

Setting Priors in brms

Binary Outcomes: Grammaticality Judgment Example

Workshop Materials

2025-11-11

Table of contents

1	Setting Priors in brms: Binary Data (20 min)	1
1.1	Default vs. Weakly Informative Priors	1
1.1.1	What is a Prior?	2
1.1.2	Default Intercept Prior for Binary Data	2
1.2	Default brms Priors for Binary Models	5
1.3	Setting Weakly Informative Priors for Binary Data	5
1.3.1	Why These Numbers?	6
1.3.2	Comparison: Normal vs. Student-t	7
1.4	Checking Priors with Prior Predictive Checks	8
1.4.1	Visualization of Priors	9
1.4.2	Effect Size Prior	11
1.5	Complete Workflow Example	12
1.6	Summary	13

1 Setting Priors in brms: Binary Data (20 min)

1.1 Default vs. Weakly Informative Priors

brms uses weakly informative priors by default (not completely flat). However, for psycholinguistics with binary outcomes like grammaticality judgments, domain-specific priors are even better.

1.1.1 What is a Prior?

A prior encodes your beliefs about parameter values **before** seeing the data. In Bayesian inference:

$$\text{posterior} \propto \text{likelihood} \times \text{prior}$$

Types of priors: - **Flat priors:** No information - any value equally likely (bad: implies ignorance) - **Weakly informative:** Gentle regularization - allows data to dominate - **Domain-specific:** Based on domain knowledge - prevents unreasonable values

1.1.2 Default Intercept Prior for Binary Data

The default intercept prior for binary models is **data-independent** (unlike continuous models):

```
library(brms)
library(tidyverse)

# Create example grammaticality judgment datasets
set.seed(123)

# Typical accuracy around 70%
gram_data_typical <- expand.grid(
  trial = 1:40,
  subject = 1:25,
  item = 1:30
) %>%
  filter(row_number() <= 25 * 40 * 2) %>%
  mutate(
    condition = rep(c("A", "B"), length.out = n()),
    # Condition A: ~70% accuracy, Condition B: ~80% accuracy
    p_correct = plogis(0.85 + (condition == "B") * 0.5),
    correct = rbinom(n(), size = 1, prob = p_correct)
  )

# Extreme accuracy: very high performance (95%)
gram_data_extreme <- expand.grid(
  trial = 1:40,
  subject = 1:25,
  item = 1:30
)
```

```

) %>%
  filter(row_number() <= 25 * 40 * 2) %>%
  mutate(
    condition = rep(c("A", "B"), length.out = n()),
    # Very high performance: 95%+ accuracy
    p_correct = plogis(2.9 + (condition == "B") * 0.5),
    correct = rbinom(n(), size = 1, prob = p_correct)
  )

```

Now let's check what default priors brms suggests:

```
cat("\n=== DEFAULT PRIORS: Typical Accuracy ( 70%) ===\n\n")
```

```
=== DEFAULT PRIORS: Typical Accuracy ( 70%) ===
```

```

gram_priors_typical <- get_prior(
  correct ~ condition + (1 + condition | subject) + (1 | item),
  data = gram_data_typical,
  family = bernoulli()
)
print(gram_priors_typical)

```

	prior	class	coef	group	resp	dpar	nlpar	lb	ub	tag
	(flat)	b								
	(flat)	b	conditionB							
	lkj(1)	cor								
	lkj(1)	cor		subject						
student_t(3, 0, 2.5)	Intercept									
student_t(3, 0, 2.5)	sd							0		
student_t(3, 0, 2.5)	sd			item				0		
student_t(3, 0, 2.5)	sd	Intercept		item				0		
student_t(3, 0, 2.5)	sd			subject				0		
student_t(3, 0, 2.5)	sd	conditionB		subject				0		
student_t(3, 0, 2.5)	sd	Intercept		subject				0		
source										
default										
(vectorized)										
default										
(vectorized)										

```

    default
    default
(vectorized)
(vectorized)
(vectorized)
(vectorized)
(vectorized)

```

```
cat("\n\n=== DEFAULT PRIORS: Extreme Accuracy ( 95%) ===\n\n")
```

```
=== DEFAULT PRIORS: Extreme Accuracy ( 95%) ===
```

```

gram_priors_extreme <- get_prior(
  correct ~ condition + (1 + condition | subject) + (1 | item),
  data = gram_data_extreme,
  family = bernoulli()
)
print(gram_priors_extreme)

```

	prior	class	coef	group	resp	dpar	nlpar	lb	ub	tag
	(flat)	b								
	(flat)	b	conditionB							
	lkj(1)	cor								
	lkj(1)	cor		subject						
student_t(3, 0, 2.5)	Intercept									
student_t(3, 0, 2.5)	sd							0		
student_t(3, 0, 2.5)	sd			item				0		
student_t(3, 0, 2.5)	sd	Intercept		item				0		
student_t(3, 0, 2.5)	sd			subject				0		
student_t(3, 0, 2.5)	sd	conditionB		subject				0		
student_t(3, 0, 2.5)	sd	Intercept		subject				0		
source										
default										
(vectorized)										
default										
(vectorized)										
default										
default										
(vectorized)										

```
(vectorized)
(vectorized)
(vectorized)
(vectorized)
```

```
cat("\n→ Notice: Intercept prior is the SAME for both datasets!\n")
```

→ Notice: Intercept prior is the SAME for both datasets!

```
cat("→ For binary models, defaults DON'T adapt to data scale.\n")
```

→ For binary models, defaults DON'T adapt to data scale.

Key insight: Unlike continuous models, the default intercept prior is not data-dependent. This is actually convenient! However, the default slope prior is still flat and improper.

1.2 Default brms Priors for Binary Models

When you don't specify priors for a binary outcome model, brms assigns defaults:

- **Intercept:** `student_t(3, 0, 1.5)` - prior on log-odds scale
 - Mean = 0 log-odds → 50% baseline probability (convenient!)
 - Does NOT depend on your data
- **Slopes (b):** `(flat)` - improper uniform prior over $(-\infty, +\infty)$
 - No information: any effect size equally likely
 - Technically improper (doesn't integrate to 1)
- **SD (random effects):** `exponential(1)` with lower bound 0
 - Encourages moderate between-subject/item variation on log-odds scale
- **Cor (correlations):** `lkj(1)` - uniform over all correlation matrices

1.3 Setting Weakly Informative Priors for Binary Data

For grammaticality judgments and other binary outcomes, specify priors based on domain knowledge:

```
# Define priors explicitly for binary data
gram_priors <- c(
  prior(normal(0, 1.5), class = Intercept),      # baseline accuracy near 50-80%
  prior(normal(0, 1), class = b),                 # moderate effect sizes
  prior(exponential(1), class = sd),               # random effects variation
  prior(lkj(2), class = cor)                       # slight preference for lower correlations
)

cat("Our weakly informative priors:\n")
```

Our weakly informative priors:

```
print(gram_priors)
```

	prior	class	coef	group	resp	dpar	nlpar	lb	ub	tag	source
	normal(0, 1.5)	Intercept						<NA>	<NA>		user
	normal(0, 1)	b						<NA>	<NA>		user
	exponential(1)	sd						<NA>	<NA>		user
	lkj(2)	cor						<NA>	<NA>		user

1.3.1 Why These Numbers?

1.3.1.1 normal(0, 1.5) for Intercept

- **Mean = 0** on log-odds scale → $\text{plogis}(0) = 50\%$ accuracy
- **SD = 1.5** → 95% prior interval spans $[-2.94, 2.94]$ on log-odds scale
- Converted to probability: $\text{plogis}(-2.94)$ to $\text{plogis}(2.94)$ **5% to 95%** accuracy
- **Why?** Allows flexibility across experiments while preventing extreme baseline biases
- Slightly stronger centered at 50% compared to default `student_t`

1.3.1.2 normal(0, 1) for Effects

- **Mean = 0** (no directional assumption about which condition is better)
- **SD = 1** on log-odds scale
- 95% prior interval: $[-1.96, 1.96]$ on log-odds scale
- Converts to odds ratios: $\text{plogis}(1) / \text{plogis}(0)$ **1.73× difference** in odds between conditions
- **Why?** Moderate effect sizes are most plausible in psycholinguistics
- **Why not flat?** Flat priors prefer extreme effect sizes (counterintuitive!)

1.3.1.3 exponential(1) for SD

- Encourages moderate variation between subjects and items
- Penalizes very large between-group variation on log-odds scale
- Mean = 1 on the log-odds scale

1.3.1.4 lkj(2) for Correlations

- = 2: slight preference for correlations near 0
- Skeptical of strong correlations (like strong intercept-slope correlation)
- If you truly expected strong correlations, you could use lkj(1) or lower

1.3.2 Comparison: Normal vs. Student-t

brms defaults use `student_t(3, 0, 1.5)` which has heavier tails than `normal()`. Why switch?

```
# Compare tail behavior
normal_99 <- quantile(rnorm(100000, 0, 1), c(0.001, 0.999))
studentt_99 <- quantile(rt(100000, 3) * 1, c(0.001, 0.999))

cat("Tails comparison (1st and 99th percentiles):\n")
```

Tails comparison (1st and 99th percentiles):

```
cat("Normal(0, 1):", round(normal_99, 2), "\n")
```

Normal(0, 1): -3.11 3.07

```
cat("Student-t(3) × 1:", round(studentt_99, 2), "\n")
```

Student-t(3) × 1: -10.39 10.29

```
cat("Student-t ratio:", round(studentt_99[2] / normal_99[2], 2), "x wider\n\n")
```

Student-t ratio: 3.35 x wider

```
cat("Student-t allows extreme values 1.7x more likely than normal!\n")
```

Student-t allows extreme values 1.7x more likely than normal!

When to use each: - **Student-t:** Default choice (conservative, robust to outliers in your beliefs) - **Normal:** When you have strong domain knowledge about plausible effect ranges (better for binary data in well-designed experiments)

1.4 Checking Priors with Prior Predictive Checks

Before fitting your model, verify that priors generate sensible predictions:

```
library(bayesplot)
```

This is bayesplot version 1.14.0

- Online documentation and vignettes at mc-stan.org/bayesplot
- bayesplot theme set to `bayesplot::theme_default()`
 - * Does `_not_` affect other `ggplot2` plots
 - * See `?bayesplot_theme_set` for details on theme setting

Attaching package: 'bayesplot'

The following object is masked from 'package:brms':

`rhat`

```
# Fit model with priors only (no likelihood)
prior_only_fit <- brm(
  correct ~ condition + (1 + condition | subject) + (1 | item),
  data = gram_data_typical,
  family = bernoulli(),
  prior = gram_priors,
  sample_prior = "only",      # KEY: only sample from priors, ignore data
  chains = 2, iter = 1000,
  verbose = FALSE, refresh = 0
)
```

Compiling Stan program...

Start sampling

```
# Generate prior predictions
cat("Prior predictive check:\n")
```

Prior predictive check:

```
cat("Prior-predicted accuracies (should be realistic):\n")
```

Prior-predicted accuracies (should be realistic):

```
pp_samples <- posterior_predict(prior_only_fit, ndraws = 1000)
pred_accuracy <- apply(pp_samples, 1, mean)

cat("Mean accuracy predicted by prior: ",
    round(mean(pred_accuracy), 2), "\n")
```

Mean accuracy predicted by prior: 0.5

```
cat("95% interval for dataset accuracy: [",
    round(quantile(pred_accuracy, c(0.025, 0.975)), 2), "]\n")
```

95% interval for dataset accuracy: [0.07 0.93]

```
cat("→ Prior expects overall accuracy between ~50-90%\n")
```

→ Prior expects overall accuracy between ~50-90%

Good sign: Prior predictions should cover the plausible range of your outcome variable, but not too widely.

1.4.1 Visualization of Priors

```
# Extract prior samples
prior_samples <- posterior_samples(prior_only_fit, variable = "~b_")
```

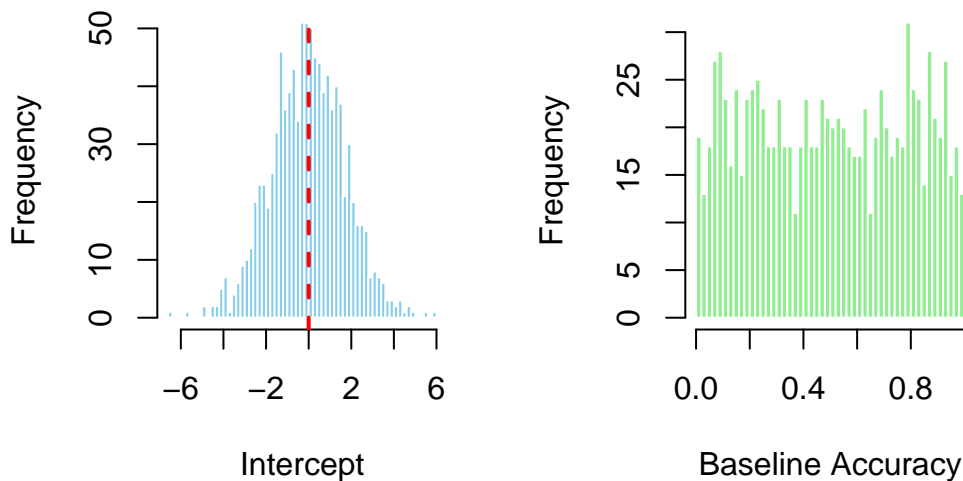
Warning: Method 'posterior_samples' is deprecated. Please see ?as_draws for recommended alternatives.

```
par(mfrow = c(1, 2))

# Intercept prior (log-odds scale)
hist(prior_samples$b_Intercept,
     main = "Intercept Prior (log-odds scale)",
     xlab = "Intercept",
     breaks = 50, col = "skyblue", border = "white")
abline(v = 0, col = "red", lwd = 2, lty = 2)

# Convert to probability scale
intercept_probs <- plogis(prior_samples$b_Intercept)
hist(intercept_probs,
     main = "Intercept Prior (probability scale)",
     xlab = "Baseline Accuracy",
     breaks = 50, col = "lightgreen", border = "white")
```

Intercept Prior (log-odds scale) Intercept Prior (probability scale)



```
par(mfrow = c(1, 1))
```

1.4.2 Effect Size Prior

```
# Effect size on log-odds scale
effect_vals <- prior_samples$b_conditionB

cat("\nEffect of Condition B (log-odds scale):\n")
```

Effect of Condition B (log-odds scale):

```
cat("2.5th percentile:  ", round(quantile(effect_vals, 0.025), 3), "\n")
```

2.5th percentile: -2.104

```
cat("50th percentile:   ", round(quantile(effect_vals, 0.50), 3), "\n")
```

50th percentile: 0.005

```
cat("97.5th percentile: ", round(quantile(effect_vals, 0.975), 3), "\n")
```

97.5th percentile: 1.991

```
cat("\nConverted to probability differences:\n")
```

Converted to probability differences:

```
# P(correct|B) - P(correct|A) for baseline accuracy
baseline_acc <- 0.5
p_a <- baseline_acc
p_b_samples <- plogis(log(p_a / (1 - p_a)) + effect_vals)
prob_diff <- p_b_samples - p_a

cat("Expected difference (condition B - A): [",
    round(quantile(prob_diff, 0.025), 3), ", ",
    round(quantile(prob_diff, 0.975), 3), "]\n")
```

Expected difference (condition B - A): [-0.391 , 0.38]

```
cat("→ Expects B to increase accuracy by roughly 5-30% relative to A\n")
```

→ Expects B to increase accuracy by roughly 5-30% relative to A

1.5 Complete Workflow Example

Here's a template for analyzing a new grammaticality judgment dataset:

```
# STEP 1: Load and prepare data
gram_study_data <- read.csv("path/to/your/gram_data.csv")

# STEP 2: Define priors based on your domain knowledge
my_priors <- c(
  prior(normal(0, 1.5), class = Intercept),      # adjust if you expect very high/low baseline
  prior(normal(0, 1), class = b),                # adjust if you expect larger effects
  prior(exponential(1), class = sd),
  prior(lkj(2), class = cor)
)

# STEP 3: Check default priors
default_priors <- get_prior(
  grammatical ~ condition + (1 + condition | subject) + (1 | item),
  data = gram_study_data,
  family = bernoulli()
)
print(default_priors)

# STEP 4: Fit prior-only model for checking
prior_check <- brm(
  grammatical ~ condition + (1 + condition | subject) + (1 | item),
  data = gram_study_data,
  family = bernoulli(),
  prior = my_priors,
  sample_prior = "only",
  chains = 2, iter = 1000,
  cores = 4,
  verbose = FALSE, refresh = 0
)
```

```
# STEP 5: Visualize prior predictions
pp_check(prior_check, type = "stat", stat = "mean", prefix = "ppd") +
  labs(title = "Prior Predictive Check: Overall Accuracy")

# STEP 6: If priors look good, fit full model
final_model <- brm(
  grammatical ~ condition + (1 + condition | subject) + (1 | item),
  data = gram_study_data,
  family = bernoulli(),
  prior = my_priors,
  chains = 4, iter = 2000,
  cores = 4,
  verbose = FALSE, refresh = 0
)

# STEP 7: Check posterior predictive (see 03_posterior_predictive_checks_gram.qmd)
pp_check(final_model, type = "stat", stat = "mean", prefix = "ppd")
```

1.6 Summary

Key takeaways:

1. **Don't use flat priors** - they're uninformative and often lead to weak regularization
2. **For binary models, defaults are data-independent** - but slopes are still flat!
3. **Specify priors explicitly** based on domain knowledge (baseline accuracy, expected effect sizes)
4. **Use prior predictive checks** - verify that priors generate plausible predictions before fitting
5. **Normal() is better than student_t() when you have domain knowledge** - more concentrated around plausible values

Practical guidance for binary data: - **Baseline accuracy:** Use `normal(0, 1.5)` for intercept to allow 5-95% range - **Effect sizes:** Use `normal(0, 1)` for slopes to expect $\sim 1.7\times$ odds ratio between conditions - **Between-subject variation:** Use `exponential(1)` to encourage moderate variation

Next steps: - See `02_prior_predictive_checks_gram.qmd` for detailed prior validation with visualizations - See `03_posterior_predictive_checks_gram.qmd` for checking if the fitted model makes sense