# Module 3: Semi-supervised learning (PheCAP)

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```
# Load helper functions.
source(".../Rscripts/helper_function.R")
source(".../Rscripts/ex2_helper_function.R")
```

```
load('../data/CAD_norm_pub.rda')
```

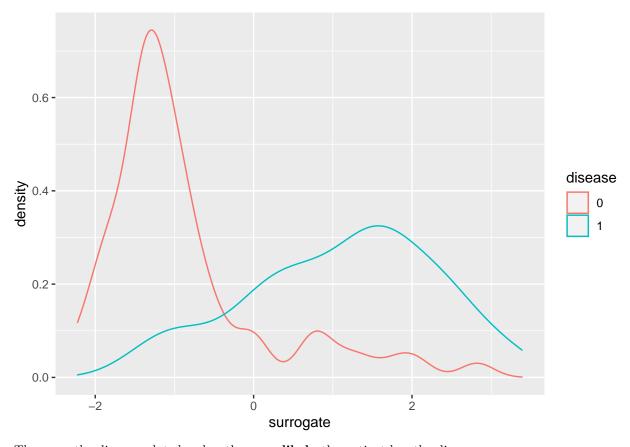
#### Feature selection

#### How to select features?

Can leverage some clinical-meaningful features that are related to Y.

e.g. Feature "surrogate" = the total number of the disease-related billing codes + disease-specific NLP mentions.

```
cbind(label = y, x) %>%
  filter(!is.na(label)) %>%
  mutate(disease = factor(label)) %>%
  ggplot(aes(x = surrogate)) +
  geom_density(aes(color = disease))
```



The more the disease-related codes, the more likely the patient has the disease.

```
nonmissing_index <- which(!is.na(y))
surrogate <- x$surrogate
get_auc(y[nonmissing_index], surrogate[nonmissing_index])</pre>
```

## [1] 0.8877745

We call these highly predictive features of the true disease status "surrogates".

## Opportunities of using surrogate features

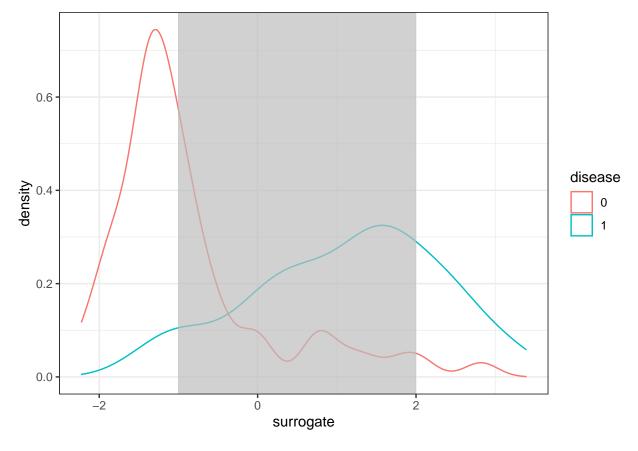
- 1. Feature selection to reduce p
- 2. Algorithm development with limited Y
- 3. Algorithm validation with limited Y

Opportunity 2 and 3 will be covered in the next module!

# Feature selection method

Motivation (Extreme assumption):

- Patients with high main ICD or NLP mentions generally have the phenotype.
- Patients with **extremely** low counts are unlikely to have the phenotype.



- Left white rect: patients not having the disease.
- Right white rect: patients having the disease.

### Prepare data for feature selection

#### Prepare surrogates

Surrogates are available for all the patients!

```
# Prepare 3 surrogates.
surrogate <- x$surrogate

# Prepare features to be selected.
features <- data.matrix(x %>% select(starts_with("COD") | starts_with("NLP")))
```

Run surrogate-assisted feature extraction (SAFE) and show result.

```
# Truncated at 2 and -1.
SAFE <- extreme_method(surrogate, features, u_bound = 2, 1_bound = -1)
SAFE_feature <- colnames(features)[SAFE$beta_select]
SAFE_feature</pre>
```

```
## [1] "NLP56" "NLP93" "NLP160" "NLP161" "NLP176" "NLP231" "NLP304" "NLP306" "HP403" "NLP309" "NLP321" "NLP349" "NLP403" "NLP434" "NLP446" "NLP456" "NLP46" "
```

We select features that occur 50% among the three different surrogate-selected feature sets. This is the idea of majority voting.

Train phenotyping model and show the AUC on the testing set.

- Split data into training and testing set
- Training 60% (n = 106), Testing 40% (n = 75)

```
# Split index.
set.seed(1234)
training_set <- sample(nonmissing_index, size = 106, replace = FALSE)
testing_set <- setdiff(nonmissing_index, training_set)

# Training set.
train_x <- as.matrix(x[training_set, ])
train_y <- y[training_set]

# Testing set.
test_x <- as.matrix(x[testing_set, ])
test_y <- y[testing_set]</pre>
```

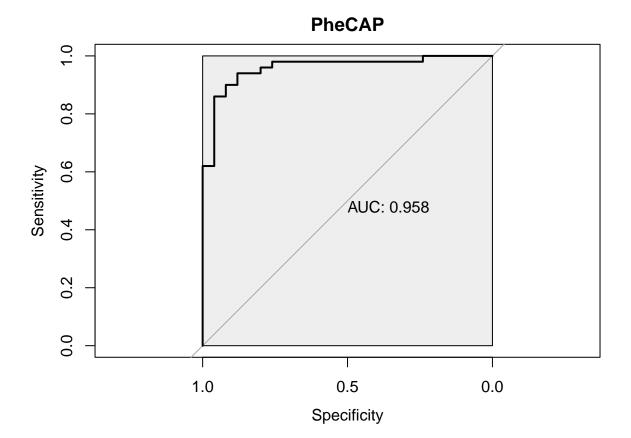
```
selected_features <- c("surrogate", "healthcare_utilization", SAFE_feature)

modelssl_lr <- fit_alasso_bic(
    y = train_y,
    x = train_x[, selected_features]
)

newx <- cbind(1, test_x[, selected_features])

# Prediction on testing set.
y_hat <- expit(newx %*% modelssl_lr$beta_hat) # Inverse Logit

plot(roc(test_y, y_hat),
    print.auc = TRUE,
    max.auc.polygon = TRUE, main = "PheCAP"
)</pre>
```



```
head(get_roc(test_y, y_hat))
```

```
## [1,] 0.9940930 0.006666667 0.00 0.3003125 1.000000 0.4167752 0.4619082 ## [2,] 0.9526009 0.293333333 0.00 0.4601562 1.000000 0.4808415 0.6302836 ## [3,] 0.9111088 0.420000000 0.02 0.6200000 0.9841270 0.5632184 0.7607362 ## [4,] 0.9023323 0.426666667 0.04 0.6800000 0.9714286 0.6000000 0.8000000 ## [5,] 0.8935558 0.433333333 0.04 0.7400000 0.9736842 0.6486486 0.8409091 ## [6,] 0.8326341 0.493333333 0.04 0.8000000 0.9756098 0.7058824 0.8791209
```

```
test_auc <- c()
for (i in round(seq(0.2, 0.7, 0.1) * length(nonmissing_index))) {
    set.seed(123456)
    train_idx <- sample(nonmissing_index, i)
    test_idx <- setdiff(nonmissing_index, train_idx)
    train_data <- data.frame(label = y[train_idx], x[train_idx, ])
    test_data <- data.frame(label = y[test_idx], x[test_idx, ])

train_data <- as.matrix(train_data)
    test_data <- as.matrix(test_data)

# LASSO
metric <- validate_model(
    train_y = train_data$label,
    test_y = test_data$label,
    test_y_hat = lasso_pred(train_data, test_data),</pre>
```

```
train_y_hat = lasso_pred(train_data, train_data)
  )
  # Formatting
  test_auc <- rbind(test_auc, data.frame(</pre>
   n_training = i,
   method = "LASSO",
   median = metric$test AUC,
   L = as.numeric(sub(".*?(\d+\.\d+).*", "\1", metric$test_CI)),
   U = as.numeric(sub(".*\b(\d+\.\d+).*", "\1", metric$test_CI))
  ))
  # ALASSO
  metric <- validate_model(</pre>
   train_y = train_data$label,
   test_y = test_data$label,
   test_y_hat = alasso_pred(train_data, test_data),
   train_y_hat = alasso_pred(train_data, train_data)
  )
  test_auc <- rbind(test_auc, data.frame(</pre>
   n_training = i,
   method = "ALASSO",
   median = metric$test_AUC,
   L = as.numeric(sub(".*?(\d+\.\d+).*", "\1", metric$test_CI)),
   U = as.numeric(sub(".*\\b(\\d+\\.\\d+).*", "\\1", metric$test_CI))
  ))
  # PheCAP
  metric <- validate_model(</pre>
   train_y = train_data$label,
   test_y = test_data$label,
   test_y_hat = phe_pred(train_data, test_data),
   train_y_hat = phe_pred(train_data, train_data)
  )
 test_auc <- rbind(test_auc, data.frame(</pre>
   n_training = i,
   method = "PheCAP",
   median = metric$test_AUC,
   L = as.numeric(sub(".*?(\d+\.\d+).*", "\1", metric$test_CI)),
   U = as.numeric(sub(".*\\b(\\d+\\.\\d+).*", "\\1", metric$test_CI))
 ))
}
# Facet Plot
test_auc %>% ggplot(aes(
 x = n_training, y = median,
 group = method, color = method
)) +
 geom_point() +
 geom_line() +
  geom_errorbar(aes(ymin = L, ymax = U)) +
```

```
facet_grid(. ~ method)
```

Save the data and feature selected for module 4 and model fitting.

```
save(list = ls(), file = "../module4/environment.RData")
```