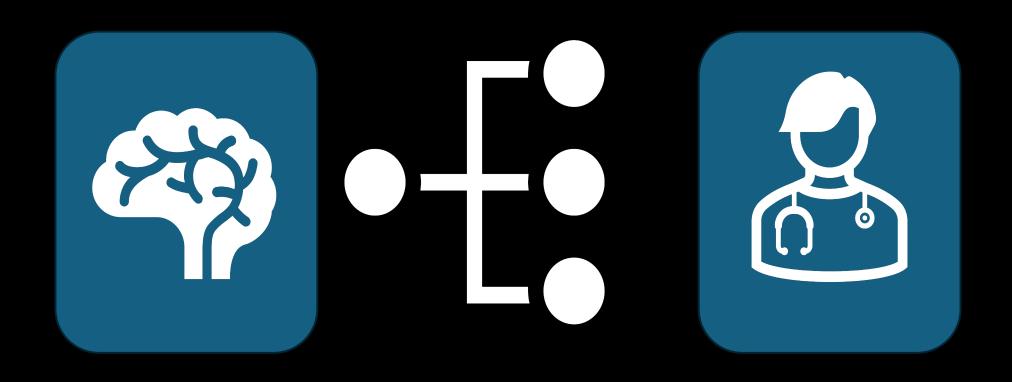
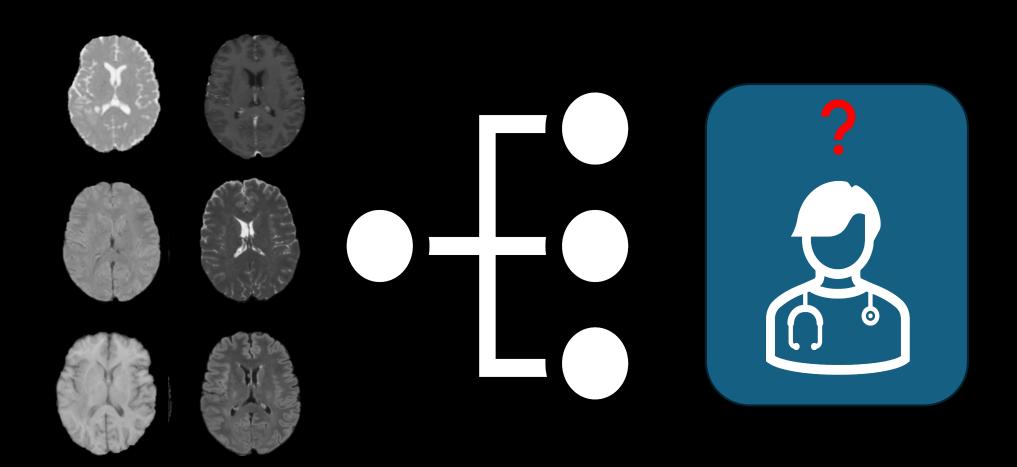
Classifying Multimodal Brain MRI Using a 2D Convolutional Neural Network

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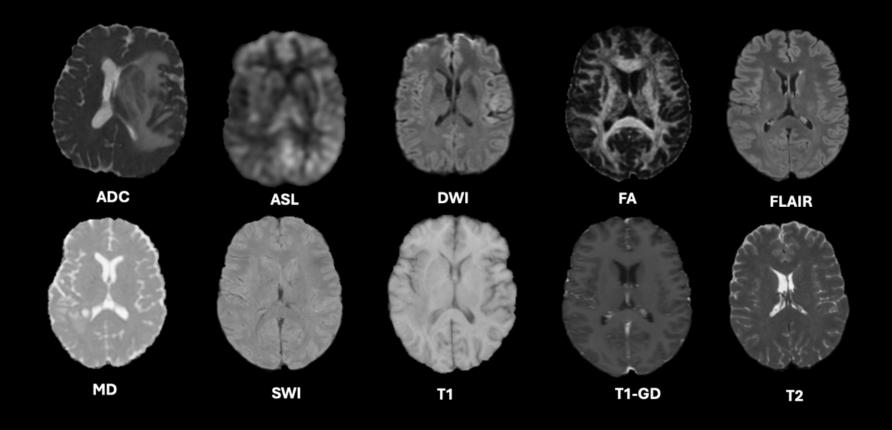


Introduction:



Purpose:

The purpose of this study was to develop a CNN that would automatically classify and label 10 common brain MRI sequences using single slice 2D images.



Methods - Dataset:

The data used was was comprised of **5,010** multiparametric images collected from **501** patients with glioblastoma. The data comes from The Cancer Imaging Archive (TCIA) University of California San Francisco Preoperative Diffuse Glioma MRI (UCSF-PDGM) dataset. Link provided below.



The University of California San Francisco Preoperative Diffuse Glioma MRI (UCSF-PDGM)

Created by Tracy Nolan, last modified on Dec 07, 2023

https://wiki.cancerimagingarchive.net/pages/viewpage.action?pageId=119705830

```
# Make all directories and subdirectories
import os
import numpy as np
import nibabel as nib
import pandas as pd
from sklearn.model selection import StratifiedShuffleSplit
basedir = '/project/radiology/ANSIR lab/shared/s175064 workspace/UCSF IDH Trial/image picker'
os.mkdir(str(basedir) + str('/train'))
os.mkdir(str(basedir) + str('/val'))
os.mkdir(str(basedir) + str('/test'))
image list = ['t1', 't1gd', 't2', 'flair', 'swi', 'adc', 'fa', 'md', 'asl', 'dwi']
for i in range(len(image list)):
    os.mkdir(str(basedir) + str('/train/') + str(image list[i]))
    os.mkdir(str(basedir) + str('/val/') + str(image list[i]))
    os.mkdir(str(basedir) + str('/test/') + str(image list[i]))
```

```
# read in all image paths from csv
base = '/project/radiology/ANSIR lab/shared/s175064 workspace/UCSF IDH Trial/Results'
ADC = pd.read csv(str(base) + str('/ADC.csv'))
   = pd.read csv(str(base) + str('/ASL.csv'))
FA = pd.read csv(str(base) + str('/DTI eddy FA.csv'))
  = pd.read csv(str(base) + str('/DTI eddy MD.csv'))
DWI = pd.read csv(str(base) + str('/DWI bias.csv'))
FLAIR = pd.read csv(str(base) + str('/FLAIR bias.csv'))
SWI = pd.read csv(str(base) + str('/SWI bias.csv'))
T1 = pd.read csv(str(base) + str('/T1 bias.csv'))
T1GD = pd.read csv(str(base) + str('/T1gad bias.csv'))
T2 = pd.read csv(str(base) + str('/T2 bias.csv'))
# get brain mask paths so each image will be completely dark in non-brain areas
mask = ADC['Brain Mask'].to list()
ADC = ADC['Image'].to list()
ASL = ASL['Image'].to list()
FA = FA['Image'].to list()
MD = MD['Image'].to list()
DWI = DWI['Image'].to list()
FLAIR = FLAIR['Image'].to list()
SWI = SWI['Image'].to list()
T1 = T1['Image'].to list()
T1GD = T1GD['Image'].to list()
T2 = T2['Image'].to list()
```

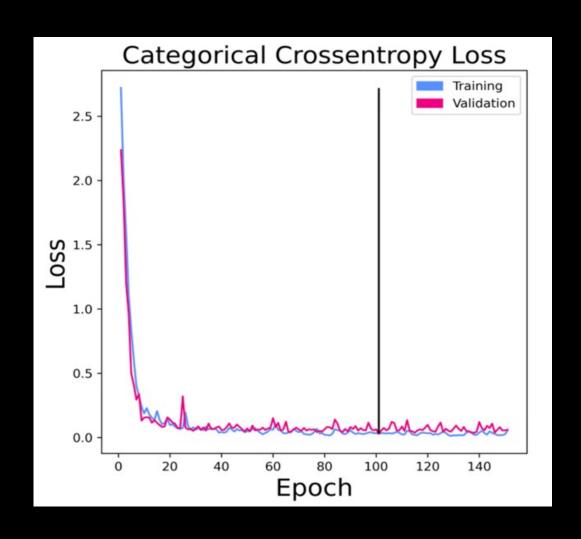
```
# manually define cutoffs for training, validation, and testing
num_subs = len(ADC)
# where validation indices start - use first 80% of data for training
val_cutoff = round(num_subs*0.8)
# where test indices start
test_cutoff = val_cutoff + round((num_subs-val_cutoff)/2)
print(val_cutoff)
print(test_cutoff)
401
451
```

```
# process all images - read in a nifti file and convert to an anxial slice png
def preprocess img(img, i, val cutoff, test cutoff, subfolder, mask):
   from PIL import Image as im
   outdir = '/project/radiology/ANSIR lab/shared/s175064 workspace/UCSF IDH Trial/image picker'
   sub id = img.split('/')[-7]
    mask = nib.load(mask[:])
   mask = np.array(mask.dataobj)
   mask = np.rot90(mask[:, :, 95], 3)
   img = nib.load(img[:])
   img = np.array(img.dataobj)
   # normalize image intensities
    min = np.amin(img[:, :, :])
    max = np.amax(img[:, :, :])
   img = ((img[:, :, :] - min) * (1/(max - min) * 255.0)).astype('uint8')
   img = np.rot90(img[:, :, 95], 3)
   img = im.fromarray(np.multiply(mask, img))
   img = img.convert("L")
   if i <= val cutoff -1:</pre>
        end path = str('/train/') + str(subfolder) + str('/') + str(sub id) + str(' ') + str(subfolder) + str('.png')
   elif i > test cutoff-1:
        end path = str('/test/') + str(subfolder) + str('/') + str(sub id) + str(' ') + str(subfolder) + str('.png')
        end_path = str('/val/') + str(subfolder) + str('/') + str(sub_id) + str('_') + str(subfolder) + str('.png')
   outpath = str(outdir) + str(end path)
   img.save(outpath)
import nibabel as nib
from PIL import Image as im
for i in range(num subs):
   preprocess img(ADC[i], i, val cutoff, test cutoff, 'adc', mask[i])
   preprocess img(ASL[i], i, val cutoff, test cutoff, 'asl', mask[i])
   preprocess img(FA[i], i, val cutoff, test cutoff, 'fa', mask[i])
   preprocess img(MD[i], i, val cutoff, test cutoff, 'md', mask[i])
   preprocess img(DWI[i], i, val cutoff, test cutoff, 'dwi', mask[i])
   preprocess img(FLAIR[i], i, val cutoff, test cutoff, 'flair', mask[i])
   preprocess img(SWI[i], i, val cutoff, test cutoff, 'swi', mask[i])
   preprocess img(T1[i], i, val cutoff, test cutoff, 't1', mask[i])
   preprocess img(TIGD[i], i, val cutoff, test cutoff, 'tlqd', mask[i])
   preprocess img(T2[i], i, val cutoff, test cutoff, 't2', mask[i])
```

```
# Build model and create training/validation batches
from tensorflow import keras
from tensorflow.keras.models import Sequential
from tensorflow.keras.layers import Activation, Dense, Flatten, BatchNormalization, Conv2D, MaxPool2D, Dropout
from tensorflow.keras.optimizers import Adam
from tensorflow.keras.metrics import categorical crossentropy
from tensorflow.keras.preprocessing.image import ImageDataGenerator
from sklearn.metrics import confusion matrix
import tensorflow as tf
from tensorflow.keras.callbacks import EarlyStopping
outdir = '/project/radiology/ANSIR lab/shared/s175064 workspace/UCSF IDH Trial/image picker'
train path = str(outdir) + str("/train")
val path = str(outdir) + str("/val")
img datagen = ImageDataGenerator(rescale=1./255,
                                    fill mode="nearest",
                                    rotation range=180,
                                    width shift range=0.1,
                                    height shift range=0.1,
                                    shear range=0.0,
                                    zoom range=0.8,
                                    vertical flip=1,
                                    horizontal flip=1,
                                    validation split=0.2
train batches = img datagen.flow from directory(directory=train path, target size=(224,224),
                    classes=['adc', 'asl', 'dwi', 'fa', 'flair', 'md', 'swi', 'tl', 'tlqd', 't2'], batch size=128)
img datagen2 = ImageDataGenerator(rescale=1./255, fill mode="nearest")
val batches = img datagen2.flow from directory(directory=val path, target size=(224, 224),
                    classes=['adc', 'asl', 'dwi', 'fa', 'flair', 'md', 'swi', 'tl', 'tlqd', 't2'], batch size=128)
vgg16 model = tf.keras.applications.vgg16.VGG16()
model = Sequential()
for layer in vgg16 model.layers[:-1]:
    model.add(layer)
for layer in model.layers:
    layer.trainable = True
model.add(Dense(10, activation='softmax'))
# View final model architecture
print(model.summary())
```

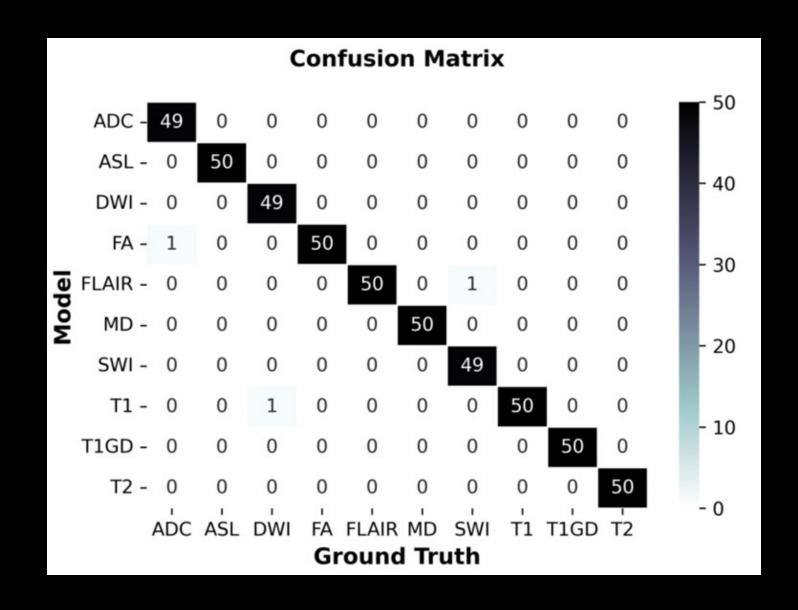
```
# set optimization and how to monitor training
# fit model, save losses, and save model
opt = Adam(learning rate=le-4, decay=le-4 / 500)
model.compile(optimizer=opt, loss='categorical crossentropy', metrics = [tf.keras.metrics.CategoricalCrossentropy()]
monitor = EarlyStopping(monitor='val loss', min delta=le-4, patience=50, verbose=1, mode='auto',
                        restore best weights=True)
history = model.fit(x=train batches,
            steps per epoch=len(train batches),
           validation data=val batches,
            validation steps=len(val batches),
           callbacks = monitor,
            epochs=500,
           verbose=2)
model path = str(outdir) + str("/image picker") + str(".h5")
model.save(model path)
hist df = pd.DataFrame(history.history)
csv path = str(outdir) + str("/image picker") + str(".csv")
hist df.to csv(csv path)
```

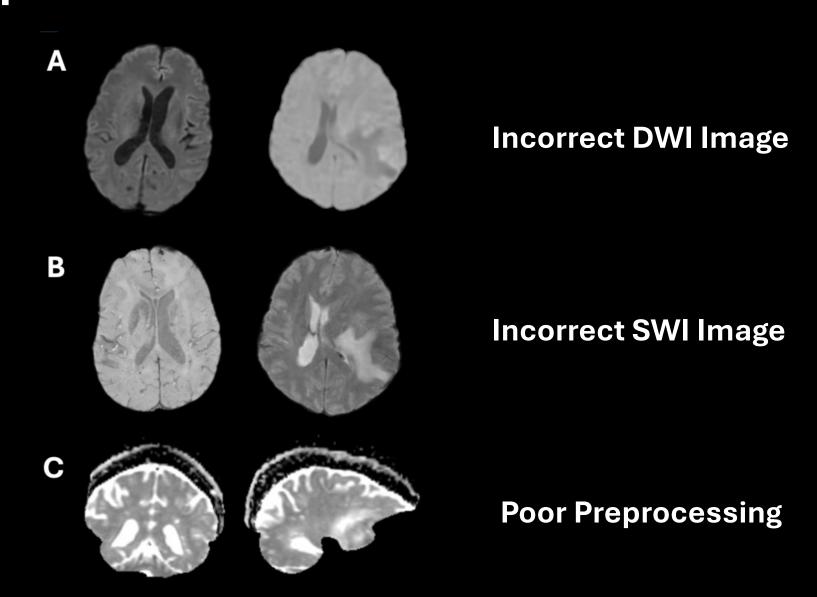
```
# view the losses to make sure the model is learning
import matplotlib.pyplot as plt
import seaborn as sns
import matplotlib.patches as mpatches
f = plt.figure(figsize=(6, 6))
losses = pd.read csv('/project/radiology/ANSIR lab/shared/s175064 workspace/UCSF IDH Trial/image picker/image picker.
color palette = ['#648fff', '#dc267f']
sns.set palette(color palette)
a = sns.lineplot(y=losses['categorical crossentropy'], x=range(1, len(losses)+1))
a.axes.set title("Categorical Crossentropy Loss", fontsize=20)
a = sns.lineplot(y=losses['val categorical crossentropy'], x=range(1, len(losses)+1))
a.set xlabel('Epoch', fontsize=20)
a.set ylabel('Loss', fontsize=20)
plt.vlines(len(losses)-50, min(losses['val categorical crossentropy']),
          max(losses['categorical crossentropy']), color='k')
train = mpatches.Patch(color='#648fff', label='Training')
val = mpatches.Patch(color='#dc267f', label='Validation')
plt.legend(handles=[train, val])
plt.savefig('losses.tiff', dpi=600, format="tiff")
```



```
# reload the model if needed
import keras
model = keras.models.load model("/project/radiology/ANSIR lab/shared/s175064 workspace/UCSF IDH Trial/image picker/im
# Get all test image paths and store in test imagePaths list
import os
test imagePaths = []
# traverse whole directory
for root, dirs, files in os.walk(r'/project/radiology/ANSIR_lab/shared/s175064_workspace/UCSF_IDH_Trial/image_picker/
   # select file name
   for file in files:
        # check the extension of files
        if file.endswith('.png'):
            test imagePaths.append((os.path.join(root, file)))
# sort the list for easy interpretation
test imagePaths.sort()
# Check image paths to make sure they make sense
print(test imagePaths)
```

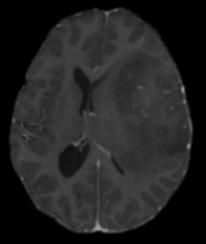
```
import tensorflow as tf
test generator = tf.keras.preprocessing.image.ImageDataGenerator(
    rescale=1./255, fill mode = "nearest")
import keras
test images = test generator.flow from dataframe(
    dataframe=test df,
    x col='id',
    y col='score',
    target size=(224, 224),
    color mode='rgb',
   class mode='raw',
    batch size=1,
    shuffle=False)
Found 500 validated image filenames.
# Get probabilities for each class and predictions
probs = model.predict(test images).tolist()
preds = np.argmax(probs, axis=1)
print(preds)
# Calculate Accuracy
from sklearn.metrics import accuracy score
accuracy = accuracy score(np.array(preds), np.array(test y))
print("Overall Accuracy: ", accuracy)
Overall Accuracy: 0.994
```



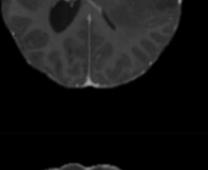


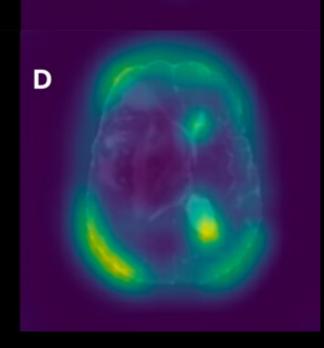
Results GradCAM:

Vessels Used to Classify T1GD Image

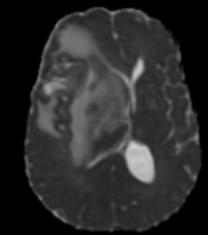








Tumor ignored on ADC image



Conclusion:

- When including poorly curated data, my model is 99.4% accurate.
- When excluding poorly curated data, this model is 100% accurate.
- Both accuracies > all CNN brain MRI sequence classification results currently reported in the literature.
- Image categories are also **greater** than anything currently published in the literature.

Limitations and Future Work:

- My model was trained on imaging from one scanner at one hospital. Future work should compile images from multiple scanners and institutions for training.
- My model is dependent on high quality data free of artifact. A motion correction algorithm or a quality filter may improve model performance in practice.
- My model was only trained for ten categories of structural images. An eleventh "Other" category would need to be added and appropriate examples would need to be included in the training.
- Lastly, some image types cannot be identified on appearance alone. My model would thus benefit by incorporating large language models to read DICOM information.

Thank you!

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