Id	Name/Reaction	Information	Kinetics
GLUT2	Facilitated glucose transporter member 2	Mechanism	Km(3-O-MG)=42.3±4.1mM (human liver) (Gould, et al., 1991;
	D-glucose (disse) [$\mathbf{glc_dis}$] \leftrightarrow D-glucose	TCDB:2.A.1.1 (glucose transporter	Walmsley, et al., 1998)
	(cytosol) [glc]	subfamily)	$Km(3-O-Methyl glc)=17.3 \pm 4.3mM$ (rat liver) (Ciaraldi, et al.,
	D-galactose (disse) [gal_dis] \leftrightarrow D-galactose	Protein/Structure	1986)
	(cytosol) [gal]	UniProt: <u>P11168</u> (GTR2_HUMAN)	Km(D-glc)=21.7 ± 1.8mM (rat liver) (Ciaraldi, et al., 1986)
		Gene	Km(D-glc)=66±14mM (rat? hepatocytes) (Elliott and Craik, 1982)
	"Free galactose is transported into cells through	SLC2A2, GLUT2	
	a variety of transporters that are members of	Disease	Km(D-fruct) = 66mM (Walmsley, et al., 1998)
	the glucose transporter (GLUT) family.	MIM:227810 (Fanconi-Bickel	Km(D-fruct)=212±32mM (rat? hepatocytes) (Elliott and Craik,
	Deficient transport of galactose into	syndrome; FBS)	1982)
	hepatocytes is evident in human patients with		
	defective GLUT2 transporters (Fanconi-Bickel		Km(D-gal)=174±48mM (rat? hepatocytes) (Elliott and Craik, 1982)
	syndrome) resulting in galactose		Km(D-gal) > 50mM (GLUT2 enderocytes) (Walmsley, et al., 1998)
	malabsorption/intolerance (Brown, 2000;		$Km(D-gal) = 85.5 \pm 10.7 mM$ (human, liver-type GLUT2) (Colville,
	Leslie, 2003)."		et al., 1993)
	"GLUT2 is not completely specific for glucose		$Km(D-gal) = 92 \pm 8.4mM$ (human, liver-type GLUT2) (Arbuckle,
	and is probably also involved in galactose		et al., 1996)
	transport (Brown, 2000)."		
	"Data suggests that all transported sugars are		accumulation rate (human GLUT2):
	likely to use the same outward-facing sugar-		$v(deoxy-D-glc) = 4.33\pm0.15 \text{ pmol/min/oocyte}$
	binding site on the transporter molecule. This is		v(gal) = 1.68±0.09 pmol/min/oocyte
	further indicated by the observation that each		$v(\text{fru}) = 0.78 \pm 0.09 \text{ pmol/min/oocyte}$
	of the alternative substrates for each isoform is		
	a competitive inhibitor of deGlc and that D-		v(D-glc)=220±19mmol/min/l of cell H2O (rat? hepatocytes) (Elliott
	glucose is a competitive inhibitor of the		and Craik, 1982)
	transport of the alternative substrates. (Colville,		v(D-gal)=288±48 mmol/min/l of cell H2O (rat? hepatocytes) (Elliott

	et al., 1993)"		and Craik, 1982)
			v(D-fruct)=291±26 mmol/min/l of cell H2O (rat? hepatocytes)
			(Elliott and Craik, 1982)
			Competitive inhibitor kinetics (Colville, et al., 1993)
GALK	Galactokinase	Protein	SabioRK:P51570
	D-galactose [gal] + ATP [atp] ↔ D-galactose	UniProt:P51570 (GALK1_HUMAN	two-substrate ordered, ternary complex reaction (Timson and
	1-phosphate [gal1p] + ADP [adp])	Reece, 2003)
	EC:2.7.1.6	homodimer P51570*2	kcat(gal) = 8.7±5 1/s (SABIORK:14785)(Timson and Reece, 2003)
	RHEA:13556	Gene	km(atp) = 0.034±0.004mM (SABIORK:14792)(Timson and Reece,
	<u>KEGG:R01092</u>	GALK, GALK1	2003)
		Disease	km(gal)=0.97±0.22 mM (SABIORK:14785) (Timson and Reece,
		MIM:230200 (GALCT2	2003)
		Galactosemia II)	
			km(gal) = 0.436mM (SABIORK:45367), (Sangiuolo, et al., 2004)
			Uncompetitive product inhibition of GALK (adult rat liver) by
			gal1p with both 1mM and 5mM gal1p altering the Km for galactose
			from 0.150mM to 0.800mM. (Cuatrecasas and Segal, 1965)
			ki(gal1p) = 5.3mM (5.0-5.7mM) (Cuatrecasas and Segal, 1965)
			(1mM gal1p caused 15% inhibition, 5mM gal1p 50% inhibition)
			Galactokinase being rate limiting for galactose clearance (Schirmer,
			et al., 1986) -> [27,28, 50]
IMP	Inositol monophosphatase	Protein	Competitive inhibition model:
	D-galactose 1-phosphate [gal1p] ↔ D-	<u>UniProt:P29218</u> (IMPA1_HUMAN)	$km(ino1p) = 0.320\pm0.050mM$ (Slepak, et al., 2007)
	galactose [gal] + phosphate [pi]	Homodimer P29218*2	km(gal1p) = 0.35mM (similar kinetics gal1p to ino1p in vitro)

	IMPA1, IMPA	
		"Our kinetic analysis demonstrated that gal1p competitively
		inhibited human IMPase1 by increasing its Km for inositol-1p
		(ino1p) from 320±50μM to 980±70μM without changing the Vmax
		(Slepak, et al., 2007)."
Galactose-1-phosphate uridyl transferase	Protein	Mutation analysis (Quimby, et al., 1996)
UDP-D-glucose [udpglc] + D-galactose 1-	UniProt: <u>P07902</u> (GALT_HUMAN)	$km(gal1p) = 0.57\pm0.14mM$ (human, wildtype) (Quimby, et al.,
phosphate [gal1p] \leftrightarrow D-glucose 1-	homodimer P07902*2	1996)
phosphate [glc1p] + UDP-D-	Gene	km(udpglc) = 0.21±0.04mM (human, wildtype) (Quimby, et al.,
galactose [udpgal].	GALT	1996)
EC:2.7.7.12	Disease	
RHEA:13992	MIM:230400 (GALCT	Mutation analysis (Tang, et al., 2012)
<u>KEGG:R00955</u>	Galactosemia)	km(gal1p) = 1.25±0.36mM (human, wildtype) (Tang, et al., 2012)
		km(udpglc) = 0.43±0.09mM (human, wildtype) (Tang, et al., 2012)
The catalytic mechanism of GALT is ping-		
pong kinetics with covalent intermediate		(?species, 4°C) (Geeganage and Frey, 1998)
UMP-enzyme (Facchiano and Marabotti,		$km(udpglc) = 0.5\pm0.1 mM$
2010).		$v(glc1p) = 281 \pm 18 \ 1/s$
		$km(glc1p) = 0.37 \pm 0.18 mM$
		$v(glc1p) = 226 \pm 10 \text{ 1/s}$
		$km(gal1p) = 0.061 \pm 0.020 mM$
		$v(glc1p) = 166 \pm 13 \text{ 1/s}$
		Potent linear competent inhibitors UTP and UDP of UDP-
		glucose (Segal and Rogers, 1971):
		Ki(UTP) = 0.13mM (rat, liver)
	UDP-D-glucose [udpglc] + D-galactose 1- phosphate [gal1p] ↔ D-glucose 1- phosphate [glc1p] + UDP-D- galactose [udpgal]. EC:2.7.7.12 RHEA:13992 KEGG:R00955 The catalytic mechanism of GALT is ping- pong kinetics with covalent intermediate UMP-enzyme (Facchiano and Marabotti,	UDP-D-glucose [udpglc] + D-galactose 1- phosphate [gal1p] ↔ D-glucose 1- phosphate [glc1p] + UDP-D- galactose [udpgal]. EC:2.7.7.12 RHEA:13992 KEGG:R00955 The catalytic mechanism of GALT is ping- pong kinetics with covalent intermediate UMP-enzyme (Facchiano and Marabotti, UniProt:P07902 (GALT_HUMAN) homodimer P07902*2 Gene GALT Disease MIM:230400 (GALCT Galactosemia)

		Ki(UDP) = 0.35mM (rat, liver)
		Ki(UMP) = 2.3mM (rat, liver)
		Ki(UDP-glucuronic acid)=0.40mM (rat, liver)
UDP-glucose 4-epimerase	Protein	Mutation analysis(Timson, 2005)
UDP-D-glucose [udpglc] \leftrightarrow UDP-D-	UniProt:Q14376 (GALE_HUMAN)	km(udpgal)=0.069±0.012mM (human, wildtype) (Timson, 2005)
galactose [udpgal]	homodimer Q14376*2	$kcat(udpgal) = 36\pm1.4 \text{ 1/s} \text{ (human, wildtype) (Timson, 2005)}$
EC:5.1.3.2	Gene	
RHEA:22171	GALE	$km(udpgal) = 0.15 \pm 0.02mM$ (human, wildtype) (Wohlers and
KEGG:R00291	Disease	Fridovich-Keil, 2000)
	MIM:230350	km(udpgal, V94M) = 0.27 ± 0.01 mM (human, V94M) (Wohlers and
Alternative activity with ! GlcNAc ?		Fridovich-Keil, 2000)
UDP-GalNAc <-> UDP-GlcNAc		km(udpgal)=0.140± 0.007mM (human, wildtype)
		(SABIORK:19823) (Winans and Bertozzi, 2002)
		km(udpgal)=0.120± 0.04mM (human, wildtype)
		(SABIORK:46260) (Wasilenko, et al., 2005)
		kcat= 33.8±11.2 (human, wildtype) (SABIORK:16222) (Thoden, et
		al., 2002)
		$km(udpgal) = 0.230\pm0.06mM$ (human, wildtype)
		(SABIORK:46263) (Quimby, et al., 1997)
		"Ethanol treatment increases the NADH/NAD ratio in liver
		(Keppler, et al., 1970) -> [2-4] and by this inhibits the GALE -
		>[3,5]. Under these conditions oxidation and elimination ->[6-8] of
		galactose are impaired. Combined galactose+ethanol treatment
		results in accumulation of gallp and udpgal in rat liver. The
		formation of high amounts of udpgal leads to a change in the
		distribution of liver uracil nucleotides. A marked decrease of udpglc,
	UDP-D-glucose [udpglc] ↔ UDP-D-galactose [udpgal] EC:5.1.3.2 RHEA:22171 KEGG:R00291 Alternative activity with ! GlcNAc?	UDP-D-glucose [udpglc] ↔ UDP-D- galactose [udpgal] EC:5.1.3.2 RHEA:22171 KEGG:R00291 Alternative activity with ! GlcNAc ? UniProt:Q14376 (GALE_HUMAN) homodimer Q14376*2 Gene GALE Disease MIM:230350

	UDP-galactose pyrophosphorylase	"Previously, we showed that galactose-1-phosphate competitively	$ki(utp) = 0.643 \pm 0.047$ mM (competitive inhibition with respect to UDP-glc)
	(Chang, et al., 1996)	[Lai2002]	$km(utp) = 0.563 \pm 0.115 \text{ mM}$
	competitive inhibition with respect to UDP-Glc	yeast from ``galactose toxicity."	$km(glc1p) = 0.172\pm0.010 \text{ mM}$
	MgUTP is a product inhibitor that shows	uridyltransferase (GALT)-deficient	$km(pp) = 0.166 \pm 0.013 \text{ mM}$
	al., 1996)	rescued galactose-1-phosphate	$km(udpglc) = 0.049\pm0.004mM$
	simple Michaelis-Menten kinetics (Chang, et	pyrophosphorylase (hUGP2)	(human, liver, wildtype) (Duggleby, et al., 1996)
	in both reaction directions, the enzyme displays	transfection of human UDP-glucose	$V_{\text{fwd}}/V_{\text{rev}} = 0.260$
		"Previously we reported that stable	UDP-glc)
	<u>KEGG:R00289</u>		$ki(utp) = 0.477 \pm 41$ mM (competitive inhibition with respect to
	RHEA:19892	UGP2, UGP1	km(utp) = [0.563 - 0.692] mM
	EC:2.7.7.9	Gene	km(glc1p) = [0.172 - 0.174] mM
	UDP-glucose [udglc]+ diphosphate [pp]	homooctamer Q16851*8	km(pp) = [0.172 - 0.210] mM
UGALP	D-glucose 1-phosphate [glc1p] + UTP [utp] \leftrightarrow	UniProt:Q16851 (UGPA_HUMAN)	km(udpglc) = [0.031 - 0.051]mM
UGP	UDP-glucose pyrophosphorylase	Protein	(human, liver, wildtype) (Chang, et al., 1996)
			GALE (Keppler, et al., 1970)"
			contents in the liver, which are intensified by an inhibition of the
			"Galactose provokes pronounced alterations of the uracil nucleotide
			GALE (Keppler, et al., 1970)".
			a galactose load demonstrate the ethanol-induced inhibition of the
			stronger drop of the udpglc content in the ethanol treated liver avter
			"The almost 4-fold increase of gal1p and updgal and the even
			metabolites (Keppler, et al., 1970)".
			galactose metabolism in rat liver by the ratio of galactose
			"The GALE reaction is indicated as the rate-limiting step of
			nucleotides. (Keppler, et al., 1970)
			utp, udp and ump is followed by an increase of the sum of uracil

+ ↔ UDP-D-galactose [udpgal] pyrophosphate [pp] EC:2.7.7.10 RHEA:14212 KEGG:R00502 Could circumvent GALT deficiency (Isselbacher) Appears to have very low activity, several magnitudes lower than GALDH [Segal1968] Activity locacted in glucose dependent enzyme. 1-2 % activity with gallp (Km gal ~10-20mM)	inhibited UDP-glucose pyrophosphorylase, leading to 66% reduction in UDP- glucose/galactose contents in GALT-deficient cells under galactose challenge [Slepak2007- >Lai2002]." The formation of UDP-glucose is the major physiological function of UGP, however at slow rates, the enzyme also catalyzes the phosphorylation of UDP-galactose. (Knop and Hansen, 1970) "Under normal physiological conditions, this may not be significant, but, in a galactosemic patient, in which the normal galactose metabolism is impaired, the pyrophosphorylase may participate in an abnormal role.	ki(udpglc) = 0.013± 4 mM (competitive inhibition with respect to UTP?) (human, liver, wildtype) (Knop and Hansen, 1970) keq([udpglc][pp]/([UTP][glc1p])) = 0.15 – 0.16 km(udpglc) = 50mM km(utp) = 48 mM km(glc1p) = 95±10 mM keq([UTP][glc1p]/([udpglc][pp])) = 4.55±0.1 (Guynn, et al., 1974) (0.22) The saturating concentration for UDP-galactose is 10 times that of UDP-glucose: km(udpgal) = 10*km(udpglc) ~ 0.5mM (human, liver, wildtype) (Knop and Hansen, 1970) km(udpgal) = 0.420mM (rabbit, liver, wildtype) (Turnquist, et al., 1974) udpgal was an adequate substrate at 10 times the concentration ofudpglc, showing 14.3% of udpglc (Calf) and 12.0% (Human). activity with udpgal 2-12% of udpglc (12% with 3mM udpgal) (human liver) (Turnquist, et al., 1974) The activity of UDPG:galactose-1-phosphate uridylyltransferase from rat liver under optimal conditions in vitro is less than 5% of the UDPG pyrophosphorylase activity (Keppler, et al., 1970)[Keppler1970 ->39,40]" SabioRK:P15121
	<u>UniProt:P15121</u> (ALDR_HUMAN)	km(gal) = 40.0mM (human brain) (SABIORK: 22893) (Wermuth, et

	D-galactose [gal] + NADPH [nadph] + H ↔	monomer P15121*1	al., 1982)
	galactitol [galtol] + NADP [nadp]	Gene	kcat(gal) = 0.40 1/s (human brain) (SABIORK: 22893) (Wermuth,
	EC:1.1.1.21	AKR1B1, ALDR1	et al., 1982)
	RHEA:12792 -> RHEA:37967		
	<u>KEGG:R01095</u>	Tissue distribution: Aldolase	km(gal) = 110.0mM (human brain) (SABIORK:15695) (Wermuth
	Aldolase reductase is specific for NADPH as	reductase activity has been	and von Wartburg, 1982)
	cofactor (NADH ~10% of NADPH-dependent	demonstrated in brain, kidney,	
	activity) (Wermuth and von Wartburg, 1982).	placenta, testis, lens, lung, heart,	
		and pancreas, but appears to be	
	"Aldolase reductase catalyzes the conversion of	absent from the liver. The enzyme is	
	aldoses and a number of other aldehydes to the	localized in the cytoplasm of the	
	corresponding alcohol metabolites. It is one of	cell. [Wermuth1982].	
	several cytosolic, monomeric, NADPH-		
	dependent aldehyde and ketone reductases of		
	wide substrate specificity (Wermuth, et al.,		
	1982)".		
PGM1	Phosphoglucomutase-1	Multiple isoforms (PGM1, PGM2)	The equilibrium lies strongly toward G6P (Keq) and the reaction
	D-glucose 1-phosphate [$\mathbf{glc1p}$] \leftrightarrow D-glucose 6-	Protein	proceeds through a ping-pong mechanism involving aspartyl-
	phosphate [glc6p]	UniProt:P36871 (PGM1_HUMAN)	phosphoenzyme and glucose-1,6-bisphosphate intermediates.
	EC:5.4.2.2	monomer P36871*1	
	<u>KEGG:R00959</u>	main isoform for glc1p ↔ glc6p	[glc6p]/[glc1p] ~10-12 (Guynn, et al., 1974)
	RHEA:23539	reaction	DeltaG =-7.1 kJ/mol [Koenig2012]
		Gene	
		PGM1	km(glc1p) = 0.049mM (human, RBC) (Quick, et al., 1974)
		Disease	
		Glycogen storage disease 14	km(glc1p) = 0.045mM (rat, heart) (Kashiwaya, et al., 1994)
		(GSD14) [MIM: <u>612934</u>]	km(glc6p) = 0.67mM (rat, heart) (Kashiwaya, et al., 1994)

		1T (CDG1T) [MIM:614921]: A multisystem disorder caused by a defect in glycoprotein biosynthesis and characterized by underglycosylated serum glycoproteins.	km(glc1p) = 0.083mM (human, RBC, PGM1) (Accorsi, et al., 1989) ki(fru16bp) = 0.092mM (human, RBC, PGM1) (Accorsi, et al., 1989) The kinetic properties of PGM1 and PGM2 are essentially the same. PGM1 is specific for mutation of glucose, whereas PGM2 also has phosphoribomutase activities. (human, RBC) (Accorsi, et al., 1989)
		defect in glycoprotein biosynthesis and characterized by underglycosylated serum glycoproteins.	1989) The kinetic properties of PGM1 and PGM2 are essentially the same. PGM1 is specific for mutation of glucose, whereas PGM2 also has
		and characterized by under- glycosylated serum glycoproteins.	The kinetic properties of PGM1 and PGM2 are essentially the same. PGM1 is specific for mutation of glucose, whereas PGM2 also has
		glycosylated serum glycoproteins.	PGM1 is specific for mutation of glucose, whereas PGM2 also has
			phosphorihamutase activities (human PBC) (Accorsi et al. 1080)
			phosphoriounidase activities. (human, RDC) (Accorsi, et al., 1989)
		Protein	
		UniProt:Q96G03	
		(PGM2_HUMAN)	
		Gene	
		PGM2	
PPASE	Pyrophosphatase	Protein	km(pp) = 0.005mM (rat liver) (Yoshida, et al., 1982)
	Pyrophosphate $[\mathbf{pp}] + H2O \rightarrow 2$ phosphate $[\mathbf{pi}]$	<u>UniProt:Q15181</u> (IPYR_HUMAN)	km(pp) = 0.14mM (human erythrocyte) (Thuillier, 1978)
	EC:3.6.1.1	homodimer Q15181*2	km(pp) = 0.07mM (rat liver) (Irie, et al., 1970)
	RHEA:24579	Gene	Delta G0 = -23.56 kJ/mol (Thuillier, 1978)
	KEGG:R00004	PPA1, IOPPP, PP	Delta G0 = -19.2 kJ/mol (Guynn, et al., 1974)
NDKU	Nucleoside diphosphokinase (ATP:UDP	Multiple isoforms	Compulsory-order substituted-enzyme (Ping Pong Bi Bi)
	phosphotransferase)		mechanism (Lam and Packham, 1986)
	$ATP [atp] + UDP [udp] \leftrightarrow ADP [adp] +$		km(atp) = 0.38mM (human, platelets) (Lam and Packham, 1986)
	UTP [udp]		km(adp) = 0.024mM (human, platelets) (Lam and Packham, 1986)
	EC: 2.7.4.6		km(gtp) = 0.12mM (human, platelets) (Lam and Packham, 1986)
	RHEA:25101		
	KEGG:R00156		km(atp) = 1.33mM (rat, liver) (Kimura and Shimada, 1988)
			km(adp) = 0.042mM (rat, liver) (Kimura and Shimada, 1988)
1			km(udp) = 0.19mM(rat, liver) (Kimura and Shimada, 1988)
NDKU	RHEA:24579 KEGG:R00004 Nucleoside diphosphokinase (ATP:UDP phosphotransferase) ATP [atp] + UDP [udp] ↔ ADP [adp] + UTP [udp] EC: 2.7.4.6 RHEA:25101	Gene PPA1, IOPPP, PP	Delta G0 = -23.56 kJ/mol (Thuillier, 1978) Delta G0 = -19.2 kJ/mol (Guynn, et al., 1974) Compulsory-order substituted-enzyme (Ping Pong Bi Bi) mechanism (Lam and Packham, 1986) km(atp) = 0.38mM (human, platelets) (Lam and Packham, I km(adp) = 0.024mM (human, platelets) (Lam and Packham, km(gtp) = 0.12mM (human, platelets) (Lam and Packham, I km(atp) = 1.33mM (rat, liver) (Kimura and Shimada, 1988) km(adp) = 0.042mM (rat, liver) (Kimura and Shimada, 1988)

			km(atp) = 1.80 mM (rat, liver) (Fukuchi, et al., 1994)
			km(adp) = 0.066 mM (rat, liver) (Fukuchi, et al., 1994)
			km(utp) = 27.00mM (rat, liver) (Fukuchi, et al., 1994)
			km(gtp) = 0.15mM (rat, liver) (Fukuchi, et al., 1994)
			km(gdp) = 0.049mM (rat, liver) (Fukuchi, et al., 1994)
NADPR	NADP reductase	Protein	Delta G0 = -19.6 kJ/mol [Schuster1995]
	NADP [nadp] + H2→NADPH [nadph]	UniProt:P11413 (G6PD_HUMAN)	
		homotetramer (dimer of dimer)	$km(glc6p) = 0.040\pm0.008 \text{ mM}$ (human, placenta) (Ozer, et al., 2001)
	Modeled via pentose phosphate pathway	P11413*4	$km(nadp) = 0.020\pm0.010 \text{ mM}$ (human, placenta) (Ozer, et al., 2001)
	glucose-6-phosphate dehydrogenase	Gene	ki(nadph) = 0.0171±0.0032 mM (human, placenta) (Ozer, et al.,
	D-glucose 6-phosphate [glc6p] +	G6PD	2001)
	NADP [nadp] → 6-phospho-D-glucono-1,5-		
	lactone + NADPH [nadph] + H		km(glc6p) = 0.072 mM (human, RBC) (Bautista, et al., 1992)
	EC: 1.1.1.49		$km(glc6p) = 0.069\pm0.003 \text{ mM}$ (human, recombinant) (Bautista, et
	RHEA:15844		al., 1992)
	<u>KEGG:R00835</u>		km(nadp) = 0.013 mM (human, RBC) (Bautista, et al., 1992)
			$km(nadp) = 0.012\pm0.002 \text{ mM}$ (human, recombinant) (Bautista, et
			al., 1992)
			$km(nadph) = 0.015\pm0.002 \text{ mM}$ (human, RBC) (Bautista, et al.,
			1992)
			$km(nadph) = 0.014\pm0.003 \text{ mM}$ (human, recombinant) (Bautista, et
			al., 1992)
			km(glc6p) = 0.326mM (rat, liver)
			km(glc6p) = 0.157mM (rat, liver)
			(Corpas, et al., 1995; Corpas, et al., 1995)
			km(nadp) = 0.108 mM (rat, liver)

km(nadp) = 0.258 mM (rat, liver) (Corpas, et al., 1995; Corpas, et
al., 1995)
ki(nadhp) = 0.010 mM(rat, liver)
ki(nadhp) = 0.021 mM(rat, liver) (Corpas, et al., 1995; Corpas, et al., 1995; Corp
al., 1995)

References

Accorsi, A., et al. (1989) Isoenzymes of phosphoglucomutase from human red blood cells: isolation and kinetic properties, *Preparative biochemistry*, **19**, 251-271.

Arbuckle, M.I., *et al.* (1996) Structure-function analysis of liver-type (GLUT2) and brain-type (GLUT3) glucose transporters: expression of chimeric transporters in Xenopus oocytes suggests an important role for putative transmembrane helix 7 in determining substrate selectivity, *Biochemistry*, **35**, 16519-16527.

Bautista, J.M., Mason, P.J. and Luzzatto, L. (1992) Purification and properties of human glucose-6-phosphate dehydrogenase made in E. coli, *Biochimica et biophysica acta*, **1119**, 74-80.

Brown, G.K. (2000) Glucose transporters: structure, function and consequences of deficiency, *Journal of inherited metabolic disease*, **23**, 237-246.

Chang, H.Y., et al. (1996) The importance of conserved residues in human liver UDPglucose pyrophosphorylase, European journal of biochemistry / FEBS, 236, 723-728.

Ciaraldi, T.P., Horuk, R. and Matthaei, S. (1986) Biochemical and functional characterization of the rat liver glucose-transport system. Comparisons with the adipocyte glucose-transport system, *The Biochemical journal*, **240**, 115-123.

Colville, C.A., *et al.* (1993) Kinetic analysis of the liver-type (GLUT2) and brain-type (GLUT3) glucose transporters in Xenopus oocytes: substrate specificities and effects of transport inhibitors, *The Biochemical journal*, **290** (**Pt 3**), 701-706.

Corpas, F.J., et al. (1995) Kinetic properties of hexose-monophosphate dehydrogenases. II. Isolation and partial purification of 6-phosphogluconate dehydrogenase from rat liver and kidney cortex, *Molecular and cellular biochemistry*, **144**, 97-104.

Corpas, F.J., *et al.* (1995) Kinetic properties of hexose-monophosphate dehydrogenases. I. Isolation and partial purification of glucose-6-phosphate dehydrogenase from rat liver and kidney cortex, *Life sciences*, **56**, 179-189.

Cuatrecasas, P. and Segal, S. (1965) Mammalian Galactokinase. Developmental and Adaptive Characteristics in the Rat Liver, *The Journal of biological chemistry*, **240**, 2382-2388.

Duggleby, R.G., *et al.* (1996) Sequence differences between human muscle and liver cDNAs for UDPglucose pyrophosphorylase and kinetic properties of the recombinant enzymes expressed in Escherichia coli, *European journal of biochemistry / FEBS*, **235**, 173-179.

Elliott, K.R. and Craik, J.D. (1982) Sugar transport across the hepatocyte plasma membrane, *Biochemical Society transactions*, 10, 12-13.

Facchiano, A. and Marabotti, A. (2010) Analysis of galactosemia-linked mutations of GALT enzyme using a computational biology approach, *Protein engineering, design & selection : PEDS*, **23**, 103-113.

Fukuchi, T., et al. (1994) Recombinant rat nucleoside diphosphate kinase isoforms (alpha and beta): purification, properties and application to immunological detection of native isoforms in rat tissues, *Biochimica et biophysica acta*, **1205**, 113-122.

Geeganage, S. and Frey, P.A. (1998) Transient kinetics of formation and reaction of the uridylyl-enzyme form of galactose-1-P uridylyltransferase and its Q168R-variant: insight into the molecular basis of galactosemia, *Biochemistry*, **37**, 14500-14507.

Gould, G.W., et al. (1991) Expression of human glucose transporters in Xenopus oocytes: kinetic characterization and substrate specificities of the erythrocyte, liver, and brain isoforms, *Biochemistry*, **30**, 5139-5145.

Guynn, R.W., et al. (1974) The concentration and control of cytoplasmic free inorganic pyrophosphate in rat liver in vivo, *The Biochemical journal*, **140**, 369-375.

Irie, M., et al. (1970) Distribution and properties of alkaline pyrophosphatases of rat liver, Journal of biochemistry, 67, 47-58.

Kashiwaya, Y., et al. (1994) Control of glucose utilization in working perfused rat heart, *The Journal of biological chemistry*, **269**, 25502-25514.

Keppler, D., Rudigier, J. and Decker, K. (1970) Trapping of uridine phosphates by D-galactose in ethanol-treated liver, *FEBS letters*, **11**, 193-196.

Keppler, D.O., et al. (1970) The trapping of uridine phosphates by D-galactosamine. D-glucosamine, and 2-deoxy-D-galactose. A study on the mechanism of galactosamine hepatitis, European journal of biochemistry / FEBS, 17, 246-253.

Kimura, N. and Shimada, N. (1988) Membrane-associated nucleoside diphosphate kinase from rat liver. Purification, characterization, and comparison with cytosolic enzyme, *The Journal of biological chemistry*, **263**, 4647-4653.

Knop, J.K. and Hansen, R.G. (1970) Uridine diphosphate glucose pyrophosphorylase. IV. Crystallization and properties of the enzyme from human liver, *The Journal of biological chemistry*, **245**, 2499-2504.

Lam, S.C. and Packham, M.A. (1986) Isolation and kinetic studies of nucleoside diphosphokinase from human platelets and effects of cAMP phosphodiesterase inhibitors, *Biochemical pharmacology*, **35**, 4449-4455.

Leslie, N.D. (2003) Insights into the pathogenesis of galactosemia, *Annual review of nutrition*, 23, 59-80.

Ozer, N., Aksoy, Y. and Ogus, I.H. (2001) Kinetic properties of human placental glucose-6-phosphate dehydrogenase, *The international journal of biochemistry & cell biology*, **33**, 221-226.

Parthasarathy, R., Parthasarathy, L. and Vadnal, R. (1997) Brain inositol monophosphatase identified as a galactose 1-phosphatase, *Brain research*, **778**, 99-106.

Quick, C.B., Fisher, R.A. and Harris, H. (1974) A kinetic study of the isozymes determined by the three human phosphoglucomutase loci PGM1, PGM2, and PGM3, *European journal of biochemistry / FEBS*, **42**, 511-517.

Quimby, B.B., et al. (1997) Characterization of two mutations associated with epimerase-deficiency galactosemia, by use of a yeast expression system for human UDP-galactose-4-epimerase, *American journal of human genetics*, **61**, 590-598.

Quimby, B.B., et al. (1996) Functional requirements of the active site position 185 in the human enzyme galactose-1-phosphate uridylyltransferase, *The Journal of biological chemistry*, **271**, 26835-26842.

Sangiuolo, F., et al. (2004) Biochemical characterization of two GALK1 mutations in patients with galactokinase deficiency, *Human mutation*, **23**, 396.

Schirmer, W.J., et al. (1986) Galactose clearance as an estimate of effective hepatic blood flow: validation and limitations, *The Journal of surgical research*, **41**, 543-556.

Segal, S. and Rogers, S. (1971) Nucleotide inhibition of mammalian liver galactose-I-phosphate uridylyltransferase, *Biochimica et biophysica acta*, **250**, 351-360.

Slepak, T.I., et al. (2007) Involvement of endoplasmic reticulum stress in a novel Classic Galactosemia model, *Molecular genetics and metabolism*, **92**, 78-87.

Tang, M., et al. (2012) Correlation assessment among clinical phenotypes, expression analysis and molecular modeling of 14 novel variations in the human galactose-1-phosphate uridylyltransferase gene, *Human mutation*, **33**, 1107-1115.

Thoden, J.B., et al. (2002) Structural analysis of the Y299C mutant of Escherichia coli UDP-galactose 4-epimerase. Teaching an old dog new tricks, *The Journal of biological chemistry*, **277**, 27528-27534.

Thuillier, L. (1978) Purification and kinetic properties of human erythrocyte Mg2+-dependent inorganic pyrophosphatase, *Biochimica et biophysica acta*, **524**, 198-206.

Timson, D.J. (2005) Functional analysis of disease-causing mutations in human UDP-galactose 4-epimerase, *The FEBS journal*, **272**, 6170-6177.

Timson, D.J. and Reece, R.J. (2003) Functional analysis of disease-causing mutations in human galactokinase, *European journal of biochemistry / FEBS*, **270**, 1767-1774.

Turnquist, R.L., et al. (1974) Uridine diphosphate glucose pyrophosphorylase: differential heat inactivation and further characterization of human liver enzyme, *Biochimica et biophysica acta*, **364**, 59-67.

Walmsley, A.R., et al. (1998) Sugar transporters from bacteria, parasites and mammals: structure-activity relationships, *Trends in biochemical sciences*, **23**, 476-481.

Wasilenko, J., et al. (2005) Functional characterization of the K257R and G319E-hGALE alleles found in patients with ostensibly peripheral epimerase deficiency galactosemia, *Molecular genetics and metabolism*, **84**, 32-38.

Wermuth, B., et al. (1982) Purification and characterization of human-brain aldose reductase, European journal of biochemistry / FEBS, 127, 279-284.

Wermuth, B. and von Wartburg, J.P. (1982) Aldose reductase from human tissues, Methods in enzymology, 89 Pt D, 181-186.

Winans, K.A. and Bertozzi, C.R. (2002) An inhibitor of the human UDP-GlcNAc 4-epimerase identified from a uridine-based library: a strategy to inhibit O-linked glycosylation, *Chemistry & biology*, **9**, 113-129.

Wohlers, T.M. and Fridovich-Keil, J.L. (2000) Studies of the V94M-substituted human UDPgalactose-4-epimerase enzyme associated with generalized epimerase-deficiency galactosaemia, *Journal of inherited metabolic disease*, **23**, 713-729.

Yoshida, C., Shah, H. and Weinhouse, S. (1982) Purification and properties of inorganic pyrophosphatase of rat liver and hepatoma 3924A, *Cancer research*, **42**, 3526-3531.