

Soilpad analysis

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```
knitr::opts_chunk$set(echo = TRUE,  
  cache = TRUE,  
  fig.width = 12,  
  fig.height = 12)
```

```
library("lme4")
```

```
## Loading required package: Matrix
```

```
library("lmerTest")
```

```
##  
## Attaching package: 'lmerTest'
```

```
## The following object is masked from 'package:lme4':  
##  
## lmer
```

```
## The following object is masked from 'package:stats':  
##  
## step
```

```
library("tidyverse"); theme_set(theme_bw())
```

```
## -- Attaching packages ----- tidyverse 1.3.1 --
```

```
## v ggplot2 3.3.5      v purrr  0.3.4
## v tibble  3.1.4      v dplyr  1.0.7
## v tidyr   1.1.3      v stringr 1.4.0
## v readr   2.0.1      v forcats 0.5.1

## -- Conflicts ----- tidyverse_conflicts() --
## x tidyr::expand() masks Matrix::expand()
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()    masks stats::lag()
## x tidyr::pack()   masks Matrix::pack()
## x tidyr::unpack() masks Matrix::unpack()

library("emmeans")
library("ggResidpanel")
library("data.table")

##
## Attaching package: 'data.table'

## The following objects are masked from 'package:dplyr':
##
##   between, first, last

## The following object is masked from 'package:purrr':
##
##   transpose

library("stringr")
#install.packages("rstatix")
library("ggpubr")

## Warning: package 'ggpubr' was built under R version 4.1.3

library("rstatix")

## Warning: package 'rstatix' was built under R version 4.1.3

##
## Attaching package: 'rstatix'

## The following object is masked from 'package:stats':
##
##   filter

library("broom")

## Warning: package 'broom' was built under R version 4.1.3
```

```
options(width = 120)
```

```
dir.create("fig", showWarnings = FALSE)
```

```
sessionInfo()
```

```
## R version 4.1.1 (2021-08-10)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 19044)
##
## Matrix products: default
##
## locale:
## [1] LC_COLLATE=English_United States.1252 LC_CTYPE=English_United States.1252 LC_MONETARY=English_United States.1252
## [4] LC_NUMERIC=C LC_TIME=English_United States.1252
##
## attached base packages:
## [1] stats graphics grDevices utils datasets methods base
##
## other attached packages:
## [1] broom_0.7.12 rstatix_0.7.0 ggpubr_0.4.0 data.table_1.14.0 ggResidpanel_0.3.0
## [7] forcats_0.5.1 stringr_1.4.0 dplyr_1.0.7 purrr_0.3.4 readr_2.0.1
## [13] tibble_3.1.4 ggplot2_3.3.5 tidyverse_1.3.1 lmerTest_3.1-3 lme4_1.1-27.1
##
## loaded via a namespace (and not attached):
## [1] nlme_3.1-152 fs_1.5.0 lubridate_1.7.10 http_1.4.2 numDeriv_2016.8
## [6] tools_4.1.1 backports_1.2.1 utf8_1.2.2 R6_2.5.1 DBI_1.1.1
## [11] lazyeval_0.2.2 colorspace_2.0-2 withr_2.4.2 tidyselct_1.1.1 curl_4.3.2
## [16] compiler_4.1.1 cli_3.0.1 rvest_1.0.1 xml2_1.3.2 plotly_4.9.4.1
## [21] scales_1.1.1 DEoptimR_1.0-9 mvtnorm_1.1-2 robustbase_0.93-8 digest_0.6.27
## [26] foreign_0.8-81 minqa_1.2.4 rmarkdown_2.10 qqplotr_0.0.5 rio_0.5.27
## [31] pkgconfig_2.0.3 htmltools_0.5.1.1 dbplyr_2.1.1 htmlwidgets_1.5.3 rlang_0.4.11
## [36] readxl_1.3.1 rstudioapi_0.13 generics_0.1.0 jsonlite_1.7.2 zip_2.2.0
## [41] car_3.0-11 magrittr_2.0.1 Rcpp_1.0.7 munsell_0.5.0 fansi_0.5.0
## [46] abind_1.4-5 lifecycle_1.0.0 stringi_1.7.3 yaml_2.2.1 carData_3.0-4
## [51] MASS_7.3-54 grid_4.1.1 crayon_1.4.1 lattice_0.20-44 haven_2.4.3
## [56] cowplot_1.1.1 splines_4.1.1 hms_1.1.0 knitr_1.33 pillar_1.6.2
## [61] boot_1.3-28 estimability_1.3 ggsignif_0.6.3 reprex_2.0.1 glue_1.4.2
## [66] evaluate_0.14 modelr_0.1.8 vctrs_0.3.8 nloptr_1.2.2.2 tzdb_0.1.2
## [71] cellranger_1.1.0 gtable_0.3.0 assertthat_0.2.1 openxlsx_4.2.4 xfun_0.25
## [76] xtable_1.8-4 viridisLite_0.4.0 ellipsis_0.3.2
```

Read in data

```
library("tidyverse")
```

```
flume <- read_csv("../data/tidy/flume_event_data612_UPDATE.csv") %>%
  mutate(Year = factor(Year)) %>%
  subset(subtreatment != 'grass strip') %>%
  subset(SiteID != 'MCN') %>%
```

```

subset(subset=!(SiteID=="RHO" & Year == 2016)) %>%
subset(subset=!(SiteID=="RHO" & Year == 2017))

## Rows: 432 Columns: 19

## -- Column specification -----
## Delimiter: ","
## chr (7): SiteID, subtreatment, Treatment, sampleID, random, crop, f_loc
## dbl (12): precipitation, rain_time, rf_event, sample_event, ro_event, Year, flow_time, flow, tss_sum

##
## i Use 'spec()' to retrieve the full column specification for this data.
## i Specify the column types or set 'show_col_types = FALSE' to quiet this message.

flume_sum <- flume %>%
  group_by(Treatment, Year, SiteID, sample_event, tss_sum) %>%
  summarize(tss_load = tss_sum,
            ln_tss_load = log(tss_load+0.000198)) %>%
  distinct()

## 'summarise()' has grouped output by 'Treatment', 'Year', 'SiteID', 'sample_event', 'tss_sum'. You can

ppt_sum <- flume %>%
  group_by(Treatment, Year, SiteID, sample_event) %>%
  summarize(ppt_sum = sum(precipitation)) %>%
  ungroup() %>%
  filter(!duplicated(cbind(Year, SiteID, sample_event)))

## 'summarise()' has grouped output by 'Treatment', 'Year', 'SiteID'. You can override using the '.group

sample_anova <- flume_sum %>%
  filter(!is.na(tss_sum)) %>%
  select(Year, SiteID, Treatment, sample_event, tss_sum) %>%
  group_by(SiteID, Year, Treatment, sample_event) %>%
  summarize(tss_load = sum(tss_sum)) %>%
  ungroup() %>%
  select(Year, SiteID, Treatment, sample_event, tss_load) %>%
  pivot_wider(names_from = Treatment, values_from = tss_load)

## 'summarise()' has grouped output by 'SiteID', 'Year', 'Treatment'. You can override using the '.group

pivot_sample <- sample_anova %>%
  inner_join(ppt_sum, by=c("SiteID", "Year", "sample_event")) %>%
  filter(!is.na(strips)) %>%
  mutate(ln_ppt = log(ppt_sum),
         diff = strips-control,
         ln_diff = log(abs(diff)+0.00165),
         ln_ctl = log(control+0.005322915),
         ln_trt = log(strips+0.0104)) %>%
  subset(select = -c(Treatment))

```

```

long_load <- pivot_sample %>%
  gather(Treatment, tss_load, control:strips) %>%
  arrange(Treatment, tss_load) %>%
  filter(!is.na(diff)) %>%
  select(SiteID, Treatment, Year, sample_event, tss_load, diff, ppt_sum)

```

```

rf_ro_pivot <- long_load %>%
  mutate(random = (ifelse(SiteID == 'ARM', 'NR',
    ifelse(SiteID == 'EIA', 'R',
    ifelse(SiteID == 'MCN', 'R',
    ifelse(SiteID == 'HOE', 'NR',
    ifelse(SiteID == 'MAR', 'NR',
    ifelse(SiteID == 'RHO', 'R',
    ifelse(SiteID == 'WHI', 'NR',
    ifelse(SiteID == 'WOR', 'R', 0))))))))))

```

```

long_load <- long_load %>%
  mutate(random = (ifelse(SiteID == 'ARM', 'NR',
    ifelse(SiteID == 'EIA', 'R',
    ifelse(SiteID == 'MCN', 'R',
    ifelse(SiteID == 'HOE', 'NR',
    ifelse(SiteID == 'MAR', 'NR',
    ifelse(SiteID == 'RHO', 'R',
    ifelse(SiteID == 'WHI', 'NR',
    ifelse(SiteID == 'WOR', 'R', 0))))))))))

```

```

full_df <- rf_ro_pivot %>%
  inner_join(ppt_sum, by=c("SiteID", "Year", "sample_event")) %>%
  drop_na(tss_load) %>%
  mutate(ppt_sum = ppt_sum.x,
    ln_ppt = log(ppt_sum),
    ln_tss_load = log(tss_load+0.005322915),
    Treatment = Treatment.x) %>%
  subset(select = -c(Treatment.y, Treatment.x, ppt_sum.x, ppt_sum.y)) %>%
  arrange(Year, SiteID, Treatment, sample_event)

```

```

#write.csv(full_df, "D:/ISU/ResearchProject/flume_analysis/data/tidy/full_df.csv", row.names = FALSE)

```

```

load("full_df.RData")

```

Exploratory analysis

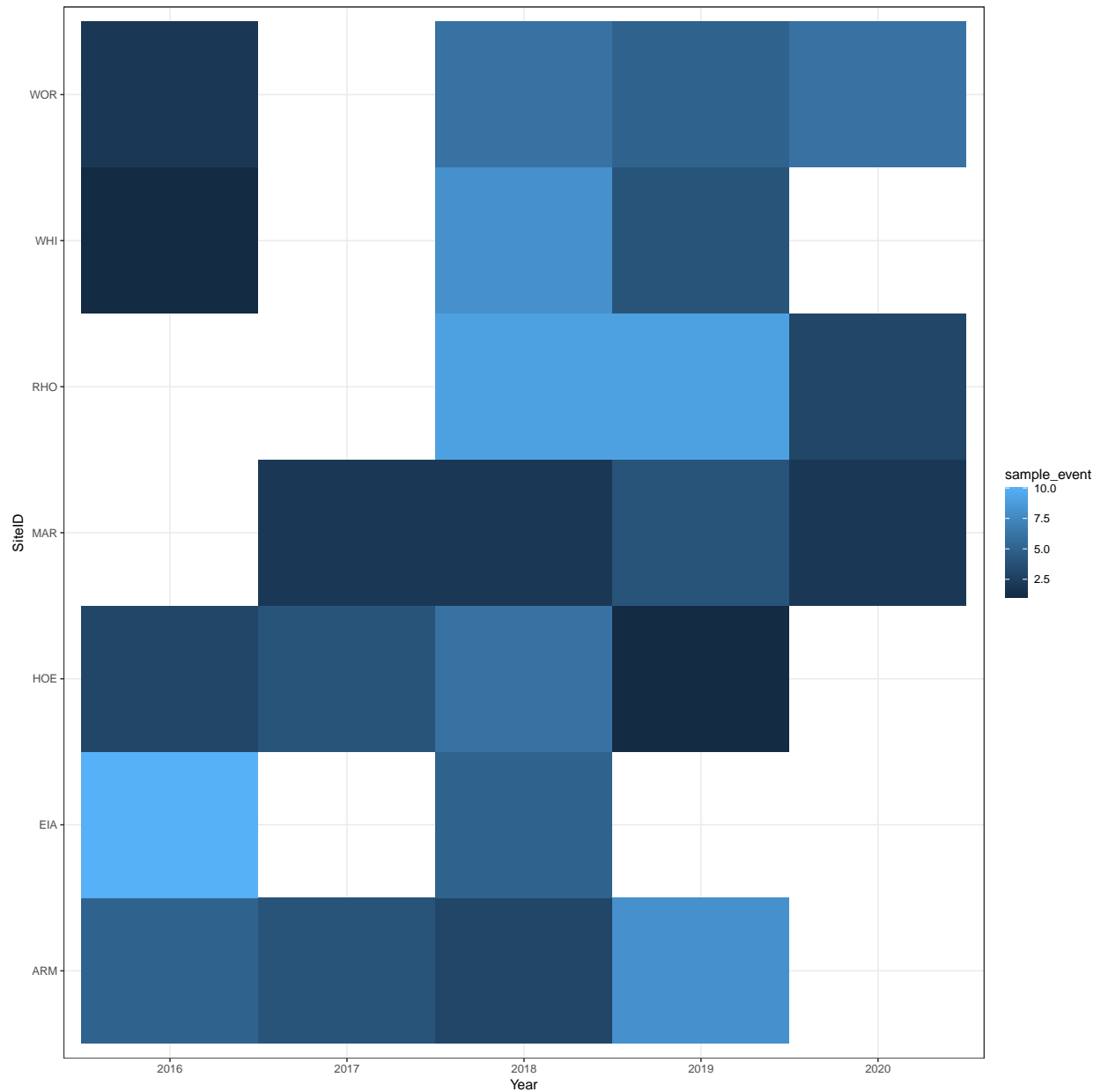
Site-year with rainfall event

```

site_year_rfevent <- full_df %>%
  select(SiteID, Year, sample_event) %>%
  unique()

ggplot(site_year_rfevent, aes(Year, SiteID, fill=sample_event)) +
  geom_tile()

```



Number of samples

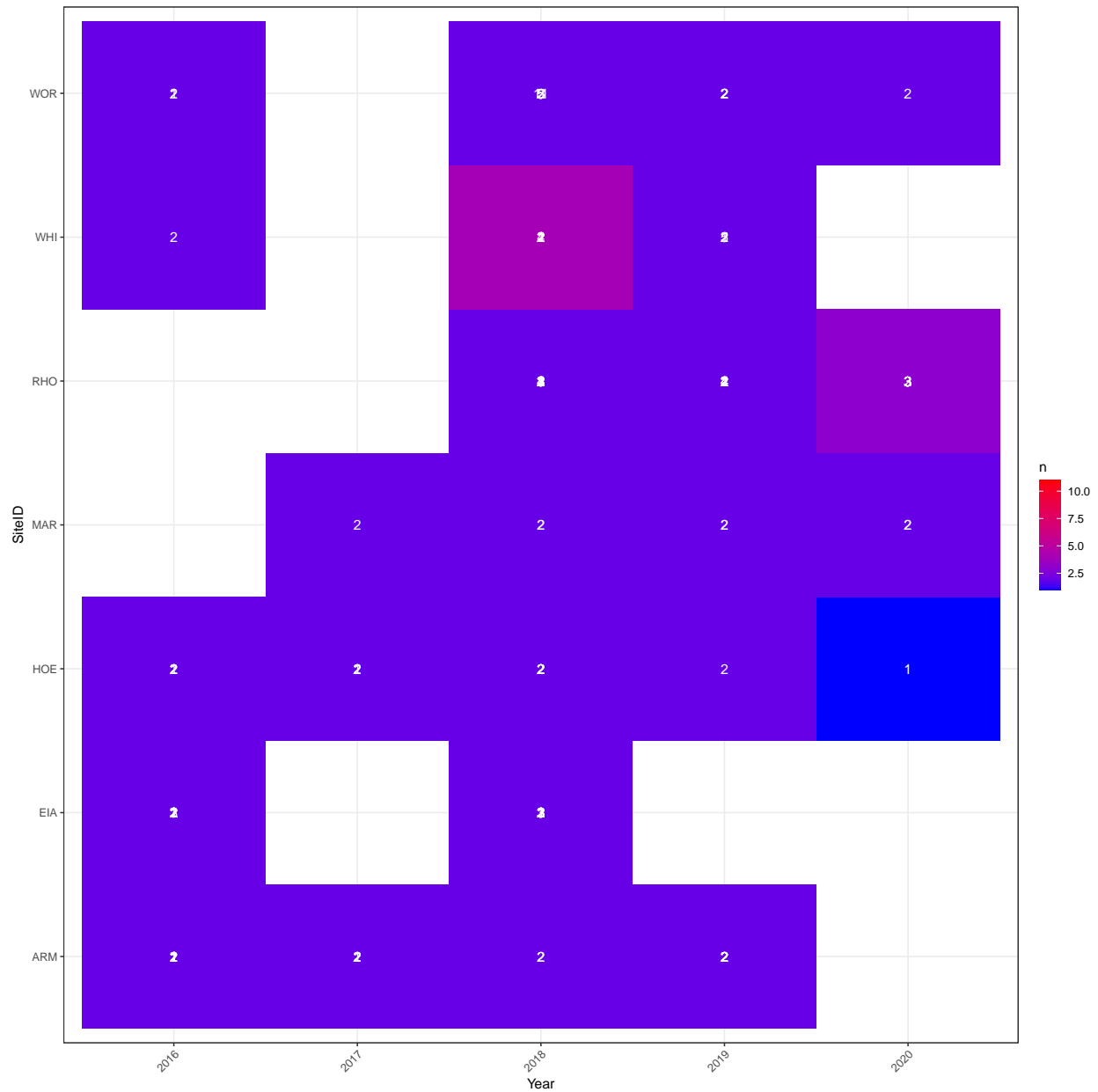
Calculate the number of observations for each treatment-position-year-site-time combination.

```
TSS_counts <- flume %>%
  group_by(Year, SiteID, rf_event) %>%
  distinct() %>%
  summarize(n = n(), .groups = "drop")
```

Plot the number of observations for each combination.

```
g <- ggplot(TSS_counts, aes(x = Year, y= SiteID, fill = n)) +
  geom_tile() +
  geom_text(aes(label = n), color = "white") +
  scale_fill_gradient(low = "blue", high = "red") +
  theme(axis.text.x = element_text(angle = 45, hjust = 1))
```

g



```
##ggsave("fig/soilpad_counts_no_diversion.png", g, width = 12, height = 12)
```

Data visualization

```
#h <- ggplot(full_df, aes(x=ln_ppt, y=ln_tss_load, color=Treatment), inherit.aes = FALSE) +  
# geom_point() +  
# geom_smooth(method=lm, se=FALSE, fullrange=TRUE) +  
# ggtitle("Log-log relationship between TSS load and rainfall accumulation \n(full dataset)") +  
# theme(plot.title = element_text(size=14, face="bold", hjust = 0.5))  
  
#ggsave("fig/randReg_ppt_load.png", h, width = 12, height = 12)  
  
#pivot_sample %>%  
# anova_test(ln_trt ~ ln_ppt*ln_ctl)  
## purr https://stackoverflow.com/questions/50702152/compare-models-via-anova-with-purrr-or-dplyr  
## anova() and may need an linear model built up.
```

Main Analyses

There are three main analyses of interest:

- confirmatory, design-based analysis
- exploratory, covariate analysis
- relationship of sediment flow to sediment loss

Confirmatory, design-based analysis

Treatment effect

```
m_flume <- lmerTest::lmer(log(tss_load+0.005322915) ~  
  (1 | SiteID) +  
  (1 | SiteID:Treatment) +  
  #(1|SiteID:Treatment:sample_event) +  
  
  Treatment*ln_ppt +  
  Treatment*crop +  
  
  Year,  
  data = full_df)
```

```
## boundary (singular) fit: see ?isSingular
```

```
summary(m_flume)
```

```
## Linear mixed model fit by REML. t-tests use Satterthwaite's method ['lmerModLmerTest']  
## Formula: log(tss_load + 0.005322915) ~ (1 | SiteID) + (1 | SiteID:Treatment) +  
##      Treatment * ln_ppt + Treatment * crop + Year  
##      Data: full_df
```



```
##
## REML criterion at convergence: 965.3
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -3.3778 -0.5772  0.0148  0.6231  2.4739
##
## Random effects:
##   Groups             Name             Variance Std.Dev.
## SiteID:Treatment (Intercept) 2.785e+00 1.669e+00
## SiteID              (Intercept) 5.469e-09 7.396e-05
## Residual                        6.292e+00 2.508e+00
## Number of obs: 204, groups: SiteID:Treatment, 14; SiteID, 7
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)      5.0023     1.2967 100.4190   3.858 0.000202 ***
## Treatmentstrips    -4.7733     1.7434  89.0443  -2.738 0.007465 **
## ln_ppt             1.6719     0.3141 184.6241   5.323 2.93e-07 ***
## cropsoybean       -1.3309     0.6687 192.8648  -1.990 0.047968 *
## Year2017           1.3665     0.7684 189.8879   1.778 0.076928 .
## Year2018           1.1635     0.5474 188.9205   2.126 0.034827 *
## Year2019           2.0982     0.5994 191.2516   3.501 0.000578 ***
## Year2020          -0.4797     0.9251 192.7128  -0.518 0.604715
## Treatmentstrips:ln_ppt -0.6660     0.4426 184.7193  -1.505 0.134094
## Treatmentstrips:cropsoybean 1.5244     0.9361 191.5921   1.629 0.105062
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) Trtmnt ln_ppt crpsyb Yr2017 Yr2018 Yr2019 Yr2020 Trtm:_
## Trtmntstrps -0.672
## ln_ppt       0.759 -0.572
## cropsoybean -0.226  0.140 -0.012
## Year2017     -0.163  0.000  0.047  0.021
## Year2018     -0.300  0.000  0.019  0.126  0.453
## Year2019     -0.249  0.000  0.069  0.116  0.448  0.685
## Year2020     -0.196  0.000  0.017  0.054  0.293  0.475  0.478
## Trtmntstr:_ -0.546  0.812 -0.705  0.012  0.000  0.000  0.000  0.000
## Trtmntstrp:  0.134 -0.200  0.012 -0.700  0.000  0.000  0.000  0.000 -0.017
## optimizer (nloptwrap) convergence code: 0 (OK)
## boundary (singular) fit: see ?isSingular
```

```
m_flume_step <- step(m_flume, reduce.random = FALSE, alpha.fixed = 0.1)
```

```
## boundary (singular) fit: see ?isSingular
## boundary (singular) fit: see ?isSingular
## boundary (singular) fit: see ?isSingular
```

```
m_flume_model <- get_model(m_flume_step)
summary(m_flume_model)
```

```
## Linear mixed model fit by REML. t-tests use Satterthwaite's method ['lmerModLmerTest']
```

```
## Formula: log(tss_load + 0.005322915) ~ (1 | SiteID) + (1 | SiteID:Treatment) + Treatment + ln_p
## Data: full_df
##
## REML criterion at convergence: 973.8
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -3.1861 -0.6003  0.0621  0.6738  2.3623
##
## Random effects:
## Groups           Name             Variance Std.Dev.
## SiteID:Treatment (Intercept) 2.675      1.636
## SiteID            (Intercept) 0.000      0.000
## Residual                                6.415      2.533
## Number of obs: 204, groups: SiteID:Treatment, 14; SiteID, 7
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)      3.3971      1.0457  57.2941   3.249 0.001942 **
## Treatmentstrips  -2.1234      0.9497  11.6541  -2.236 0.045760 *
## ln_ppt            1.3379      0.2250  187.4208   5.946 1.32e-08 ***
## Year2017          1.3846      0.7750  193.4757   1.787 0.075546 .
## Year2018          1.2787      0.5436  195.0865   2.352 0.019651 *
## Year2019          2.2130      0.5964  196.7014   3.711 0.000269 ***
## Year2020         -0.4016      0.9304  196.5760  -0.432 0.666478
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) Trtmnt ln_ppt Yr2017 Yr2018 Yr2019
## Trtmntstrps -0.454
## ln_ppt       0.658  0.000
## Year2017     -0.196  0.000  0.067
## Year2018     -0.340  0.000  0.029  0.455
## Year2019     -0.277  0.000  0.100  0.449  0.675
## Year2020     -0.228  0.000  0.026  0.291  0.470  0.473
## optimizer (nloptwrap) convergence code: 0 (OK)
## boundary (singular) fit: see ?isSingular
```

<https://campus.datacamp.com/courses/hierarchical-and-mixed-effects-models-in-r/linear-mixed-effect-models>

```
trt_yr = emmeans(m_flume, pairwise ~ Treatment|Year,
                  type = "response",
                  lmer.df = "asymptotic")
confint(trt_yr)$contrasts
```

```
## Year = 2016:
## contrast      ratio    SE df asymp.LCL asymp.UCL
## control / strips 6.73 6.57 Inf    0.995    45.6
##
## Year = 2017:
## contrast      ratio    SE df asymp.LCL asymp.UCL
## control / strips 6.73 6.57 Inf    0.995    45.6
```

```
##
## Year = 2018:
## contrast      ratio    SE  df asymp.LCL asymp.UCL
## control / strips 6.73 6.57 Inf      0.995      45.6
##
## Year = 2019:
## contrast      ratio    SE  df asymp.LCL asymp.UCL
## control / strips 6.73 6.57 Inf      0.995      45.6
##
## Year = 2020:
## contrast      ratio    SE  df asymp.LCL asymp.UCL
## control / strips 6.73 6.57 Inf      0.995      45.6
##
## Results are averaged over the levels of: crop
## Degrees-of-freedom method: asymptotic
## Confidence level used: 0.95
## Intervals are back-transformed from the log scale
```

```
trt = emmeans(m_flume, pairwise ~ Treatment,
              type = "response",
              lmer.df = "asymptotic")
```

```
## NOTE: Results may be misleading due to involvement in interactions
```

```
confint(trt)
```

```
## $emmeans
## Treatment response      SE  df asymp.LCL asymp.UCL
## control      0.885 0.6249 Inf      0.2198      3.518
## strips      0.127 0.0928 Inf      0.0281      0.518
##
## Results are averaged over the levels of: crop, Year
## Degrees-of-freedom method: asymptotic
## Confidence level used: 0.95
## Intervals are back-transformed from the log(mu + 0.005) scale
##
## $contrasts
## contrast      ratio    SE  df asymp.LCL asymp.UCL
## control / strips 6.73 6.57 Inf      0.995      45.6
##
## Results are averaged over the levels of: crop, Year
## Degrees-of-freedom method: asymptotic
## Confidence level used: 0.95
## Intervals are back-transformed from the log scale
```

```
year = emmeans(m_flume, ~ Year,
              type = "response",
              lmer.df = "asymptotic")
confint(year)
```

```
## Year response      SE  df asymp.LCL asymp.UCL
## 2016  0.1444 0.0946 Inf      0.0381      0.511
```

```
## 2017 0.5818 0.4563 Inf 0.1227 2.688
## 2018 0.4739 0.2615 Inf 0.1592 1.391
## 2019 1.2150 0.7074 Inf 0.3865 3.796
## 2020 0.0873 0.0821 Inf 0.0110 0.521
##
## Results are averaged over the levels of: Treatment, crop
## Degrees-of-freedom method: asymptotic
## Confidence level used: 0.95
## Intervals are back-transformed from the log(mu + 0.005) scale
```

```
trt_ppt = emmeans(m_flume, pairwise ~ Treatment|ln_ppt,
                  at=list(ln_ppt=c(-4,-3,-2)),
                  type = "response",
                  lmer.df = "asymptotic")

confint(trt_ppt)$contrasts ## exp. the values
```

```
## ln_ppt = -4:
## contrast      ratio    SE df asymp.LCL asymp.UCL
## control / strips 3.85 4.00 Inf      0.50      29.6
##
## ln_ppt = -3:
## contrast      ratio    SE df asymp.LCL asymp.UCL
## control / strips 7.49 7.33 Inf      1.10      51.0
##
## ln_ppt = -2:
## contrast      ratio    SE df asymp.LCL asymp.UCL
## control / strips 14.57 16.13 Inf      1.67     127.5
##
## Results are averaged over the levels of: crop, Year
## Degrees-of-freedom method: asymptotic
## Confidence level used: 0.95
## Intervals are back-transformed from the log scale
```

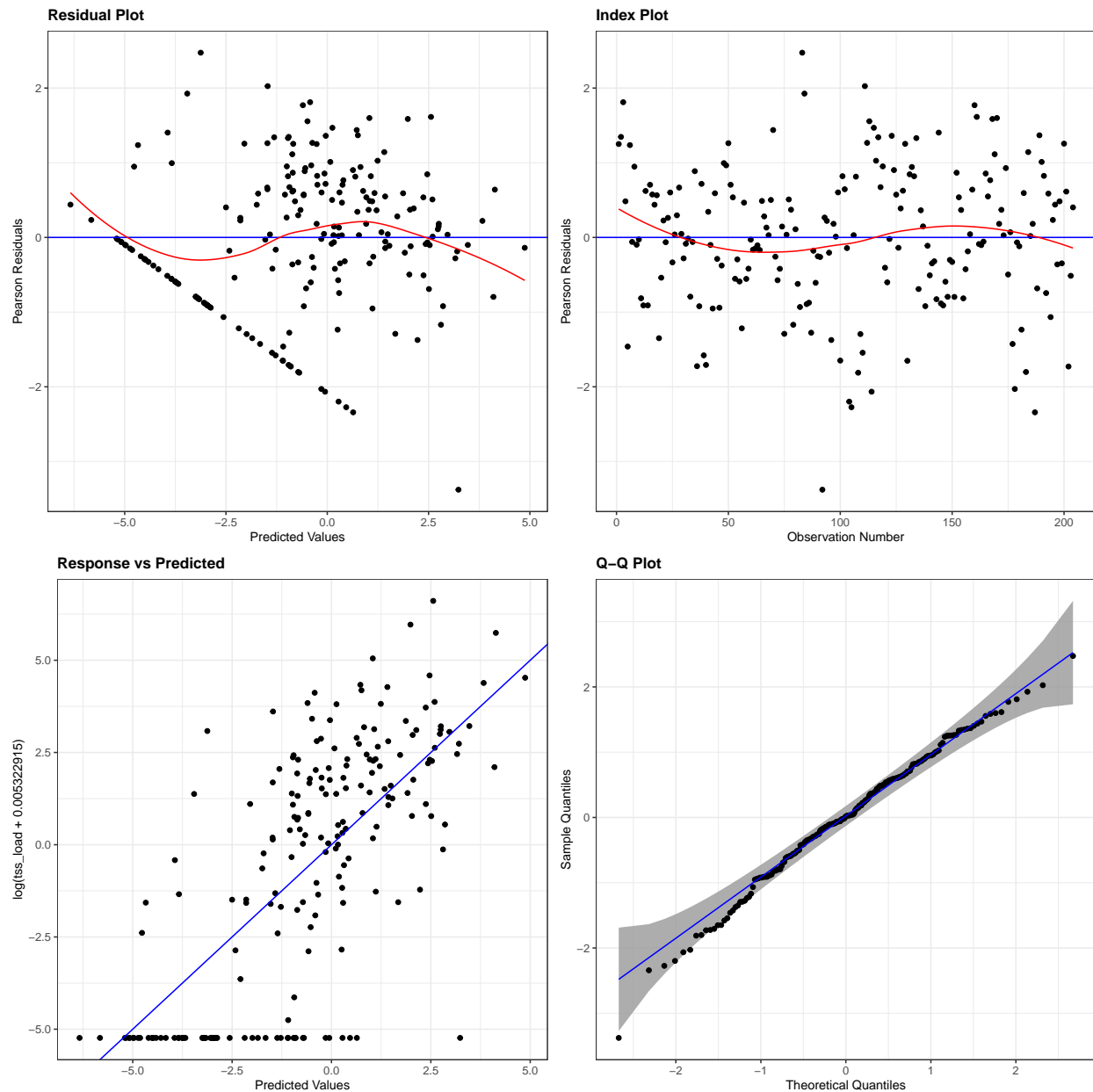
```
crop      = emmeans(m_flume, pairwise ~ Treatment|crop,
                    type = "response",
                    lmer.df = "asymptotic")

confint(crop)$contrasts
```

```
## crop = corn:
## contrast      ratio    SE df asymp.LCL asymp.UCL
## control / strips 14.43 14.68 Inf      1.962     106.1
##
## crop = soybean:
## contrast      ratio    SE df asymp.LCL asymp.UCL
## control / strips 3.14 3.59 Inf      0.335      29.5
##
## Results are averaged over the levels of: Year
## Degrees-of-freedom method: asymptotic
## Confidence level used: 0.95
## Intervals are back-transformed from the log scale
```

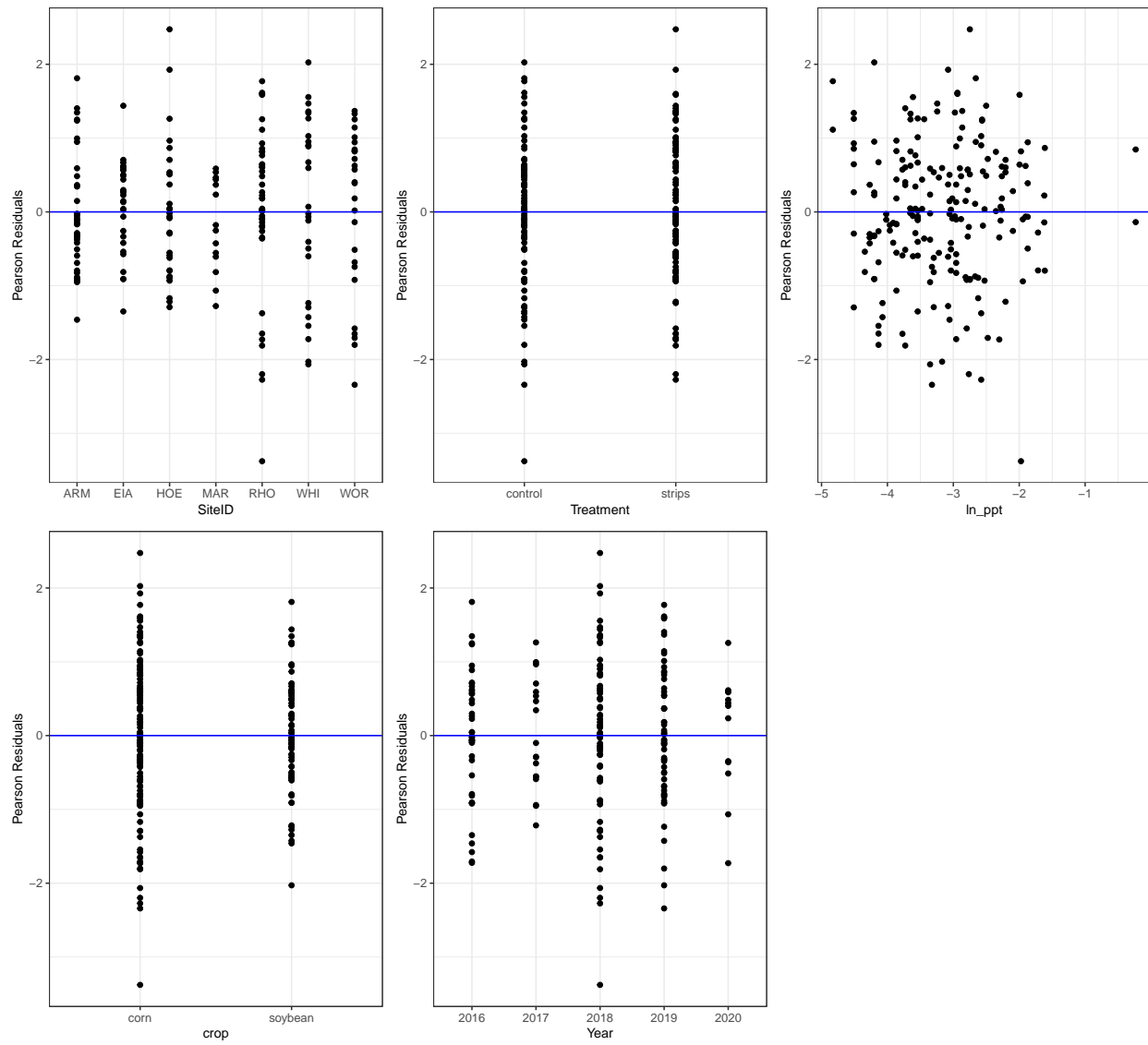
```
resid_panel(m_flume,
            plots = c("resid", "index", "yvp", "qq"),
            smoother = TRUE, qqbands = TRUE)
```

```
## 'geom_smooth()' using formula 'y ~ x'
## 'geom_smooth()' using formula 'y ~ x'
```



```
resid_xpanel(m_flume)
```

Plots of Residuals vs Predictor Variables



Possibly heavy-tailed residuals.

Non-constant variance amongst Site and Diversion.

Ratio comparison

List of PadIDs that had a diversion pair.