Class 7

We will import the UK_foods.csv dataset

```
url <- "https://tinyurl.com/UK-foods"
x <- read.csv(url)</pre>
```

Q1. How many rows and columns are in your new data frame named x? What R functions could you use to answer this questions?

```
dim(x)
```

[1] 17 5

We will want to check that the data has imported correctly

```
head(x)
```

```
X England Wales Scotland N.Ireland
          Cheese
                      105
                            103
                                     103
                                                 66
1
2
  Carcass_meat
                     245
                            227
                                     242
                                                267
3
     Other_meat
                     685
                            803
                                     750
                                                586
4
            Fish
                     147
                            160
                                     122
                                                93
5 Fats_and_oils
                     193
                            235
                                     184
                                                209
6
          Sugars
                     156
                            175
                                     147
                                                139
```

It seems that the names for the rows was incorrectly counted as a column, let's fix that

```
rownames(x) <- x[,1]
x <- x[,-1]
head(x)
```

	England	Wales	Scotland	N.Ireland
Cheese	105	103	103	66
Carcass_meat	245	227	242	267
Other_meat	685	803	750	586
Fish	147	160	122	93
Fats_and_oils	193	235	184	209
Sugars	156	175	147	139

Or alternatively:

```
x <- read.csv(url, row.names=1)
head(x)</pre>
```

	England	Wales	Scotland	N.Ireland
Cheese	105	103	103	66
Carcass_meat	245	227	242	267
Other_meat	685	803	750	586
Fish	147	160	122	93
	400		404	

Fats_and_oils 193 235 184 209
Sugars 156 175 147 139

We see that the problem seems to have been resolved, let's check with dim()

dim(x)

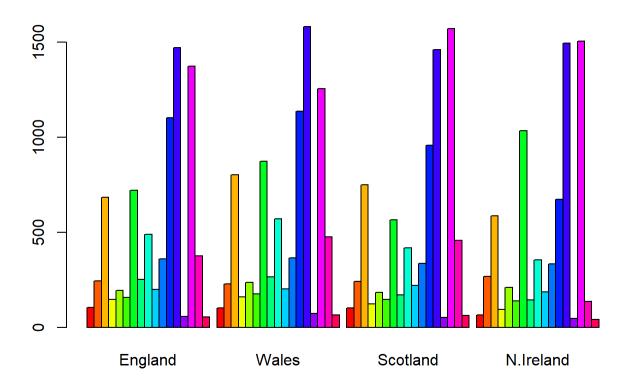
[1] 17 4

Q2. Which approach to solving the 'row-names problem' mentioned above do you prefer and why? Is one approach more robust than another under certain circumstances?

The second approach is better because it applies the change to the entire dataset without having to redefine x, which may become a problem if there are other aspects of the imported dataset that you would like to change

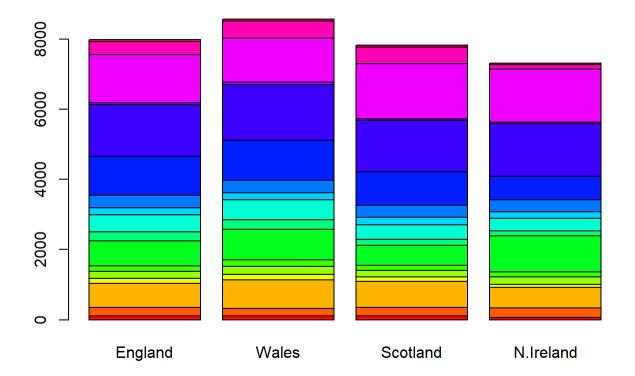
As a table this data does not show us much. Let's graph it:

barplot(as.matrix(x), beside=T, col=rainbow(nrow(x)))



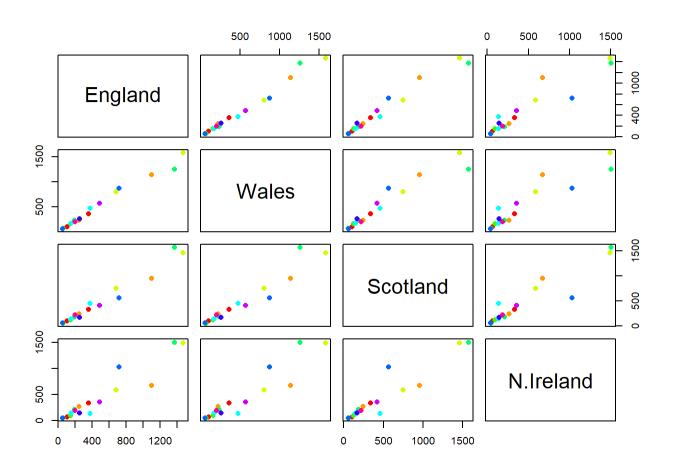
Q3: Changing what optional argument in the above barplot() function results in a stacked barplot?

#the beside portion is shown as true, needs to be false to stack the bars barplot(as.matrix(x), beside=F, col=rainbow(nrow(x)))



Q5: Generating all pairwise plots may help somewhat. Can you make sense of the following code and resulting figure? What does it mean if a given point lies on the diagonal for a given plot?

pairs(x, col=rainbow(10), pch=16)



The pairs shows graphs of each combination of observations plotted against each other: i.e England against itself, England vs Wales, etc. The closer the dots to the diagonal line the more similarities there are, the further the more differences

Q6. What is the main differences between N. Ireland and the other countries of the UK in terms of this data-set?

The first cluster of dots deviate from the diagonal line slightly more than the other plots, it is hard to really say what this means, or even see the differences clearly in the first place b/c we are forced to visually compare with 9 other graphs

First, the data needs to be transposed to work with PCA

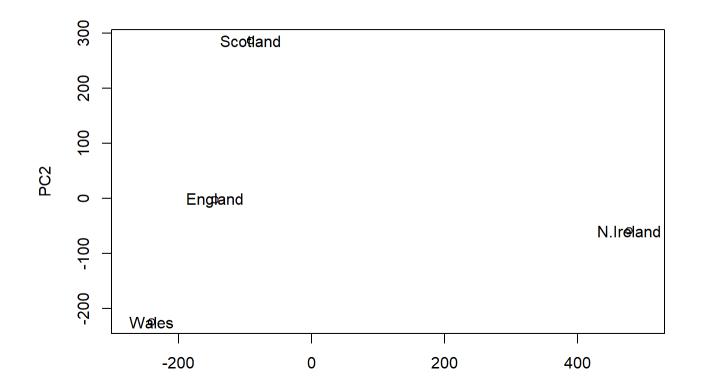
```
pca <- prcomp( t(x) )
summary(pca)</pre>
```

Importance of components:

```
PC1 PC2 PC3 PC4
Standard deviation 324.1502 212.7478 73.87622 3.176e-14
Proportion of Variance 0.6744 0.2905 0.03503 0.000e+00
Cumulative Proportion 0.6744 0.9650 1.00000 1.000e+00
```

Q7. Complete the code below to generate a plot of PC1 vs PC2. The second line adds text labels over the data points.

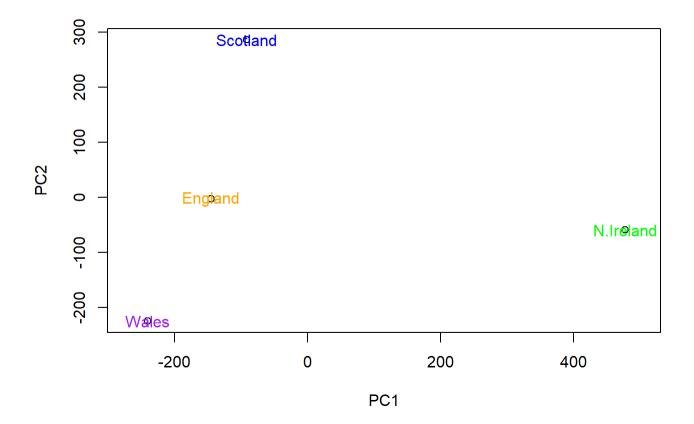
```
plot(pca$x[,1], pca$x[,2], xlab="PC1", ylab="PC2", xlim=c(-270,500))
text(pca$x[,1], pca$x[,2], colnames(x))
```



Q8. Customize your plot so that the colors of the country names match the colors in our UK and Ireland map and table at start of this document.

```
my_colors <- c("orange", "purple", "blue", "green")

plot(pca$x[,1], pca$x[,2], xlab="PC1", ylab="PC2", xlim=c(-270,500))
text(pca$x[,1], pca$x[,2], colnames(x), col = my_colors)</pre>
```



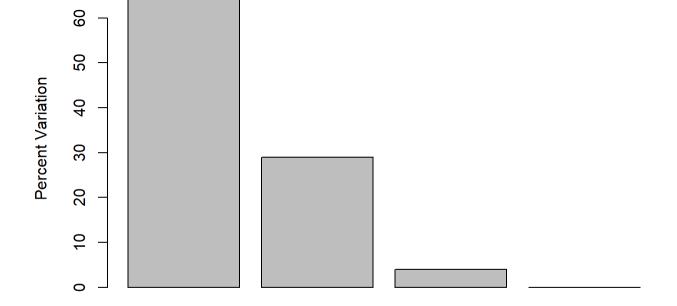
To figure out the variance in the data for each PC we need the standard deviation:

```
v <- round( pca$sdev^2/sum(pca$sdev^2) * 100 )
v</pre>
```

[1] 67 29 4 0

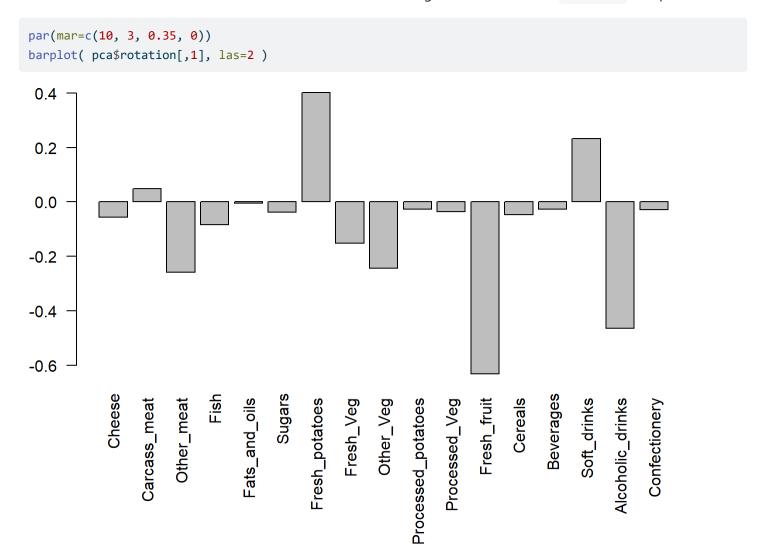
Now to plot the variances

```
barplot(v, xlab="Principal Component", ylab="Percent Variation")
```



Principal Component

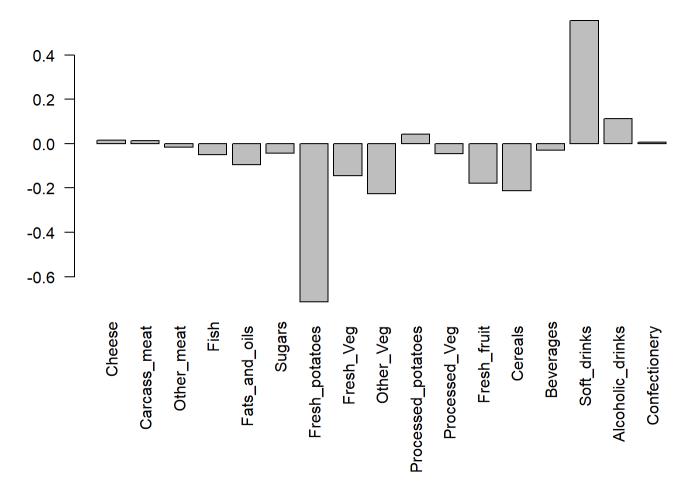
To see the influence of each variable on the PCs, aka the loading scores, we use the \$rotation component:



The highest positive values are what cause skewing to the right in the PC1 vs PC2 plot, which is potatoes and soft drinks here. The lowest negativee values are what cause skewing to the right in the PC1 vs PC2 plot, which is

Q9: Generate a similar 'loadings plot' for PC2. What two food groups feature prominantly and what does PC2 maninly tell us about?

```
par(mar=c(10, 3, 0.35, 0))
barplot( pca$rotation[,2], las=2 )
```



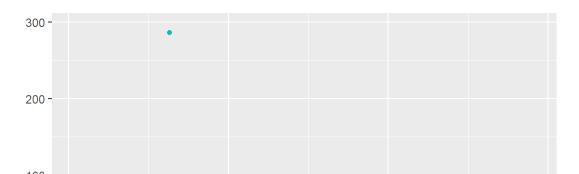
Now we will try to plot the same thing on ggplot2

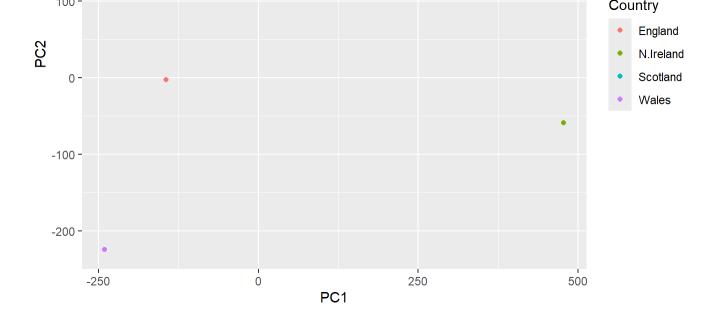
```
library(ggplot2)

df <- as.data.frame(pca$x)

df_lab <- tibble::rownames_to_column(df, "Country")

ggplot(df_lab) +
  aes(PC1, PC2, col=Country) +
  geom_point()</pre>
```

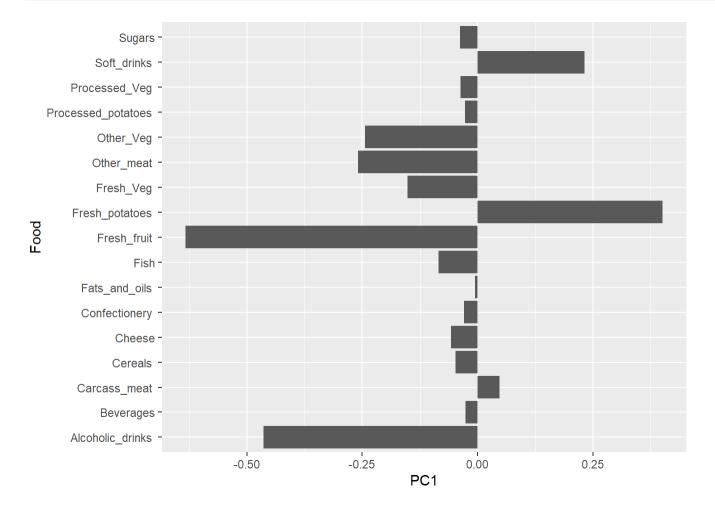




Now to plot the loading plot:

```
ld <- as.data.frame(pca$rotation)
ld_lab <- tibble::rownames_to_column(ld, "Food")

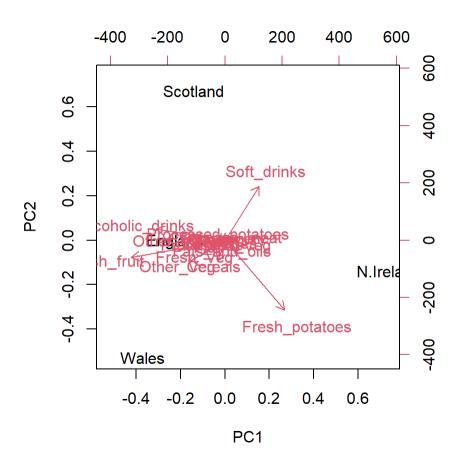
ggplot(ld_lab) +
  aes(PC1, Food) +
  geom_col()</pre>
```



Another way to see this information together with the main PCA plot is in a so-called biplot:

L2.1.4/....

pipiot(bca)



PCA of RNA seq data

```
url2 <- "https://tinyurl.com/expression-CSV"</pre>
rna.data <- read.csv(url2, row.names=1)</pre>
head(rna.data)
                wt3
                     wt4 wt5 ko1 ko2 ko3 ko4 ko5
                     429 420 90 88 86 90 93
gene1 439 458
                408
gene2 219 200
                204
                     210 187 427 423 434 433 426
gene3 1006 989 1030 1017 973 252 237 238 226 210
       783 792
                829
                     856 760 849 856 835 885 894
gene5
       181 249
                204
                     244 225 277 305 272 270 279
                491 491 493 612 594 577 618 638
gene6
       460 502
#Again we have to take the transpose of our data
 pca <- prcomp(t(rna.data), scale=TRUE)</pre>
 summary(pca)
```

Importance of components:

```
PC1 PC2 PC3 PC4 PC5 PC6 PC7

Standard deviation 9.6237 1.5198 1.05787 1.05203 0.88062 0.82545 0.80111

Proportion of Variance 0.9262 0.0231 0.01119 0.01107 0.00775 0.00681 0.00642

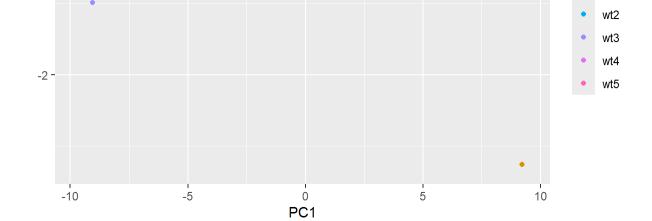
Cumulative Proportion 0.9262 0.9493 0.96045 0.97152 0.97928 0.98609 0.99251
```

```
PC9
                        PC8
Standard deviation
                     0.62065 0.60342 3.457e-15
Proportion of Variance 0.00385 0.00364 0.000e+00
Cumulative Proportion 0.99636 1.00000 1.000e+00
  Q How many genes in this dataset?
 nrow(rna.data)
[1] 100
 head(pca$x)
         PC1
                   PC2
                             PC3
                                       PC4
                                                 PC5
                                                           PC6
wt1 -9.697374 1.5233313 -0.2753567 0.7322391 -0.6749398
wt2 -9.138950 0.3748504 1.0867958 -1.9461655 0.7571209 -0.4369228
wt4 -8.731483 -0.7468371 0.5875748 0.2268129 -1.5404775 -1.2723618
wt5 -9.006312 -0.2945307 -1.8498101 -0.4303812 0.8666124 -0.2496025
ko1 8.846999 2.2345475 -0.1462750 -1.1544333 -0.6947862 0.7128021
           PC7
                      PC8
                                 PC9
                                           PC10
wt1 -0.24446614 1.03519396 0.07010231 3.073930e-15
wt2 -0.03275370 0.26622249 0.72780448 1.963707e-15
wt3 -0.03578383 -1.05851494 0.52979799 2.893519e-15
wt4 -0.52795595 -0.20995085 -0.50325679 2.872702e-15
ko1 -0.07864392 -0.94652648 -0.24613776 4.052314e-15
i will make a main result figure use ggplot:
 library(ggplot2)
 res <- as.data.frame(pca$x)
 ggplot(res) +
   aes(x=PC1, y=PC2, col=rownames(res))+
  geom point()
   2 -
                                                                       rownames(res)
                                                                           ko1
                                                                           ko2
```

0 -

ko3

ko4 ko5 wt1



colnames(rna.data)

[1] "wt1" "wt2" "wt3" "wt4" "wt5" "ko1" "ko2" "ko3" "ko4" "ko5"