Wk 6 - coding exercise answers

Christine Hawkes

2/16/2022

Coding Exercises

Please submit as a knitted html markdown to GitHub due on 2/23 ##1. run PCoA on clr transformed otus in phyloseq

- Use microbiome::transform for clr transform on ps object
 - This function adds a pseudocount if there are zeros in data
 - Resulting transform will differ from compositions::clr
- Run ordination via phyloseq::ordinate with option PCoA
 - Use euclidean distance on clr == Aitchison's distance
 - Note: alternatively calculate Aitchison's distance via robCompositions::aDist function
- Use phyloseq::plot_scree to evaluate variance explained by each axis
- Plot ordination results
- Access sample scores and eigenvalues for export

##2. run PCoA with clr transformed otus + environmental data using an alternative method

- Select a method for PCoA outside of phyloseq
 - Examples: vegan::wcmdscale,vegan::rda, FactoMineR::PCA, ade4::dudi.pca, stats::prcomp, stats::princomp, ecodist::pco, ape::pcoa
- Examine eigenvalues
- Plot results
- Describe how the clr-based ordination results here and in #1 above differ from the vst results and what this means for analysis
- Access sample scores and eigenvalues for export

```
# retrieve otu matrix
otu clr <- otu table(ps clr)
otu_clr <- as.data.frame(t(otu_clr))</pre>
# calculate dissimilarities
dist_clr <- vegan::vegdist(otu_clr, "euclidean")</pre>
# PCoA in vegan using wcmdscale
ord_clr2 <- vegan::wcmdscale(dist_clr, k=2, eig=TRUE)</pre>
ord_clr2
summary(vegan::eigenvals(ord_clr2))
# envfit
ord_clr2_env <- vegan::envfit(ord_clr2, sam.new, permutations = 99, strata = NULL, choices=c(1,2))
ord_clr2_env
# many plot options
plot(ord_clr2)
ggordiplots::gg_ordiplot(ord=ord_clr2, groups=sam.old$SampleType,
                         kind = "sd", conf = 0.95, pt.size=4)
ggordiplots::gg_envfit(ord=ord_clr2, env=sam.new, perm=99,
                       pt.size=4, alpha= 0.1)
ggordiplots::gg_ordisurf(ord=ord_clr2, env.var=sam.new$pH,
                         binwidth=0.5, pt.size=1, var.label="pH")
ggordiplots::gg_ordisurf(ord=ord_clr2, env.var=sam.new$salinity,
                         binwidth=10, pt.size=1, var.label="salinity")
# accessing results for export
# sample scores
ord_clr2$points
# eigenvalues
ord clr2$eig
# correlations
ord_clr2_env$vectors
```

##3. Examine beta-diversity in a phylogenetic context using DPCoA

- This will take ~10 min to run
 - if longer, consider further reducing GP dataset to top 100-200 taxa
- Analyze the Global Patterns bacteria data with DPCoA in phyloseq::DPCoA
 - https://rdrr.io/bioc/phyloseq/man/DPCoA.html
- Access the resulting list object using \$ to get the Axis 1 and 2 coordinates
- Use data.frame to combine the following into one file for plotting:
 - sample names from SAM\$X.SampleID
 - Axis1 and Axis2 coordinates
 - sample types from SAM\$SampleType
- Plot eigenvalues by axis with phyloseq::plot_scree
- Plot results by samples with plot_ordination using color="SampleType"
- Plot results by species with plot_ordination using color="Phylum"

```
# run DPCoA
ord_dpcoa <- phyloseq::DPCoA(ps_gp_bact)</pre>
# alternative
# ord_dpcoa <- ordinate(ps_qp_bact, "DPCoA")</pre>
# extract data from resulting ord5 list for plotting
str(ord_dpcoa)
Axis1 <- ord_dpcoa$li[["Axis1"]]</pre>
Axis2 <- ord_dpcoa$li[["Axis2"]]</pre>
ord dpcoa data <- data.frame(SAM$X.SampleID, Axis1, Axis2, SAM$SampleType)
# can write the above to csv
# eig plot
phyloseq::plot_scree(ord_dpcoa)
# samples plot
phyloseq::plot_ordination(ps_gp_bact, ord_dpcoa,
                           type="samples", color="SampleType") +
                           ggplot2::geom_point(size=4)
# species plot
phyloseq::plot_ordination(ps_gp_bact, ord_dpcoa,
                           type="species", color="Phylum", title="DPCoA") +
                           ggplot2::geom_point(size=4)
# note: I used separate plots to get around the limit of 6 shapes
```