PROTOCOL EXCHANGE | ***COMMUNITY CONTRIBUTED***

Isolation of mouse bone marrow-derived monocytes

Shicheng Su & **Jiang Liu**

Erwei Song's lab

Abstract

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Subject terms: **Cell biology**

Keywords: **mice** **monocytes**

Reagents

Materials
PBS powder (Sigma, P3813)
Pen/Strep, 100U/100□g /ml (Invitrogen, 15140-122)
recombinant mouse M-CSF (PeproTech, 315-02 or AF-315-02)
75% ethanol solution
DMEM (Sigma, D5648)
Sodium bicarbonate
Sodium chloride
FBS (Invitrogen, 10437-028)
Scissors, sterile
Forceps, sterile
10-mL Syringe with 25-G needle, sterile
Sterile Dishes, 10cm
24-well plates(Costar, Cambridge, MA)
Rotator
Centrifuge polypropylene tubes 15ml, 50ml
Centrifuge(Beckman Allegra X-15R)
Cell Strainer, 70μm (BD Falcon cat#352350)
EDTA

Media
D-MEM (makes 1 L)
D-MEM powder
3.7g sodium bicarbonate
Filter sterilize
10ml pen/strep (100 U/ml penicillin,100 □g/ml streptomycin)
Note: Be sure to check new packages of DMEM for recommended amounts of NaHCO3 and other additives

Antibiotic PBS, 1L
Prepare PBS per package instructions, then supplement with:
10ml pen/strep
Filter sterilize (0.2 μ)

Procedure

Protocol:
1. Sacrifice mouse by cervical dislocation method and immerse whole body thoroughly with 75% ethanol solution 10min.
2. Pull both back legs apart until cracks are heard. With sterile scissors and forceps, cut skin around one of the back legs. Pull skin down towards paw and remove it. Pierce leg with scissors and tear muscle alongside bone by opening scissors. Repeat this on both sides of femur and tibia until both bones are roughly clean. Cut ligaments between femur and hip. Cut bone below the ankle joint.
3. Store femurs and tibias in DMEM on ice before use.
4. Place bones in 75% ethanol for 1 min to ensure sterility, wash twice in sterile PBS.
5. Place femur and tibia in a tube of ice-cold, sterile PBS. Wipe femur and tibia by rubbing with low-lint tissues to remove attached tissue.
6. Separate tibia from femur by bending slightly at the knee joint. Hold femur/tibia with sterile forceps and then remove both epiphyses with sterile scissors.
7. Insert a 25-G needle through the cutted end and flush bone marrow cells into a 50ml sterile tube with medium. While flushing, move the needle up and down while scraping the inside of the bone. Do this until the bone appears white.
8. Mechanically disrupt the marrow plugs by passing through a 19-G needle twice, filter the resulting cell suspension using a 70-μM cell strainer (optional) and centrifuge at 250g for 5 min.
9. Discard the supernatant and replace with DMEM, supplemented with 10% FBS and 100ng/ml recombinant mouse M-CSF. Pipet up and down several times to disaggregate pellet.
10. Plate BM cells in plastic 10cm-plates and culture in 10% FBS DMEM medium supplemented with 100 ng/ml recombinant mouse M-CSF.

11. Replace medium on day 3 and 6 with DMEM, supplemented with 10% FBS and 100ng/ml recombinant mouse M-CSF.
12. On day 7, harvest the BMDMφ by incubating with 10 mL PBS containing 10 mM EDTA, followed by vigorous pipetting.
13. Collect the cell suspension in a 50-mL polypropylene tube and quench with an equal volume of DMEM, supplemented with 10% FBS and 100ng/ml recombinant mouse M-CSF.
14. Centrifuge the cells at 250g for 5 min and plate at 2×10^5 Mφ/1ml media per well of a 24-well plate.

Author information

Affiliations

. **erwei song's lab**
Shicheng Su

. **Unaffiliated**
Jiang Liu

Competing financial interests

All the authors declare no conflict of interest.

Corresponding author

Correspondence to: Shicheng Su (seasonso@163.com)

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Protocol Exchange ISSN 2043-0116