demand (Junker et al. 2020), as such, we predicted total annual OM fluxes to consumers to scale with among stream patterns in primary production and consumer energy demand, and therefore, increase with temperature across streams. We, further expected that increasing temperature would reduce consumer species (**ogorman2019?**), thereby altering how OM fluxes are distributed within and across communities. The distribution of OM fluxes to consumers will shift towards relatively faster, higher turnover consumer at higher temperatures, ‘speeding up’ consumer dynamics both through direct effects on consumer turnover and through a decrease in mean body size. Seasonally, warmer streams will lead to earlier food web fluxes in warmer compared to colder streams.

# Methods

We studied six streams within the Hengill geothermal field of southwestern Iceland (64 03’N 021 18’W) that varied in mean annual temperature. Hengill is characterized by indirect geothermal heating of groundwater (Arnason et al. 1969), leading to a natural variability in water temperatures (4.5–54.0 C), but similar solute chemistries (Friberg et al. 2009). These conditions create a “natural laboratory” for isolating the effects of temperature on ecosystem processes (O’Gorman et al. 2014, Nelson et al. 2017b). We selected streams to maximize the temperature range, while minimizing differences in the structural aspects of primary producers. In each stream, we measured temperature and water depth every 15 min from July 2010 through August 2012 (U20-001-01 water-level logger, Onset Computer Corp. Pocasset, MA, USA). Light availability in the watershed was measured every 15 min from atmospheric stations (HOBO pendant temperature/light UA-002-64, Onset Computer Corp. Pocasset, MA, USA).

## Invertebrate sampling

We sampled macroinvertebrate communities approximately monthly from July 2011 to August 2012 in four streams and from October 2010 to October 2011 in two streams used in a previous study (*n* = 6 streams). The two streams were part of an experiment beginning October 2011, therefore overlapping years were not used to exclude the impact of experimental manipulation (see Nelson et al. 2017a, 2017b). Inter-annual comparisons of primary and secondary production in previous studies showed minimal differences among years in unmanipulated streams, suggesting that combining data from different years would not significantly bias our results (Nelson et al. 2017a, Hood et al. 2018). We collected fiver Surber samples (0.023 m2, 250 m mesh) from randomly selected locations within each stream. Within the sampler, inorganic substrates were disturbed to ~10 cm depth and invertebrates and organic matter were removed from stones with a brush. Samples were then preserved with 5% formaldehyde until laboratory analysis. In the laboratory, we split samples into coarse (>1 mm) and fine (<1 mm but >250 m) fractions using nested sieves and then removed invertebrates form each fraction under a dissecting microscope (10–15 x magnification). For particularly large samples, fine fractions were subsampled (1/2–1/16th) using a modified Folsom plankton splitter prior to removal of invertebrates. Subsamples were scaled to the rest of the sample assuming similar abundance and body size distributions. Macroinvertebrates were identified to the lowest practical taxonomic level (usually genus) with taxonomic keys (Peterson 1977, Merritt et al. 2008, Andersen et al. 2013). Taxon-specific abundance and biomass were scaled to a per meter basis by dividing by the Surber sampler area.

## Secondary Production

Daily secondary production of invertebrate taxa was calculated using the instantaneous growth rate method [IGR; Benke and Huryn (2017)]. Growth rates were determined using taxon appropriate approaches described in (Junker et al. 2020). Briefly, growth rates of common taxa (e.g., Chironomidae spp., Radix balthica, etc.) were determined using *in situ* chambers (Huryn and Wallace 1986). Multiple individuals (*n* = 5–15) within small size categories (~1 mm length range) were photographed next to a field micrometer, placed into the stream within pre-conditioned chambers for 7–15 days, after which they were again photographed. Individual lengths were measured from field pictures using image analysis software (Schindelin et al. 2012), and body lengths were converted to mass (mg ash-free dry mass [AFDM]) using published length-mass regressions (Benke et al. 1999, O’Gorman et al. 2012, Hannesdóttir et al. 2013). Growth rates (*g*, d-1) were calculated by the changes in mean body size (*W*) over a given time interval (*t*) with the following equation:

Variability in growth rates was estimated by bootstrapping through repeated resampling of individual lengths with replacement (*n* = 500). For taxa which exhibit synchronous growth and development (e.g., Simuliidae spp., some Chironomidae spp., etc.), we examined temporal changes in length-frequency distributions and calculated growth rates and uncertainty using a bootstrap technique similar to that described in Benke and Huryn (2017). Individual Lengths were converted to mg AFDM using published length-mass regression cited above and size-frequency histograms were visually inspected for directional changes in body size through time. For each date, size-frequency distributions were resampled with replacement (*n* = 500) and growth rates estimated from equation 1. We prevented the calculation of negative growth rates by requiring > . To estimate growth rates of taxa for which growth could not be estimated empirically, we developed stream-specific growth rate models by constructing multivariate linear regressions of empirical growth data against body size and temperature within each stream. To estimate uncertainty in production of each taxon, we used a bootstrapping technique that resampled measured growth rates, in addition to abundance and size distributions from individual samples. For each iteration (*n* = 1000), size-specific growth rates were multiplied by mean interval biomass for each size class and the number of days between sample dates to estimate size class-specific production. For each interval, size classes were summed for each taxon to calculate total population-level production. Intervals were summed to estimate annual secondary production.

## **Diet analysis**

**Come back and confirm this section** Macroinvertebrate diets were quantified for dominant taxa in each stream. We focused on numerically abundant taxa and/or taxa with relatively high annual production. A minimum of five individuals were selected from samples, and, when possible included individuals of different size classes to account for ontogenetic shifts in diet. We included individuals from different seasons to capture concurrent ontogenetic and seasonal changes. For small-bodied taxa, we combined multiple individuals (*n* = 3–5) to ensure samples contained enough material for quantification. We used methods outlined in Rosi-Marshall (2016) to remove gut tracts and prepare gut content slides. Briefly, we removed the foregut from each individual or collection of individuals and sonicated gut contents in water for 30 seconds. Gut content slurries were filtered onto gridded nitrocellulose membrane filters (Metricel GN-6, 25 mm, 0.45 m pore size; Gelman Sciences, Ann Arbor, MI, USA), dried at 60 C for 15 min, placed on a microscope slide, cleared with Type B immersion old, and covered with a cover slip. We took 5–10 random photographs under 200–400x magnification, depending on the density of particles, using a digital camera mounted on a compound microscope. From these photographs we identified all particles within each field and measured the relative area of particles using image analysis software (Schindelin et al. 2012). We identified particles as diatoms, green and filamentous algae, cyanobacteria, amorphous detritus, vascular and non-vascular plants (bryophytes), and animal material. We calculated the proportion of each food category in the gut by dividing their summed area by the total area of all particles. Gut contents of many predators were empty or contained unidentifiable, macerated prey. For these taxa, we assumed 100% animal material.

To estimate variability in diet compositions and to impute missing values for non-dominant, yet present, taxa, we modeled the diet proportions within each stream using a hierarchical multivariate model (Fordyce et al. 2011). Here, the diet of a consumer population, *i*, is a multinomial vector,, of

where, , is a vector of consumer diet proportions, is a vector of the population’s diet proportions and is a concentration parameter of the Dirichlet process. We used uniform priors for and ,

where, is a vector of ones the same length of basal resource types and is the presumed maximum value the concentration value. **add in site-level model parameter and model fitting language**. Models were fit in Stan with the ‘brms’ package in R (Bürkner 2017). For non-dominant taxa, diet proportions were imputed from the hierarchical model by resampling from posterior distributions. Importantly, this process allowed to maintain the hierarchical structure of the data when imputing missing values.

## Organic Matter Consumption Estimates

Consumption fluxes (g m-2 t-1) were calculated using the trophic basis of production method (TBP; (Benke and Wallace 1980). Taxon-specific secondary production estimates were combined with diet proportions, diet-specific assimilation efficiencies, , and assumed net production efficiencies, , to estimate consumption of organic matter. For each food category, , diet proportions were multiplied by the gross growth efficiency () to calculate the relative production attributable to each food category. The relative production from each food type was then multiplied by the interval-level production and finally divided by to estimate consumption of organic matter from each food category by a consumer (Benke and Wallace 1980). Consumption was calculated for each taxon across sampling intervals (typically ~1 month). Total interval consumption was calculated by summing across all taxa, while annual consumption was calculated by summing across all taxa and inte(rvals. Variability in consumption estimates was estimated through a Monte Carlo approach, wherein bootstrapped vectors of secondary production for each taxon (see *Secondary production* methods above) were resampled and consumption estimated with the TBP method using simulated diet proportions (see *Diet analysis* above), diet-specific assimilation efficiencies, and net production efficiency. Variability in was incoporated by resampling values from beta distributions fit to median and 2.5% and 97.5% percentiles for each diet item: diatoms = 0.30 (95% percentile interval (PI): 0.24-0.36), filamentous and green algae = 0.30 (95% PI: 0.24-0.36), cyanobacteria = 0.10 (95% PI: 0.08-0.12), amorphous detritus = 0.10 (95% PI: 0.08-0.12), vascular and non-vascular plants (bryophytes) = 0.1 (95% PI:: 0.08-0.12), and animal material = 0.7 (95% PI: 0.56-0.84)(Welch 1968, Benke and Wallace 1980, 1997, Cross et al. 2007, 2011). Variability in was incorporated by resampling values from an assumed beta distribution with median = 0.45 (95% PI = 0.4-0.5). Beta distributions were fit using the ‘get.beta.par()’ function within the *rriskDistributions* package (Belgorodski et al. 2017).

## Quantifying the distribution of food web fluxes

### Evenness Among consumers

To visualize and quantify how evenly OM fluxes were distributed among consumers within a stream, we constructed Lorenz curves (Lorenz 1905) on ordered relative consumption fluxes, such that in a community with species and the relative consumption of species *i*, , is ordered . The Lorenz curve plots how a value, in this case OM flux, accumulates with increasing cumulative proportion of species. In a community with perfectly equal distribution of OM consumption among species, the Lorenz curve is simply a straight diagonal line. Deviation from perfect equality was calculated as the Gini coefficient (Gini 1921), normalized for differences in among streams, (Solomon 1975, Chao and Ricotta 2019):

### Distribution along species’ trait axes

We were interested in how OM fluxes were distributed (randomly vs. non-randomly) in relation to species traits (i.e., body size, ratio, population biomass) across the temperature gradient. We used a permutation analyses to test for non-random patterns in OM flux with species traits. First, we ordered species based on within-stream ranking of annual population traits (i.e, , ) and measured the asymmetry in the distribution of annual OM flux with species traits. To determine the probability of observing the empirical asymmetry we constructed a random distribution of asymmetry values. The number of unique orderings of species increases to computationally intractable numbers very quickly (e.g., ), therefore chose to permute estimated this by randomly ordering species 10,000,000 times and calculating the skew in the cumulative distribution of annual OM fluxes. We then use this random distribution to estimate the probability of observing

## Statistical Analyses

# Results

## -- Attaching packages --------------------------------------- tidyverse 1.3.0 --

## v ggplot2 3.3.3 v purrr 0.3.4  
## v tibble 3.0.6 v dplyr 1.0.3  
## v tidyr 1.1.2 v stringr 1.4.0  
## v readr 1.4.0 v forcats 0.5.0

## -- Conflicts ------------------------------------------ tidyverse\_conflicts() --  
## x dplyr::filter() masks stats::filter()  
## x dplyr::lag() masks stats::lag()

## Community OM fluxes

Annual community energy demand mirrored patterns of secondary production previously reported (Junker et al. 2020). Community energy demand varied ~ XXX-fold across streams () and was positively related to temperature. Total OM flux through the consumer community was strongly associated with community energy demand and varied from 3.91 [2.1 - 6.24] to 177.36[125.59 - 236.61] (mean 95% percentile interval [PI]).

Patterns of OM flux among streams were most closely tied to total energy demands as consumer diets exhibited high similarity among streams (Figure SX). Diets composition among all streams were dominated by diatoms (% )

# Discussion

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# References

Allen, A. P., J. F. Gillooly, and J. H. Brown. 2005. Linking the global carbon cycle to individual metabolism. Functional Ecology 19:202–213.

Andersen, T., P. S. Cranston, and J. H. Epler. 2013. Chironomidae of the Holarctic region: Keys and diagnoses, Part 1. Media Tryck, Lund, Sweden.

Arnason, B., P. Theodorsson, S. Björnsson, and K. Saemundsson. 1969. Hengill, a high temperature thermal area in Iceland. Bulletin Volcanologique 33:245–259.

Baiser, B., D. Gravel, A. R. Cirtwill, J. A. Dunne, A. K. Fahimipour, L. J. Gilarranz, J. A. Grochow, D. Li, N. D. Martinez, A. McGrew, T. Poisot, T. N. Romanuk, D. B. Stouffer, L. B. Trotta, F. S. Valdovinos, R. J. Williams, S. A. Wood, and J. D. Yeakel. 2019. Ecogeographical rules and the macroecology of food webs. Global Ecology and Biogeography 28:1204–1218.

Barnes, A. D., M. Jochum, J. S. Lefcheck, N. Eisenhauer, C. Scherber, M. I. O’Connor, P. de Ruiter, and U. Brose. 2018. Energy Flux: The Link between Multitrophic Biodiversity and Ecosystem Functioning. Trends in Ecology & Evolution 33:186–197.

Belgorodski, N., M. Greiner, K. Tolksdorf, and K. Schueller. 2017. rriskDistributions: Fitting Distributions to Given Data or Known Quantiles.

Benke, A. C., and A. D. Huryn. 2017. Secondary production and quantitative food webs. Pages 235–254 Methods in Stream Ecology. Elsevier.

Benke, A. C., A. D. Huryn, L. A. Smock, and J. B. Wallace. 1999. Length-Mass Relationships for Freshwater Macroinvertebrates in North America with Particular Reference to the Southeastern United States. Journal of the North American Benthological Society 18:308–343.

Benke, A. C., and J. B. Wallace. 1980. Trophic basis of production among net-spinning caddisflies in a southern appalachain stream. Ecology 61:108–118.

Benke, A. C., and J. B. Wallace. 1997. Trophic Basis of Production Among Riverine Caddisflies: Implications for Food Web Analysis. Ecology 78:1132–1145.

Brose, U., J. A. Dunne, Montoya José M., O. L. Petchey, F. D. Schneider, and U. Jacob. 2012. Climate change in size-structured ecosystems. Philosophical Transactions of the Royal Society B: Biological Sciences 367:2903–2912.

Brown, J. H., J. F. Gillooly, A. P. Allen, V. M. Savage, and G. B. West. 2004. Toward a Metabolic Theory of Ecology. Ecology 85:1771–1789.

Bürkner, P.-C. 2017. Brms: An R Package for Bayesian Multilevel Models Using Stan. Journal of Statistical Software 80:1–28.

Chao, A., and C. Ricotta. 2019. Quantifying evenness and linking it to diversity, beta diversity, and similarity. Ecology 100:e02852.

Cross, W. F., C. V. Baxter, K. C. Donner, E. J. Rosi-Marshall, T. A. Kennedy, R. O. Hall, H. A. W. Kelly, and R. S. Rogers. 2011. Ecosystem ecology meets adaptive management: Food web response to a controlled flood on the Colorado River, Glen Canyon. Ecological Applications 21:2016–2033.

Cross, W. F., J. B. Wallace, and A. D. Rosemond. 2007. Nutrient Enrichment Reduces Constraints on Material Flows in a Detritus-Based Food Web. Ecology 88:2563–2575.

de Ruiter, P. C., A.-M. Neutel, and J. C. Moore. 1995. Energetics, Patterns of Interaction Strengths, and Stability in Real Ecosystems. Science 269:1257–1260.

Dell, A. I., S. Pawar, and V. M. Savage. 2014. Temperature dependence of trophic interactions are driven by asymmetry of species responses and foraging strategy. Journal of Animal Ecology 83:70–84.

Fordyce, J. A., Z. Gompert, M. L. Forister, and C. C. Nice. 2011. A Hierarchical Bayesian Approach to Ecological Count Data: A Flexible Tool for Ecologists. PLOS ONE 6:e26785.

Friberg, N., J. B. Dybkjær, J. S. Olafsson, G. M. Gislason, S. E. Larsen, and T. L. Lauridsen. 2009. Relationships between structure and function in streams contrasting in temperature. Freshwater Biology 54:2051–2068.

Gibert, J. P. 2019. Temperature directly and indirectly influences food web structure. Scientific Reports 9:5312.

Gibert, J. P., and J. P. DeLong. 2017. Phenotypic variation explains food web structural patterns. Proceedings of the National Academy of Sciences 114:11187–11192.

Gillooly, J. F., J. H. Brown, G. B. West, V. M. Savage, and E. L. Charnov. 2001. Effects of size and temperature on metabolic rate. Science (New York, N.Y.) 293:2248–2251.

Gini, C. 1921. Measurement of Inequality of Incomes. The Economic Journal 31:124–126.

Hannesdóttir, E. R., G. M. Gíslason, J. S. Ólafsson, Ó. P. Ólafsson, and E. J. O’Gorman. 2013. Increased Stream Productivity with Warming Supports Higher Trophic Levels. Advances in Ecological Research 48:285–342.

Hood, J. M., J. P. Benstead, W. F. Cross, A. D. Huryn, P. W. Johnson, G. M. Gíslason, J. R. Junker, D. Nelson, J. S. Ólafsson, and C. Tran. 2018. Increased resource use efficiency amplifies positive response of aquatic primary production to experimental warming. Global Change Biology 24:1069–1084.

Huryn, A. D., and J. B. Wallace. 1986. A method for obtaining in situ growth rates of larval Chironomidae (Diptera) and its application to studies of secondary Production1. Limnology and Oceanography 31:216–221.

Junker, J. R., W. F. Cross, J. P. Benstead, A. D. Huryn, J. M. Hood, D. Nelson, G. M. Gíslason, and J. S. Ólafsson. 2020. Resource supply governs the apparent temperature dependence of animal production in stream ecosystems. Ecology Letters 23:1809–1819.

Lorenz, M. O. 1905. Methods of Measuring the Concentration of Wealth. Publications of the American Statistical Association 9:209.

May, R. M. 1972. Will a Large Complex System be Stable? Nature 238:413–414.

McCann, K., A. Hastings, and G. R. Huxel. 1998. Weak trophic interactions and the balance of nature. Nature 395:794–798.

Merritt, R. W., K. W. Cummins, and M. B. Berg, editors. 2008. An Introduction to the Aquatic Insects of North America. Fourth. Kendall/Hunt Publishing Co., Dubuque, IA.

Nelson, D., J. P. Benstead, A. D. Huryn, W. F. Cross, J. M. Hood, P. W. Johnson, J. R. Junker, G. M. Gíslason, and J. S. Ólafsson. 2017a. Shifts in community size structure drive temperature invariance of secondary production in a stream-warming experiment. Ecology 98:1797–1806.

Nelson, D., J. P. Benstead, A. D. Huryn, W. F. Cross, J. M. Hood, P. W. Johnson, J. R. Junker, G. M. Gíslason, and J. S. Ólafsson. 2017b. Experimental whole-stream warming alters community size structure. Global Change Biology 23:2618–2628.

O’Connor, M. I., M. F. Piehler, D. M. Leech, A. Anton, and J. F. Bruno. 2009. Warming and Resource Availability Shift Food Web Structure and Metabolism. PLOS Biology 7:e1000178.

O’Gorman, E. J., J. P. Benstead, W. F. Cross, N. Friberg, J. M. Hood, P. W. Johnson, B. D. Sigurdsson, and G. Woodward. 2014. Climate change and geothermal ecosystems: Natural laboratories, sentinel systems, and future refugia. Global Change Biology 20:3291–3299.

O’Gorman, E. J., D. E. Pichler, G. Adams, J. P. Benstead, H. Cohen, N. Craig, W. F. Cross, B. O. L. Demars, N. Friberg, G. M. Gíslason, R. Gudmundsdóttir, A. Hawczak, J. M. Hood, L. N. Hudson, L. Johansson, M. P. Johansson, J. R. Junker, A. Laurila, J. R. Manson, E. Mavromati, D. Nelson, J. S. Ólafsson, D. M. Perkins, O. L. Petchey, M. Plebani, D. C. Reuman, B. C. Rall, R. Stewart, M. S. A. Thompson, and G. Woodward. 2012. Impacts of Warming on the Structure and Functioning of Aquatic Communities. Pages 81–176 Advances in Ecological Research. Elsevier.

Padfield, D., C. Lowe, A. Buckling, R. Ffrench-Constant, S. Jennings, F. Shelley, J. S. Ólafsson, and G. Yvon-Durocher. 2017. Metabolic compensation constrains the temperature dependence of gross primary production. Ecology Letters 20:1250–1260.

Peterson, B. V. 1977. Black flies of Iceland (Diptera-Simuliidae). Canadian Entomologist 109:449–472.

Rosi-Marshall, E. J., H. A. Wellard Kelly, R. O. Hall, and K. A. Vallis. 2016. Methods for quantifying aquatic macroinvertebrate diets. Freshwater Science 35:229–236.

Schindelin, J., I. Arganda-Carreras, E. Frise, V. Kaynig, M. Longair, T. Pietzsch, S. Preibisch, C. Rueden, S. Saalfeld, B. Schmid, J.-Y. Tinevez, D. J. White, V. Hartenstein, K. Eliceiri, P. Tomancak, and A. Cardona. 2012. Fiji: An open-source platform for biological-image analysis. Nature Methods 9:676–682.

Solomon, D. L. 1975. A comparative approach to species diversity:7.

Thompson, R. M., U. Brose, J. A. Dunne, R. O. Hall, S. Hladyz, R. L. Kitching, N. D. Martinez, H. Rantala, T. N. Romanuk, D. B. Stouffer, and J. M. Tylianakis. 2012. Food webs: Reconciling the structure and function of biodiversity. Trends in Ecology & Evolution 27:689–697.

Vasseur, D. A., and K. S. McCann. 2005. A Mechanistic Approach for Modeling Temperature-Dependent Consumer-Resource Dynamics. The American Naturalist 166:184–198.

Walther, G.-R., E. Post, P. Convey, A. Menzel, C. Parmesan, T. J. C. Beebee, J.-M. Fromentin, O. Hoegh-Guldberg, and F. Bairlein. 2002. Ecological responses to recent climate change. Nature 416:389–395.

Welch, H. E. 1968. Relationships between Assimiliation Efficiencies and Growth Efficiencies for Aquatic Consumers. Ecology 49:755–759.

Woodward, G., J. P. Benstead, O. S. Beveridge, J. Blanchard, T. Brey, L. E. Brown, W. F. Cross, N. Friberg, T. C. Ings, U. Jacob, S. Jennings, M. E. Ledger, A. M. Milner, J. M. Montoya, E. O’Gorman, J. M. Olesen, O. L. Petchey, D. E. Pichler, D. C. Reuman, M. S. A. Thompson, F. J. F. Van Veen, and G. Yvon-Durocher. 2010. Ecological Networks in a Changing Climate. Pages 71–138 Advances in Ecological Research. Elsevier.

Zhang, L., D. Takahashi, M. Hartvig, and K. H. Andersen. 2017. Food-web dynamics under climate change. Proceedings of the Royal Society B: Biological Sciences 284:20171772.