Significance and Background

A large proportion of common diseases (i.e. prevalence > (XX) %) are genetically complex. In contrast to Mendelian diseases, genetic causation of complex diseases is not straightforward, with genetic risk for any given disease spread across a large number of genes, each individually of relatively minor effect. While successful efforts to map the genetics of Mendelian diseases date well back into the 20^{th} century, only in the past decade with the maturation of the genome wide association study (GWAS) approach has some understanding of the genetic basis of complex disease begun to emerge. 1,? It's become clear now that estimates of heritability based on the twin study design were broadly accurate (if slightly overestimated in some cases), and that the heritabilities of common complex diseases are generally high. GWAS and related approaches have also shown that a substantial proportion of the variance in risk can be attributed to a very large number of common alleles.^{2,3,4,5} while sequencing and exome studies indicate a role for rare variants of large effect as well.^{6,7,8} The totality of the evidence therefore suggests that thousands or perhaps even tens of thousands of individual genetic variants play a role in determining susceptibility to any given complex disease. Less clear however is our understanding of the forces which govern the prevalence and genetic architecture (i.e. the relationship between allele frequency and effect size) of complex disease. In contrast to Mendelian diseases, where nearly a century of theory explain how mutation rate, dominance, selection⁹ and demography? combine to influence the prevalence of these diseases, there is relatively little in the way of quantitative theory on the evolution of complex disease.

Nonetheless, several qualitative mechanisms have been proposed which may potentially explain the observed patterns. Perhaps the most straightforward is that the present distribution of genetic risk reflects a simple balance between mutation and selection. 10 The prevalence of common disease may therefore be ascribed to relatively large mutational target sizes for many diseases. Variation among diseases in their prevalence and genetic architectures may simply reflect difference in features such as the mutational target size, distribution of effects, and fitness cost of the disease. The direct selection experienced by a given allele is likely only part of the story, however, as the extreme polygenicity of complex diseases indicate that pleiotropy must almost surely be extensive. 11,12 Indirect selection due to these pleiotropic effects may therefore play a major role in determining architecture and prevalence, both in steady-state scenarios, where pleiotropic effects may alter the stregth of selection for or against disease alleles or under more dynamic hypotheses, where recent positive selection on a trait genetically correlated to disease may have altered prevalence and/or architecture, as has been argued in a number of cases with conflicting evidence. 13, 14, 15, 16, 17, 18 Alternative explanations invoke the recent and profound changes in the human environment (diet, lifestyle, etc.) and the possibility that the mismatch between the ancestral human environment and present conditions that have resulted in a large increase in disease prevalence. 19,20 Such was the original basis for the "thrifty genotype" hypothesis, ^{21,22} and subsequent observation of large scale oscilations in type 2 diabetes incidence in response to food shortage and economic crisis and subsequent recoverv in Cuba in the 1990s²³ indicate that this effect can be profound indeed. While some of these ideas likely apply to some diseases (and they are also not mutually exclusive), more often than not they have been put forward as verbal models only, and so it is difficult to quantitatively check their predictions.

Each of these hypotheses are ultimately statements about population and quantitative genetic processes. While some models relating population genetic processes with the kind of data emerging from GWAS have been proposed, they generally rely on various ad hoc or limiting assumptions that make easily interpretable inference difficult. Nonetheless, two studies in particular have been especially influential. Pritchard, in 2001,²⁴ considered a model in which the effect on disease risk is completely uncoupled from their fitness cost, i.e. an extreme pleiotropy limit. While this paper was enormously influential in grounding the debate surrounding the common disease-common variant hypothesis in population genetic theory,²⁵ it seems unrealistic that a mutation's effect on disease risk should not have any consequences for its fitness. A second study which has been influential is that of Eyre-Walker in 2010, 26 who posited that all mutations are deleterious a priori, but arbitrarily assumed a relationship between effect size and selection coefficient of the form $\alpha = \delta S^{\tau}(1+\epsilon)$, where S is the selection coefficient, δ is a randomly chosen sign (i.e. 1 or -1), ϵ is a noise term, and τ is a "coupling parameter" meant to capture the effect of pleiotropy: where $\tau = 1$ means no pleiotropy and $\tau = 0$ gives Pritchard's²⁴ extreme pleiotropy limit. While this model has seen empirical application (see Specific Aim 2 below for more background on the Eyre-Walker model in this context), it does not have any obvious theoretical justification, and indeed possesses some odd features, such as fitenss equivalence of mutations with opposing effects on disease. While these studies have both been extremely influential, it seems worth reemphasizing the fact that neither posesses any concept of fitness surface

relating an explicit disease phenotype to natural selection, and in light of our rapidly accumulating knowledge of the genetic architecture of complex disease, a fresh take on the problem seems due.

Here I propose to develope generative models for the way that genetic architecture and disease prevalence will be affected by evolutionary parameters such as mutation, natural selection, pleiotropy and demography. I will develop these models into statistical inference approaches which take advantage of the rich information present in GWAS data to infer the underlying parameters which govern the evolution of complex disease genetic architecture.

Approach

Preliminary Results Our simplest model and point of departure considers the impact of mutation and selection on a single disease in a constant environment. In this model, each individual's risk of developing disease is a non-linear transform of an underlying (and generally unobserved) disease liability trait, such that

$$R = \ell(Z), \qquad Z = \sum_{i} \alpha_{i} g_{i} + \epsilon$$
 (1)

where an individual's liability for disease (Z) is an additive trait, α_i and g_i are liability scale effect size and genotype respectively at site i, and ϵ is a normally distributed deviate which captures stochastic variation in risk among genotypes with the same mean liability, (i.e. the "environment" of classical quantitative genetics). An individual's probability of developing disease (i.e. their "risk": R) is a monotonic function of their liability. This form covers a range of standard models for the genetics of binary traits, including Wright's liabity threshold model, $^{27,28,?,29}$ the logistic model commonly employed in GWAS, 30 and the exponential of Risch's multiplicative model, 31 among other possibilities (the difference among these is simply in the choice of the monotonic function ℓ). Previous work 32,33 suggests that the exact choice of ℓ is unlikely to be particularly important, and our preliminary investigations (omitted due to space constraints) support this conclusion. As such I will take ℓ to be the probit link function of Wright's liability threshold model for the remainder of this proposal, though I will explore other choices to understand what impact if any they might have on our results.

The number, frequencies and effect sizes of the sites contributing to variation in liability arise from population genetics processes. In our model, liability increasing and decreasing mutations arise according to an infinite sites model with free recombination among sites at genome wide rates μ^+ and μ^- , with effect size distributions f^+ (α) and f^- (α) respectively. Individuals with the disease have a reduced fitness of 1-S (disease free individuals have fitness 1).

The genetic architecture of the disease under this simple model is shaped by mutation-selection-drift balance, and can be related to standard results from quantitative genetics. The steady state is reached when

$$U^{+} = U^{-} + V_{A} \underbrace{S\phi\left(\Phi^{-1}\left(1 - P\right)\right)}_{\text{selection gradient}}$$
 (2)

where $U^+ = \mu^+ \int_0^\infty \alpha f^+ (\alpha) \, \mathrm{d}\alpha$ is the total per generation mutational increase in liabilty (with U^- defined similarly as mutational pressure toward decreased liability). The final term accounts for the selection pressure toward lower liability; V_A is the additive genetic variance of liability, P is the disease prevalence, and ϕ and Φ are the Gaussian pdf and cdf repsectively. The compound term multiplying V_A is a selection gradient in the standard quantitative genetics sense, and is a simple generalization of the gradient exerted by truncation selection to cases where the "truncated" individuals have fitness greater than zero.? An individual allele in this model with effect size α on disease liability will experience a selection coefficient

$$s = -2\alpha S\phi \left(\Phi^{-1} \left(1 - P\right)\right) \tag{3}$$

against the liability increasing homozygote, and will evolve under fitness additivity so long as α is small.

Equations (2) and (3) provide a path to solving the model, as the appearance of S and P in both equations couple together the behavior of the population at the macroscopic level with the microscopic dynamics of individual allele frequencies, and it is these microscropic dynamics that determine V_A . These dynamics can be described by standard results from diffusion theory,?,34,35,? and the integration of all of this together allows us to solve for the additive genetic variance and disease prevalence given the specification of the mutational inputs and the

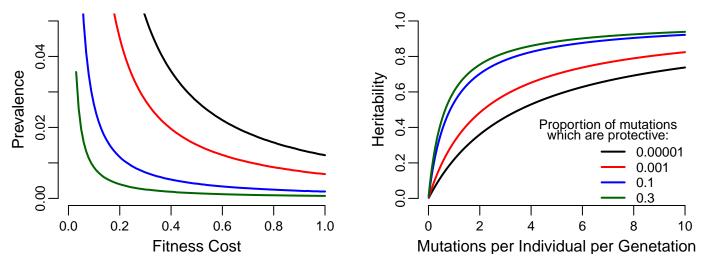


Figure 1: Solutions to the model specified by equations (2) and (3). The left panel shows the disease prevalence at equilibrium as a function of the fitness cost for various different choices of the strength of mutation bias. The right panel shows the heritability of disease liability as a function of the total size of the mutational target, again for different strengths of mutational bias. The independence relationships described can be seen here by noting that the plot in the left panel is insensitive to variation alone the x axis of the right plot, and vice versa.

fitness cost. The solution provides a few curious insights. The first is that while disease prevalence depends on the fitness cost and the mutational bias, it is insensitive to the total volume of mutational input. Conversely, while the heritability of liability is quite sensitive to both the extent of mutational bias and the total mutational input, it is independent of the fitness cost of the disease. These independence relationships have potentially significant implications for how we interpret studies of the genetic architecture of disease, and their existence is to my knowledge not recognized in the field.

Specific Aim 1: Relating Population Genetic Processes with the Architecture of Complex Diseases

I will generalize the basic model from our preliminary results to include the impacts of environmental and demographic change, pleiotropy, and I will thoroughly study the robustness of our results to a range of different assumptions about the biology of complex disease. When possible, I will solve these generalized models to obtain analytical expressions for the genetic architecture an disease prevalence. In cases where analysitical treatment proves intractable, I will use simulations and other numerical methods to obtain solutions, and all anlytical results will be verified in this manner as well.

Environmental Change

The most straightforward way to incorporate environmental change may also be the most relevant to human disease. Such models are those in which the mean or the variance of the environmental component of the phenotype shifts or increases suddenly. Given the recent and rapid changes to human environments (e.g. diet and lifestyle on type 2 diabetes prevalence), we are most interested in the scenario where an environmental shift has just occurred, but there has not been sufficient time for allele frequencies to evolve away from their previous equilibria. In the case of a shift in the enironmental mean, the effect is simply an increase in prevalence such that $P_{new} = 1 - \Phi\left(\Phi^{-1}\left(1 - P_{old}\right) - \delta\right)$, where δ is the shift in the environmental contribution to liability measured in units of the phenotypic standard deviation (Φ , again, is the Gaussian cdf).

The result of a change in the environmental variance are slightly more complex, as it impacts both the prevalence and the heritability of the disease. The most straightforward impact is on heritability. If the environmental variance is increased by an amount ψ , then heritability is decreased such that $h_{g,new}^2 = \frac{h_{g,old}^2}{1+\psi}$ (where again, ψ is given here in units of the pre-change phenotypic variance). Prevalence, on the other hand, will increase to $P_{new} = 1 - \Phi\left(\frac{\Phi^{-1}(1-P_{old})}{1+\psi}\right)$. In both of these simple environmental change scenarios, the a given mutation's effect on liability is unchanged. The effect on risk, however, is increased, as a given mutation's contribution to risk conditional on its contribution to liability also depends on the prevalence of the disease (expressions omitted due to space). The result is that the while the additive genetic variance for liability is unchanged under either

scenario, on the *risk* scale it will likely be increased. A shift in the mean of the environmental contribution should actually cause an increase in heritability on the risk scale, while the impact of an increase in the variance of the environmental contribution on the heritability of risk are not immediately obvious, and may be architecture dependent.

In addition to these simple scenarios, I will also study how more complicated patterns of environmental change influence disease prevalence and the genetic architecture of risk. Such scenarios include environmental change which is ancient enough for the subsequent response to selection to influence architecture and prevalence (e.g. change coinciding with the Out of Africa migration event), large environmental changes which only influence part of the population (therefore violating the assumption of normality), and any others which we can identify as biologically plausible.

Pleiotropy As noted in the background above, mounting evidence suggests that genetic variation is often highly pleiotropic. ^{11, 12} We will consider pleiotropic effects of two kinds. The first includes pleiotropic effects on other disease traits. Pleiotropy of this form will generally act to modulate the additive selection coefficient felt by a given allele. Liability increasing mutations which have protective effects on other diseases will be less deleterious than in the preliminary model (or may even be beneficial on average), while those which increase liability for multiple diseases will incur additional selective cost beyond that due to direct selection on the focal disease.

The second form of pleiotropy considered will be due to stabilizing selection on continuously distributed (i.e. non-disease) quantitative traits. In models of stabilizing selection on quantitative traits at equilibrium, there is no directional component to the selection felt by individual alleles. Rather, variance reducing selection for individuals to cluster near the optimum causes individual alleles to experience symetric underdominance with respect to fitness (i.e. the minor allele is always selected against), where the strength of selection depends on the effect size of the mutation on the quantitative trait relative to the strength of stabilizing selection.³⁶

This observation suggests an interesting relationship which will form the foundation for our modeling of pleiotropy. When a mutation effects only disease, selection is directional and additive, whereas when a mutation effects only quantitative traits, selection is underdominant. Mutations which have large effects on disease and smaller impacts on quantitative traits will experience directional selection with the disease causing mutation being partially dominant for fitness, while mutations which have large effects on quantitative traits and smaller effects on disease will be underdominant for fitness, but asymetrically so, with the liability increasing homozygote less fit than the liability decreasing homozygote.

The major objective here is to derive an expression for joint distribution of effect size and allele frequency (i.e. the genetic architecture) as a function of the parameters (Θ_A) which describe the nature of pleiotropy impacting the disease architecture. These two quantities are conditionally independent of one another given the specification of the selection coefficient(s), which suggests a tractable approach for theoretical analysis and for inference (aim 2). The expression for the genetic architecture can be written as an integral over the selection coefficients

$$p(\alpha, x \mid \Theta_A) = \int \int p(\alpha \mid s, h, \Theta_A) p(x \mid s, h) p(s, h) \, ds dh. \tag{4}$$

The expression for the distribution of allele frequencies $(p(x \mid s, h))$ can be computed from standard diffusion theory,? and this fact can be leveraged in an inference context (aim 2) to learn the distribution of the selection coefficients (p(s,h)) directly from the data (alternately, plausible distribution can be specified in a theoretical context to understand how differences in the distribution of selection coefficients impact architecture). This leaves specifying the distribution of effect sizes for a given set of selection coefficients and under a given set of parameters governing the effects of pleiotropy $(p(\alpha \mid s, h, \Theta_A))$ as the primary theoretical task.

I will approach this problem first by considering simple isotropic models of pleiotropy in which mutations may affect more than one disease/trait, but which lack mutational covariance among diseases or traits. In particulary, I will investigate how the number of pleiotropically related diseases/traits, the strength of selection on them, and the degree of functional overlap among diseases/traits all impact the genetic architecture of disease. I will then consider extensions of the isotropic model in order to understand how covariance among traits affects the dynamics.

This result of this theoretical work will be a broad framework for studying the effects of pleiotropy which I expect to be applicable beyond the stated aims of this proposal. In addition studying how pleiotropy impacts the architecture of one disease, this work will form a basis for the explicit study of the joint architecture of multiple traits

together in a unified framework, and it provies a natural way of examining popular models in medical genetics such as the idea that some common diseases may be in fact be collections of multiple distinct but biologically related disorders which all present similar symptoms.

Non-Equilibrium Demography It is clear from population genetic work over the last decade and a half that demographic events such as the Out of Africa bottleneck and recent explosive population growth have had a significant impact on allele frequencies and therefore potentially on genetic architecture. Recent work from Dr Sella's lab,³⁷ and others^{?,?,?} suggests that both the bottleneck and recent growth may have impacted genetic architecture, though the relative importance of each depends on the (as yet unknown) selection coefficients of disease alleles. However, what most work to date on this problem has been done in the context of a single site in a vaccum with a fixed selection coefficient, with no explicit disease phenotype, and therefore no model of the relationship between effect size and selection coefficient.

I will use simulation based approaches, together with the theory of pleiotropy developed above, to study how the particular course of human demographic history has impacted the evolution of complex disease architecture and prevalence. Population size changes impact architecture through their modulation of the relationship between genetic drift and natural selection. All else being equal, genetic architectures in larger populations will be composed of more rare alleles, due to the increased efficiency of selection, though the precise impact depends on the details of the model. My investigation of the preliminary model also suggests that disease prevalence at equilibrium is fairly sensitive to population size. This is significant, specifically because the selection coefficient experienced by a particular allele depends on the disease prevalence. If prevalence evolves over time in response to changes in population size, then selection coefficients will change over time as well.

Additional Complications In addition to these major issues, I will study how our results depend on the fidelity of our baseline assumptions. (expand)

A final theoretical goal will be to obtain an understanding of the extent to which we should expect the various parameters which arise in the investigations described above to be uniquely identifiable, and on the basis of what kinds of data. It seems likely that we will find that there are qualititatively distinct phenomena which lead to similar genetic architecture and disease prevalence. A major product of this theoretical work will therefore be thorough guidelines about what can and cannot be inferred about the population genetics of complex disease from GWAS data, guidelines which we will be able to apply immediately in the development of our inference approaches below.

Specific Aim 2: Inference of Model Parameters from Complex Disease GWAS

Background The existing work which comes closest to that proposed in this aim is recent work from the Reich and Altshuler labs applying the Eyre-Walker model (discussed in intro above) to investigate the genetic architecture of complex disease. Three studies in particular have applied this approach.^{38, 39, 40} In each, the authors simulated data under a range of parameter choices for the Eyre-Walker model, and used statistical summaries of the genetic architecture (such as the number of significant variants discovered or the proportion of variance explained by SNPs with minor allele frequency < 5%) in an approximate Bayesian computation approach to determine what sort of pleiotropy was most consistent with GWAS data on a partiulcar complex disease.

(this needs work) In general, these studies have been able to exclude extreme models, ruling out scenarios with either perfect coupling (limited/no pleiotropy) or no coupling (i.e. extreme pleiotropy) between a mutation's effect and selection coefficient, as well as providing estimates of the mutational target size. However, the existing approach leaves much to be desired, as further interpretation of inferences based on the Eyre-Walker model in general remains difficult. In part this is because the distribution of selection coefficients is usually taken to be gamma (rather than inferred from the data), likely with little real justification. 42,43 It is also clear that the statistical summaries used for inference by these authors discard a great deal of potentially useful information about the genetic architecture. The Eyre-Walker model is also fundamental not extensible to inference with multiple traits together, which we expect to particularly important in the future as multi-trait GWAS methods continue to mature.?,?

It is therefore clear that there is a need for an inference approach which A) uses as much of the information available in GWAS data as possible, and B) allows for the estimation of parameters which have biologically meaningful interpretations.

Approach Intuitively, the number of variants discovered (K) in a GWAS of a given sample size places some bounds on the plausible size of the mutational target and on the distribution of effect sizes, and specific knowledge of their effect sizes and frequencies $(\alpha_i$ and $x_i)$ provides information about their selection coefficients and the impact of pleiotropy on the genetic architecture (Fig 2). Unfortunately, even in very large GWAS, a significant number of variants will remain undetected, limiting our inference in part to only variants with sufficiently large effect sizes, or with sufficiently intermediate frequencies. We can overcome this limitation by leveraging recently developed methods that estimate the proportion of the genetic variance

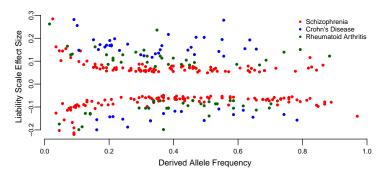


Figure 2: The relationship between allele frequency and effect size at known GWAS loci for three different complex diseases. Differences among diseases reflect differences in the underlying parameters which govern their evolution.

attributable to alleles with frequencies in a given range.^{?,?} Intuitively, diseases with genetic architectures dominated by large selection coefficients will have more of the genetic variance explained by rare alleles, though this relationship will be moderated by additional factors such as pleiotropy and demography.

I will formalize the above intuition into a composite likelihood based inference approach to infer the salient parameters which govern genetic architecture. A general version of this likelihood can be written as

$$L\left(\Theta_{A}\mid\{(\alpha_{i},x_{i})\}_{i=1}^{K},\{V_{G,b}^{*}\}_{b=1}^{B},\Theta_{S}\right) \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihood of observing K variants under model} \\ = \underbrace{Pr\left(K\mid\Theta_{A},\Theta_{S}\right)}_{\begin{subarray}{l} likelihoo$$

The first two terms account for indivivdual genome wide significant variants. The third term upweights and down-weights individual alleles in proportion to our confidence that the represent a true association rather than a false positive, and the final term accounts for the proportion of the genetic variance $(V_{G,b}^*)$ which can be attributed to alleles in each of B different allele frequency bins, and is not captured by the genome-wide significant variants. Here, Θ_A represents the parameters to be inferred, which determine the genetic architecture and prevalence, while Θ_S represents the known parameters of the GWAS, such as the numbers of cases and controls.

An advantage of the likelihood approach to inference is that it facilitates model comparison. I will derive expressions for the likelihood under a range of models, and then use model comparison metrics like the widely applicable infromation criteria (WAIC) or leave one out cross validation? to determine which model is most consistent with the genetic architecture of the disease. One potential pitfall is that in the case that GWAS variants are in linkage disequilibrium with one another, our approach is a composite rather than full likelihood. Such approaches are commonplace in population genetics, and provide statistically consistent parameter estimates,? but they underestimate the uncertainty of those estimates, and can wrongly favor more complex models with more parameters. We can circumvent this obstacle by evaluating model fit and uncertainty via approaches based on the bootstrap. If this proves computationally prohibitive, we may also be able to proceed by adapting approaches from composite likelihood based demographic inference? to our case.

An additional advantage of pursuing inference in this likelihood framework is that we will be able to make predictions about the number of variants we should expect to discover, and the distribution of their effect sizes and frequencies, as future GWAS samples sizes are increased. This will allow for continued reassessment of our models of disease into the future, and in cases where multiple qualititatively distinct models fit the available data, we will be able to make forecasts about what sort of sample sizes will be necessary in order to distinguish different models.

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