Codes used in Microbiome Section Report (G2)

a. START in Ubuntu

```
# import my files into qiime2 and create the artefact
qiime tools import \
--input-path demux.qza \
--output-path sequences
```

b. In R Studio

```
# Load required packages
```{r}
library(dada2)
Load sequences
```{r}
path <- "sequences"
list.files(path)
# Read in file names
```{r}
Forward and reverse fastq filenames have format: SAMPLENAME_R1_001.fastq
and SAMPLENAME_R2_001.fastq
fnFs <- sort(list.files(path, pattern="_R1_001.fastq", full.names = TRUE))
fnRs <- sort(list.files(path, pattern="_R2_001.fastq", full.names = TRUE))
Extract sample names, assuming filenames have format:
SAMPLENAME_XXX.fastq
sample.names <- sapply(strsplit(basename(fnFs), "_"), `[`, 1)</pre>
Inspect read quality
```{r}
plotQualityProfile(fnFs[1:2])
```{r}
plotQualityProfile(fnRs[1:2])
```

# c. In Ubuntu using qiime2

--i-table table.qza \

--o-visualization table.qzv \

```
quality control of forward and reverse reads based on histogram
it uses the demux.qza file to trim and truncate reads
the output a rep-seqs file, a table of the sequences, and stats
qiime dada2 denoise-single \
qiime dada2 denoise-paired --i-demultiplexed-segs demux.qza --p-trim-left-f 15 --
p-trunc-len-f 150 --p-trim-left-r 0 --p-trunc-len-r 120 --o-representative-sequences
rep-seqs.qza --o-table table.qza --o-denoising-stats stats.qza
the code creates a visualizations for the Table and Sequences
it tabulates the stats file
an output would be a visualization of the stats file
qiime metadata tabulate \
--m-input-file stats.qza \
--o-visualization stats.qzv
creating a table summarizing the data
it uses the table files
results in an output of a metadata file
giime feature-table summarize \
--i-table table.qza \
--o-visualization table.qzv \
--m-sample-metadata-file metadata.tsv
creating a table of the sequences, output a visualization
it uses the rep-seqs file
results in a visualization of the rep-seqs file
qiime feature-table tabulate-seqs \
--i-data rep-seqs.qza \
--o-visualization rep-seqs.qzv
adjusted the following code by adjusting the names of the files I personalized
giime feature-table summarize \
```

```
--m-sample-metadata-file metadata.tsv
qiime feature-table tabulate-seqs \
--i-data rep-seqs.qza \
--o-visualization rep-seqs.qzv
qiime uses a specific database as a classifier
input the reads
output of the taxonomy
qiime feature-classifier classify-sklearn \
--i-classifier gg-13-8-99-515-806-nb-classifier.qza \
--i-reads rep-seqs.qza \
--o-classification taxonomy.qza
making a visualization of the taxonomy.qza file to a qzv
qiime metadata tabulate \
--m-input-file taxonomy.qza \
--o-visualization taxonomy.qzv
filters the taxonomy, excluding contaminants of mitochondria and chloroplast
genetic material
giime taxa filter-table \
--i-table table.qza \
--i-taxonomy taxonomy.qza \
--p-exclude mitochondria, chloroplast \
--o-filtered-table table.qza
making a visualization of the table.qza file to a qzv (now excluding mitochondria
and chloroplast DNA)
qiime metadata tabulate \
--m-input-file table.qza \
--o-visualization table.qzv
input the table.qza of the data (now excluding mitochondria and chloroplast DNA)
input the metadata file
output of the taxonomy bar plots through a .qzv file
qiime taxa barplot \
--i-table table.qza \
--i-taxonomy taxonomy.qza \
```

- --m-metadata-file metadata.tsv \
- --o-visualization taxa-bar-plots.qzv
- # generating a phylogenetic tree for diversity analysis
- # input sequences
- # output of alignment, masked aligned, unrooted and rooted phylo trees. qiime phylogeny align-to-tree-mafft-fasttree \
- --i-sequences rep-seqs.qza \
- --o-alignment aligned-rep-seqs.qza \
- --o-masked-alignment masked-aligned-rep-seqs.qza \
- --o-tree unrooted-tree.qza \
- --o-rooted-tree rooted-tree.qza
- #generating the phylogenetic core metrics
- # input the phylogenetic tree
- # input the table excluding mitochondria and chloroplast DNA
- # setting parameters for the sampling-depth based on the visualization of the table.qzv file
- # using the metadata file to make an output of the phylogenetic core metrics qiime diversity core-metrics-phylogenetic \
- --i-phylogeny rooted-tree.qza \
- --i-table table.qza \
- --p-sampling-depth \
- --m-metadata-file metadata.tsv \
- --output-dir core-metrics-results
- # generate alpha/beta diversity numbers using a sampling of 1103 qiime diversity core-metrics-phylogenetic \
- --i-phylogeny rooted-tree.qza \
- --i-table table.qza \
- --p-sampling-depth 6741 \
- --m-metadata-file metadata.tsv \
- --output-dir core-metrics-results
- # analysis of the alpha-group-significance, specific to Shannon Diversity and Observed Features
- # input specific core-metric outputs
- # output visualizations of core-metric outputs
- qiime diversity alpha-group-significance \

- --i-alpha-diversity core-metrics-results/shannon vector.gza \
- --m-metadata-file metadata.tsv \
- --o-visualization core-metrics-results/shannon\_vector.qzv

### qiime diversity alpha-group-significance \

- --i-alpha-diversity core-metrics-results/observed\_features\_vector.qza \
- --m-metadata-file metadata.tsv \
- --o-visualization core-metrics-results/observed\_features\_vector.qzv
- # analysis of beta-group-significance, specific to the metadata columns sex, population, and flock
- # input core-metric output from beta analysis
- # specific metadata file
- # specify what column on the metadata file should be used as data
- # output of visualization of the specific core-metric result
- # pair-wise parameters

# qiime diversity beta-group-significance \

- --i-distance-matrix core-metrics-results/bray\_curtis\_distance\_matrix.qza \
- --m-metadata-file metadata.tsv \
- --m-metadata-column sex \
- --o-visualization core-metrics-results/bray curtis distance sex.gzv \
- --p-pairwise

# qiime diversity beta-group-significance \

- --i-distance-matrix core-metrics-results/bray curtis distance matrix.gza \
- --m-metadata-file metadata.tsv \
- --m-metadata-column population \
- --o-visualization core-metrics-results/bray\_curtis\_distance\_population.qzv \
- --p-pairwise

#### qiime diversity beta-group-significance \

- --i-distance-matrix core-metrics-results/bray curtis distance matrix.gza \
- --m-metadata-file metadata.tsv \
- --m-metadata-column flock \
- --o-visualization core-metrics-results/bray curtis distance flock.gza \
- --p-pairwise