

# Lung function

Michael Cho

October 7, 2012

## Contents

<b>I</b>	<b>COPDGene</b>	<b>1</b>
1	Phenotypes	2
2	Qqplots	3
<b>II</b>	<b>ECLIPSE</b>	<b>10</b>
3	Phenotypes	10
4	Qqplots	12
<b>III</b>	<b>Norway</b>	<b>19</b>
5	Phenotypes	19
6	Qqplots	20
<b>IV</b>	<b>Meta-analysis</b>	<b>27</b>

```
options(stringsAsFactors = F)
.libPaths("/udd/remhc/bin/R-2.15.0/library")
source("/udd/remhc/bin/gwasFunctions.r")
source("http://faculty.ucr.edu/~tgirke/Documents/R_BioCond/My_R_Scripts/vennDia.R")
require(xtable)
```

# Part I

## COPDGene

Run in recge00. Files created are sAa.txt and sNhw.txt.

### 1 Phenotypes

```
nhw <- read.table("sNhw.txt", header = T, na = "-9")
summary(nhw$fev1pp_utah)
```

##	Min.	1st Qu.	Median	Mean	3rd Qu.	Max.	NA's
##	8.9	54.8	77.9	73.5	93.1	151.0	19

```
summary(nhw$FEV1_FVC_utah)
```

##	Min.	1st Qu.	Median	Mean	3rd Qu.	Max.	NA's
##	0.15	0.53	0.69	0.64	0.77	1.00	19

```
summary(nhw$FEV1_utah)
```

##	Min.	1st Qu.	Median	Mean	3rd Qu.	Max.	NA's
##	0.22	1.50	2.20	2.21	2.86	5.50	19

```
summary(nhw$fev1pp_utah[nhw$finalgold %in% c(2, 3, 4)])
```

##	Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
##	8.9	34.7	50.1	49.6	65.6	79.9

```
summary(nhw$FEV1_FVC_utah[nhw$finalgold %in% c(2, 3, 4)])
```

##	Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
##	0.150	0.380	0.490	0.486	0.600	0.690

```
summary(nhw$FEV1_utah[nhw$finalgold %in% c(2, 3, 4)])
```

##	Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
##	0.220	0.949	1.380	1.450	1.870	3.570

QQPlots and lambdas.

## 2 Qqplots

```
makePlot <- FALSE
qqNames <- as.vector(sapply(c("fev1", "fevip", "ratio"), function(x) paste(x,
  c("Nh", "NhCa"), sep = "")))
for (n in qqNames) {
  if (makePlot) {
    pdf(paste("lungFx-Cg", n, ".pdf", sep = ""))
    tab <- read.table(paste("Cg/", n, "All.assoc.linear.gz", sep = ""),
      header = T, colClasses = c(rep("NULL", 8), "numeric", rep("NULL",
        5), "numeric"))
    fMyQqplot(tab)
    dev.off()
  }
  cat(paste("\\begin{figure}\\n\\caption{", n, "}\\n\\includegraphics{lungFx-Cg",
    n, ".pdf}\\n\\end{figure}\\n", sep = ""))
}
```

Figure 1: fev1Nhw

**Lambda = 1.02258 :**

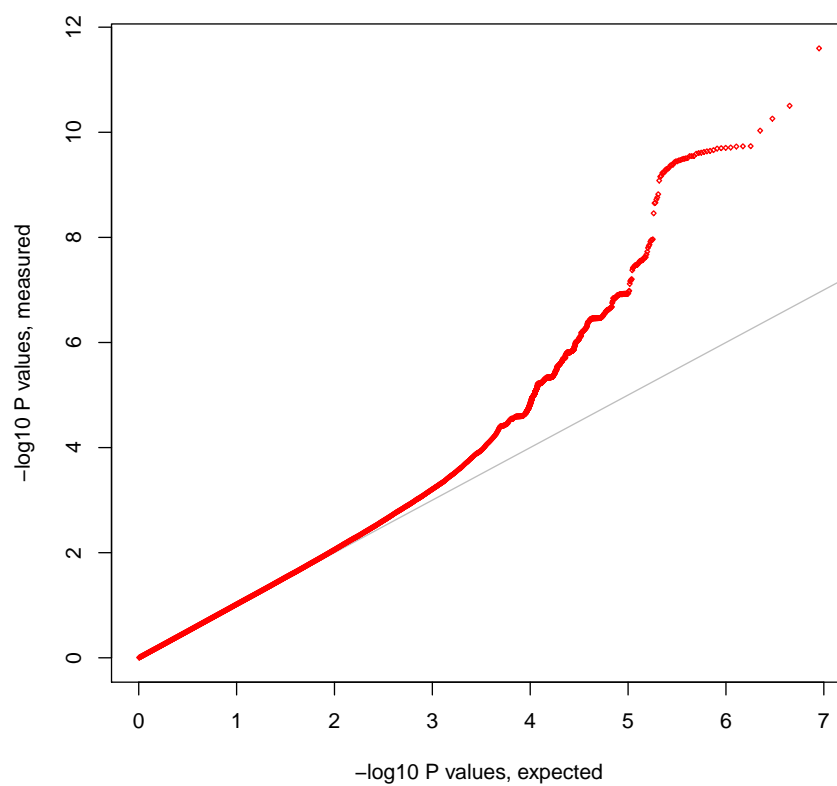


Figure 2: fev1NhwCa

**Lambda = 0.99674 :**

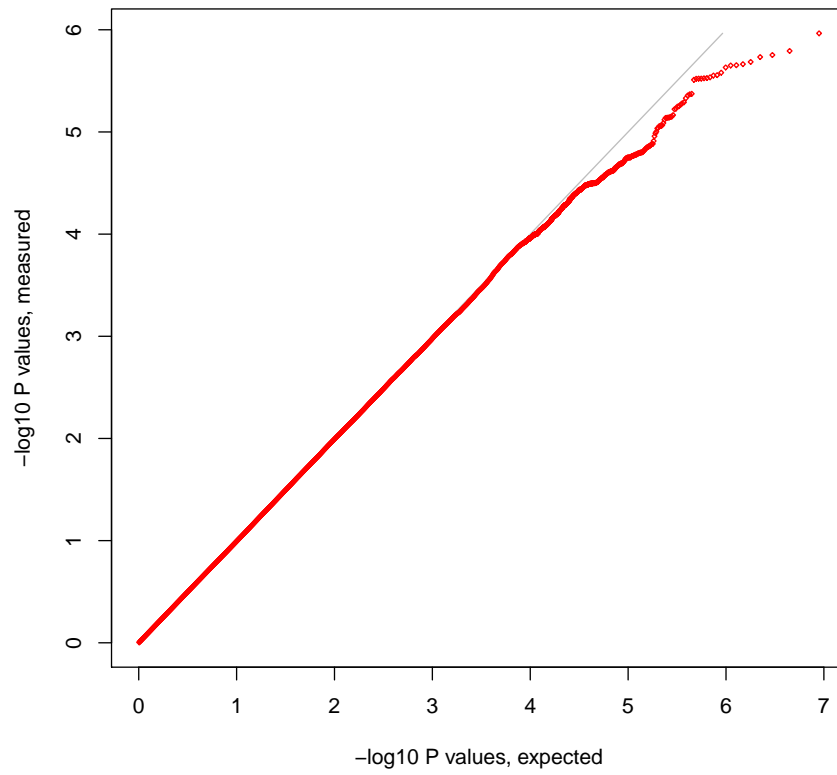


Figure 3: fev1ppNhw

**Lambda = 1.02495 :**

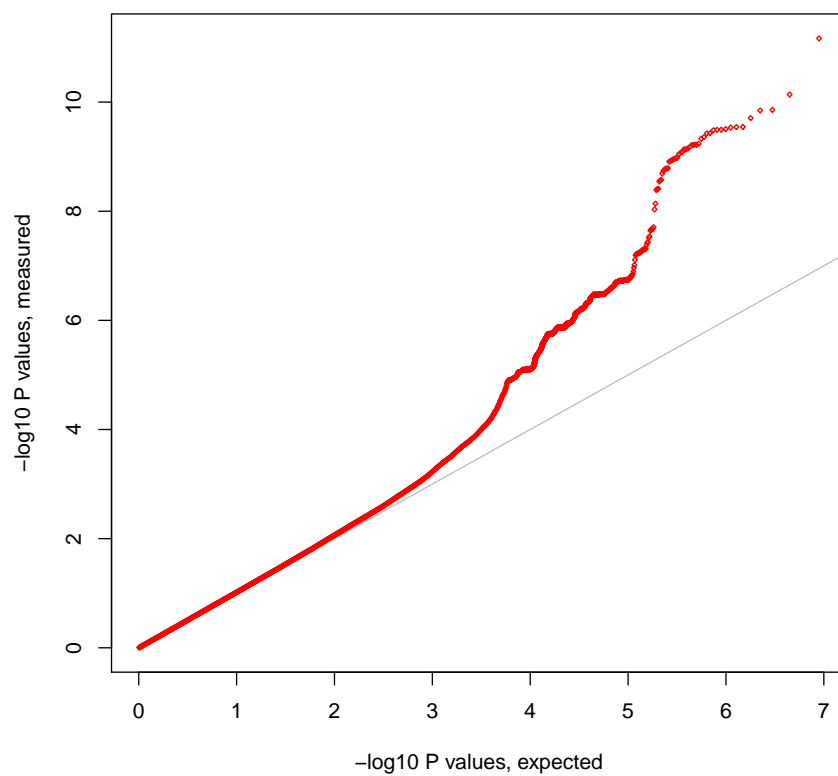


Figure 4: fev1ppNhwCa

**Lambda = 1.00047 :**

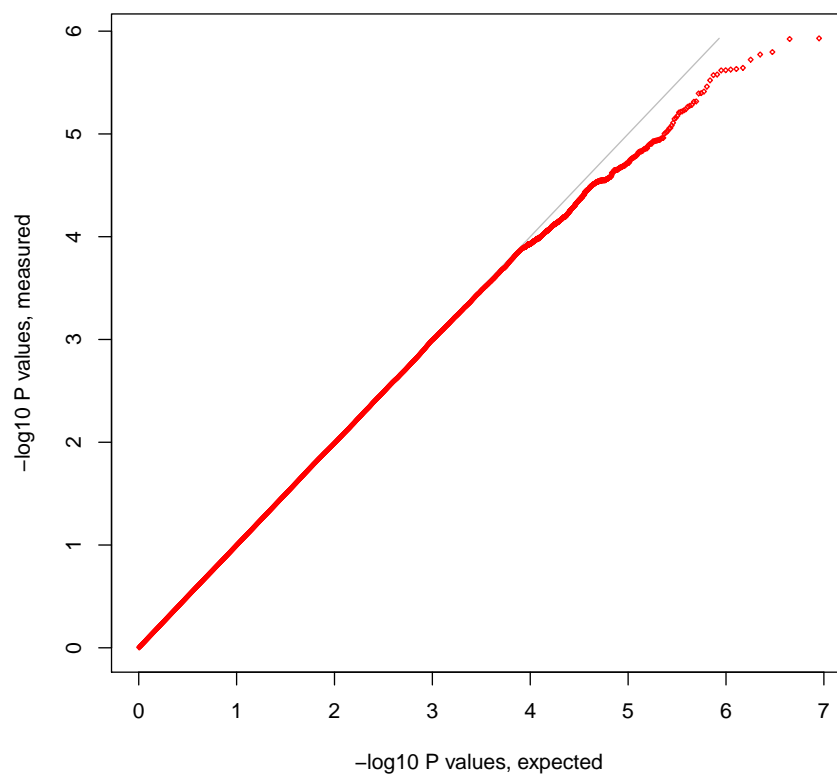


Figure 5: ratioNhw

**Lambda = 1.02543 :**

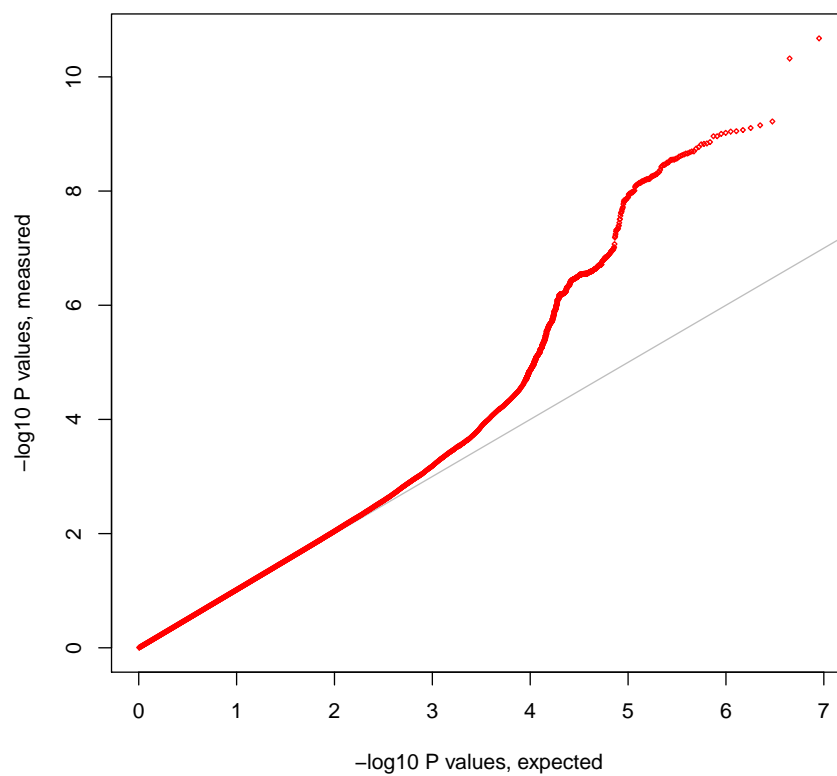
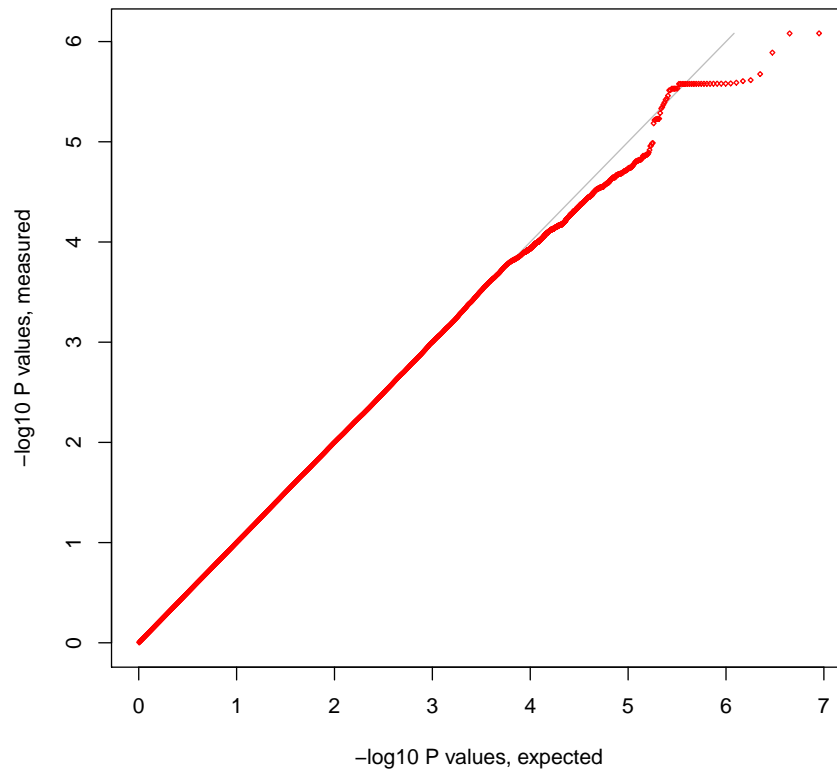




Figure 6: ratioNhwCa

**Lambda = 1.0028 :**



## Part II

# ECLIPSE

### 3 Phenotypes

```
eclCohort <- read.table("~/GWAS/eclipsePrune01IterCovPcas.txt", header = T,
  na.strings = "-9")
temp <- read.csv("~/GWAS/ECLIPSEFiles/genetics_phenotype20090810.csv")
temp <- temp[, c("Global.Subject.ID", "Forced.expiratory.volume..1.second...L.",
  "Post.dose.FEV1", "Height..cm.")]
names(temp) <- c("FID", "preFev1", "fev1", "height")
temp$FID <- paste("EC", temp$FID, sep = "")
any(is.na(eclCohort$post_fev1pp2))

## [1] TRUE

## use the postbd values to keep the sample size the same
eclCohort$post_fev1pp2 <- ifelse(is.na(eclCohort$post_fev1pp), eclCohort$pre_fev1pp,
  eclCohort$post_fev1pp)
any(is.na(eclCohort$post_fev1pp2))

## [1] FALSE

eclCohort$post_ratio2 <- ifelse(is.na(eclCohort$post_ratio), eclCohort$pre_ratio,
  eclCohort$post_ratio)
eclCohort <- merge(eclCohort, temp, by = "FID", all.x = T)
eclCohort$fev1 <- ifelse(is.na(eclCohort$fev1), eclCohort$preFev1, eclCohort$fev1)
##
## write.table(eclCohort, 'eclCohortLungFx.txt', row.names=F, quote=F, na='-9')
eclCase <- read.table("~/GWAS/eclipseCovPcsCases.txt", header = T, na.strings = "-9")
eclCase$post_fev1pp2 <- ifelse(is.na(eclCase$post_fev1pp), eclCase$pre_fev1pp,
  eclCase$post_fev1pp)
any(is.na(eclCase$post_fev1pp2))

## [1] FALSE

eclCase$post_ratio2 <- ifelse(is.na(eclCase$post_ratio), eclCase$pre_ratio,
  eclCase$post_ratio)
eclCase <- merge(eclCase, temp, by = "FID", all.x = T)
eclCase$fev1 <- ifelse(is.na(eclCase$fev1), eclCase$preFev1, eclCase$fev1)
## write.table(eclCase, 'eclCaseLungFx.txt', row.names=F, quote=F, na='-9')
```

Show the distribution of the phenotypes.

```
ecl <- read.table("eclCohortLungFx.txt", header = T, na = "-9")
ecl <- ecl[!is.na(ecl$pca1), ]
summary(ecl$post_fev1pp2)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##      13.4   36.9   48.6   53.2   64.7   147.0
```

```
summary(ecl$post_ratio2)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##      0.172  0.367   0.458   0.478   0.569   0.986
```

```
summary(ecl$fev1)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##      0.387  0.964   1.320   1.510   1.840   5.300
```

```
ecl <- read.table("eclCaseLungFx.txt", header = T, na = "-9")
summary(ecl$post_fev1pp2)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##      13.4   35.5   46.3   47.6   59.8   80.6
```

```
summary(ecl$post_ratio2)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##      0.172  0.359   0.441   0.447   0.535   0.839
```

```
summary(ecl$fev1)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##      0.387  0.938   1.250   1.330   1.650   3.320
```

```
## eclFev1pp.param
```

```
/proj/reglms/regln00/GWAS/Imputation/eclipse
1000g
```

```
--bfile /proj/reglms/regln00/GWAS/COPD_ECLIPSE_2009sorted
```

```
--linear
```

```
--covar eclCohortLungFx.txt
```

```
--covar-name pca1-pca8,age,py
```

```
--sex
```

```
--pheno eclCohortLungFx.txt
```

```
--pheno-name post_fev1pp2
```

```
--out eclFev1pp
```

```
## same covariates; alternative outcomes are post_ratio (eclRatio) and fev1 (eclFev1) with the
## for cases
```

```
/proj/reglms/regln00/GWAS/Imputation/eclipse
```

```

1000g
--bfile /proj/reglms/regln00/GWAS/COPD_ECLIPSE_2009sorted
--linear
--covar eclCaseLungFx.txt
--covar-name pca1-pca6,age,py
--sex
--pheno eclCaseLungFx.txt
--pheno-name post_fev1pp2
--out eclCaFev1pp

```

## 4 Qqplots

ECLIPSE Qq plots etc.

```

makePlot <- FALSE
qqNames <- as.vector(sapply(c("Fev1", "Fev1pp", "Ratio"), function(x) paste(c("Ca",
  "Ca"), x, sep = "")))
for (n in qqNames) {
  if (makePlot) {
    pdf(paste("lungFx-ecl", n, ".pdf", sep = ""))
    tab <- read.table(paste("ecl", n, ".assoc.linear.gz", sep = ""), header = T,
      colClasses = c(rep("NULL", 8), "numeric", rep("NULL", 5), "numeric"))
    fMyQqplot(tab)
    dev.off()
  }
  cat(paste("\\begin{figure}\\n\\caption{" , n, "}\\n\\includegraphics{lungFx-ecl",
    n, ".pdf}\\n\\end{figure}\\n", sep = ""))
}

```

Figure 7: Fev1

**Lambda = 1.00842 :**

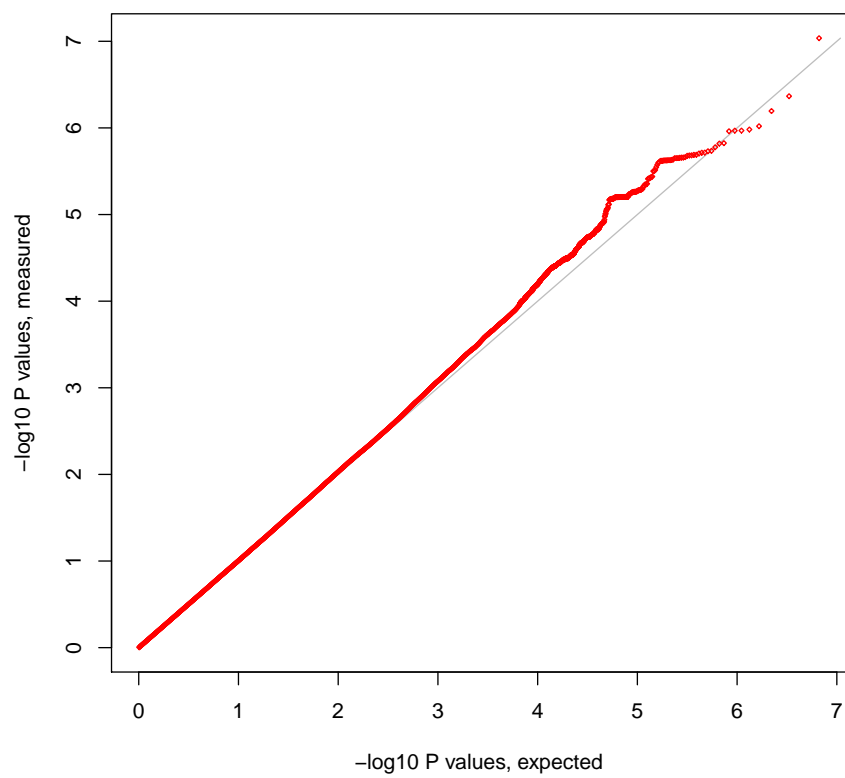


Figure 8: CaFev1

**Lambda = 0.99348 :**

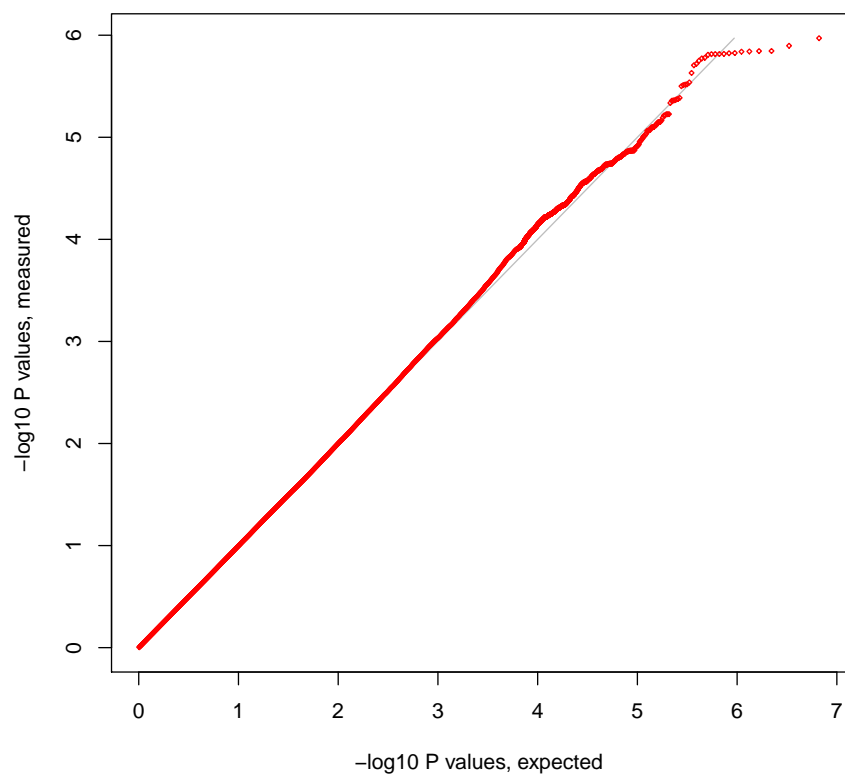


Figure 9: Fev1pp

**Lambda = 1.00655 :**

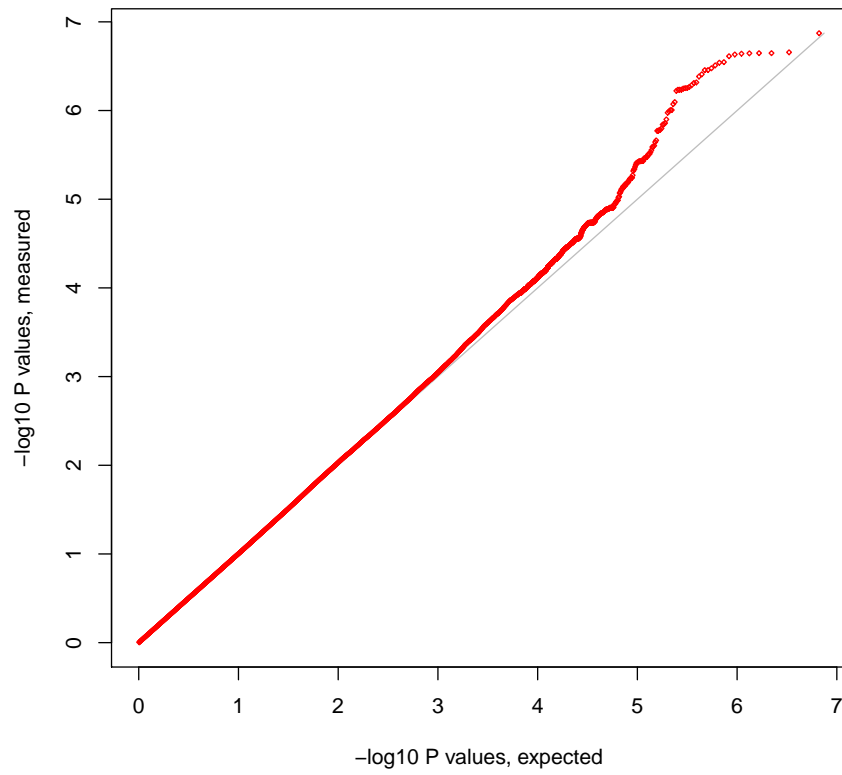


Figure 10: CaFev1pp

**Lambda = 0.99209 :**

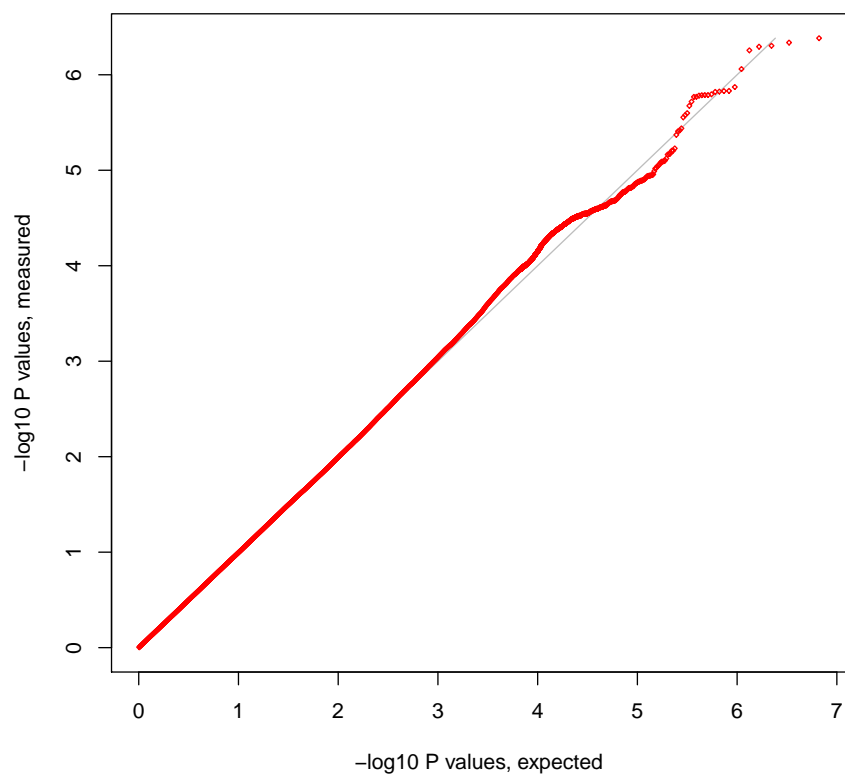




Figure 11: Ratio

**Lambda = 1.00187 :**

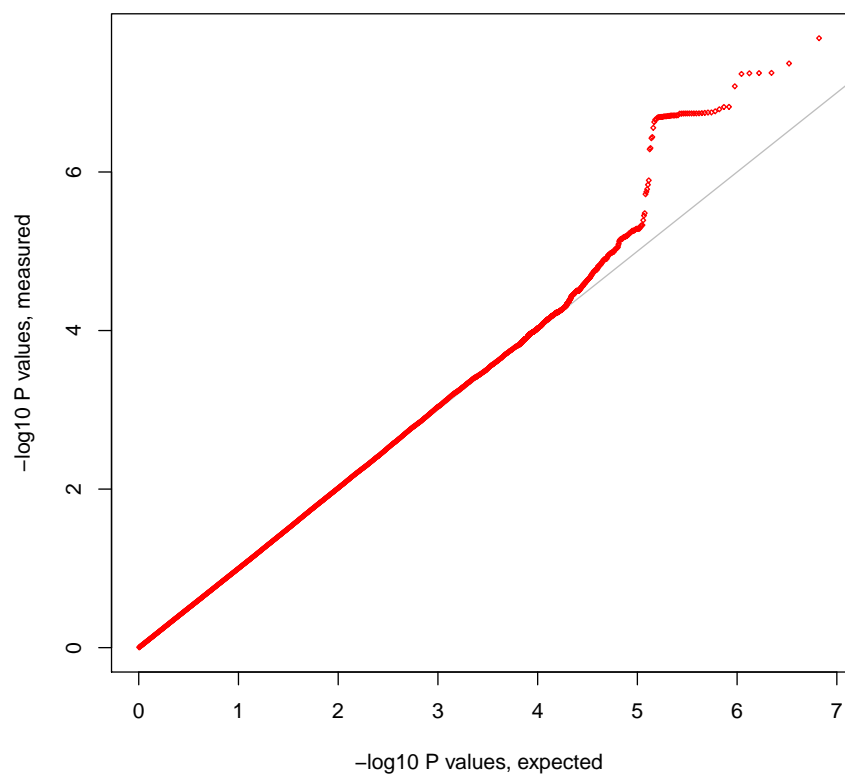
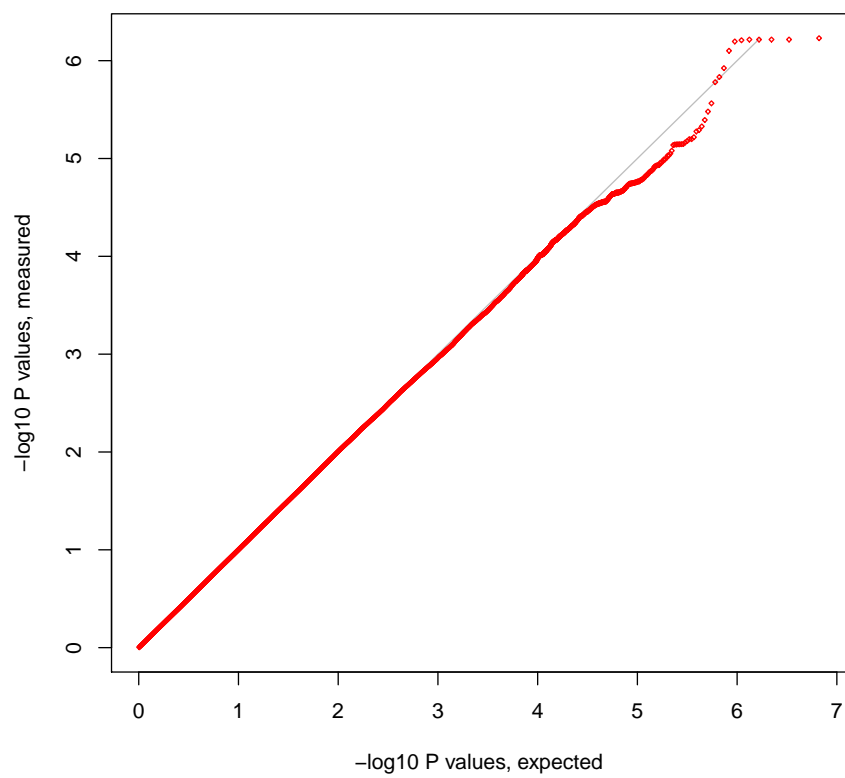


Figure 12: CaRatio

**Lambda = 0.99814 :**



## Part III

# Norway

## 5 Phenotypes

```

nor <- read.table("~/GWAS/norwayIterCovPcs.txt", header = T, na = "-9")
nor <- nor[!is.na(nor$pca1), ]
summary(nor$post_fev1pp)

##      Min. 1st Qu.  Median      Mean 3rd Qu.      Max.
##      8.3   52.5   77.8   72.2   93.1   139.0

summary(nor$post_maxrat)

##      Min. 1st Qu.  Median      Mean 3rd Qu.      Max.
##      0.170  0.530  0.690  0.648  0.790  1.000

summary(nor$post_fev1max)

##      Min. 1st Qu.  Median      Mean 3rd Qu.      Max.
##      0.25   1.46   2.42   2.38   3.18   5.57

nor <- read.table("~/GWAS/norwayCovPcCaseIter.txt", header = T, na = "-9")
nor <- nor[!is.na(nor$pca1), ]
summary(nor$post_fev1pp)

##      Min. 1st Qu.  Median      Mean 3rd Qu.      Max.
##      8.3   37.0   53.2   50.7   65.0   79.9

summary(nor$post_maxrat)

##      Min. 1st Qu.  Median      Mean 3rd Qu.      Max.
##      0.170  0.410  0.540  0.514  0.625  0.700

summary(nor$post_fev1max)

##      Min. 1st Qu.  Median      Mean 3rd Qu.      Max.
##      0.25   1.04   1.49   1.57   1.99   3.78

/proj/reglns/regln00/GWAS/Imputation/norway
1000g
--bfile /proj/reglns/regln00/GWAS/NORWAY_GSK_COPD_2009sorted
--linear
--covar ~/GWAS/norwayIterCovPcs.txt
--covar-name pca1-pca14,age,py

```

```

--sex
--pheno ~/GWAS/norwayIterCovPcs.txt
--pheno-name post_fev1pp
--out norFev1pp
## ratio is post_maxrat and norRatio; fev1 is post_fev1max using height as a covariate
## for cases
/proj/reglins/regln00/GWAS/Imputation/norway
1000g
--bfile /proj/reglins/regln00/GWAS/NORWAY_GSK_COPD_2009sorted
--linear
--covar ~/GWAS/norCovPcCaseIter.txt
--covar-name pca1-pca5,age,py
--sex
--pheno ~/GWAS/norCovPcCaseIter.txt
--pheno-name post_fev1pp
--out norCaFev1pp

```

## 6 Qqplots

```

makePlot <- FALSE
qqNames <- as.vector(sapply(c("Fev1", "Fev1pp", "Ratio"), function(x) paste(c("",
  "Ca"), x, sep = "")))
for (n in qqNames) {
  if (makePlot) {
    pdf(paste("lungFx-nor", n, ".pdf", sep = ""))
    tab <- read.table(paste("nor", n, ".assoc.linear.gz", sep = ""), header = T,
      colClasses = c(rep("NULL", 8), "numeric", rep("NULL", 5), "numeric"))
    fMyQqplot(tab)
    dev.off()
  }
  cat(paste("\\begin{figure}\\n\\caption{", n, "}\\n\\includegraphics{lungFx-nor",
    n, ".pdf}\\n\\end{figure}\\n", sep = ""))
}

```

Figure 13: Fev1

**Lambda = 1.01501 :**

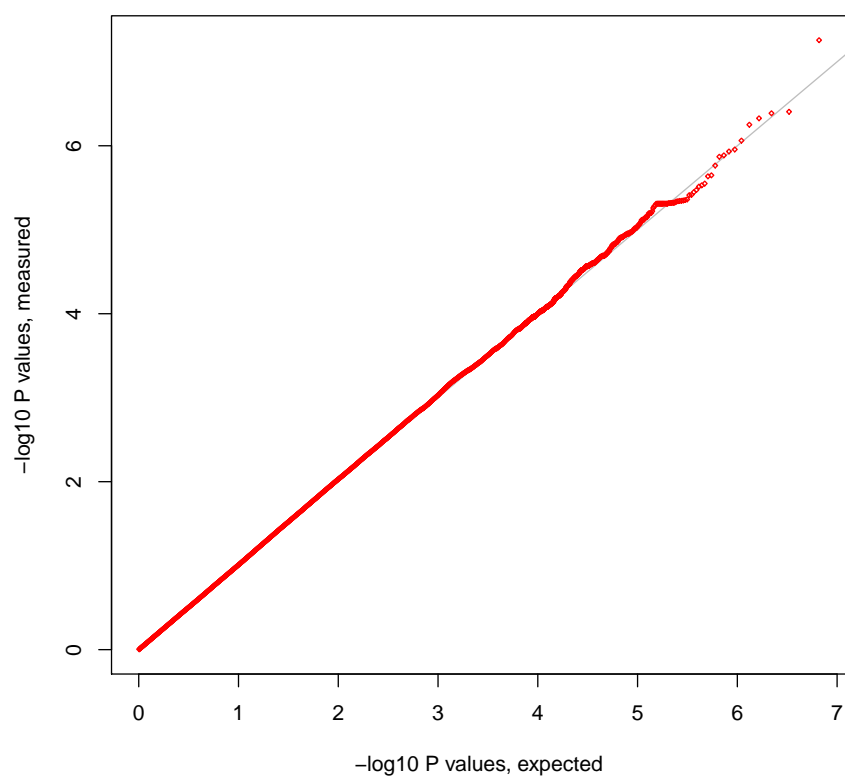


Figure 14: CaFev1

**Lambda = 1.00467 :**

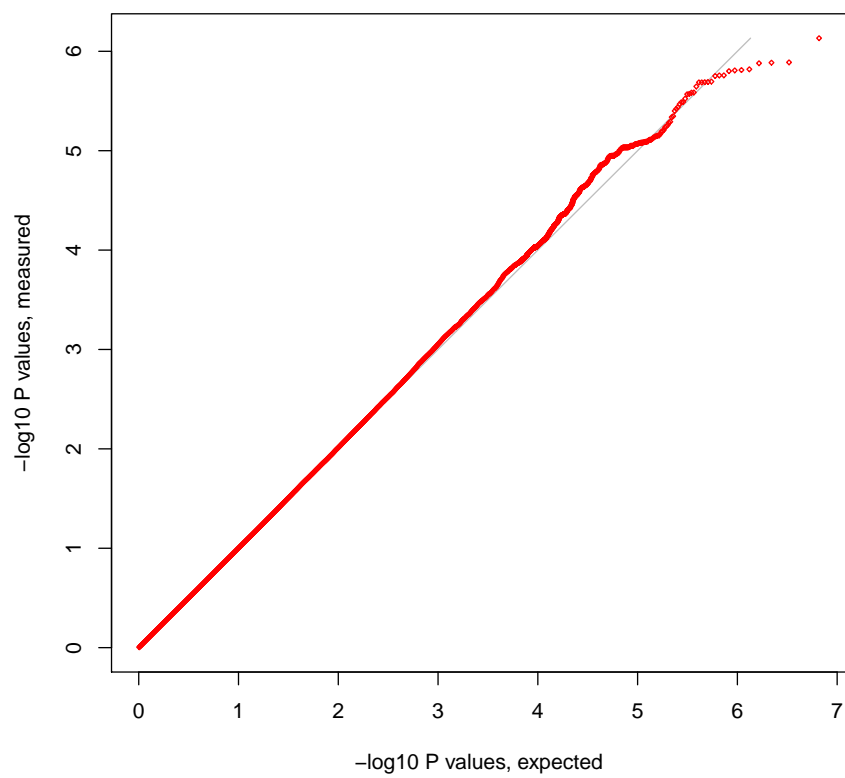


Figure 15: Fev1pp

**Lambda = 1.00983 :**

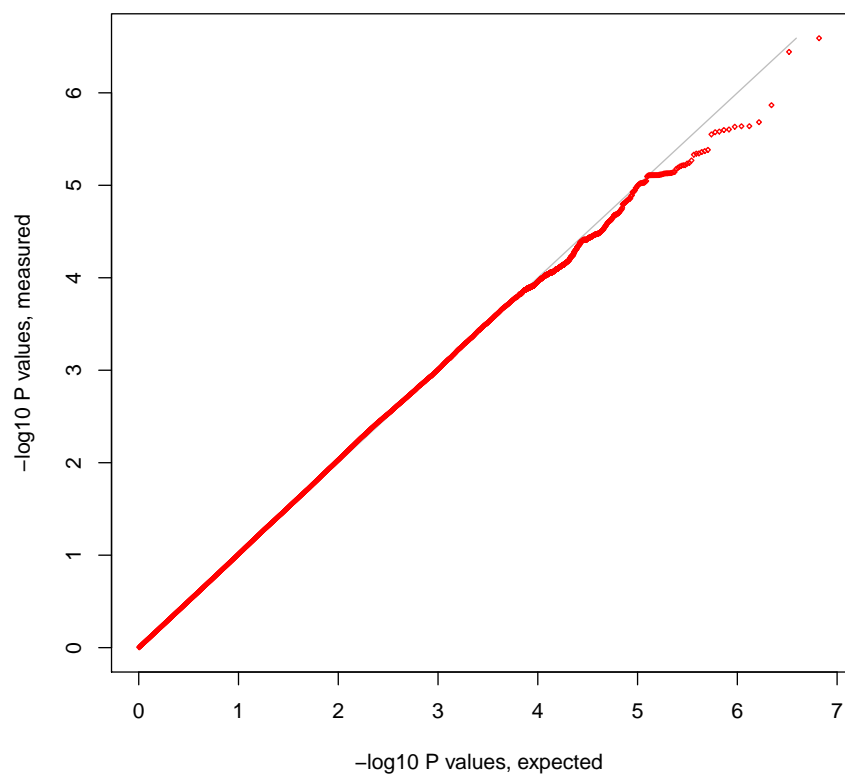


Figure 16: CaFev1pp

**Lambda = 1.00655 :**

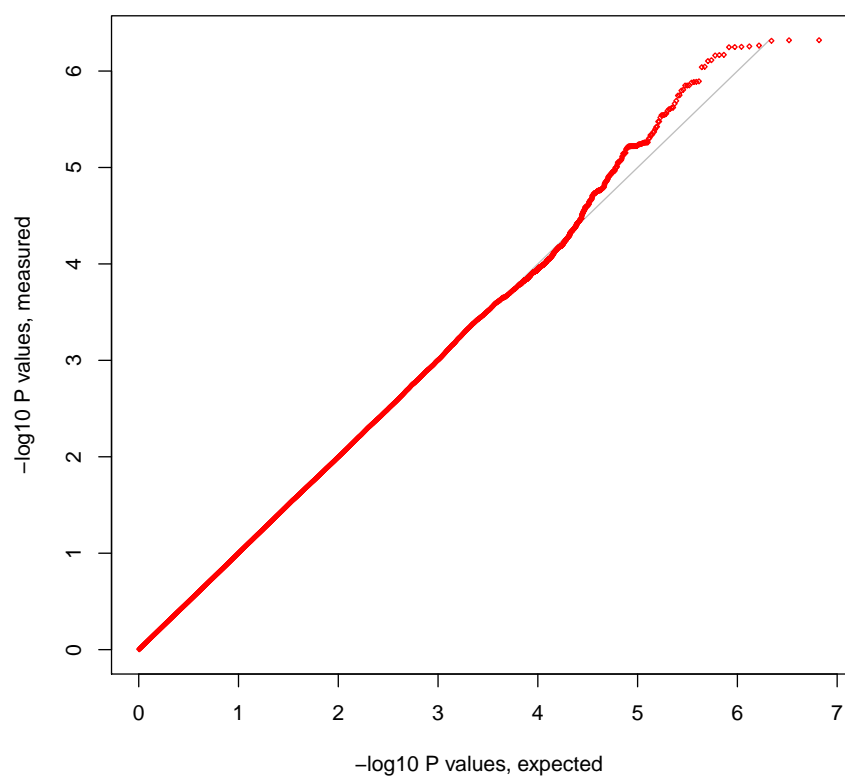




Figure 17: Ratio

**Lambda = 1.01218 :**

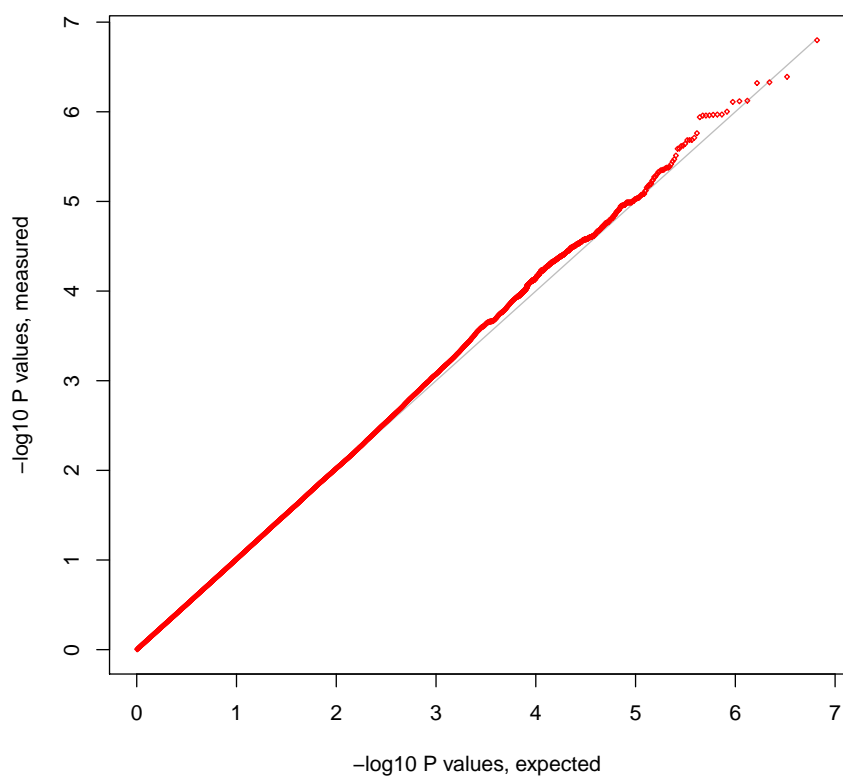
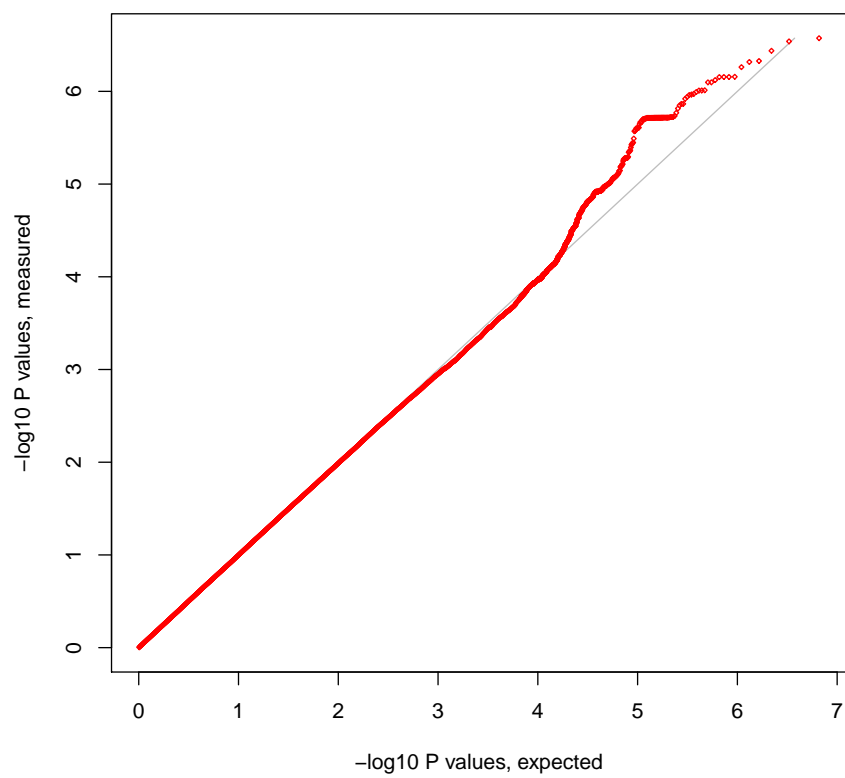


Figure 18: CaRatio

**Lambda = 1.00327 :**



## Part IV

# Meta-analysis

```

VERBOSE OFF
## SCHEME STDERR
AVERAGEFREQ ON
MINMAXFREQ ON
MARKER SNP
STDERRLABEL SE
ALLELE A1 A2
EFFECT BETA
PVAL P
FREQLABEL FRQ
DEFAULT 5351
PROCESS Cg/fev1ppNhwAll.assoc.logistic.gz
DEFAULT 1912
PROCESS eclFev1pp.assoc.linear.gz
DEFAULT 1659
PROCESS norFev1pp.assoc.linear.gz
GENOMICCONTROL ON
OUTFILE metal201209Fev1pp.tbl
ANALYZE HETEROGENEITY
## for cases CaFev1pp
DEFAULT 2819
PROCESS Cg/fev1ppNhwCaAll.assoc.linear.gz
DEFAULT 1736
PROCESS eclCaFev1pp.assoc.linear.gz
DEFAULT 854
PROCESS norCaFev1pp.assoc.linear.gz
## Run the metal analyses - create a small shell script to deal with the results
##### ##
## We first do: join -1 2 -2 1 temp2 temp1 -a 1 | awk '!$6{$6="."}; {print $0}'
## to create the joined marker file
## head -1 metal201209CaFev1pp1.tbl > tempHeader
## echo 'Chr Bp Gene' >> tempHeader
## ### ##processMetal.sh
grep -v '?' metal201209${1}1.tbl | sort -k10,10g | gzip -c > metal201209${1}1NoMiss.gz
zcat metal201209${1}1NoMiss.gz | head -5000 | sort -k1,1 > temp1${1}
join -1 1 -2 1 temp1${1} /udd/remhc/Resources/cgNhwAllSnpGeneLoc.txt | sort -k10,10g | awk 'BE

res <- as.vector(sapply(c("Fev1pp", "Fev1", "Ratio"), function(x) paste(c("",
"Ca"), x, sep = "")))
for (r in res) {
  cmd <- paste("sh processMetal.sh", r)

```

```

## system(cmd,wait=FALSE)
res <- read.table(paste("top", r, "1NoMissDet.txt", sep = ""), header = T)
res2 <- NULL
while (nrow(res) > 1) {
  ## simple pruning - ignore LD structure, just go by distance...
  res2 <- rbind(res2, res[1, ])
  res <- res[!(res$Chr == res$Chr[1] & abs(res$Bp - res$Bp[1]) < 250000),
    ]
}
## shorten res2$gene
res2$Gene <- sapply(res2$Gene, function(x) substr(x, 1, 15))
print(xtable(res2[1:10, c("Chr", "MarkerName", "Bp", "P.value", "Gene",
  "Direction", "HetISq")], caption = r, display = c("s", "d", "s", "d",
  "e", "s", "s", "f")), include.rownames = FALSE)
}

```

Chr	MarkerName	Bp	P.value	Gene	Direction	HetISq
15	rs8040868	78911181	7.28e-15	CHRNA3(0.0)	+++	-26.30
4	rs1512281	145434901	6.46e-13	HHIP-AS1(129.0)	—	0.00
4	rs6837671	89873092	3.54e-11	FAM13A(0.0)	+++	0.00
19	rs7937	41302706	1.56e-08	RAB4B-EGLN2(0.0)	—	0.00
3	rs2811520	128012277	6.43e-08	EEFSEC(0.0)	—	-34.50
4	rs7664805	106843958	8.96e-08	NPNT(0.0)	—	34.20
11	rs737693	102726142	6.14e-07	MMP12(7.0)	—	-22.70
9	rs1108581	136505241	7.53e-07	DBH(0.0)	—	0.00
10	rs7100689	82222178	8.81e-07	TSPAN14(0.0)	+++	15.50
17	rs7224296	44800046	9.33e-07	NSF(0.0),MIR431	+++	0.00

Table 1: Fev1pp

Reviewing these results:

For FEV1pp and FEV1, recapitulates known loci; NPNT locus is near GSTCD.

Other comments: EEFSEC is stronger here than in the case control analysis. Other loci seem to be novel: TSPAN1 is near but still a few MB away from C10orf11 (prior GWAS loci) on 10q, KNTC1 is also new. To do: Look at heterogeneity - meta-regression?

Chr	MarkerName	Bp	P.value	Gene	Direction	HetISq
13	rs9536463	54132247	7.51e-07	LINC00558(257.0)	—	0.00
15	rs2656065	78750549	8.04e-07	IREB2(0.0)	—	0.00
20	rs34306377	59597631	1.63e-06	CDH4(229.0)	—	2.60
9	rs7875024	16288424	3.52e-06	BNC2(121.0)	—	0.00
9	rs551517	113636793	4.06e-06	LPAR1(0.0)	—	0.00
1	rs412967	196813716	5.50e-06	CFHR1(12.0)	+++	0.00
5	rs25777	156682890	6.97e-06	ITK(0.0)	—	-51.80
12	rs887357	3474645	8.21e-06	PRMT8(15.0)	+++	-13.70
8	rs6473873	54535433	8.39e-06	ATP6V1H(92.0)	—	0.00
10	rs12783688	134246240	1.16e-05	C10orf91(12.0)	—	-65.20

Table 2: CaFev1pp

Chr	MarkerName	Bp	P.value	Gene	Direction	HetISq
15	rs8040868	78911181	3.31e-15	CHRNA3(0.0)	+++	4.10
4	rs1512281	145434901	9.98e-13	HHIP-AS1(129.0)	—	0.00
4	rs6837671	89873092	4.34e-10	FAM13A(0.0)	+++	0.00
19	rs12461383	41370338	9.07e-09	CYP2A7(11.0)	+++	0.00
10	rs7100689	82222178	4.09e-08	TSPAN14(0.0)	+++	48.10
4	rs7664805	106843958	2.32e-07	NPNT(0.0)	—	41.70
3	rs2811524	127999955	2.46e-07	EEFSEC(0.0)	—	1.70
9	rs1108581	136505241	3.71e-07	DBH(0.0)	—	0.00
17	rs7224296	44800046	1.75e-06	NSF(0.0),MIR431	+++	0.00
13	rs9531098	36401576	1.76e-06	MIR548F5(0.0),D	—	0.00

Table 3: Fev1

Chr	MarkerName	Bp	P.value	Gene	Direction	HetISq
13	rs9536463	54132247	2.01e-07	LINC00558(257.0)	—	0.00
20	rs34306377	59597631	2.76e-06	CDH4(229.0)	—	35.70
15	rs2656065	78750549	4.54e-06	IREB2(0.0)	—	0.00
12	rs650466	118646661	4.70e-06	TAOK3(0.0)	—	0.00
9	rs551517	113636793	5.85e-06	LPAR1(0.0)	—	0.00
5	rs29545	132759324	6.75e-06	FSTL4(0.0)	—	0.00
1	rs412967	196813716	9.38e-06	CFHR1(12.0)	+++	0.00
13	rs12874564	69133985	9.72e-06	LINC00550(301.0)	+++	-33.10
9	rs7875024	16288424	1.13e-05	BNC2(121.0)	—	0.00
12	rs12833560	30424221	1.26e-05	IPO8(357.0)	+++	-40.00

Table 4: CaFev1

Chr	MarkerName	Bp	P.value	Gene	Direction	HetISq
4	rs1512281	145434901	6.35e-18	HHIP-AS1(129.0)	—	-84.10
15	rs1317286	78896129	4.94e-15	CHRNA3(0.0)	+++	-92.70
4	rs6837671	89873092	3.31e-13	FAM13A(0.0)	+++	-21.50
11	rs17361668	102720344	4.73e-09	MMP3(6.0)	+++	0.00
1	rs622912	218670357	9.68e-09	TGFB2(52.0)	+++	0.00
19	rs12461383	41370338	5.45e-08	CYP2A7(11.0)	+++	0.00
14	rs35629566	93072317	1.39e-07	RIN3(0.0)	—	-13.20
3	rs2811524	127999955	4.64e-07	EEFSEC(0.0)	—	6.50
4	rs7664805	106843958	4.91e-07	NPNT(0.0)	—	0.00
4	rs6842499	145910286	6.17e-07	ANAPC10(5.0)	+++	23.70

Table 5: Ratio

Chr	MarkerName	Bp	P.value	Gene	Direction	HetISq
12	rs11615433	123018475	5.68e-08	KNTC1(0.0)	—	-27.20
15	rs8040868	78911181	2.63e-07	CHRNA3(0.0)	+++	0.00
11	rs494963	102715826	3.84e-07	MMP3(1.0)	+++	0.00
1	rs1329427	196704559	9.27e-07	CFH(0.0)	+++	1.90
4	rs13141641	145506456	9.70e-07	HHIP-AS1(57.0)	—	-35.60
1	rs4457591	222078600	1.04e-06	DUSP10(163.0)	—	-55.40
5	rs12657392	78963323	2.50e-06	PAPD4(0.0)	—	0.00
1	rs9970786	239090514	3.64e-06	LOC339535(441.0)	—	0.00
5	rs281143	70969032	3.83e-06	MCCC2(14.0)	+++	0.00
11	rs947950	100407990	4.60e-06	ARHGAP42(150.0)	+++	0.00

Table 6: CaRatio