

Supplementary Table 1 - Reactions and transporters in human galactose metabolism and kinetic parameters.

Id	Information	Kinetics
GLUT2	<p>Facilitated glucose transporter member 2</p> <p>D-glucose (disse) [glc_dis] ↔ D-glucose (cytosol) [glc] D-galactose (disse) [gal_dis] ↔ D-galactose (cytosol) [gal]</p> <p>Mechanism TCDB:2.A.1.1 (glucose transporter subfamily)</p> <p>Protein/Structure UniProt:P11168 (GTR2_HUMAN)</p> <p>Gene SLC2A2, GLUT2</p> <p>Disease OcMIM:227810 (Fanconi-Bickel syndrome; FBS)</p> <p>Galactose and glucose transported via GLUT2 (competitive inhibition kinetics) (Brown, 2000; Colville, et al., 1993)</p> <p>Deficient transport of galactose into hepatocytes in human patients with defective GLUT2 transporters (Fanconi-Bickel syndrome) resulting in galactose malabsorption/intolerance (Brown, 2000; Leslie, 2003).</p>	<p>km(D-glc)=21.7 ± 1.8mM (rat liver) (Ciaraldi, et al., 1986) km(D-glc)=66±14mM (rat hepatocytes) (Elliott and Craik, 1982) km(D-glc)=17mM (perfused rat liver, cited) (Elliott and Craik, 1982) km(D-glc)=30mM (rat hepatocytes, cited) (Elliott and Craik, 1982) km(3-O-MG)=42.3±4.1mM (human liver) (Gould, et al., 1991; Walmsley, et al., 1998) km(3-O-Methyl glc)=17.3 ± 4.3mM (rat liver) (Ciaraldi, et al., 1986) V_{max}(D-glc)=220±19mmol/min/l of cell H2O (rat hepatocytes) (Elliott and Craik, 1982) V_{max}(D-glc)=345mmol/min/l of cell H2O (perfused rat liver, cited) (Elliott and Craik, 1982) V_{max}(D-glc)=70mmol/min/l of cell H2O (rat hepatocytes, cited) (Elliott and Craik, 1982)</p> <p>km(D-gal)=174±48mM (rat hepatocytes) (Elliott and Craik, 1982) km(D-gal)=100mM (rat hepatocytes, cited) (Elliott and Craik, 1982) km(D-gal)>50mM (GLUT2 enderocytes) (Walmsley, et al., 1998) km(D-gal)=85.5 ± 10.7mM (human, liver-type GLUT2) (Colville, et al., 1993) km(D-gal)=92 ± 8.4mM (human, liver-type GLUT2) (Arbuckle, et al., 1996) km(D-gal)~27.7mM (dog liver, multiple indicator dilution curves) (Goresky, et al., 1973) V_{max} (D-gal)=288±48 mmol/min/l of cell H2O (rat hepatocytes) (Elliott and Craik, 1982) V_{max} (D-gal)=160mmol/min/l of cell H2O (rat hepatocytes, cited) (Elliott and Craik, 1982)</p> <p>Km(D-fru)=66mM (Walmsley, et al., 1998) Km(D-fru)=67mM (perfused rat liver, cited) (Elliott and Craik, 1982) Km(D-fru)=>100mM (rat hepatocytes, cited) (Elliott and Craik, 1982) v(D-fru)=291±26 mmol/min/l of cell H2O (rat hepatocytes) (Elliott and Craik, 1982) V_{max} (D-fru)=50mmol/min/l of cell H2O (perfused rat liver, cited) (Elliott and Craik, 1982) V_{max} (D-fru)=>160mmol/min/l of cell H2O (rat hepatocytes, cited) (Elliott and Craik, 1982)</p> <p>Accumulation rate (human GLUT2)</p>

		<p>v(deoxy-D-glc) = 4.33±0.15 pmol/min/oocyte v(D-gal) = 1.68±0.09 pmol/min/oocyte v(D-fru) = 0.78±0.09 pmol/min/oocyte</p>
GALK	<p>Galactokinase D-galactose [gal] + ATP [atp] ↔ D-galactose 1-phosphate [gal1p] + ADP [adp] + H⁺ [hydron]</p> <p>Reaction EC:2.7.1.6 RHEA:13556 KEGG:R01092 MetaCyc:GALACTOKIN-RXN</p> <p>Protein UniProt:P51570 (GALK1_HUMAN) homodimer P51570*2</p> <p>Gene GALK, GALK1</p> <p>Disease MIM:230200 (GALCT2 Galactosemia II)</p> <p>Galactokinase being rate limiting for galactose clearance (Schirmer, et al., 1986)</p>	<p>Two-substrate ordered, ternary complex reaction (Timson and Reece, 2003)</p> <p>kcat(gal) = 8.7±5 1/s (SABIORK:14785)(Timson and Reece, 2003)</p> <p>km(atp) = 0.034±0.004mM (SABIORK:14792)(Timson and Reece, 2003) km(atp) = 0.12mM (adult, rat liver){Cuatrecasas1965}</p> <p>km(gal)=0.97±0.22mM (SABIORK:14785) (Timson and Reece, 2003) km(gal) = 0.436mM (SABIORK:45367), (Sanguolo, et al., 2004) km(gal) = 0.15mM (adult, rat liver){Cuatrecasas1965} km(gal) = 0.65mM (newborn, rat liver){Cuatrecasas1965} km(gal) = 0.91mM (18 day fetal, rat liver){Cuatrecasas1965} km(gal) = 0.14±0.01mM (SEM, N=6, adult rat liver) {Walker1968} km(gal) = 0.15±0.01mM (SEM, N=4, neonatal rat liver) {Walker1968} km(gal) = 0.14±0.01mM (SEM, N=4, foetal rat liver) {Walker1968}</p> <p>Uncompetitive product inhibition of GALK (adult rat liver) by gal1p with both 1mM and 5mM gal1p altering the Km for galactose from 0.150mM to 0.800mM (1mM gal1p caused 15% inhibition, 5mM gal1p 50% inhibition) ki(gal1p) = 5.3mM (5.0-5.7mM) (adult rat liver) (Cuatrecasas and Segal, 1965)</p> <p>km(gal)<0.83mM (dog liver, multiple indicator dilution curves) (Goresky, et al., 1973)</p>
IMP	<p>Inositol monophosphatase D-galactose 1-phosphate [gal1p] ↔ D-galactose [gal] + phosphate [pi]</p> <p>Reaction EC:3.1.3.25</p> <p>Protein UniProt:P29218 (IMPA1_HUMAN) homodimer P29218*2</p> <p>Gene IMPA1, IMPA</p>	<p>Competitive inhibition model Kinetic analysis demonstrated that gal1p competitively inhibited human IMP1 by increasing Km for inositol-1p (ino1p) from 320±50µM to 980±70µM without changing the Vmax (Slepak, et al., 2007) km(ino1p) = 0.320±0.050mM (Slepak, et al., 2007) km(gal1p) = 0.35mM (similar kinetics gal1p to ino1p in vitro) (Parthasarathy, et al., 1997)</p>

<hr/>		
	Normal substrate inositol-1p (ino1p)	
GALT	<p>Galactose-1-phosphate uridyl transferase UDP-D-glucose [udpglc] + D-galactose 1-phosphate [gal1p] ↔ D-glucose 1-phosphate [glc1p] + UDP-D-galactose [udpgal].</p> <p>Reaction EC:2.7.7.12 RHEA:13992 KEGG:R00955</p> <p>Protein UniProt:P07902 (GALT_HUMAN) homodimer P07902*2</p> <p>Gene GALT</p> <p>Disease OMIM:230400 (GALCT Galactosemia)</p>	<p>The catalytic mechanism of GALT is ping-pong kinetics with covalent intermediate UMP-enzyme (Facchiano and Marabotti, 2010).</p> <p>Mutation analysis (Quimby, et al., 1996) km(gal1p) = 0.57±0.14mM (human, wildtype) (Quimby, et al., 1996) km(udpglc) = 0.21±0.04mM (human, wildtype) (Quimby, et al., 1996)</p> <p>Mutation analysis (Tang, et al., 2012) km(gal1p) = 1.25±0.36mM (human, wildtype) (Tang, et al., 2012) km(udpglc) = 0.43±0.09mM (human, wildtype) (Tang, et al., 2012)</p> <p>(?species, 4°C) (Geeganage and Frey, 1998) km(udpglc) = 0.5±0.1mM v(glc1p) = 281± 18 1/s km(glc1p) = 0.37±0.18mM v(glc1p) = 226± 10 1/s km(gal1p) = 0.061±0.020mM v(glc1p) = 166± 13 1/s</p> <p>Potent linear competent inhibitors UTP and UDP of UDP-glucose (Segal and Rogers, 1971): Ki(UTP) = 0.13mM (rat, liver) Ki(UDP) = 0.35mM (rat, liver) Ki(UMP) = 2.3mM (rat, liver) Ki(UDP-glucuronic acid)=0.40mM (rat, liver)</p>
GALE	<p>UDP-glucose 4-epimerase UDP-D-glucose [udpglc] ↔ UDP-D-galactose [udpgal]</p> <p>Reaction EC:5.1.3.2 RHEA:22171 KEGG:R00291</p> <p>Protein UniProt:Q14376 (GALE_HUMAN) homodimer Q14376*2</p> <p>Gene GALE</p> <p>Disease OMIM:230350 (GALE deficiency)</p>	<p>Mutation analysis (Timson, 2005) km(udpgal)=0.069±0.012mM (human, wildtype) (Timson, 2005) kcat(udpgal) = 36±1.4 1/s (human, wildtype) (Timson, 2005)</p> <p>km(udpgal) = 0.15 ± 0.02mM (human, wildtype) (Wohlers and Fridovich-Keil, 2000) km(udpgal, V94M) = 0.27 ± 0.01mM (human, V94M) (Wohlers and Fridovich-Keil, 2000) km(udpgal)=0.140± 0.007mM (human, wildtype) (SABIORK:19823) (Winans and Bertozzi, 2002) km(udpgal)=0.120± 0.04mM (human, wildtype) (SABIORK:46260) (Wasilenko, et al., 2005) kcat= 33.8±11.2 (human, wildtype) (SABIORK:16222) (Thoden, et al., 2002) km(udpgal) = 0.230±0.06mM (human, wildtype) (SABIORK:46263) (Quimby, et</p>
<hr/>		

	Alternative activity with GlcNAc: UDP-GalNAc ↔ UDP-GlcNAc	al., 1997)
	<p>“Ethanol treatment increases the NADH/NAD ratio in liver and by this inhibits the GALE. Under these conditions oxidation and elimination of galactose are impaired. Combined galactose+ethanol treatment results in accumulation of gal1p and udpgal in rat liver. The formation of high amounts of udpgal leads to a change in the distribution of liver uracil nucleotides. A marked decrease of udpglc, utp, udp and ump is followed by an increase of the sum of uracil nucleotides.” (Keppler, et al., 1970)</p> <p>“The GALE reaction is indicated as the rate-limiting step of galactose metabolism in rat liver by the ratio of galactose metabolites.” (Keppler, et al., 1970)</p> <p>“The almost 4-fold increase of gal1p and udpgal and the even stronger drop of the udpglc content in the ethanol treated liver after a galactose load demonstrates the ethanol-induced inhibition of the GALE.” (Keppler, et al., 1970).</p> <p>“Galactose provokes pronounced alterations of the uracil nucleotide contents in the liver, which are intensified by an inhibition of the GALE” (Keppler, et al., 1970)</p>	
UGP	<p>UDP-glucose pyrophosphorylase D-glucose 1-phosphate [glc1p] + UTP [utp] + H+[hydron] ↔ UDP-glucose [udglc] + diphosphate [ppi]</p> <p>Reaction EC:2.7.7.9 RHEA:19892 KEGG:R00289</p> <p>Protein UniProt:Q16851 (UGPA_HUMAN) homooctamer Q16851*8</p> <p>Gene UGP2, UGP1</p>	<p>Enzyme displays simple Michaelis-Menten kinetics in both directions (Chang, et al., 1996)</p> <p>MgUTP is a product inhibitor that shows competitive inhibition with respect to UDP-Glc (Chang, et al., 1996)</p> <p>(human, liver, wildtype) (Chang, et al., 1996)</p> <p>km(udpglc) = [0.031 - 0.051]mM km(pp) = [0.172 - 0.210] mM km(glc1p) = [0.172 - 0.174] mM km(utp) = [0.563 - 0.692] mM ki(utp) = 0.477± 41 mM (competitive inhibition with respect to UDP-glc) V_{fwd}/V_{rev} = 0.260 (human, liver, wildtype) (Duggleby, et al., 1996)</p> <p>km(udpglc) = 0.049±0.004mM km(pp) = 0.166±0.013 mM km(glc1p) = 0.172±0.010 mM km(utp) = 0.563±0.115 mM ki(utp) = 0.643± 0.047 mM (competitive inhibition with respect to UDP-glc)</p>
UGALP	<p>UDP-galactose pyrophosphorylase D-galactose-1-phosphate [gal1p] + UTP [utp] + H+[hydron] ↔ UDP-D-galactose [udpgal] pyrophosphate [pp]</p>	

	<p>Reaction EC:2.7.7.10 RHEA:14212 KEGG:R00502</p> <p>Protein UniProt:Q16851 (UGPA_HUMAN) homooctamer Q16851*8</p> <p>Gene UGP2, UGP1</p> <p>“The formation of UDP-glucose is the major physiological function of UGP, however at slow rates, the enzyme also catalyzes the phosphorylation of UDP-galactose.” (Knop and Hansen, 1970) [Segal1968].</p> <p>“Not significant in normal physiological conditions, but in galactosemic patients could circumvent GALT deficiency. Stable transfection of human UGP (hUGP2) rescued galactose GALT deficient yeast from “galactose toxicity.” [Lai2002].</p>	<p>ki(udpglc) = 0.013± 4 mM (competitive inhibition with respect to UTP?) (human, liver, wildtype) (Knop and Hansen, 1970)</p> <p>keq([udpglc][pp]/([UTP][glc1p])) = 0.15 – 0.16</p> <p>km(udpglc) = 50mM</p> <p>km(utp) = 48 mM</p> <p>km(glc1p) = 95±10 mM</p> <p>keq([UTP][glc1p]/([udpglc][pp])) = 4.55±0.1 (Guynn, et al., 1974) (0.22)</p> <p>The saturating concentration for UDP-galactose is 10 times that of UDP-glucose: km(udpgal) = 10*km(udpglc) ~ 0.5mM (human, liver, wildtype) (Knop and Hansen, 1970)</p> <p>km(udpgal) = 0.420mM (rabbit, liver, wildtype) (Turnquist, et al., 1974)</p> <p>udpgal was an adequate substrate at 10 times the concentration of udpglc, showing 14.3% of udpglc (Calf) and 12.0% (Human).</p> <p>activity with udpgal 2-12% of udpglc (12% with 3mM udpgal) (human liver) (Turnquist, et al., 1974)</p> <p>“The activity of UDPG:galactose-1-phosphate uridylyltransferase from rat liver under optimal conditions in vitro is less than 5% of the UDPG pyrophosphorylase activity.” (Keppler, et al., 1970)[Keppler1970 ->39,40]</p> <p>gal1p as competitive inhibitor of glc1p</p> <p>“Previously, we showed that galactose-1-phosphate competitively inhibited UDP-glucose pyrophosphorylase, leading to 66% reduction in UDP-glucose/galactose contents in GALT-deficient cells under galactose challenge” [Slepek2007->Lai2002].</p>
ALDR	<p>Aldose reductase (galactitol NAD 1-oxidoreductase) D-galactose [gal] + NADPH [nadph] + H ↔ galactitol [galtol] + NADP [nadp]</p> <p>Reaction EC:1.1.1.21 RHEA:37967 KEGG:R01095</p> <p>Protein UniProt:P15121 (ALDR_HUMAN) monomer P15121*1</p> <p>Gene AKR1B1, ALDR1</p>	<p>km(gal) = 40.0mM (human brain) (SABIORK:22893) (Wermuth, et al., 1982)</p> <p>kcat(gal) = 0.40 1/s (human brain) (SABIORK:22893) (Wermuth, et al., 1982)</p> <p>km(gal) = 110.0mM (human brain) (SABIORK:15695) (Wermuth and von Wartburg, 1982)</p>

Aldolase reductase is specific for NADPH as cofactor (NADH ~10% of NADPH-dependent activity) ([Wermuth and von Wartburg, 1982](#)).

“Aldolase reductase catalyzes the conversion of aldoses and a number of other aldehydes to the corresponding alcohol metabolites. It is one of several cytosolic, monomeric, NADPH-dependent aldehyde and ketone reductases of wide substrate specificity” ([Wermuth, et al., 1982](#))”.

PGM1

Phosphoglucomutase-1

D-glucose 1-phosphate [**glc1p**] ↔ D-glucose 6-phosphate [**glc6p**]

Reaction

EC:5.4.2.2

RHEA:23539

KEGG:R00959

Protein (multiple isoforms PGM1, PGM2)

UniProt:P36871 (PGM1_HUMAN)

monomer P36871*1

main isoform for glc1p ↔ glc6p reaction

Gene

PGM1

Disease

OMIM:612934 (Glycogen storage disease 14)

OMIM:614921 (Congenital disorder of glycosylation 1T CDG1T)

Protein

UniProt:Q96G03 (PGM2_HUMAN)

Gene

PGM2

CDG1T - A multisystem disorder caused by a defect in glycoprotein biosynthesis and characterized by under-glycosylated serum glycoproteins.

The equilibrium lies strongly toward glc6p and reaction proceeds through **ping-pong mechanism** ([Guynn, et al., 1974](#))

The kinetic properties of PGM1 and PGM2 are essentially the same. PGM1 is specific for mutation of glucose, whereas PGM2 also has phosphoribomutase activities. (human, RBC) ([Accorsi, et al., 1989](#))

[**glc6p**]/[**glc1p**] ~10-12 ([Guynn, et al., 1974](#))

DeltaG = -7.1 kJ/mol ([König, et al., 2012](#))

km(glc1p) = 0.049mM (human, RBC) ([Quick, et al., 1974](#))

km(glc1p) = 0.045mM (rat, heart) ([Kashiwaya, et al., 1994](#))

km(glc6p) = 0.67mM (rat, heart) ([Kashiwaya, et al., 1994](#))

km(glc1p) = 0.083mM (human, RBC, PGM1) ([Accorsi, et al., 1989](#))

ki(fru16bp) = 0.092mM (human, RBC, PGM1) ([Accorsi, et al., 1989](#))

PPASE

Pyrophosphatase

Pyrophosphate [**pp**] + H₂O [**h2o**] → 2 phosphate [**phos**] + H⁺ [**hydron**]

km(pp) = 0.005mM (rat liver) ([Yoshida, et al., 1982](#))

km(pp) = 0.14mM (human erythrocyte) ([Thuillier, 1978](#))

km(pp) = 0.07mM (rat liver) ([Irie, et al., 1970](#))

Delta G0 = -23.56 kJ/mol ([Thuillier, 1978](#))

	Reaction EC:3.6.1.1 RHEA:24579 KEGG:R00004 Protein UniProt:Q15181 (IPYR_HUMAN) homodimer Q15181*2 Gene PPA1, IOPPP, PP	Delta G0 = -19.2 kJ/mol (Guynn, et al., 1974)
NDKU	Nucleoside diphosphokinase (ATP:UDP phosphotransferase) ATP [atp] + UDP [udp] ↔ ADP [adp] + UTP [utp] Reaction EC: 2.7.4.6 RHEA:25101 KEGG:R00156 Protein Multitude of isoforms	Compulsory-order substituted-enzyme (Ping Pong Bi Bi) mechanism (Lam and Packham, 1986) km(atp) = 0.38mM (human, platelets) (Lam and Packham, 1986) km(adp) = 0.024mM (human, platelets) (Lam and Packham, 1986) km(gtp) = 0.12mM (human, platelets) (Lam and Packham, 1986) km(atp) = 1.33mM (rat, liver) (Kimura and Shimada, 1988) km(adp) = 0.042mM (rat, liver) (Kimura and Shimada, 1988) km(udp) = 0.19mM (rat, liver) (Kimura and Shimada, 1988) km(atp) = 1.80 mM (rat, liver) (Fukuchi, et al., 1994) km(adp) = 0.066 mM (rat, liver) (Fukuchi, et al., 1994) km(utp) = 27.00mM (rat, liver) (Fukuchi, et al., 1994) km(gtp) = 0.15mM (rat, liver) (Fukuchi, et al., 1994) km(gdp) = 0.049mM (rat, liver) (Fukuchi, et al., 1994)
NADPR	NADP reductase NADP [nadp] + H2 → NADPH [nadph] Modeled via glucose-6-phosphate dehydrogenase in pentose phosphate pathway D-glucose 6-phosphate [glc6p] + NADP [nadp] → 6-phospho-D-glucono-1,5-lactone + NADPH [nadph] + H Reaction EC: 1.1.1.49 RHEA:15844 KEGG:R00835 Protein UniProt:P11413 (G6PD_HUMAN) homotetramer (dimer of dimer) P11413*4	Delta G0 = -19.6 kJ/mol [Schuster1995] km(glc6p) = 0.040±0.008 mM (human, placenta) (Ozer, et al., 2001) km(nadp) = 0.020±0.010 mM (human, placenta) (Ozer, et al., 2001) ki(nadph) = 0.0171±0.0032 mM (human, placenta) (Ozer, et al., 2001) km(glc6p) = 0.072 mM (human, RBC) (Bautista, et al., 1992) km(glc6p) = 0.069±0.003 mM (human, recombinant) (Bautista, et al., 1992) km(nadp) = 0.013 mM (human, RBC) (Bautista, et al., 1992) km(nadp) = 0.012±0.002 mM (human, recombinant) (Bautista, et al., 1992) km(nadph) = 0.015±0.002 mM (human, RBC) (Bautista, et al., 1992) km(nadph) = 0.014±0.003 mM (human, recombinant) (Bautista, et al., 1992) km(glc6p) = 0.326mM (rat, liver) km(glc6p) = 0.157mM (rat, liver)

	<p>Gene G6PD</p>	<p>(Corpas, et al., 1995; Corpas, et al., 1995) km(nadp) = 0.108 mM (rat, liver) km(nadp) = 0.258 mM (rat, liver) (Corpas, et al., 1995; Corpas, et al., 1995) ki(nadhp) = 0.010 mM(rat, liver) ki(nadhp) = 0.021 mM(rat, liver) (Corpas, et al., 1995; Corpas, et al., 1995)</p>
ATPS	<p>ATP synthesis ADP [adp] + phosphate [phos] + H⁺ [hydron] → ATP [atp] + H₂O [h2o]</p> <p>Reaction RHEA:13068 KEGG:R00086</p> <p>Modelled via general ATP producing reaction representative for ATP production via glycolysis and oxidative phosphorylation</p>	
GTFGAL GTFGLC	<p>Glycosyltransferase Acceptor + UDP-glucose [udpglc] → Acceptor-glucose + UDP [udp]</p> <p>Acceptor + UDP-glucose [udpgal] → Acceptor-glucose + UDP [udp]</p> <p>Enzymes that transfer mono- or oligosaccharides from donor molecules to growing oligosaccharide chains or proteins are called <u>glycosyltransferases</u> (Gtfs)</p>	
GLY	<p>Glycolysis D-glucose 6-phosphate [glc6p] + 6 O₂ [o2] → phosphate [phos] + 6 CO₂ [co2] + 5 H₂O [h2o]</p> <p>Pseudo-reaction for using galactose in glycolysis freeing the phosphate.</p>	
GALDH	<p>Galactose 1-dehydrogenase D-galactose + NAD⁺ ↔ D-galactono-1,4-lactone + NADH + H⁺ EC.1.1.1.48 (Brenda only bacteria) D-galactose -> galactonate (first enzyme in oxidative pathway) [Segal1968 -> Cuatrecasas1966,15] Alternative pathway to xylulose.</p> <p>D-Galactose + Oxygen + H₂O <=> D-Galactonate + Hydrogen</p>	

peroxide
EC:1.1.3.9
[KEGG:R01098](#)
(only bacteria)

REFERENCES

- Accorsi, A., *et al.* (1989) Isoenzymes of phosphoglucomutase from human red blood cells: isolation and kinetic properties, *Preparative biochemistry*, **19**, 251-271.
- Arbuckle, M.I., *et al.* (1996) Structure-function analysis of liver-type (GLUT2) and brain-type (GLUT3) glucose transporters: expression of chimeric transporters in *Xenopus* oocytes suggests an important role for putative transmembrane helix 7 in determining substrate selectivity, *Biochemistry*, **35**, 16519-16527.
- Bautista, J.M., Mason, P.J. and Luzzatto, L. (1992) Purification and properties of human glucose-6-phosphate dehydrogenase made in *E. coli*, *Biochimica et biophysica acta*, **1119**, 74-80.
- Brown, G.K. (2000) Glucose transporters: structure, function and consequences of deficiency, *Journal of inherited metabolic disease*, **23**, 237-246.
- Chang, H.Y., *et al.* (1996) The importance of conserved residues in human liver UDPglucose pyrophosphorylase, *European journal of biochemistry / FEBS*, **236**, 723-728.
- Ciaraldi, T.P., Horuk, R. and Matthaei, S. (1986) Biochemical and functional characterization of the rat liver glucose-transport system. Comparisons with the adipocyte glucose-transport system, *The Biochemical journal*, **240**, 115-123.
- Colville, C.A., *et al.* (1993) Kinetic analysis of the liver-type (GLUT2) and brain-type (GLUT3) glucose transporters in *Xenopus* oocytes: substrate specificities and effects of transport inhibitors, *The Biochemical journal*, **290 (Pt 3)**, 701-706.
- Corpas, F.J., *et al.* (1995) Kinetic properties of hexose-monophosphate dehydrogenases. II. Isolation and partial purification of 6-phosphogluconate dehydrogenase from rat liver and kidney cortex, *Molecular and cellular biochemistry*, **144**, 97-104.
- Corpas, F.J., *et al.* (1995) Kinetic properties of hexose-monophosphate dehydrogenases. I. Isolation and partial purification of glucose-6-phosphate dehydrogenase from rat liver and kidney cortex, *Life sciences*, **56**, 179-189.
- Cuatrecasas, P. and Segal, S. (1965) Mammalian Galactokinase. Developmental and Adaptive Characteristics in the Rat Liver, *The Journal of biological chemistry*, **240**, 2382-2388.
- Duggleby, R.G., *et al.* (1996) Sequence differences between human muscle and liver cDNAs for UDPglucose pyrophosphorylase and kinetic properties of the recombinant enzymes expressed in *Escherichia coli*, *European journal of biochemistry / FEBS*, **235**, 173-179.
- Elliott, K.R. and Craik, J.D. (1982) Sugar transport across the hepatocyte plasma membrane, *Biochemical Society transactions*, **10**, 12-13.
- Facchiano, A. and Marabotti, A. (2010) Analysis of galactosemia-linked mutations of GALT enzyme using a computational biology

approach, *Protein engineering, design & selection : PEDS*, **23**, 103-113.

Fukuchi, T., *et al.* (1994) Recombinant rat nucleoside diphosphate kinase isoforms (alpha and beta): purification, properties and application to immunological detection of native isoforms in rat tissues, *Biochimica et biophysica acta*, **1205**, 113-122.

Geeganage, S. and Frey, P.A. (1998) Transient kinetics of formation and reaction of the uridylyl-enzyme form of galactose-1-P uridylyltransferase and its Q168R-variant: insight into the molecular basis of galactosemia, *Biochemistry*, **37**, 14500-14507.

Goresky, C.A., Bach, G.G. and Nadeau, B.E. (1973) On the uptake of materials by the intact liver. The transport and net removal of galactose, *The Journal of clinical investigation*, **52**, 991-1009.

Gould, G.W., *et al.* (1991) Expression of human glucose transporters in *Xenopus* oocytes: kinetic characterization and substrate specificities of the erythrocyte, liver, and brain isoforms, *Biochemistry*, **30**, 5139-5145.

Guynn, R.W., *et al.* (1974) The concentration and control of cytoplasmic free inorganic pyrophosphate in rat liver in vivo, *The Biochemical journal*, **140**, 369-375.

Irie, M., *et al.* (1970) Distribution and properties of alkaline pyrophosphatases of rat liver, *Journal of biochemistry*, **67**, 47-58.

Kashiwaya, Y., *et al.* (1994) Control of glucose utilization in working perfused rat heart, *The Journal of biological chemistry*, **269**, 25502-25514.

Keppler, D., Rudigier, J. and Decker, K. (1970) Trapping of uridine phosphates by D-galactose in ethanol-treated liver, *FEBS letters*, **11**, 193-196.

Keppler, D.O., *et al.* (1970) The trapping of uridine phosphates by D-galactosamine. D-glucosamine, and 2-deoxy-D-galactose. A study on the mechanism of galactosamine hepatitis, *European journal of biochemistry / FEBS*, **17**, 246-253.

Kimura, N. and Shimada, N. (1988) Membrane-associated nucleoside diphosphate kinase from rat liver. Purification, characterization, and comparison with cytosolic enzyme, *The Journal of biological chemistry*, **263**, 4647-4653.

Knop, J.K. and Hansen, R.G. (1970) Uridine diphosphate glucose pyrophosphorylase. IV. Crystallization and properties of the enzyme from human liver, *The Journal of biological chemistry*, **245**, 2499-2504.

König, M., Bulik, S. and Holzhütter, H.G. (2012) Quantifying the contribution of the liver to glucose homeostasis: a detailed kinetic model of human hepatic glucose metabolism, *PLoS computational biology*, **8**, e1002577.

Lam, S.C. and Packham, M.A. (1986) Isolation and kinetic studies of nucleoside diphosphokinase from human platelets and effects of cAMP phosphodiesterase inhibitors, *Biochemical pharmacology*, **35**, 4449-4455.

Leslie, N.D. (2003) Insights into the pathogenesis of galactosemia, *Annual review of nutrition*, **23**, 59-80.

Ozer, N., Aksoy, Y. and Ogus, I.H. (2001) Kinetic properties of human placental glucose-6-phosphate dehydrogenase, *The international journal of biochemistry & cell biology*, **33**, 221-226.

Parthasarathy, R., Parthasarathy, L. and Vadnal, R. (1997) Brain inositol monophosphatase identified as a galactose 1-phosphatase, *Brain research*, **778**, 99-106.

Quick, C.B., Fisher, R.A. and Harris, H. (1974) A kinetic study of the isozymes determined by the three human phosphoglucomutase loci PGM1, PGM2, and PGM3, *European journal of biochemistry / FEBS*, **42**, 511-517.

Quimby, B.B., *et al.* (1997) Characterization of two mutations associated with epimerase-deficiency galactosemia, by use of a yeast

expression system for human UDP-galactose-4-epimerase, *American journal of human genetics*, **61**, 590-598.

Quimby, B.B., *et al.* (1996) Functional requirements of the active site position 185 in the human enzyme galactose-1-phosphate uridylyltransferase, *The Journal of biological chemistry*, **271**, 26835-26842.

Sangiuolo, F., *et al.* (2004) Biochemical characterization of two GALK1 mutations in patients with galactokinase deficiency, *Human mutation*, **23**, 396.

Schirmer, W.J., *et al.* (1986) Galactose clearance as an estimate of effective hepatic blood flow: validation and limitations, *The Journal of surgical research*, **41**, 543-556.

Segal, S. and Rogers, S. (1971) Nucleotide inhibition of mammalian liver galactose-1-phosphate uridylyltransferase, *Biochimica et biophysica acta*, **250**, 351-360.

Slepek, T.I., *et al.* (2007) Involvement of endoplasmic reticulum stress in a novel Classic Galactosemia model, *Molecular genetics and metabolism*, **92**, 78-87.

Tang, M., *et al.* (2012) Correlation assessment among clinical phenotypes, expression analysis and molecular modeling of 14 novel variations in the human galactose-1-phosphate uridylyltransferase gene, *Human mutation*, **33**, 1107-1115.

Thoden, J.B., *et al.* (2002) Structural analysis of the Y299C mutant of Escherichia coli UDP-galactose 4-epimerase. Teaching an old dog new tricks, *The Journal of biological chemistry*, **277**, 27528-27534.

Thuillier, L. (1978) Purification and kinetic properties of human erythrocyte Mg²⁺-dependent inorganic pyrophosphatase, *Biochimica et biophysica acta*, **524**, 198-206.

Timson, D.J. (2005) Functional analysis of disease-causing mutations in human UDP-galactose 4-epimerase, *The FEBS journal*, **272**, 6170-6177.

Timson, D.J. and Reece, R.J. (2003) Functional analysis of disease-causing mutations in human galactokinase, *European journal of biochemistry / FEBS*, **270**, 1767-1774.

Turnquist, R.L., *et al.* (1974) Uridine diphosphate glucose pyrophosphorylase: differential heat inactivation and further characterization of human liver enzyme, *Biochimica et biophysica acta*, **364**, 59-67.

Walmsley, A.R., *et al.* (1998) Sugar transporters from bacteria, parasites and mammals: structure-activity relationships, *Trends in biochemical sciences*, **23**, 476-481.

Wasilenko, J., *et al.* (2005) Functional characterization of the K257R and G319E-hGALE alleles found in patients with ostensibly peripheral epimerase deficiency galactosemia, *Molecular genetics and metabolism*, **84**, 32-38.

Wermuth, B., *et al.* (1982) Purification and characterization of human-brain aldose reductase, *European journal of biochemistry / FEBS*, **127**, 279-284.

Wermuth, B. and von Wartburg, J.P. (1982) Aldose reductase from human tissues, *Methods in enzymology*, **89 Pt D**, 181-186.

Winans, K.A. and Bertozzi, C.R. (2002) An inhibitor of the human UDP-GlcNAc 4-epimerase identified from a uridine-based library: a strategy to inhibit O-linked glycosylation, *Chemistry & biology*, **9**, 113-129.

Wohlers, T.M. and Fridovich-Keil, J.L. (2000) Studies of the V94M-substituted human UDPgalactose-4-epimerase enzyme associated with generalized epimerase-deficiency galactosaemia, *Journal of inherited metabolic disease*, **23**, 713-729.

Yoshida, C., Shah, H. and Weinhouse, S. (1982) Purification and properties of inorganic pyrophosphatase of rat liver and hepatoma 3924A, *Cancer research*, **42**, 3526-3531.