Developing an Antimicrobial Strategy for Sepsis in Malawi

_

Thesis submitted in accordance with the requirements of the Liverpool School of Tropical Medicine for the degree of Doctor in Philosophy by Joseph Michael Lewis

August 2019

Contents

Pı	refac	e	9
1	Intr	\mathbf{r} oduction	11
	1.1	Chapter Overview	13
	1.2	Sepsis in sub-Saharan Africa	13
	1.3	ESBL-E in sub-Saharan Africa	13
	1.4	Conclusions	13
	1.5	Thesis overview	13
	1.6	Appendix	13
	1.7	References	13
2	Met	chods	15
	2.1	Chapter Overview	17
	2.2	Study site	17
	2.3	Clinical Study	17
	2.4	Diagnostic Laboratory Procedures	17
	2.5	Molecular methods	17
	2.6	Bioinformatics	17
	2.7	Statistical Analysis	17
	2.8	Study Team	17
	2.9	Data Collection and Storage	17
	2.10	Ethical Approval, Consent and Participant Remuneration	17
3	My	cobacterium tuberculosis BSI: an IPD meta analysis	19
4	Sep	sis in Blantyre, Malawi	21
	4.1	Chapter overview	21
	4.2	Methods	21
	43	Results	21

4	CONTENTS
±	CONTENTS

6	Gut	mucosal carriage of ESBL-E in Blantyre, Malawi	4
7	Wh	ole genome sequencing of ESBL E. coli carriage isolates	4
	7.1	Chapter overview	4
	7.2	Methods	4
	7.3	Results	4
	7.4	Discussion	4
	7.5	Appendix	4

List of Tables

4.1	Participant Characteristics	22
4.2	Prehospital heathcare seeking and antimicrobial exposure	26
4.3	Admission physiology, haematology and biochemistry	28
4.4	Final diagnosis of all participants	30
4.5	Positive diagnostic tests for all participants, stratified by HIV status	30
4.6	Door-to-antimicrobial times	33
4.7	Day 28, 90 and 180 mortality stratified by HIV status	35
4.8	Bivariate associations with death by 28 days	38
4.9	Bivariate associations with diagnosis	39

6 LIST OF TABLES

List of Figures

4.1	Study recruitment and follow up	25
4.2	Symptoms of recruited participants	27
4.3	Pathogenic isolates recovered from aerobic blood culture	31
4.4	Venn diagram of positive diagnostic tests	31
4.5	UpSet plot of positive diagnostic tests	32
4.6	Antimicrobial and fluid administration as a function of time	34
4.7	Kaplan-Meier survival curves	35
4.8	Health-related QoL following admission with sepsis	36
4.9	Hypothesised causal structre of mortality in sepsis	37

8 LIST OF FIGURES

Preface

Placeholder

10 LIST OF FIGURES

Chapter 1

Introduction

Placeholder

1.1 Chapter Overview

- 1.2 Sepsis in sub-Saharan Africa
- 1.2.1 Search strategy
- 1.2.2 Defining sepsis
- 1.2.3 Applicability of sepsis-3 definitions in sub-Saharan Africa
- 1.2.4 Sepsis epidemiology in sub-Sahara Africa
- 1.2.4.1 Incidence
- 1.2.4.2 Risk factors: the sepsis population in sub-Saharan Africa
- 1.2.4.3 **Outcomes**
- 1.2.5 Sepsis aetiology in sub-Saharan Africa
- 1.2.5.1 Bacterial zoonoses, Rickettsioses and arboviruses
- 1.2.5.2 HIV opportunistic infections: PCP, histoplasmosis and cryptococcal disease
- 1.2.6 Sepsis management
- 1.2.6.1 Early goal directed therapy
- 1.2.6.2 Evidence to guide antimicrobial therapy in sSA
- 1.2.6.3 Evidence to guide intravenous fluid therapy in sub-Saharan Africa
- 1.3 ESBL-E in sub-Saharan Africa
- 1.3.1 Search strategy
- 1.3.2 Introduction: definition and classification of ESBL-E
- 1.3.3 Global molecular epidemiology of ESBL-E: an overview
- 1.3.3.1 1980s-1990s: First identification of ESBL in nosocomial pathogens
- 1.3.3.2 1990s-2010s: Emergence and globalisation of CTX-M

Chapter 2

Methods

Placeholder

2.1 Chapter Overview

- 2.2 Study site
- 2.2.1 Malawi
- 2.2.2 Queen Elizabeth Central Hospital
- 2.2.3 Participating Laboratories
- 2.2.3.1 Malawi-Liverpool-Wellcome Clinical Research Programme
- 2.2.3.2 Malawi College of Medicine Tuberculosis Laboratory
- 2.2.3.3 Wellcome Trust Sanger Institute
- 2.3 Clinical Study
- 2.3.1 Entry Criteria
- 2.3.2 Study Visits and Patient Sampling
- 2.3.2.1 Enrollment assessment and first six hours
- 2.3.2.2 Subsequent visits
- 2.3.2.3 Blood, urine, and stool, sputum and CSF collection
- 2.3.2.4 Imaging: chest x-ray and ultrasound scanning
- 2.3.3 Outcomes and sample size calculations
- 2.4 Diagnostic Laboratory Procedures
- 2.4.1 Point of care diagnostics
- 2.4.2 Laboratory diagnostics
- 2.4.2.1 Haematology and biochemistry
- 2.4.2.2 Aerobic blood and CSF culture
- 2.4.2.3 Mycobacterial blood culture

Chapter 3

Mycobacterium tuberculosis BSI: an IPD meta analysis

Chapter 4

Sepsis in Blantyre, Malawi

4.1 Chapter overview

4.2 Methods

blah blah

4.3 Results

4.3.1 Study population

Figure 4.1 shows flow through the study. 225 participants were recruited in 20 months between 19th February 2017 and 2nd October 2018. Participants were recuited, in general, soon after arrival in hospital, a median (IQR) of 1.5 (0.8-2.6) hours after fist attendance. In total, 4 participants (2%) were lost to follow up over the 180-day study period; 5 participants (2%) withdrew; and 7 participants (3%) transferred out of the study area before 180 days. Four of the five participants who withdrew gave a reason for their wish to withdraw, all that they no longer wished the inconvenience of being involved in the study. 15/225 (7%) participants had their final study visit before 180 days, and so were not included in the 180-day outcome analysis.

4.3.2 Symptoms and health-seeking behaviour

Table 4.1 shows the baseline characteristics of the recruited participants. They were young (median [IQR] age 36 [28-44]) and predominantly HIV-infected. Of those who were HIVinfected, the majority (117/143 [82%]) were on ART, almost exclusively the Malawian first-line regimen of efavirenz, lamivudine and tenofovir, and 88/117 (75%) had been taking ART for more than three months. Figure 4.2 shows the presenting symptoms of the participants. Almost all (221/225 [98%] of participants) experienced subjective fever. Participants had been unwell for some time, a median (IQR) of 7 (3-14) days; 32/225 (14%) of participants had been unwell for more than 4 weeks. 18/225 (8%) of participants had been admitted to hospital within the last 4 weeks. Over half (123/225 [55%]) of participants had sought care for their current ilness (Table 4.2), most commonly (101/123 [82%] of participants) at a government health centre, a median (IQR) of 2 (1-6) days previously. 60/225 (27%) of all participants had recieved an antimicrobial for their current illness: 7/60 (12%) of all prehospital antimicrobials were antimalarials, the remainder anitbacterial, most commonly co-trimoxazole or ciprofloxacin. Prehopsital intravenous or intramuscular antimirobials were administered in 16/60 (27%) participants recieving antimicrobials: ceftriaxone (n=6), benzylpenicillin (n=4), gentamicin (n=3) and artesunate (n=3).

Table 4.1: Participant Characteristics

Variable	Value
Demographics	
Age (years)	36 (28-44)
Male sex	114/225 (51%)
HIV/TB status	
HIV Reactive	143/225 (64%)
HIV Non Reactive	70/225 (31%)
HIV Unknown	12/225 (5%)
Ever treated for TB	$37/225 \ (16\%)$
Of those, current TB treatment	10/37~(27%)
ART status*	
Current ART	117/143 (82%)
Months on ART	29 (4-73)
ART regimen: EFV/3TC/TDF	110/117 (94%)
ART regimen: other	7/117 (6%)
Current CPT^{\dagger}	98/141 (70%)
Tobacco/alcohol use	
Never tobacco	196/225 (87%)
Ex tobacco	17/225 (8%)
Current tobacco	12/225 (5%)

Table 4.1: Participant Characteristics (continued)

Variable	Value
Current alcohol	51/225 (23%)
Education Primary incomplete or complete Secondary school complete Some secondary education College or higher No formal schooling	97/225 (43%) 48/225 (21%) 47/225 (21%) 17/225 (8%) 16/225 (7%)
Employment	10/229 (170)
Unemployed Currently employed Self-employed Student Retired	82/225 (36%) 65/225 (29%) 56/225 (25%) 21/225 (9%) 1/225 (0%)
Toilet facilities	
Pit latrine with slab +/- foot rest Hanging toilet/latrine Pit latrine with slab and cover +/- foot rest Flush Toliet (any type) No toilet Composting toilet	104/225 (46%) 59/225 (26%) 45/225 (20%) 14/225 (6%) 2/225 (1%) 1/225 (0%)
Main water source Piped outside dwelling Tube well/borehole Public tap/standpipe Piped into dwelling Unprotected well/spring Surface water (including rainwater collection) Tube well with powered pump	69/225 (31%) 64/225 (28%) 51/225 (23%) 30/225 (13%) 5/225 (2%) 4/225 (2%) 2/225 (1%)
Electricty	
Electricity available in house	$119/225 \ (53\%)$
Main cooking fuel Charcoal Wood Electricity	161/225 (72%) 61/225 (27%) 3/225 (1%)
Animals at home? Any animal Poultry Dogs Goats Dogs Other	71/225 (32%) 46/71 (65%) 18/71 (25%) 12/71 (17%) 18/71 (25%) 11/71 (15%)

Table 4.1: Participant Characteristics (continued)

Variable	Value

Note:

ART = Antiretroviral therapy, CPT = Co-trimoxazole preventative therapy, EFV: Efavirenz, 3TC: Lamivudine, TDF: Tenofovir. Numeric values are median (IQR)) unless otherwise stated.

Table 4.2: Prehospital heathcare seeking and antimicrobial exposure

Variable	Value
Pre-hospital healthcare seeking	
Sought care prior to attendance at hospital	123/225 (55%)
At health centre	101/123 (82%)
At hospital	$16/123 \ (13\%)$
At private doctor	$8/123 \ (7\%)$
Somewhere else	1/123~(1%)
Days prior to today that participant sought care	2'(1-6)
Prehospital antimicrobial exposure	
Recieved any antimicrobial prior to attendance at hospital	60/225~(27%)
Co-trimoxazole	12/60(20%)
Ciprofloxacin	10/60~(17%)
Amoxicillin	$9/60 \ (15\%)$
Ceftriaxone	6/60 (10%)
Metronidazole	5/60 (8%)
Benzylpenicillin	4/60~(7%)
Artesunate	3/60~(5%)
Gentamicin	3/60~(5%)
Erythromycin	2/60 (3%)
$ m L \mathring{A}$	2/60~(3%)
SP	2/60~(3%)
Azithromycin	1/60(2%)
Flucloxacillin	1/60~(2%)
Days prior to today that antimicrobials started	2 (1-5)
Method of transport to hospital	
Minibus	78/225 (35%)
Taxi	65/225~(29%)
Private car/truck	42/225~(19%)
Ambulance	37/225~(16%)
Other	$2/225 \ (1\%)$
Walk	1/225~(0%)
Cost (MWK) of transport to hospital	$1000 \ (275-3000)$

Note:

 $\label{eq:LA} LA = Lume fantrine-artemether, SP = Sulfamethoxazole-pyrimethamine, MWK = Malawian Kwacha. Numeric values are median (IQR)) unless otherwise stated.$

^{*} ART status includes HIV reactive only as denominator

 $^{^\}dagger$ Missing CPT data for two participants.

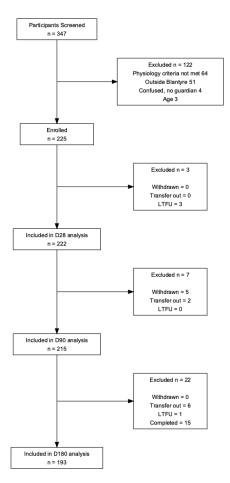


Figure 4.1: Study recruitment and follow up.

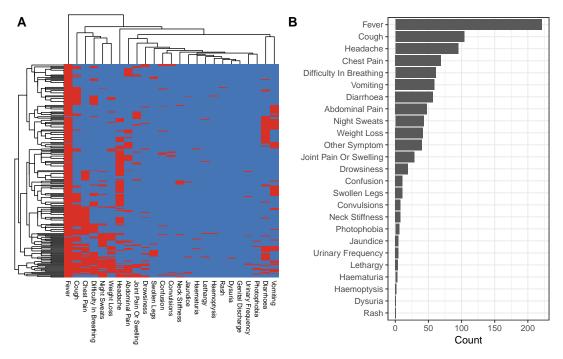


Figure 4.2: Symptoms of recruited participants. A: Row and column clustered heatmap of participant symptoms. Each row represents a patient. Red = presence, blue = absence. B: Frequency of occurence of symptoms

4.3.3 Admission physiology and laboratory investigations

Admission vital signs and laboratory investigations are shown in Table 4.3. Despite high ART coverage (117/143 [82%]) amongst HIV-infected participants for a median of 29 months, the median (IQR) CD4 count was low at 98 (31-236) cells μ L⁻¹. 108/141 (70%) of participants had a CD4 count below 200 cells μ L⁻¹. CD4 count was similar in participants who had started ART more than 6 months ago as compared to less than three months ago (median [IQR] 99 [27-260] vs 93 [39-137] cells μ L⁻¹ respectively) and 42/83 (51%) of participants who had been taking ART for more than 6 months had a CD4 count of less than 100 cells μ L⁻¹, and would fulfil a WHO definition of immunological failure.

Table 4.3: Admission physiology, haematology and biochemistry

Variable	Value
Admission physiology	
Temperature (°C)	38.5 (37.9-39.0)
Heart rate (min ⁻¹))	121 (102-132)
Systolic blood pressure (mmHg)	99 (85-119)
Diatsolic blood pressure (mmHg)	66 (56-76)
Respiratory rate (min ⁻¹)	34 (32-38)
Oxygen saturation (%)	96 (94-98)
GCS	004/005 (0107)
15 11-14	204/225 (91%)
< 11 < 11	$16/225 \ (7\%) \ 5/225 \ (2\%)$
Admission CD4 count	0/220 (270)
CD4 count* (μL^{-1})	00 (21 226)
• /	98 (31-236)
Admission haematology	10.0 (0.0.10.0)
Haemoglobin ($x10^9 \text{ g dL}^{-1}$)	10.8 (8.2-13.2)
White cell count $(x10^9 L^{-1})$	6.5 (4.4-11.4)
Neutrophil count (x10 ⁹ L ⁻¹)	4.0 (2.1-7.5)
Platelet count count ($x10^9 L^{-1}$)	218 (146-297)
Admission biochemistry	
Sodium (mmol L ⁻¹)	$134 \ (130-137)$
Potassium (mmol L ⁻¹)	4.0 (3.6-4.4)
Bicarbonate (mmol L ⁻¹)	19 (17-22)
Chloride (mmol L^{-1})	101 (97-104)
Urea (mmol L ⁻¹)	4.8 (3.5-8.0)
Creatinine (mmol L ⁻¹)	76 (59-103)
Lactate (mmol L ⁻¹)	3.4 (2.3-5.2)

Note:

 $\mathrm{GCS}=\mathrm{Glasgow}$ coma scale. Numeric values are median (IQR)) unless otherwise stated.

^{*} CD4 count includes only HIV-infected participants; 2 values were missing.

4.3.4 Aetiology

In total, 51% (114/225) of the 225 participants had at least one infectious agent identified (Table 4.4), most commonly tuberculosis (76/225 [34%]) followed by bloodstream infection (24/225 [11%]) and malaria (21/225 [9%]). Table 4.5 shows the availability of test and proportion of positive tests across the cohort, stratified by HIV status. 2/225 patients (1%) had a missing aerobic blood culture; the remaining 223 patients had a total of 259 blood cultures performed. 15/259 (6%) blood cultures grew at least one contaminant, but 26 blood cultures from 24 patients were positive for a total of 28 pathogenic bacteria (Figure 4.3): Salmonella Typhi was the most commonly isolated pathogenic bacterium, and seemed to show an association with HIV-negative participants: all (8/8) of the participants from whom S. Typhi was isolated and whose HIV status was known were HIV noninfected. Of the 18 Gram negative bacteria isolated, 3/18 (17%) were cefpodoxime resistant on AST via disc diffusion testing, and likely ESBL producers: one K. pneumoniae and one E. coli (both from the same blood culture and same patient) and one Acinetobacter baumannii. Both Staphyloccus aureus isolates were oxacillin sensitive. The one Streptococcus pneumoniae cultured was penicillin intermediate on AST.

Lumbar puncture and CSF culture was carried out in 44 participants: 5/44 (11%) of samples grew a contaminent and no pathogenic bacteria were recovered from any sample. 4/44 (9%) had a detectable cryptococcal antigen (CRAG) in CSF. Malaria testing was missing for 6/225 (3%) of participants, but of the remainder, a positive malaria test was more likely in the HIV-uninfected (12/69 [17%] vs 6/138 [4%], p = 0.01 on pairwise Fisher's exact test). Positive aerobic blood culture showed no statistically significant association with HIV, nor did positive CSF testing, though in the latter case numbers were small and all positive tests (all positive CRAG) were in fact in the HIV-infected (Table 4.5).

Testing for TB, with the exception of sputum Xpert testing, was restricted to HIV-infected participants. Sputum Xpert was carried out in 44/225 (20%) of participants, and was more commonly carried out in the HIV-infected: 35/143 [24%] of HIV-infected participants had sputum testing performed vs 8/70 (11%) of HIV uninifected (p = 0.07 by Fisher's exact test). 53 sputum samples were sent in total from the 44 patients, and 8/44 (18%) diagnoses of TB made, all except one in HIV-infected participants. One sample identified a rifampicin resistance gene; the remainder of infections were rifampcin-sensitive.

155 participants were eligible for urinary lipoarabinomannan (uLAM) and mycobacterial blood culture testing, being either HIV-infected (n=143) or of unknown HIV status (n=12). Urine was available for 145/155 (94%) of those eligible, and 74/145 (51%) of samples were positive for uLAM. 150/155 (97%) of eligible participants had blood samples collected and cultured

Diagnosis	Proportion of participants
Tuberculosis	76/225 (34%)
Bloodstream infection	$24/225\ (11\%)$
Malaria	21/225 (9%)
Meningitis	4/225 ($2%$)
No diagnosis	111/225 (49%)

Table 4.4: Final diagnosis of all participants

Table 4.5: Positive diagnostic tests for all participants, stratified by HIV status.

		HIV status			
Test	Positive	Negative	Unknown	All	p
Number of participants	143	70	12	225	_
TB diagnostics Urinary LAM Sputum Xpert TB blood culture	70/136 (51%) 7/35 (20%) 7/128 (5%)	- 1/8 (12%) -	4/9 (44%) 0/1 (0%) 1/10 (10%)	$74/145 (51\%) \ 8/44 (18\%) \ 8/138 (6\%)$	- 0.835 -
Other diagnostics Aerobic blood culture CSF culture or CRAG Malaria RDT	13/141 (9%) 4/31 (13%) 6/138 (4%)	9/70 (13%) 0/12 (0%) 12/69 (17%)	2/12 (17%) 0/1 (0%) 3/12 (25%)	$24/223 \ (11\%) \ 4/44 \ (9\%) \ 21/219 \ (10\%)$	0.647 0.445 0.007

Note:

LAM = Lipoarabinomannan, CSF = Cerebrospinal fluid, CRAG = Cryptococcal antigen, RDT = Rapid diagnostic test. p-values are chi-squared test across the three HIV status strata, and hence may be different from the pairwise exact Fisher's tests presented in the text. Urinary LAM and TB blood culture were not carried out in HIV negative participants.

for mycobacteria. 12/150 (8%) grew contaminents and are excluded from the denominators in Table 4.5; of the remainder 8/138 (6%) grew mycobacteria, all M. tuberculosis.

Figures 4.5 and 4.4 show the overlap of positive tests form the different diagnostic modalities. Of the 114 patients with at least one positive diagnosic test, 90/114 (79%) had only one positive diagnostic test. The exceptions to this were mycobacterial blood culture and sputum Xpert: patients who had TB diagnosed by these tests tended to also have a positive uLAM. 2/4 (50%) of patients with positive CSF testing (all of whom had detectible CRAG) had also grew Cryptococcus neoformans in aerobic blood culture. 111/225 (49%) of patients remained with no diagnosis.

4.3.5 Treatment

At least one antimicrobial drug was recieved by 95% (214/225) of the cohort during their admission (Table @ref:(time-to-ab-table)), most commonly an antibacterial (207/225 [92%]), but also a significant minority recieved antitubercular therapy (63/225 [28%]). Of those

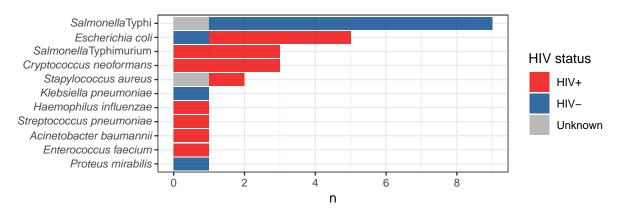


Figure 4.3: Pathogenic isolates recovered from aerobic blood culture. 26 blood cultures in 24 participants were positive for 28 pathogens in total

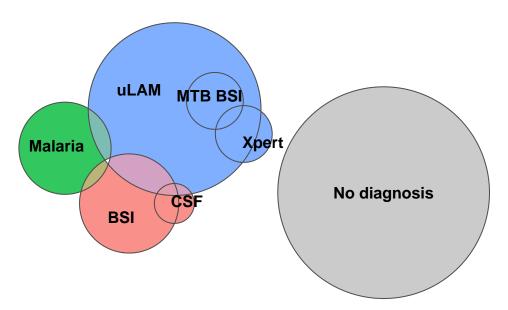


Figure 4.4: Venn diagram showing overlap of positive diagnostic tests; culture of blood and CSF shown in red, malaria in green and TB diagnostics in blue. The CSF variable in includes either a positive culture for a pathogenic bacteria or positive cryptococcal antigen, BSI a positive aerobic culture of pathogenic bacteria from blood and MTB BSI a positive mycobacterial culture of tuberculosis from blood. BSI: Bloodstream infection, CSF: Cerebrospinal fluid, CRAG: Cryptococcal antigen, mRDT: Malaria rapid diagnostic test, MTB BSI: Mycobacterium tuberculosis bloodstream infection, uLAM: urinary lipoarabinomannan.

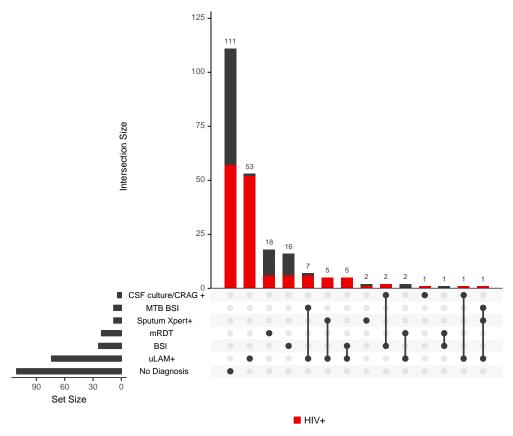


Figure 4.5: UpSet plot of overlap of positive diagnostic tests, showing that for the majority of participants, one test alone is positive. Red colour indicates HIV-infected; black is a composite of HIV-negative and unknown. The CSF variable in includes either a positive culture for a pathogenic bacteria or positive cryptococcal antigen, BSI a positive aerobic culture of pathogenic bacteria from blood and MTB BSI a positive mycobacterial culture of tuberculosis from blood. BSI: Bloodstream infection, CSF: Cerebrospinal fluid, CRAG: Cryptococcal antigen, mRDT: Malaria rapid diagnostic test, MTB BSI: Mycobacterium tuberculosis bloodstream infection, uLAM: urinary lipoarabinomannan.

Antimicrobial class	No. participants	Median [IQR] time (hours)
Antibacterial	207/225 (92%)	5.3 (3.7-10.8)
Antitubercular	63/225 (28%)*	120.9 (63.7-171.0)
Antifungal	26/225 (12%)	47.7 (27.9-73.9)
Antimalarial	12/225 (5%)	4.5 (3.1-21.7)

Table 4.6: Door-to-antimicrobial times.

receiving antitubercular therapy, 16% (10/63) were taking the medication prior to admission, and the remainder were initiated on therapy during admission. The first antibacterial agent administered was most often ceftriaxone, in 87% (181/207) of cases but ciprofloxacin (18/207 [9%] of participants), amoxicillin (6/207 [3%]) and metronidazole (2/207 [1%]) were also used. Median door to antimicrobial time was 5.3 (IQR 3.7-10.8) hours for antibacterials and 4.5 (IQR 3.1-21.7) hours for antimalarials but longer for antifungals at 47.7 (IQR 27.9-73.9) hours and longer still for antitubercular therapy at 120.9 (IQR 63.7-171.0). Cumulative incidence curves for administration of the different antimicrobial classess are shown in Figure 4.6A-D.

Of all participants, 85% (192/225) received any intravenous fluid in the first 6 hours of enrollment to the study; of these, most received 0.9% saline (160/192 [83%] of those recieving fluid) but 5% dextrose (91/192 [57%]) were also used; Ringer's lactate (6/192 [6%]) and blood (2/192 [1%]) were rarely administered. Of the 192 patients who were administered any fluid, a median of 1.5L (IQR 1-2L) was administered over the 6hr study period; fluid administration as a function of time is shown in Figure 4.6E.

4.3.6 Outcome

Median hospital stay was 4 (IQR 1-9) days. Mortality of the cohort was 18% (95% CI 13-23%) at 28 days, 24% (95% CI 18-30%) at 90 days and 31% (95% CI 25-38) at 180 days, and higher in HIV-infected participants at each time point (Table 4.7), though not statistically significant on pairwise Fisher's exact test (HIV-infected vs noninfected 19% vs 13%, [p = 0.14] at 28 days, 27% vs 17%, [p = 0.44] at 90 days and 36% vs 21% [p = 0.29] at 180 days). Kaplan-meier estimation of the survival function (Figure 4.7) showed a precipitous decline in survivorship to around day 30 but also continuous mortality therafter, to the end of the study period. Stratifying the analysis by HIV status revealed that early deaths (within the first 1-2 weeks) occur at similar rates in the two groups before the curves diverge; log-rank test suggested a significant difference in survival function between the two groups (p = 0.03).

Health related quality of life measures, as assessed by EQ-5D-3L, are shown in Figure 4.8 for

^{* 10/63} participants who received antitubercular agents during admission were taking them prior to admission; they are excluded from the calculation of median door-to-antimicrobial time for this class.

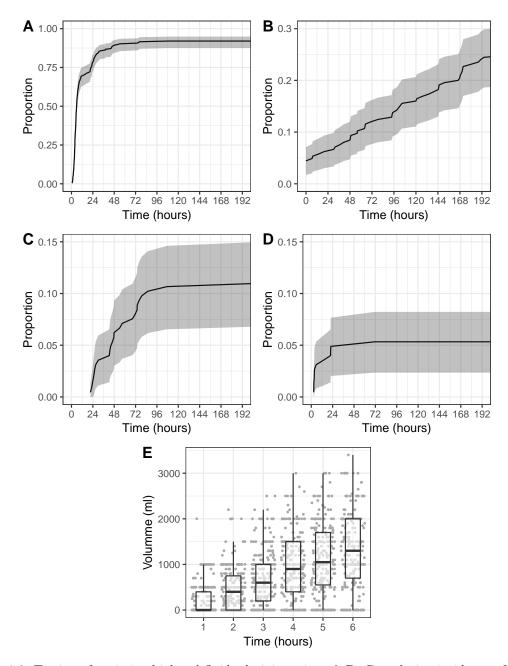


Figure 4.6: Timing of antimicrobial and fluid administration. A-D: Cumulative incidence of administration of antibacterial (A), antitubercular (B), antifungal (C) and antimalarial (D) agents as a function of time since arrival at hospital in hours. E: Total volumne of administered intavenous fluid as a function of time since enrollment to study in hours. Boxplots show median, quartiles box and 1.5 times interquartile range as whiskers. Points are jittered around the hour at which they were measured to show distribution.

	HIV+		HIV-		HI	HIV Unknown		Total	
	n	Mortality	n	Mortality	n	Mortality	\mathbf{n}	Mortality	
Day 28 Day 90 Day 180	143 139 125	19% (13-26) 27% (19-35) 36% (28-45)	67 64 58	13% (6-24) 17% (9-29) 21% (11-33)	12 12 11	25% (5-57) 25% (5-57) 27% (6-61)	222 215 194	18% (13-23) 24% (18-30) 31% (25-38)	
A _{1.0}				B 1.0					

Table 4.7: Day 28, 90 and 180 mortality stratified by HIV status

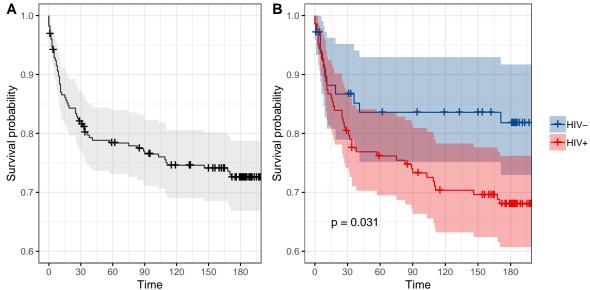


Figure 4.7: Kaplan-Meier survival curves of all included participants (A) and stratified by HIV status (B). Crosses indicate censoring. p value from log-rank test comparing survival of HIV-infected to HIV-noninfected participants shown (p = 0.03).

participants with sepsis and the community cohort as a comparator. Acutely, participants with sepsis reported were significantly disabled, reporting at least moderate impairment across all domains in the majority of cases, and over 90% of participants reporting at least moderate impairment in activities of daily living and experienceing at least moderate pain or discomfort. However, recovery following treatment in survivors was rapid. The mean EQ-5Q utility score (a measure of the weight compared to a health state compared to 1, perfect health) of healthy community controls was 0.910 (SD 0.102) at enrollment, significantly higher than participants with sepsis at enrollment (utility score 0.496 (SD 0.251), p = < 0.0001 versus community scores by t-test), but comparable to participants with sepsis at their 12 week assessment (0.913 (SD 0.147), p = 0.903 versus community enrollment scores).

4.3.6.1 Associations of mortality

Univariate associations of host and severity variables

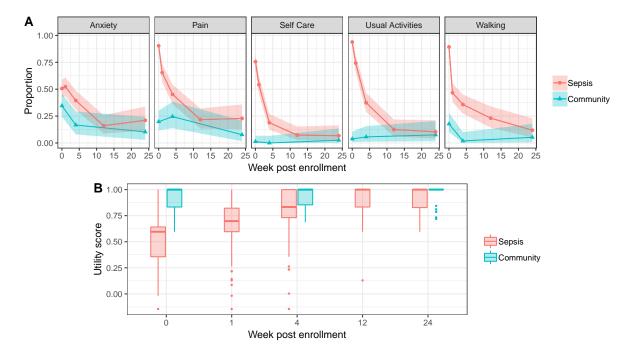


Figure 4.8: Health-related quality of life following sepsis admission, compared to community controls, showing a return to usual quality of life by 12 weeks following admission. A: proportion of participants across each of the five domains of the EQ-5D questionnaire who report at least moderate impariment. B: EQ-5D utility score derived using the Zimbabwean tariff set. The utility score is interpreted as the weight attached to a health state compared to perfect health, which is assigned a value of 1. By 12 weeks there is no statistically significant difference between sepsis and baseline community participant utility scores (p = 0.90 by t-test).

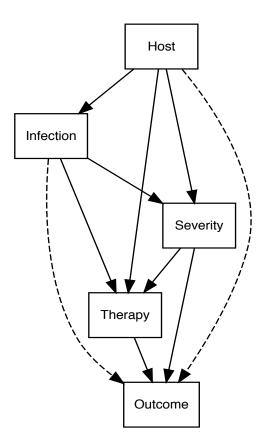


Figure 4.9: Hypothesised causal structre of death in sepsis. Host factors would theoretically include variables quantifying age, sex, immune status; infection would theoretically include causative agents, site, and variables quantifying pathogen burden; severity would include includes measures of disease severity such as shock or respiratory failure; therapy would include which antimicrobials were administered and rapidity of administration of antimicrobials. Dotted edges from host and infection to outcome are because it is not clear extita priori whether the effect of infection and host factors are entirely mediated by disease severity: see text for details.

Univariate association of

Table 4.8: Bivariate associations with death by 28 days

Variable	Died	Survived	p
Host Variables			
Age (years)	36.4 (31.5-46.0)	35.9 (27.4-42.9)	0.252
Male sex	19/39 (49%)	93/183 (51%)	0.861
$\operatorname{HIV} \operatorname{Infected}^*$	$27/36 \ (75\%)$	116/174~(67%)	0.433
${ m CD4}\;{ m count}^{\dagger}\;(\mu{ m L}^{-1})$	$28.5 \ (9.5 - 124.5)$	103.0 (43.5-251.0)	0.007
Haemoglobin $(x10^9 \text{ g dL}^{-1})$	$9.1 \ (6.0-10.4)$	$11.0 \ (8.6-13.4)$	< 0.001
Severity Variables			
Temperature (°C)	$38.1 \ (37.7 - 38.8)$	$38.5 \ (38.0 \text{-} 39.0)$	0.024
Heart rate (min ⁻¹))	123.0 (104.5-138.5)	120.0 (102.0-131.0)	0.510
Systolic BP (mmHg)	89.0 (76.0-106.0)	99.0 (86.5-118.5)	0.047
Diastolic BP (mmHg)	$59.0 \ (51.0-72.0)$	$67.0 \ (57.0-75.5)$	0.040
Mean arterial BP (mmHg)	$69.7 \; (60.0 - 81.3)$	$78.7 \; (67.0 \text{-} 89.2)$	0.035
Respiratory rate (min ⁻¹))	34.0 (32.0 - 36.5)	34.0 (32.0-38.0)	0.720
Oxygen saturation $(\%)$	$95.0 \ (89.5-97.0)$	$97.0 \ (95.0 - 98.0)$	0.019
GCS	$15.0 \ (15.0 \text{-} 15.0)$	$15.0 \; (15.0 \text{-} 15.0)$	0.044
Lactate (mmol L^{-1})	$4.6 \ (3.0 \text{-} 10.6)$	3.2 (2.1 - 4.5)	0.001
White cell count $(x10^9 L^{-1})$	5.9 (3.5-11.0)	6.9 (4.5-11.5)	0.165
Platelet count (x10 ⁹ L ⁻¹)	181.5 (86.8-300.8)	223.0 (148.0-296.5)	0.291
Sodium (mmol L ⁻¹)	131.0 (127.0-137.8)	134.0 (130.0-137.0)	0.280
Potassium (mmol L ⁻¹)	4.0 (3.4-4.5)	4.0 (3.6-4.4)	0.683
Chloride (mmol L^{-1})	101.0 (96.0-104.8)	101.0 (98.0-104.0)	0.846
Bicarbonate (mmol L ⁻¹)	$17.0 \ (14.0-21.0)$	$20.0 \ (17.0 - 22.0)$	0.007
${ m Urea} \; ({ m mmol} \; { m L}^{-1})$	$7.8 \ (4.5-14.3)$	$4.5 \ (3.2-7.0)$	< 0.001
Creatinine (mmol L ⁻¹)	90.0 (60.0-185.0)	73.0 (59.0-96.0)	0.100

Note:

 $\mathrm{BP}=\mathrm{Blood}$ pressure, $\mathrm{GCS}=\mathrm{Glasgow}$ coma scale. Numeric variables are presented as median (IQR) and categoric variables as proportions. P-values are from Kruskal-Wallace test for continuous variables and Fisher's exact test for categorical variables

 $^{^{\}ast}$ Participants with HIV status unknown not included in this row

 $^{^\}dagger$ CD4 count only measured for HIV-infected participants

Table 4.9: Bivariate associations with diagnosis

Variable	ТВ	Malaria	BSI	No Diagnosis	p
n	69	18	18	112	-
Host Variables					
Age (years)	38.0 (31.4-42.7)	32.3 (20.4-40.3)	32.8 (24.0-42.7)	36.5 (29.2-44.9)	0.289
Male sex	34/69(49%)	12/18~(67%)	9/18 (50%)	55/112 (49%)	0.587
${ m HIV~Infected}^*$	66/67~(99%)	$6/17 \ (35\%)$	8/16~(50%)	58/107~(54%)	< 0.001
CD4 count [†] (μ L ⁻¹)	100.0 (38.0-209.0)	119.0 (58.5-305.5)	19.0 (6.5-103.0)	99.5 (31.0-259.5)	0.196
Haemoglobin $(x10^9 \text{ g dL}^{-1})$	9.1 (6.6-11.3)	$12.1 \ (10.6-14.2)$	$10.8 \ (10.1-12.7)$	11.6 (9.2 - 13.9)	< 0.001
Severity Variables					
Temperature (°C)	38.5 (38.0-39.1)	38.5 (38.1-39.5)	39.0 (38.1-39.3)	38.3 (37.8-38.9)	0.224
Heart rate (min ⁻¹))	125.0 (117.0-135.0)	112.5 (95.5-131.0)	116.0 (101.5-131.8)	116.0 (98.8-130.0)	0.014
Systolic BP (mmHg)	96.5 (81.0-113.5)	96.5 (86.2-112.8)	99.0 (91.2-120.0)	99.0 (86.5-121.0)	0.777
Diastolic BP (mmHg)	67.0 (52.8-72.5)	61.0 (54.5-78.8)	69.0 (63.2-75.5)	66.0 (58.0-76.5)	0.393
Mean arterial BP (mmHg)	76.7 (62.3-85.9)	70.3 (65.6-87.8)	82.0 (71.4-90.7)	77.3 (67.0-92.0)	0.431
Respiratory rate (min ⁻¹))	$34.0 \; (32.0 \text{-} 38.0)$	$32.0\ (28.0\text{-}33.5)$	$35.5 \ (34.0 \text{-} 37.5)$	$34.0 \ (32.0 \text{-} 37.0)$	0.030
Oxygen saturation (%)	97.0 (94.0-98.0)	97.0 (95.0-98.0)	96.0 (84.8-98.0)	96.0 (95.0-98.0)	0.947
GCS	15.0 (15.0-15.0)	$15.0 \ (15.0-15.0)$	15.0 (15.0-15.0)	15.0 (15.0-15.0)	0.362
Lactate (mmol L^{-1})	$3.1\ (2.4-5.2)$	$3.1\ (2.1-4.4)$	3.9(3.0-4.8)	3.4(2.0-5.2)	0.586
White cell count $(x10^9 L^{-1})$	$6.2 \ (4.2 - 9.2)$	5.2 (3.5-6.8)	8.7 (4.2-13.0)	7.0 (4.8-11.8)	0.045
Platelet count (x10 ⁹ L ⁻¹)	223.0 (147.8-302.5)	81.5 (43.5-150.2)	160.5 (131.8-293.8)	230.0 (170.5-309.5)	< 0.001
Sodium (mmol L ⁻¹)	131.5 (127.0-135.0)	134.0 (133.0-135.0)	134.0 (132.0-136.8)	134.0 (131.8-138.0)	0.003
Potassium (mmol L ⁻¹)	4.0 (3.5-4.3)	4.2 (3.7-4.4)	3.7 (3.4-4.2)	4.1 (3.6-4.5)	0.118
Chloride (mmol L ⁻¹)	100.5 (97.0-104.0)	102.5 (101.0-104.0)	97.5 (96.0-102.8)	100.5 (98.0-104.0)	0.213
Bicarbonate (mmol L ⁻¹)	$18.0\ (14.8-20.0)$	20.0 (17.2-22.0)	20.0 (16.0-23.8)	20.0 (18.0-23.0)	0.003
Urea (mmol L ⁻¹)	4.7 (3.5-8.1)	4.7 (3.9-10.1)	6.7 (4.4-9.2)	4.6 (3.3-7.1)	0.291
Creatinine (mmol L ⁻¹)	$75.0\ (55.0-99.0)$	72.0 (61.8-89.5)	104.0 (73.0-126.8)	77.0 (59.0-97.2)	0.077

Note:

BP = Blood pressure, GCS = Glasgow coma scale. Numeric variables are presented as median (IQR) and categoric variables as proportions. P-values are from Kruskal-Wallace test for continuous variables and Fisher's exact test for categorical variables across four diganositic groups

* Participants with HIV status unknown not included in this row

[†] CD4 count only measured for HIV-infected participants

Chapter 5

Early response to resusitation in sepsis

Chapter 6

Gut mucosal carriage of ESBL-E in Blantyre, Malawi

 $42\ \ CHAPTER\ 6.\ \ GUT\ MUCOSAL\ CARRIAGE\ OF\ ESBL-E\ IN\ BLANTYRE,\ MALAWI$

Chapter 7

Whole genome sequencing of ESBL $E.\ coli$ carriage isolates

Placeholder

 $44 CHAPTER\ 7.\ \ WHOLE\ GENOME\ SEQUENCING\ OF\ ESBL\ E.\ COLI\ CARRIAGE\ ISOLATES$

7.1 Chapter overview

- 7.2 Methods
- 7.2.1 Bioinformatic pipeline
- 7.2.2 Global $E.\ coli$ collection
- 7.2.3 Statistical analysis
- 7.3 Results
- 7.3.1 Samples and quality control
- 7.3.2 Phylogroup, MLST and core genome phylogeny of study isolates
- 7.3.3 Study isolates in a global context
- 7.3.4 Antimicrobial resistance determinants
- 7.3.4.1 β -lactam resistance
- 7.3.4.2 Quinolone resistance
- 7.3.4.3 Aminoglycoside resistance
- 7.3.4.4 Chloramphenicol, co-trimoxazole, tetracycline and other resistance determinants
- 7.3.4.5 Clustering and lineage association of AMR determinants
- 7.3.5 Plasmid replicons
- 7.3.6 Testing metadata associations: SNP distance, hierBAPS sequence clusters and ESBL-clusters
- 7.3.6.1 Hierarchical BAPS clustering of core gene pseudosequences
- 7.3.6.2 ESBL-clusters
- 7.3.6.3 Assessing for healthcare-associated lineages
- 7.3.6.4 Assessing for within-patient conservation of lineage or MGE
- 7 4 Diament

 $46 CHAPTER\ 7.\ \ WHOLE\ GENOME\ SEQUENCING\ OF\ ESBL\ E.\ COLI\ CARRIAGE\ ISOLATES$

References