PROTOCOL EXCHANGE | COMMUNITY CONTRIBUTED DNA isolation protocol for Vigna

radiata with free of phenolics

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Introduction

Pulses are major constituents of human diet and are mainly grown in marginal lands under rainfed conditions. Among the pulses, blackgram and mungbean are the major pulses contributing much for the total production. Mungbean [Vigna radiata (L.) Wilczek] (2n=22), primarily an inbreeding species is cultivated throughout tropical Asia including India which accounts for 45% of the world production. Vigna species are an important source of protein for people, particularly in tropical Africa and Asia and several Vigna species have been domesticated in Asia. Species in Leguminosae (Fabaceae) genus Vigna subgenus Ceratotropis are called the Asian Vigna because of their natural distribution. Recently the taxonomy of the subgenus has been revised (Tomooka et al., 2002c). On the basis of these taxonomic revisions, a new taxonomic system was proposed for subgenus Ceratotropis that consists of 21 species, of which eight are used as food, fodder or cover crops (Tomooka et al., 2001, 2002a). In the new system, the subgenus was divided into three sections, Angulares, Ceratotropis and Aconitifoliae consisting of 12, 4 and 5 species, respectively.

Subject terms: Plant biology

Keywords: <u>DNA isolation</u> <u>Vigna sp.</u>

Reagents

The materials used for DNA isolation are agroinoculaed leaves, seeds and embryos.

a. Cetyl Trimethyl Ammonium Bromide (CTAB) Extraction Buffer

CTAB 2% W/V

Tris HCI (pH 8.0) 200 mM

Sodium Chloride 1.4 M

EDTA 20 mM

2-mercaptoethanol 0.1% v/v

(Mercaptoethanol was added immediately prior to use)

b. Tris EDTA (TE) Buffer Tris HCI (pH 8.0) 10 mM

EDTA 1 mM

- c. Ice-cold Isopropanol
- d. Chloroform: Isoamylalcohol (24:1 V/V)
- e. Phenol: chloroform (1:1)
- f. Sodium acetate (3.0 M) pH 5.2 (pH adjusted using glacial acetic acid)
- g. Ethanol (100% and 70%)
- h. PVP (1%)

Equipment

- 1.Waterbath
- 2. Centrifuge

Procedure

- 1) Grind the leaves with 600 µl of CTAB buffer, transfer into an eppendorf tube and incubate for 30 min. at 65 °C with occasional mixing after adding PVP and mercaptoethanol.
- 2) Remove the tubes from the water bath and allow to cool at room temperature.
- 3) Add equal volume of chloroform: isoamyl alcohol mixture (24:1) and mix by inversion for 15 min.
- 4) Centrifuge at 4000 rpm for 20 min and transfer the clear aqueous phase to a new sterile tube.
- 5) Add equal volume of phenol: chloroform (1:1. Centrifuged at 4000 rpm for 20 min and transfer the clear aqueous phase to a new sterile tube.
- 6) Add 1/3 volume of ice-cold isopropanol and 3M Sodium acetate and mix gently by inversion.
- 7) Then, centrifuge at 4000 rpm for 20 min to pellet the DNA and discard the supernatant.
- 8) Wash the DNA pellet with 70 per cent alcohol. After washing with 70% alcoholair dry the DNA pellet.
- 9) Discard the alcohol and air dry the DNA pellet completely. Depending upon the size of the pellet, dissolve the DNA in 100-200 μ l of 1X TE (pH 8.0) and store at 4 °C.

Timing

2 hours

Critical Steps

The supernatant in each step should be carefully transferred without disturbing the other layers.

Troubleshooting

Repeat the purification steps if there is a presence of phenolics in DNA.

Anticipated Results

PCR compatible DNA (highly purified DNA) can be isolated by this method.

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Competing financial interests

The authors declare no competing financial interests.

Readers' Comments

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