Module 2:Feature selection

```
# If a package is installed, it will be loaded. If any
## are not, the missing package(s) will be installed
## from CRAN and then loaded.
## First specify the packages of interest
packages <- c(
  "dplyr", "PheCAP", "glmnet", "randomForestSRC", "PheNorm",
  "MAP", "pROC", "mltools", "data.table", "ggplot2"
## Now load or install&load all
package.check <- lapply(</pre>
 packages,
 FUN = function(x) {
   if (!require(x, character.only = TRUE)) {
      install.packages(x, dependencies = TRUE)
      library(x, character.only = TRUE)
   }
 }
)
# load environment from example 1
load("ex1Environment.RData")
```

Prepare data for algorithm development

- Split data into training and testing set
- Training 50%, Testing 50%

```
data("ehr_data")
data <- PhecapData(PheCAP::ehr_data, "healthcare_utilization", "label", 0.5,
    patient_id = "patient_id", seed = 123
)

# Transform Features log(x + 1)
ehr_data[, 3:ncol(ehr_data)] <- log(ehr_data[, 3:ncol(ehr_data)] + 1)

# All Features
all_x <- ehr_data %>% dplyr::select(
    starts_with("COD"), starts_with("NLP"),
    starts_with("main"), healthcare_utilization
)
health_count <- ehr_data$healthcare_utilization

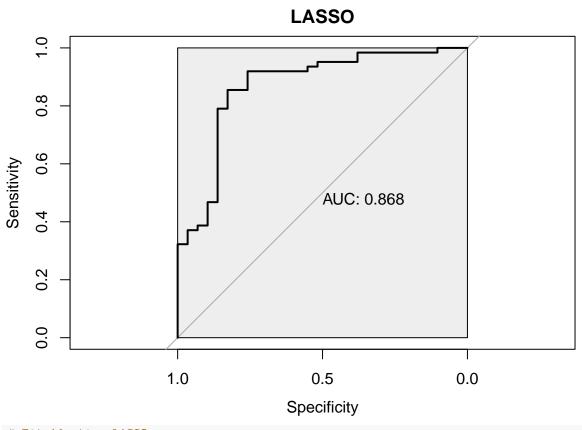
# Training Set</pre>
```

```
train_data <- ehr_data %% dplyr::filter(patient_id %in% data$training_set)
train_x <- train_data %>%
  dplyr::select(
   starts_with("COD"), starts_with("NLP"),
   starts_with("main"), healthcare_utilization
 as.matrix()
train_y <- train_data %>%
  dplyr::select(label) %>%
 pull()
# Testing Set
test_data <- ehr_data %>% dplyr::filter(patient_id %in% data$validation_set)
test_x <- test_data %>%
  dplyr::select(
   starts_with("COD"), starts_with("NLP"),
    starts_with("main"), healthcare_utilization
  ) %>%
 as.matrix()
test_y <- test_data %>%
  dplyr::select(label) %>%
 pull()
```

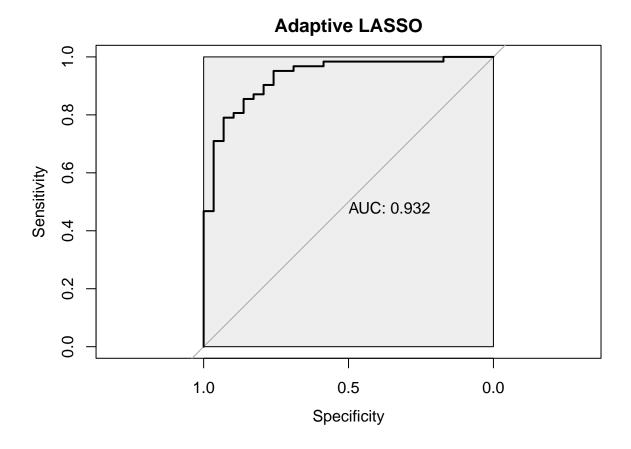
Supervised learning.

- 1. Penalized logistic regression
- Fit LASSO and Adaptive LASSO(ALASSO)

```
# Choose best lambda using CV
model_lr <- cv.glmnet(
    x = train_x,
    y = train_y,
    family = "binomial",
    alpha = 1 # default, LASSO
)
# prediction on testing set
y_hat <- predict(model_lr, newx = test_x, s = "lambda.min", type = "response")
plot(roc(test_y, y_hat),
    print.auc = TRUE,
    max.auc.polygon = TRUE, main = "LASSO"
)</pre>
```



```
# Fit Adaptive LASSO
model_alasso <- fit_alasso_bic(y = train_y, x = train_x)
y_hat <- expit(cbind(1, test_x) %*% model_alasso$beta_hat) # Inverse Logit
plot(roc(test_y, y_hat),
    print.auc = TRUE,
    max.auc.polygon = TRUE, main = "Adaptive LASSO"
)</pre>
```



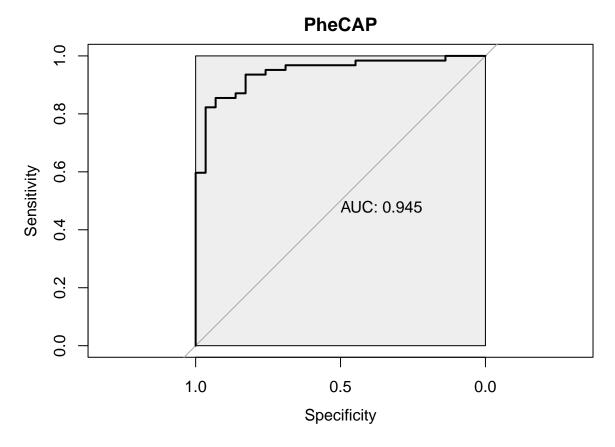
Semi-supervised learning.

1. PheCAP (fit supervised model using select features)

```
other_feature <- c("healthcare_utilization", "main_ICD", "main_NLP")

modelssl_lr <- fit_alasso_bic(
    y = train_y,
    x = train_x[, c(other_feature, SAFE_feature)] # Features selected from ex1
)

# prediction on testing set
y_hat <- expit(cbind(1, test_x[, c(other_feature, SAFE_feature)])
%*% modelssl_lr$beta_hat) # Inverse Logit
plot(roc(test_y, y_hat),
    print.auc = TRUE,
    max.auc.polygon = TRUE, main = "PheCAP"
)</pre>
```



2. Two-step semi-supervised method.

- (i) Regress the surrogate on the features with penalized least square to get the direction of beta.
- (ii) Regress the outcome on the linear predictor to get the intercept and multiplier for the beta.

Step (i):

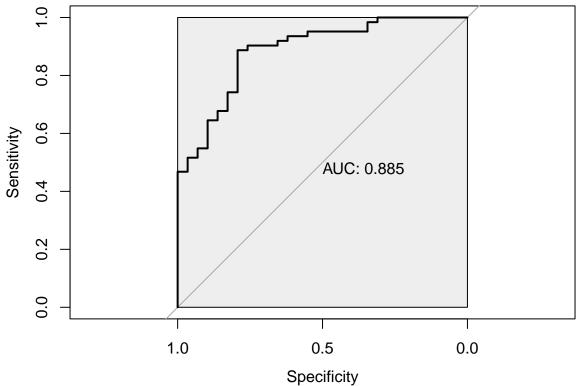
```
model2ssl_step1 <- fit_alasso_bic(
  y = sicdnlp, # surrogate
  x = x, # all X
  family = "gaussian"
)</pre>
```

Step (ii):

```
# linear predictor without intercept
bhatx <- x %*% model2ssl_step1$beta_hat[-1]
# Y ~ beta_hat * X + Main ICD + Main NLP + HU
model2ssl_step2 <- glm(
    train_y ~ bhatx[data$training_set] +
        sicdnlp[data$training_set] +
        health_count[data$training_set])
beta_step2 <- coef(model2ssl_step2)
# recover beta; beta in step 1 is proportional to true beta
beta <- beta_step2[["bhatx[data$training_set]"]] * model2ssl_step1$beta_hat[-1]
# logit = mu
# mu = intercept + beta * X + gamma * S</pre>
```

```
mu <- beta_step2[["(Intercept)"]] +
   as.numeric(x[data$validation_set, ] %*% beta) +
   as.numeric(beta_step2[["sicdnlp[data$training_set]"]] %*%
        sicdnlp[data$validation_set]) +
   as.numeric(beta_step2[["health_count[data$training_set]"]] %*%
        health_count[data$validation_set])

y_hat <- expit(mu) # Inverse logit
plot(roc(test_y, y_hat),
    print.auc = TRUE,
    max.auc.polygon = TRUE, title = "Two Step SS"
)</pre>
```



Weakly-supervised learning.

1. PheNorm

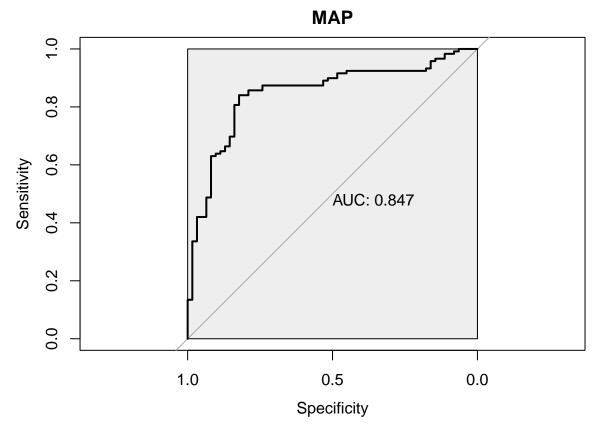
```
# reformat data
data_fit <- data.frame(
    "main_NLP" = snlp,
    "main_ICD" = sicd,
    "healthcare_utilization" = health_count
)

model_phenorm <- PheNorm.Prob(
    nm.logS.ori = c("main_ICD", "main_NLP"), # name of surrogates
    nm.utl = "healthcare_utilization", # name of HU
    nm.X = colnames(ehr_data)[-1:-5], # Other predictors X</pre>
```

```
dat = ehr_data[, -1],
    train.size = nrow(ehr_data)
)

# Since the algorithm does not use any labels to train
# all available labels can be used for validation
y_hat <- model_phenorm$probs[c(data$training_set, data$validation_set)]
plot(roc(ehr_data$label[c(data$training_set, data$validation_set)], y_hat),
    print.auc = TRUE, max.auc.polygon = TRUE, main = "PheNorm"
)</pre>
```


2. MAP



Validation

Different training size

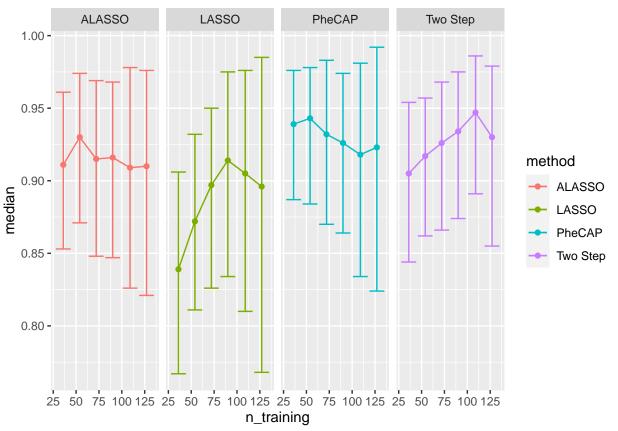
- We can try different number of labels.
- Try % of training from 0.2 to 0.7
- Calculate CI by bootstrap

```
labeled_data <- ehr_data %>% dplyr::filter(!is.na(label))

test_auc <- c()
for (i in round(seq(0.2, 0.7, 0.1) * nrow(labeled_data))) {
   set.seed(123456)
   idx <- sample(labeled_data$patient_id, i)
   train_data <- labeled_data %>% filter(patient_id %in% idx)
```

```
test_data <- labeled_data %>% filter(!(patient_id %in% idx))
idy <- test_data$patient_id</pre>
# LASSO
metric <- validate_model(</pre>
 train_y = train_data$label,
 test_y = test_data$label,
 test y hat = lasso pred(train data, test data),
 train_y_hat = lasso_pred(train_data, train_data)
# Formatting
test auc <- rbind(test auc, data.frame(</pre>
 n_training = i,
 method = "LASSO",
 median = metric$test_AUC,
 L = as.numeric(sub(".*?(\d+\.\d+).*", "\1", metric$test_CI)),
 U = as.numeric(sub(".*\b(\d+\.\d+).*", "\1", metric$test_CI))
))
# ALASSO
metric <- validate_model(</pre>
 train_y = train_data$label,
 test_y = test_data$label,
 test_y_hat = alasso_pred(train_data, test_data),
 train_y_hat = alasso_pred(train_data, train_data)
test_auc <- rbind(test_auc, data.frame(</pre>
 n_training = i,
 method = "ALASSO",
 median = metric$test_AUC,
 L = as.numeric(sub(".*?(\d+\.\d+).*", "\1", metric$test_CI)),
 U = as.numeric(sub(".*\b(\d+\.\d+).*", "\1", metric$test_CI))
))
# PheCAP
metric <- validate_model(</pre>
 train_y = train_data$label,
 test_y = test_data$label,
 test_y_hat = phe_pred(train_data, test_data),
 train_y_hat = phe_pred(train_data, train_data)
)
test_auc <- rbind(test_auc, data.frame(</pre>
 n_training = i,
 method = "PheCAP",
 median = metric$test_AUC,
 L = as.numeric(sub(".*?(\d+\.\d+).*", "\1", metric$test_CI)),
 U = as.numeric(sub(".*\b(\d+\.\d+).*", "\1", metric$test_CI))
))
# PheCAP
```

```
metric <- validate_model(</pre>
    train_y = train_data$label,
    test_y = test_data$label,
    test_y_hat = twostep_pred(train_data, test_data),
    train_y_hat = twostep_pred(train_data, train_data)
  test_auc <- rbind(test_auc, data.frame(</pre>
    n_training = i,
    method = "Two Step",
   median = metric$test_AUC,
   L = as.numeric(sub(".*?(\d+\.\d+).*", "\1", metric$test_CI)),
    U = as.numeric(sub(".*\b(\d+\.\d+).*", "\1", metric$test_CI))
  ))
}
# Facet Plot
test_auc %>% ggplot(aes(
  x = n_training, y = median,
  group = method, color = method
)) +
  geom_point() +
  geom_line() +
  geom_errorbar(aes(ymin = L, ymax = U)) +
  facet_grid(. ~ method)
```



K-fold cross validation

##

```
# To save computational time, k = 5
# Average AUC across K-fold
k_fold(dat = labeled_data)
##
        LASSO
                  ALASSO
                              PheCAP Two Step SS
   0.8970614 0.9017572 0.9299445 0.9013244
```