hw8.R

joebrew

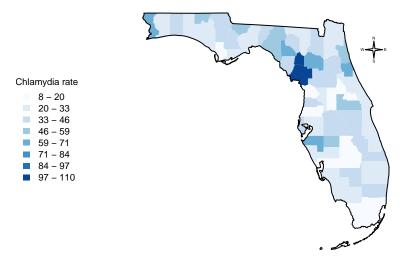
Mon Oct 27 09:23:07 2014

```
# set local working directory
setwd("/home/joebrew/Documents/uf/phc6194/hw8/")
# load some packages for mapping and colrs
library(rgdal)
## Loading required package: sp
## rgdal: version: 0.9-1, (SVN revision 518)
## Geospatial Data Abstraction Library extensions to R successfully loaded
## Loaded GDAL runtime: GDAL 1.10.1, released 2013/08/26
## Path to GDAL shared files: /usr/share/gdal/1.10
## Loaded PROJ.4 runtime: Rel. 4.8.0, 6 March 2012, [PJ_VERSION: 480]
## Path to PROJ.4 shared files: (autodetected)
library(RColorBrewer)
library(classInt)
library(sp)
############
# 1. MERGE THE DATA
############
# read in the florida shapefile (exported from arcgis)
fl <- readOGR(".", "hw8")</pre>
## OGR data source with driver: ESRI Shapefile
## Source: ".", layer: "hw8"
## with 67 features and 4 fields
## Feature type: wkbPolygon with 2 dimensions
# read in the std rate data
std <- read.csv("std.csv", stringsAsFactors = FALSE)</pre>
# compare to see if names are perfect matches
fl$NAME <- toupper(fl$NAME)</pre>
table(fl@data$NAME == std$NAME) # they're not all correct
##
## FALSE TRUE
##
      35
            32
# get the closest match for each county
fl$name <- NA # create empty vector where we'll put our names
```

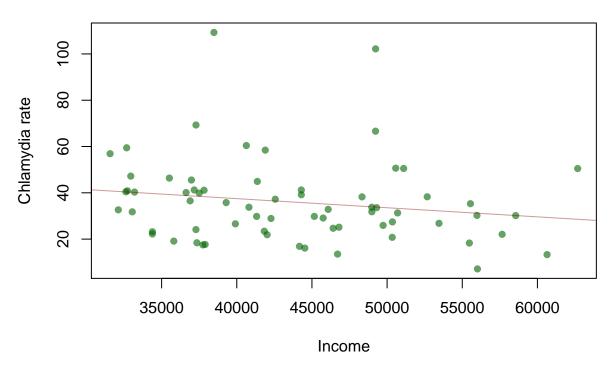
```
std$name <- std$NAME # we'll use lower cases for the matches
for (i in 1:nrow(fl)){
  # see how close (in character changes) each name in fl is to those in STD
  m <- adist(fl$NAME[i],</pre>
             std$NAME)
  # get the one with the least differences
  ind <- which.min(m)</pre>
  # get the name from std
  best <- std$NAME[ind]</pre>
  # assign to fl
  fl$name[i] <- best
}
# Merge the two datasets together
fl@data <- merge(x = fl@data,
                  y = std,
                  by = "name",
                  all.x = TRUE,
                  all.y = FALSE)
############
# 2. CHOROPLETH OF CHLAMYDIA
###########
# Create boundary shapefile of just florida
library(maptools)
## Checking rgeos availability: TRUE
boundary <- unionSpatialPolygons(fl, rep(1, length(fl@polygons)))</pre>
## Loading required package: rgeos
## rgeos version: 0.3-8, (SVN revision 460)
## GEOS runtime version: 3.4.2-CAPI-1.8.2 r3921
## Polygon checking: TRUE
# Code for compass rose
\#(from\ http://r-sig-geo.2731867.n2.nabble.com/How-to-diplasy-a-compass-rose-on-a-map-td4509034.html)
compassRose<-function(x,y,rot=0,cex=1) {</pre>
  oldcex<-par(cex=cex)</pre>
  mheight<-strheight("M")</pre>
  xylim<-par("usr")</pre>
  plotdim<-par("pin")</pre>
  xmult<-(xylim[2]-xylim[1])/(xylim[4]-xylim[3])*plotdim[2]/plotdim[1]</pre>
  point.angles<-seq(0,7*pi/4,by=pi/4)+pi*rot/180
  crspans<-rep(c(mheight*3,mheight/2),4)</pre>
  xpoints<-cos(point.angles)*crspans*xmult+x</pre>
  ypoints<-sin(point.angles)*crspans+y</pre>
  polygon(xpoints,ypoints)
  txtxpoints < -cos(point.angles[c(1,3,5,7)])*1.33*crspans[1]*xmult+x
  txtypoints < -sin(point.angles[c(1,3,5,7)])*1.33*crspans[1]+y
  text(txtxpoints,txtypoints,c("E","N","W","S"))
```

```
par(oldcex)
# Code for map (original)
TractFun <- function(var,</pre>
                      color = "Blues",
                      style = "equal", # or equal/quantile
                      nclr = 8,
                      title = NA,
                      dataPrecision = 0){
  plotclr <- brewer.pal(nclr, color)</pre>
  class <- classIntervals(var, nclr, style = style, dataPrecision=dataPrecision) #use "equal" instead</pre>
  #class <- classIntervals(0:100, nclr, style="equal")</pre>
  colcode <- findColours(class, plotclr)</pre>
  legcode <- paste0(gsub(",", " - ", gsub("[[]|[]]|[)]", "", names(attr(colcode, "table")))))</pre>
  plot(f1, border=NA, col=colcode)
  plot(boundary, add = TRUE)
  legend("left", # position
         legend = legcode, #names(attr(colcode, "table")),
         fill = attr(colcode, "palette"),
         cex = 0.6.
         border=NA,
         bty = "n",
         title = title)
  compassRose(x = -80.5, y = 30, rot=0, cex=0.25)
}
TractFun(fl$CHALMYDIA_, title = "Chlamydia rate")
title(main = "Chlamydia rate")
```

Chlamydia rate



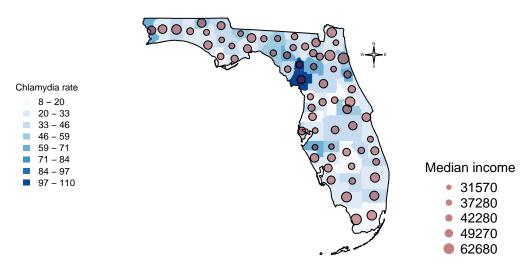
Income and Chlamydia in Florida counties



```
# First, write function to scale income to desired max
JoeScale <- function(my_vector, max_cex = 1.5){</pre>
    my_scale <- max(my_vector) / max_cex</pre>
    my_vector / my_scale
}
TractFun(f1$CHALMYDIA_,
         title = "Chlamydia rate")
points(coordinates(fl),
       cex = JoeScale(f1$INCOME),
       pch = 16,
       col = adjustcolor("darkred", alpha.f = 0.4))
points(coordinates(f1),
       cex = JoeScale(f1$INCOME))
legend("bottomright",
       pt.cex = JoeScale(quantile(f1$INCOME)),
       legend = round(quantile(f1$INCOME), digits = -1),
       pch = 16,
       col = adjustcolor("darkred", alpha.f = 0.5),
```

```
title = "Median income",
bty= "n",
cex = 0.8)
title(main = "Chlamydia and income")
```

Chlamydia and income



```
###########
# 4. MAP CHLAMYDIA, HIV, GONORRHEA AND SYPHILLIS
###########
par(mfrow = c(2,2))
par(mar= c(1,1,1,1))
TractFun(f1$CHALMYDIA_,
         title = "Chlamydia rate",
         color = "Greens")
title(main = "Chlamydia")
TractFun(f1$HIV_RATES_,
         title = "HIV rate",
         color = "Blues",
         dataPrecision = 1)
title(main = "HIV")
TractFun(f1$GONORRHEA_,
        title = "Gonorrhea rate",
         color = "Reds")
title(main = "Gonorrhea")
TractFun(f1$SYPHILIS_R,
         title = "Syphilis rate",
         color = "Purples",
         dataPrecision = 1)
title(main = "Syphilis")
```

