

Haplotype analysis

Jing Hua Zhao

Outline

- Background
- Chromosome X analysis
- Power calculation
- Associate issues

Start with a Problem ...

- 5HT2 and Schizophrenia
- HLA markers and Schizophrenia
- Further work from an early report in Lancet
- Using unrelated individuals rather than the popular TDT
- But
 - Too many analyses
 - Available program (EH) would not work





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Model-Free Analysis and Permutation Tests for Allelic Associations

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Advances and Problems ...

Advances

- Easier analysis of gene-trait association
- Model-free statistics
- Permutation tests
- Some haplotype-specific statistics (F-T)

Problems

- Too slow
- Potentially useful model-based measure of LD

Allele association studies with SSR and SNP markers at known physical distances within a 1 Mb region embracing the *ALDH2* locus in the Japanese, demonstrates linkage disequilibrium extending up to 400 kb

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Faster Haplotype Frequency Estimation Using Unrelated Subjects

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Advances and Problems...

Advances

- Faster computation
- Likelihood-based LD statistics

Problems

- Abandon model-specific statistics
- Do not handle missing data
- Still use global association test
- Do not handle covariates

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GENECOUNTING: haplotype analysis with missing genotypes

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Advances and Problems

Advances

- Provide generic algorithm for missing genotype data as well as likelihood estimation and handle multiallelic markers
- Some haplotype specific tests
- Considerable easier than HAPLO (Yale)

Problems

- Chromosome X data
- Slow for large problem

GAD2 on Chromosome 10p12 Is a Candidate Gene for Human Obesity

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www.nature.com/ejhg

ARTICLE

Haplotype construction of the FRDA gene and evaluation of its role in type II diabetes

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Genetic Polymorphisms and Weight Loss in Obesity: A Randomised Trial of Hypo-Energetic High- versus Low-Fat Diets

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2LD, GENECOUNTING and HAP: computer programs for linkage disequilibrium analysis

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ASSOCIATION ANALYSIS OF UNRELATED INDIVIDUALS USING POLYMORPHIC GENETIC MARKERS

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BACKGROUND

Association analysis of unrelated individuals using multiple genetic markers are increasingly used. This could either be a marker-marker or marker-trait analysis. Haplotype phase uncertainty needs to be taken into account.

Clayton (2001) and Qin et al. (2002) have proposed heuristic EM and MCMC algorithms, but both are limited to SNPs. Here method in Clayton (2001) is extneded to multiallelic markers.

Previous global association tests using likelihoods do not give haplotype specific statistics, which are of considerable interest. We show via example they can be obtained during likelihood-based permutation tests.

METHOD AND IMPLEMENTATION

EXTENDING CLAYTON (2001)

- The new algorithm has the same feature of Clayton algorithm but considers multiple alleles when maintaining subject and haplotype lists.
- Appropriate procedure has been implemented to use results from multiple imputation as well as producing SAS programs containing the imputed data.

GLOBAL ASSOCIATION TESTS

- Likelihood-based permutation procedure is useful for producing LDbased statistics (Zhao et al. 1999)
 - 1. MARKER-MARKER ANALYSIS

×2 Statistic = -2(l[assuming association]-l[linkage equilibrium])

- 2. CASE-CONTROL ANALYSIS
- x2 Statistic = -2(l[case+control]-l[case]-l[control])
- Haplotype frequencies can be used for haplotype specific tests.

HAPLOTYPE SPECIFIC STATISTICS

Simple Freeman-Turkey statistic for marker-marker analysis

$$FT = \sqrt{O} + \sqrt{O + 1} - \sqrt{4E + 1}$$

- O, E = haplotype counts assuming linkage disequilibrium and linkage equilibrium
- Test of proportions for case-control heterogeneity analysis

$$z = \frac{\theta_{\rm l} - \theta_{\rm 2}}{\sqrt{V(\theta_{\rm l} - \theta_{\rm 2})}}$$

 θ_1 , θ_2 =haplotype frequency parameter, V(.) = the variance function.

EXAMPLES

HLA DRB, DQA and DQB markers (25,10,15 alleles) for 94
 Schizophrenic patients and 177 controls. It shows the efficiency of polymorphic markers and use of haplotype specific tests.

Table 1. Comparison of MCMC and EM estimates

| H aplotype | Count | MCMC | $\mathbf{E}\mathbf{M}$ | $\mathbf{E}\mathbf{q}$ | FT test | Pvalue |
|------------|-------|---------|------------------------|------------------------|---------|--------|
| 22-2-12 | 62 | 11.4391 | 14.0193 | 0.4126 | 14.34 | 0.0020 |
| 4-8-1 | 62 | 11.4391 | 11.2545 | 0.5329 | 12.14 | 0.0020 |
| 9-4-1 | 46 | 8.4871 | 9.7786 | 0.3468 | 11.71 | 0.0020 |
| 1-1-7 | 41 | 7.5646 | 9.4095 | 0.2048 | 12.02 | 0.0020 |
| 6-5-3 | 34 | 6.2731 | 5.9737 | 0.2703 | 8.85 | 0.0020 |
| 8-5-3 | 27 | 4.9815 | 5.0698 | 0.1728 | 8.40 | 0.0020 |
| 14-8-2 | 20 | 3.6900 | 4.2366 | 0.1986 | 7.38 | 0.0020 |
| 6-5-2 | 18 | 3.3210 | 2.6979 | 0.3142 | 4.98 | 0.0020 |
| 17-3-13 | 13 | 2.3985 | 3.1291 | 0.0144 | 7.21 | 0.0020 |
| 10-7-6 | 12 | 2.2140 | 2.7675 | 0.0027 | 6.84 | 0.0020 |
| 21-1-9 | 10 | 1.8450 | 2.5830 | 0.0125 | 6.49 | 0.0020 |
| 18-2-14 | 9 | 1.6605 | 1.6605 | 0.0103 | 5.06 | 0.0020 |
| 3-1-7 | 8 | 1.4760 | 1.4760 | 0.0551 | 4.35 | 0.0020 |
| 9-4-4 | 8 | 1.4760 | 1.4760 | 0.067 | 4.26 | 0.0020 |
| 8-5-2 | 6 | 1.1070 | 1.3878 | 0.2009 | 3.35 | 0.0022 |
| 9-8-1 | 6 | 1.1070 | <0.0001 | 0.5745 | -2.67 | N/A |
| 12-5-4 | 6 | 1.1070 | 1.2915 | 0.0096 | 4.37 | 0.0020 |
| 16-8-2 | 6 | 1.1070 | 1.2915 | 0.0421 | 4.09 | 0.0020 |

Haplotype assignment by EM was unambiguous except for one individual with missing data.

Table 2. Comparison of individual haplotypes for HLA data

| Haplotype | Case | Control | z-test | P value | Score test | P value |
|-----------|--------|---------|--------|---------|------------|---------|
| 6-6-2 | 3.1915 | 0.0000 | 3.38 | 0.0003 | 2.95 | 0.0040 |
| 8-1-3 | 1.5957 | 8.2919 | -3.13 | 0.0002 | -3.11 | 0.0016 |
| 8-5-3 | 1.0638 | 7.2069 | -3.10 | 0.0002 | -3.05 | 0.0011 |
| 13-1-7 | 3.1915 | 0.2825 | 2.85 | 0.0001 | 2.89 | 0.0069 |
| 17-2-14 | 3.1915 | 0.5650 | 2.41 | 0.0008 | 2.45 | 0.0232 |
| 8-6-3 | 1.5957 | 0.0000 | 2.38 | 0.0012 | 2.40 | 0.0390 |
| 6-5-2 | 0.5319 | 3.8550 | -2.27 | 0.0027 | -2.14 | 0.0268 |
| 18-2-14 | 0.0000 | 2.5424 | -2.20 | 0.0003 | -2.21 | 0.0313 |
| 14-3-13 | 2.1277 | 0.2882 | 2.13 | 0.0023 | 1.38 | 0.3479 |
| 9-6-4 | 1.2395 | 0.0000 | 2.10 | 0.0014 | 1.96 | 0.1132 |
| 22-6-4 | 1.2413 | 0.0000 | 2.10 | 0.0008 | 1.96 | 0.1213 |
| 10-7-6 | 4.7872 | 1.6949 | 2.09 | 0.0025 | 2.14 | 0.0462 |
| 3-1-7 | 0.0000 | 2.2599 | -2.08 | 0.0036 | -2.08 | 0.0595 |
| 9-4-4 | 0.0000 | 2.2595 | -2.08 | 0.0005 | -2.08 | 0.0544 |

The z-statistic is comparable to score statistic, while empirical P values are due to different permutation procedures.

ALDH2 markers and 130 alcoholic patients and 133 controls. This example shows the usefulness of LD-based analysis, the effect of missing data and importance of heuristic algorithm we implemented.

Table 3. Eight ALDH2 region markers on Chromosome 12

| Marker | Distance (b) | # alleles | # of missing |
|----------|--------------|-----------|--------------|
| | | | individuals |
| D12S2070 | > 450 000 | 8 | 251 |
| D12S839 | > 450 000 | 8 | 254 |
| D12S821 | ~ 400 000 | 13 | 229 |
| D12S1344 | 83 853 | 14 | 247 |
| EXON12 | О | 2 | 261 |
| EXON1 | 37 335 | 2 | 220 |
| D12S2263 | 38 927 | 13 | 249 |
| D12S1341 | > 450 000 | 10 | 250 |

93 individuals with complete genotypes

- 1 month using only all markers by standard EM algorithm (Zhao et al. 2002)
- 6 days for 100 EM iterations using only possible haplotypes excluding two individuals with genotypes at only two loci
- 5 minutes for posterior trimming with threshold 0.001 but 8 hours with threshold 0.00001 (the new implementation)
- 9 SNPs in APOC3/A4/A5 region from 3,012 individuals to study association with CHD and triglycerides. It shows drawbacks of heuristic algorithms and need to control for covariates.
- Log-likelihoods by Qin et al. (2002), Clayton (2001), Zhao et al. (2002) were -13,988.0, -11,607.7 and -11,521.5, respectively, suggesting increasing optimality
- 30min for Qin et al. (2002) and Clayton (2001), but 5min by Zhao et al. (2002), so the raw sorting approach is less appealing, method using sufficient statistics is desirable.
- Method of Zhao et al. (2002) also gave equilibrium likelihood

CONCLUSION

- •The heuristic EM and MCMC method is able to deal with multiple multiallelic markers, but it is still difficult to use it to obtain equilibrium likelihood and sufficient statistics are necessary for large sample.
- Haplotype specific statistics an be obtained from likelihood-based implementations. They are simpler than the score statistics.

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Clayton (2001). http://www-gene.cimr.cam.ac.uk/clayton/software Qin ZS. T Niu, JS Liu (2002). Am J Hum Genet 71, 1242-7 Zhao H, Pakstis AJ, Kidd JR, Kidd KK (1999). Ann Hum Genet 63:167-179, 1999. Zhao JH, S Lissarrague, L Essioux, PC Sham (2002). Bioinformatics 18, 1694-5

Advances and Problems...

Advances

- D' and SE(D'), etc. (nontrivial since authors got wrong, and developed their MIDAS system later on)
- Kullback-Leibler information (later AJHG paper)
- Chromosome X data
- Faster algorithm for multiallelic system (extension of SNPHAP)

Problems

- Covariates, R haplo.score, hapassoc
- GEI interactions, R haplo.stats

The Latest?

- SAS/Genetics
- HTR
- WHAP/PLINK
- ZAPLO
 - Some ~60 programs
- But a synthesis is preferable, e.g., R
 - gap, hapassoc
 - snpMatrix, SNPassoc, GenABEL, pbatR



Journal of Statistical Software

December 2007, Volume 23, Issue 8.

http://www.jstatsoft.org/

gap: Genetic Analysis Package

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GWAS

- Hapmap
- IMPUTE and MACH
- HAPVLMC, HMM.map

Returning to chr. X

- Review of gene-counting
- Modification



Computer Methods and Programs in Biomedicine

Computer Methods and Programs in Biomedicine 70 (2003) 1-9

www.elsevier.com/locate/cmpb

Generic number systems and haplotype analysis

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Table 1 Genotype counts for biallelic markers

| Marker 1 | Marker 2 | | | |
|----------|----------|-------|-------|--|
| | 1/1 | 1/2 | 2/2 | |
| 1/1 | n_0 | n_1 | n_2 | |
| 1/2 | n_3 | n_4 | n_5 | |
| 2/2 | n_6 | n_7 | n_8 | |

Table 2 Genotypic probabilities for two biallelic markers

| Marker 1 | Marker 2 | | | | |
|------------|--|--|----------------------------|--|--|
| | 1/1 | 1/2 | 2/2 | | |
| 1/1 1/2 | $\begin{array}{c} h_{11}^2 \\ 2h_{21}h_{11} \end{array}$ | $\begin{array}{c} 2h_{11}h_{12} \\ 2(h_{21}h_{12} + h_{22}h_{11}) \end{array}$ | $h_{12}^2 = 2h_{22}h_{12}$ | | |
| 2/2 | h_{21}^{2} | $2h_{21}h_{22}$ | h_{22}^{2} | | |

Now Steps for X Data

- Run GENECOUNTING
 - gcx HTR2c.inp HTR2c.gc
- Run awk to extract assignment
 - awk –f HTR2c.awk HTR2c.gc > HTR2c.gco
 - HTR2c.awk has
 - ^[1\]\[2\]/{gsub(^[\]/,""); print;}
- Create indicator variables for haplotypes
 - infile id chr snp1-snp4 p hid using 4snps.gco
 - tab hid, gen(h)
 - savasas using snps4.sas7bdat,replace

We Have Two Choices

- Stata with probability weighting
 - Limited success with output of parameters
- PROC SURVEYREG of SAS
 - It has ODS system such that estimates can be stacked for all six models
- How about single-locus analysis?
 - We can use a dummy variable and go through the same procedure, effectively an allellic analysis

Any Solution from R?

Possibly with library survey

Another Example ...

LRRK2 gene G2019S mutation and SNPs [haplotypes] in subtypes of Parkinson's disease

- Biswanath Patra1, Azemat J. Parsian2, Brad A. Racette3, Jing Hua Zhao4, Joel S. Perlmutter3, Abbas Parsian*
- 1) Simmons Comprehensive Cancer Center, University of Texas Southwestern Medical Center at Dallas,
- Dallas, TX 2) Department of Radiation Oncology, University of Arkansas for Medical Sciences, Little Rock,
- AR; 3) Department of Neurology, Washington University School of Medicine, St. Louis, MO;
- 4) MRC Epidemiology Unit, Strangeways Research Laboratory, Cambridge, UK

Reviewer #1:

The major limitation of this paper is the genetic association study since the study sample size does not appear to be sufficiently powered to detect the effect size differences between cases and controls. A control size of 186 is disproportionate to the cases. The authors should provide the following information:

- 1.A power calculation to determine if their studied sample size is sufficent to detect the effect size differences (6-loci haplotypes).
- 2.Provide an explanation why and how the six SNPs were selected. Have these SNPs been examined in previous published studies? Any bioinformatic data on these SNPs?
- 3. The age of the cases and controls does not appear to be matched. Has this been taken into account in the analysis?
- 4.A comment on the potential heterogeneity of the American white population since LRRK2 mutationla frequency appear to vary across some European populations.

... the study was envisaged such that it has a good though not optimal power as compared what is available in the literature. Given findings from the study, the information regarding power would only be supplementary, esp. there are also arguments that post-hoc calculation is questionable.

Nevertheless, in the case of ordinal regression (control, sporadic, familial) ~ six-SNP haplotype analysis, (via R haplo.stats library) assuming that the global association is comparable to that of a standard chi-squared statistic with value of 20.03, df=15, for a type-I error of 0.05 and a sample size of 684 used in the final analysis there would be 83% power. As this is a rough estimate, an alternative based on the 122221 haplotype between no association (haplotype frequency 0.0145) and association (haplotype freqnency 0.00874), with type-I error 0.05, a two-sided test, roughly has 30% power (32.4% with N=759 and 29.7% with N=684) according to pwr.p.test of R package pwr.

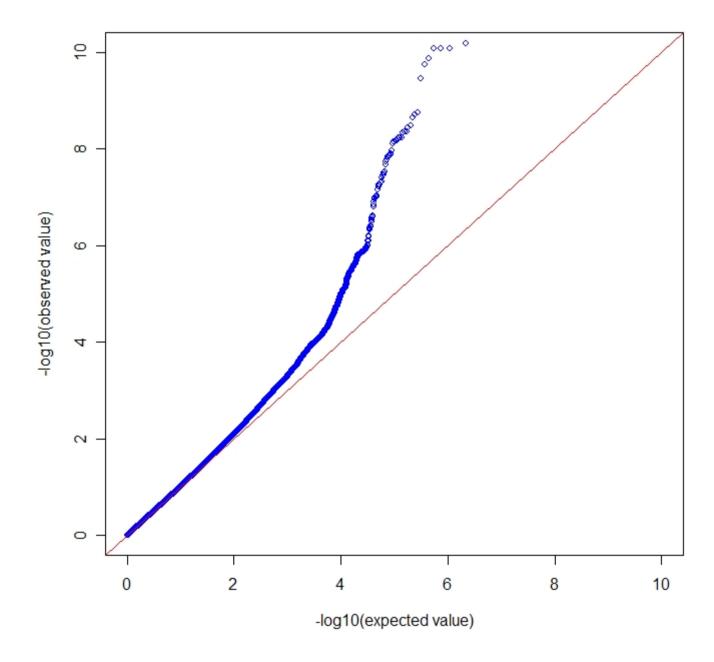
The reason to choose the six SNPs is clear from the manuscript, though not yet based on bioinformatics. The information from EMSEMBL and HapMap is consistent. In the case of HapMap which contains CEU sample (from http://www.hapmap.org/cgi-perl/gbrowse/Density_test/, type LRRK2 and search). The LRRK2 gene spans 144.3kb. Three SNPs (rs1491938, rs10784486, and rs1365763) tag 74 SNPs. The coverage is impressive although they do not cover the whole region. APART FROM rs1006151, THE OTHER TWO WERE NOT LISTED – I assume they are new relative to HapMap? Use HaploView option HapMap download for Chromosome 12 start and end positions (in Kb), Tagger, Load Includes (a column containing the six CNPs)

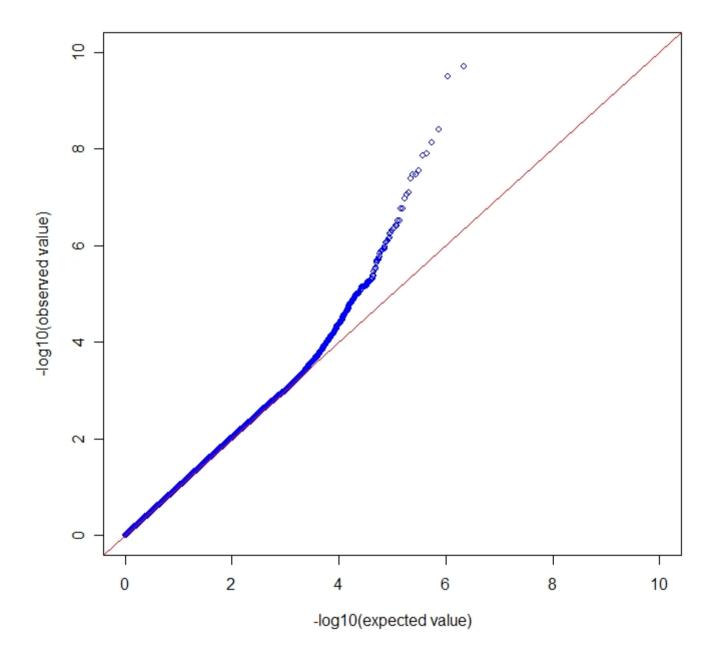
The R Trick

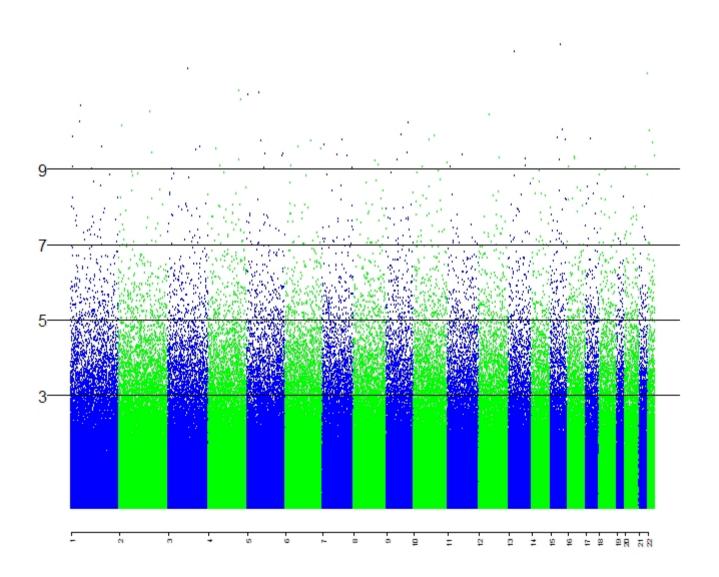
```
# 1-4-2008 MRC-Epid JHZ
# LRRK2 rely
# haplotype frequency under linkage equilibrium
p1 \leftarrow prod(c(0.15611, 0.64853, 0.84187, 0.96233, 0.58542, 0.30294))
# observed
p2 <- 0.00874
library(pwr)
p1
p2
h < -ES.h(p1,p2)
# had no missing data
pwr.p.test(h=h,n=759,sig.level=0.05,alternative="two.sided")
# as it is
pwr.p.test(h=h,n=684,sig.level=0.05,alternative="two.sided")
p1 <- 0.02093
p2 <- 0.00246
n1 <- 186
n2 <- 304+194
h<-ES.h(p1,p2)
h
pwr.2p2n.test(h=h,n1=n1,n2=n2,sig.level=0.05,alternative="two.sided")
n2 <- 350+225
pwr.2p2n.test(h=h,n1=n1,n2=n2,sig.level=0.05,alternative="two.sided")
```

Work from 2003 Onwards

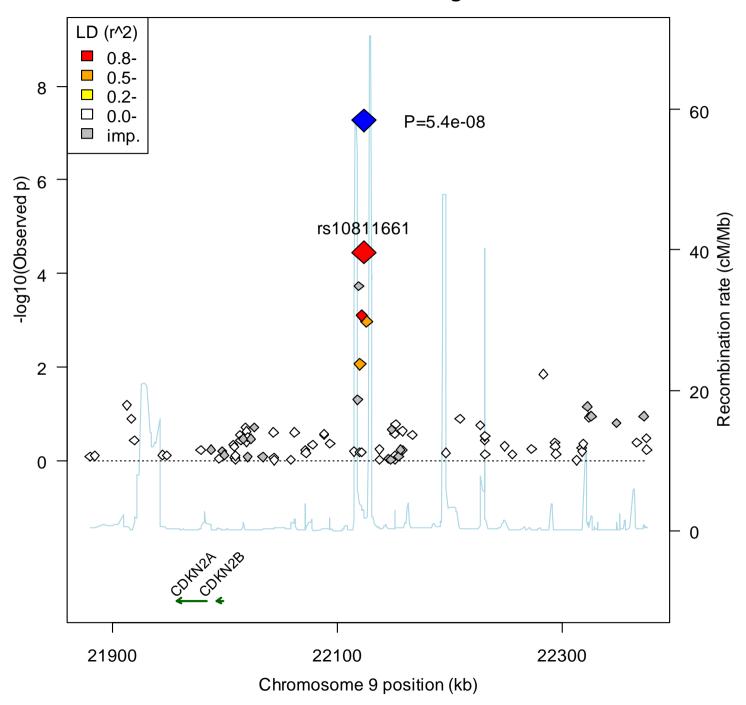
- We can do more with R ...
 - A tutorial
 - Graphics: Q-Q plot, Manhanttan plot, LD plot
 Regional Association plot



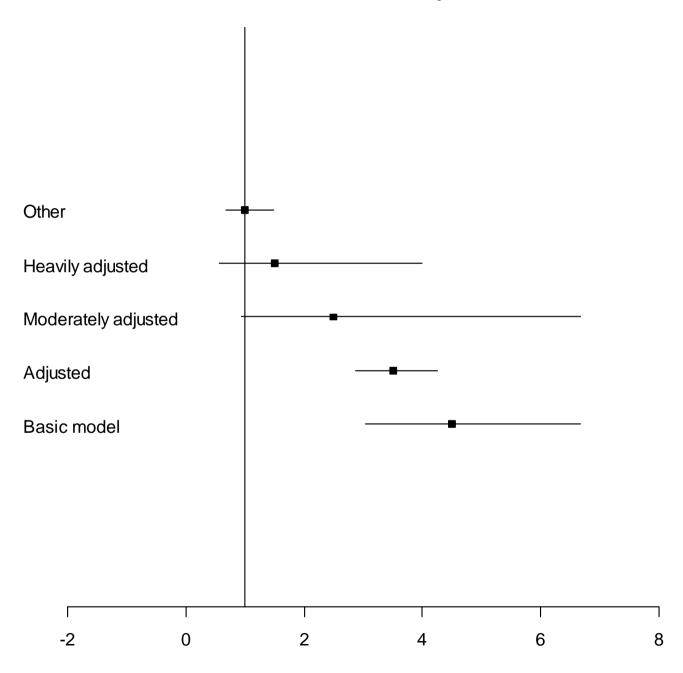




CDKN2A/CDKN2B region



This is a fictitious plot



More to go, but we stop here ...