

This is the Title of my Thesis

Your Name

August 2014

PROJECT / MASTER THESIS

Department of Production and Quality Engineering  
Norwegian University of Science and Technology

Supervisor: Professor Ask Burlefot

## Preface

Here, you give a brief introduction to your work. What it is (e.g., a Master's thesis in RAMS at NTNU as part of the study program xxx and...), when it was carried out (e.g., during the autumn semester of 2021). If the project has been carried out for a company, you should mention this and also describe the cooperation with the company. You may also describe how the idea to the project was brought up.

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Trondheim, 2012-12-16

(Your signature)

Ola Nordmann

## Acknowledgment

I would like to thank the following persons for their great help during . . .

If the project has been carried out in cooperation with an external partner (e.g., a company), you should acknowledge the contribution and give thanks to the involved persons.

You should also acknowledge the contributions made by your supervisor(s).

O.N.

(Your initials)

## Summary and Conclusions

Here you give a summary of your your work and your results. This is like a management summary and should be written in a clear and easy language, without many difficult terms and without abbreviations. Everything you present here must be treated in more detail in the main report. You should not give any references to the report in the summary – just explain what you have done and what you have found out. The Summary and Conclusions should be no more than two pages.

You may assume that you have got three minutes to present to the Rector of NTNU what you have done and what you have found out as part of your thesis. (He is an intelligent person, but does not know much about your field of expertise.)

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# Chapter 1

## Introduction

# Chapter 2

## Theory

### 2.1 Electron Microscopy

In an electron microscope, a beam of electrons are transmitted onto a material. Compared to a microscope using visible light, an EM gives a much higher resolution due to the wavelength of electrons being much smaller than that of visible light. Because of this, an EM can be used to study the atomic structure of materials.

#### 2.1.1 Electron-sample interaction

When the electron beam hits the sample, several different signals are generated. This report will only discuss two of them: Characteristic X-rays (due to inelastically scattered electrons, and elastically scattered electrons. Characteristic X-rays are produced as the incoming electrons interact with inner-core electrons in the sample, and can be used to determine which elements the sample consists of. The theory behind characteristic X-rays, and why they are useful, will be expanded upon in section 2.2. Electrons being transmitted through the sample that are elastically scattered (that is, the energy of the incoming electrons is conserved) gives rise to a diffraction pattern, which can be used to determine the crystallographic properties of the sample. Electron diffraction will be properly introduced in section 2.3 [7].



### 2.1.2 Transmission Electron Microscopy (TEM)

There are many different kinds of transmission electron microscopes (TEMs), of which only the scanning TEM (STEM), introduced in section ??, will be expanded upon here. Before discussing the STEM, however, the basics behind the TEM will now be presented. In a conventional TEM, a broad, parallel beam of electrons is transmitted through a sample. In order for a sufficient amount of electrons to be transmitted, the sample is required to be thin (a thickness of  $<100\text{ nm}$  is common). A TEM typically operates with electron energies of  $\sim 200\text{ keV}$  or  $300\text{ keV}$ , giving the electrons a velocity  $v > c/2$  which requires relativistic effects to be taken into account. Another factor that should be considered is the possibility of the electron beam damaging the material. This can happen as the chemical bonds in the material are broken (radiolysis), atoms are displaced or ejected from the crystal lattice (knock-on damage or sputtering, respectively) or as the material is changed due to heating. [7]

### 2.1.3 Scanning Transmission Electron Microscopy (STEM)

A scanning transmission electron microscope (STEM) uses an electron beam focused onto a small spot on the sample. The sample is then scanned in a raster pattern and the signal is recorded for each point. This can result in a very accurate map of the sample ++++++.

## 2.2 X-ray spectroscopy

The characteristic X-rays resulting from inelastic scattering of the electrons being transmitted through the sample can be used for both qualitative and quantitative analysis. Qualitative analysis can determine which elements are in the sample, while quantitative analysis can indicate the composition of the different elements. When X-ray spectroscopy is performed in a STEM, the electron probe is scanned over the material and focused on each pixel for a certain amount of time. During this time period, the X-rays being emitted from the sample are detected and the signal for the pixel is saved before the probe moves to the next pixel.

## 2.2.1 Energy-Dispersive X-Ray Spectroscopy (EDS)

### X-ray emission

As previously mentioned, characteristic X-rays are produced due to inelastic scattering of the incoming electrons on the sample. If an electron reaches the inner-shell electrons of an atom in the sample, and a sufficient amount of energy is transferred to the inner-shell electron, the electron will be ejected from the nucleus. That is, it reaches an energy above the Fermi level and is no longer bound to the nucleus [CHECK THIS]. The atom is now in an excited state, and will seek to return towards its ground state. This leads to an electron in an outer shell taking the place of the ejected electron in the inner shell, and in the process emitting an X-ray or an Auger electron (Auger electrons will not be discussed further). If an X-ray is emitted, its energy will be characteristic of the specific element, as different elements have different critical ionization energies. The critical ionization energy is the energy required for an inner-shell electron to be ejected from the nucleus.

There will not necessarily only be emitted one characteristic X-ray for each hole created in the inner shell of an atom. This is because the outer-shell electron filling the hole will create a new hole in its shell, which must be filled by an electron in a shell even further out, and maybe emitting another characteristic X-ray. This process continues until there is a hole in the outermost shell of the atom that can be filled by an electron in the conduction or valence band of the material. The energy of the characteristic X-rays will also depend on which transition it is emitted because of. In addition, due to quantum effects the electron shells might be split into several sub-shells, further increasing the number of different energies the emitted X-rays can have.

The probability for a specific transition to occur must also be taken into account. If a transition between two specific shells has a relatively high probability compared to other transitions in the element, more characteristic X-rays with the corresponding energy will be emitted over a certain time period. This will lead to higher peaks in the EDS spectrum, which will be discussed shortly.

### X-ray detection

When doing EDS in a TEM, an X-ray detector is positioned above the sample to detect the X-rays emitted from it. The detector then measures the energy

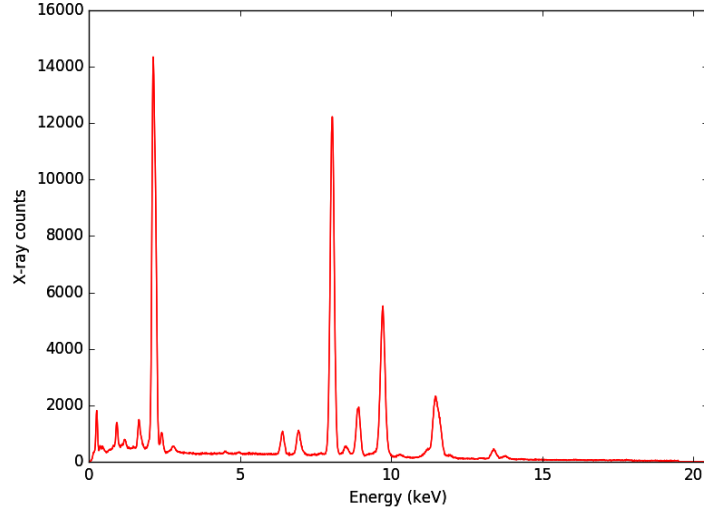


Figure 2.1: Example of EDS spectrum

of each incoming X-ray, with a certain energy resolution +++++. If the measurement is done using STEM, the electron probe is scanned over the sample and kept at each pixel for a certain amount of time during which emitted X-rays are detected. When the scanning is complete, this results in a three dimensional EDS spectrum.

### EDS spectrum

In an EDS spectrum, the intensity of the X-rays is plotted against the energy of the X-rays. The intensity at a certain energy refers to the number of counts, that is, how many X-rays with that energy that were detected by the detector. An example of an EDS spectrum is shown in Figure 2.1. By consulting a table of known X-ray energies for the emission lines from different elements, for example [2], one can determine which elements are present in the sample. However, knowing which elements are present is not enough to determine the structure of the material. A next step can be to perform quantitative analysis on the same spectrum.

## 2.2.2 Quantitative X-ray Analysis

Quantitative analysis of an EDS spectrum gives information about the composition of the different elements in the material. A quantitative analysis might result in learning that an unknown material is a compound of a few different elements. In order to determine which compound this is, quantitative analysis is necessary.

There are primarily two methods for quantitative X-ray analysis: The Cliff-Lorimer ratio technique and the  $\zeta$ -factor method. The basis for both is assuming that the concentration of an element in a material is proportional to the intensity of characteristic X-rays being emitted from it,  $C \propto I$ .

### The Cliff-Lorimer ratio technique

The Cliff-Lorimer ratio technique, first introduced in 1972 [3], assumes that the weight percent ratio of two elements is proportional to their intensity ratio,

$$\frac{C_A}{C_B} = k_{AB} \frac{I_A}{I_B}, \quad (2.1)$$

where  $C$  is the weight percent,  $I$  is the intensity of the characteristic X-rays and the proportionality factor  $k_{AB}$  is called the Cliff-Lorimer factor, or the  $k$ -factor. The  $k$ -factor depends not only on the properties of the elements  $A$  and  $B$ , but also on the instruments used, the conditions under which the experiment was done, and how the intensities are extracted from the spectrum. This technique has several drawbacks: Firstly, determining the  $k$ -factor experimentally is very time-consuming, but a theoretical calculation can have a severe error [7]. And secondly, the multi-element thin standards necessary for experimental determination are difficult to prepare.

### The $\zeta$ -factor method

The  $\zeta$ -factor method was introduced by M. Watanabe and D. B. Williams in 1996, and overcomes several of the problems with the Cliff-Lorimer technique. If the material is a thin-film, its mass-thickness can be assumed to be proportional to the intensity of the characteristic X-rays  $I$  and the composition  $C$ ,

$$\rho t = \zeta \frac{I}{C D_e}, \quad (2.2)$$

where  $\rho$  is the density of the material,  $t$  is its thickness and  $D_e$  is the total amount of electrons that hit the material during the measurement.

Using the thin-film approximation to find a theoretical expression for the intensity  $I$ , the proportionality factor  $\zeta$  can be found to be

$$\zeta = \frac{M}{N_v Q \omega a [\Omega/(4\pi)] \epsilon}. \quad (2.3)$$

Here,  $M$  is the atomic weight,  $N_v$  is Avogadro's number,  $Q$  is the ionization cross section,  $\omega$  is the fluorescence yield,  $a$  is the relative transition probability,  $\Omega/(4\pi)$  is the detector collection-angle and  $\epsilon$  is the detector efficiency [4].

If the composition and thickness of a material is known, equation (2.2) can be used to calculate the  $\zeta$ -factor for the material. ++++++

## 2.3 Electron diffraction

When electron diffraction is performed in a TEM, a beam of electrons is transmitted through the specimen. As the electrons pass through, some are scattered due to the atomic potentials in the specimen. By detecting the positions of the electrons after they have been transmitted through the specimen, a diffraction pattern is obtained. Analyzing this diffraction pattern can determine the characteristics of the specimen, for example whether it is crystalline or amorphous, if there are several phases present, and if crystalline, what its crystallographic characteristics are. This information can not be obtained through EDX.

Electron diffraction in a STEM, or scanning electron diffraction (SED) works similarly to EDX in STEM: A parallel electron probe is scanned over the specimen in a raster pattern, and the resulting 2-dimensional diffraction pattern is recorded for each pixel. The result is a 4-dimensional dataset: Two axes in real space and two in reciprocal space.

### 2.3.1 Diffraction theory

When discussing the diffraction of electrons, it is convenient to regard the electrons as waves. When a parallel electron wave with a wave vector  $\vec{k}_i$  reaches the specimen, the electrons will scatter to different angles  $\theta$ . Assuming the scattering is purely elastic, i.e. the energy of the electrons is

conserved, the wave vectors of the scattered electrons will be equal in magnitude to the wave vector of the incoming electrons,  $|\vec{k}_s| = |\vec{k}_i|$ . After being transmitted and scattered, the waves will interfere with each other. Due to the electrons being scattered to different angles, their path lengths will be different and thus they will be out of phase. Bragg's law introduces the Bragg angle  $\theta_B$ , which is the scattering angle at which there will be positive interference:

$$n\lambda = 2d \sin \theta_B \quad (2.4)$$

Here,  $n$  is an integer,  $\lambda$  is the wavelength of the electrons and  $d$  is the distance between the atoms in the lattice, as seen from the direction of the incoming electrons. At the angles  $\theta_B$  that satisfy the above equation, a bright spot will be visible in the diffraction pattern.

The equivalence of Bragg's law in reciprocal space is the Ewald sphere construction. ++++++++ Write about Ewald sphere

+++++++ Mention kinematic and dynamical scattering

## 2.3.2 Electron diffraction in STEM

### 2.3.3 Scanning Precession Electron Diffraction (SPED)

Scanning precession electron diffraction (SPED) is a modification of SED. Figure 2.2 shows the schematic of SPED. The electron beam is now tilted at a fixed angle and precessed around the vertical axis. The beam will hit the sample at the same spot during the precession, only the direction of the incoming beam will be changed. The diffracted beam is also precessed so that the location of the diffraction spots is invariant. The precession speed is set so that the beam does a full rotation at every pixel in the raster pattern.

There are several reasons why adding precession to electron diffraction gives a better results. The perhaps most important is that the Ewald sphere (see Figure +++++++) is precessed in reciprocal space, leading to more diffraction spots being visible. Another major improvement is the reduction of dynamical effects in the diffraction pattern. Double scattering results in certain diffraction spots appearing, even though these do not satisfy Bragg's law. By tilting the incoming electron beam it no longer enters the sample at the exact zone axis, resulting in fewer dynamic effects ++++++ explain this [6].

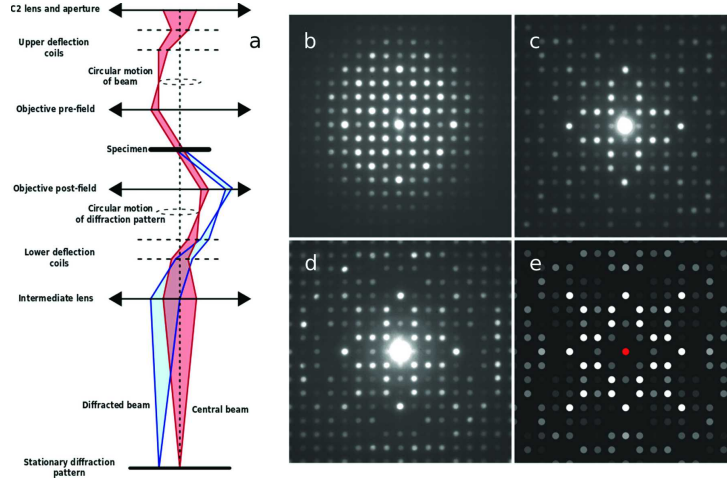


Figure 2.2

## 2.4 Data analysis

### 2.4.1 (Introduction)

### 2.4.2 (?) Multivariate analysis

### 2.4.3 Template matching

Template matching is used for finding the location in a reference image where a template image fits best. The function *MatchTemplate* in the python library OpenCV does this by sliding the template image over the reference image, and recording how well it fits at each point. This matching value is given as the normed square difference between the two images for each pixel in the reference image:

$$R(x, y) = \frac{\sum_{x', y'} (T(x', y') - I(x + x', y + y'))^2}{\sqrt{\sum_{x', y'} T(x', y')^2 \cdot \sum_{x', y'} I(x + x', y + y')^2}} \quad (2.5)$$

Here,  $(x, y)$  are the coordinates of the reference image,  $(x', y')$  the coordinates of the template,  $I(x, y)$  is the value of the pixel  $(x, y)$  in the reference image and  $T$  likewise in the template. The matching value  $R(x, y)$  can take any value between 0 and 1, where 0 means a perfect match and 1 a perfect mismatch.

# Chapter 3

## Experimental

### 3.1 Material

### 3.2 Equipment

### 3.3 Dataset matching

Several methods were attempted in order to match the SPED and EDX datasets. The goal was that the matching would be performed automatically with no human input. For this to happen, the code would have to scale, rotate and translate one of the the datasets so that it fit the other. This requires several discernible features in both images. For this reason, the EDX dataset taken at the intersection of the nanowire and the other elements deposited on it, was chosen. The SPED dataset available at the start of the project was taken over a larger area than the EDX dataset. It was therefore decided to attempt to locate the position of the EDX dataset in the SPED dataset.

The python library `imreg_dft` [?] uses image registration to match two images. Several failed attempts were made using this technique. First, no processing was made to either dataset. The mean was taken in signal space in order to convert them into 2D images, and the intensities were scaled to the range  $[0, 255]$  so they would be uint8-images. The images were then input into the function, but the result did not come close. Several other attempts were done, following the same structure, except that the datasets were processed in different ways before being converted to 2D images:



- Principal component analysis (PCA) was performed on each dataset, and only the component most related to the GaAs nanowire was kept. The nanowire in each of the images now had a high intensity while all other areas were low in intensity.
- A similar result was obtained by inserting a virtual annulus in the SPED dataset, so that only a single diffraction spot corresponding to the nanowire was visible. In the EDX dataset, the EDX spectrum was cropped to only include the characteristic peaks corresponding to the nanowire.
- A virtual high-angle annular dark field aperture (virtual HAADF) was inserted in the SPED dataset, which was then compared to the DF image taken during the EDX session.
- .. background subtraction? Thresholding

As none of these attempts gave any results, another and far more extensive python package, OpenCV [?], was used instead. The ECC image alignment algorithm [5] can transform and fit an image using one of three different motions: Translation, euclidian or affine transformation. Here also, different preprocessing steps were taken to make the 2D images of the datasets more similar to each other. The affine transformation uses rotation, translation, scale and shear to fit the images. The euclidian transformation only rotates and translates, so one of the images was rotated before this algorithm was used. The rotation angle was estimated by measuring the angle between the same straight line (the intersection between the nanowire and the deposited elements) in both datasets, and the rotation was performed using the python library sci-kit image [?]. Finally, the pure translation was attempted, and one of the images was rotated and scaled first. The scaling was performed by selecting the same two points in each datasets, and using scikit-image to scale by the relative distance between them.

Of these attempts, only the last attempt using the pure translation transformation gave results that were decent, but it was unstable and it was difficult to estimate the accuracy. It also required an unwanted amount of manual input from the user. It was then discovered that the metadata in HyperSpy contained accurate magnification numbers, so these were now used to scale the images relative to each other. It was then attempted to use another OpenCV algorithm, template matching [1], which was described in Section

?? This algorithm does not detect offset in rotation, so the SPED image was rotated using the manual technique described above.

By inserting a virtual annulus in the SPED dataset so that only the nanowire lighted up, and selecting only the peaks corresponding to the nanowire in the EDX dataset, the two images were now scaled and rotated correctly, with the nanowire bright and other areas dark. The two images were now compared, and the relative position of the scaled EDX image was found in the rotated SPED image.

The position of one of the EDX datasets was now found in the SPED dataset. This information was then used to find the positions of the remaining EDX datasets. This was done by finding the positions of the other EDX datasets relative to the dataset which was already matched to the SPED dataset. To do this, OpenCV template matching was used to locate the positions of all the datasets in the HAADF overview image, which displays a large area of the sample. It was first attempted to use the EDX datasets, converted to images using the technique described earlier. This worked well for the datasets covering a relatively large region of the sample, but not for the smaller ones. Therefore, instead of the datasets, the survey images were used. These images are taken before the EDX recording, and shows a larger region of the sample at the same scale as the EDX dataset. After image matching was complete, the matched region was cropped to the actual area of the EDX dataset. The coordinates of the top left and bottom right corners of this area were found in the survey image's metadata in HyperSpy.

++++++ Actually matching the two full datasets (including rotation of sped)

### 3.4 Quantitative analysis

The first step in the quantitative analysis was to determine the  $\zeta$ -factors of the elements in the sample. As this method had not yet been implemented into HyperSpy [?], the code was acquired from the Master's thesis of A. Garmannslund [?]. The thickness of the material was estimated by using a thickness map (see Section ???). Background subtraction was performed by marking two windows at each side of the peaks, and the intensities of the lines were calculated with an integration window of 1.2 FWHM. The quantification was then performed using functions in HyperSpy.

# Chapter 4

## Results

The results will be introduced in the same order as the experiments were conducted. First, the results of matching of one of the EDX datasets with the SPED dataset will be presented. This is followed the position matching of the various EDX datasets with the overview images. Lastly, the calculated  $\zeta$ -factors and the quantification of the samples using the  $\zeta$ -factor method and the Cliff-Lorimer technique are displayed.

### 4.1 Dataset matching

Fig. 4.1 shows the results of matching the largest EDX dataset with the SPED dataset. As explained in Section 3.3, these images have been processed to be made more similar. Fig. 4.1 shows the processed EDX image and Fig. 4.1b shows the processed SPED image with a rectangle indicating where the EDX image fit best.

As the correct result is unknown, it is difficult to quantify the error in the fit. An attempt was made by looking at the difference between the matching value for the best position, shown in Fig. 4.1b, and the higher values. The matching values are defined in Eq. (2.5). Fig. 4.2 shows the relative error of the 100 best matching values with respect to the best match.

The results from locating the EDX datasets in the HAADF overview images are displayed in Fig. 4.3 and ?? for the non-treated and the heat-treated samples, respectively. In the non-treated sample, the two datasets B and D were not correctly located in the overview image. The calculated location of dataset B was completely off, while the position of D was just

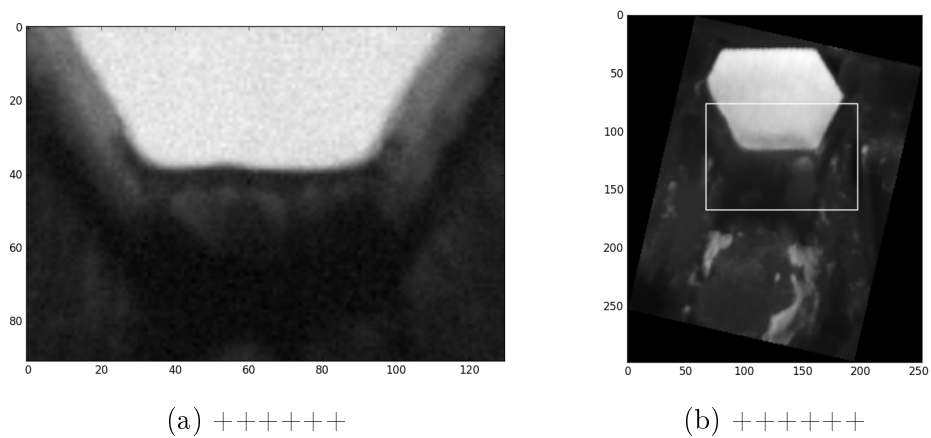


Figure 4.1: plots of...

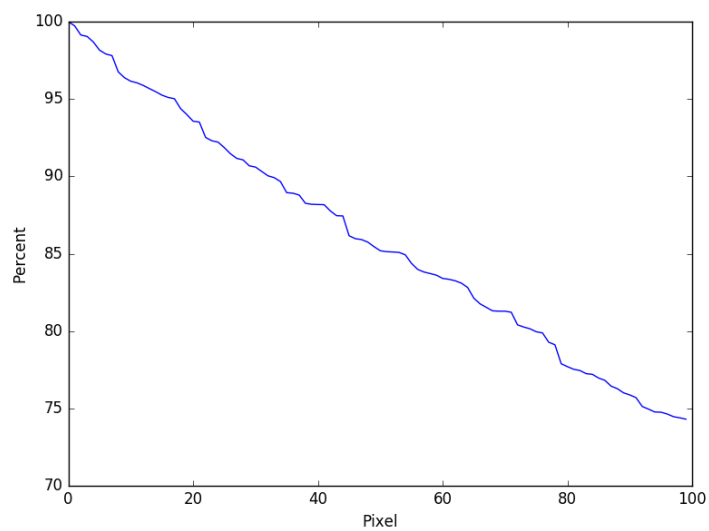


Figure 4.2: ++++++

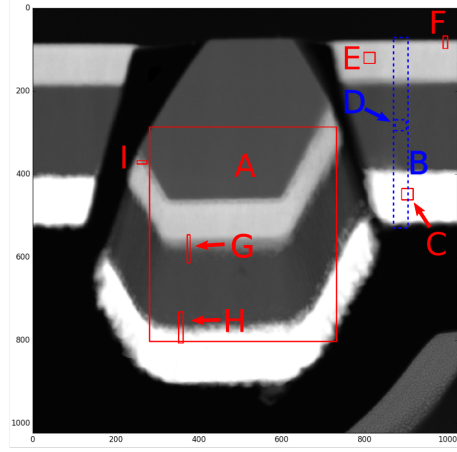


Figure 4.3

slightly offset. The actual positions of these datasets were estimated by eye and marked in Fig. 4.3 as blue dotted rectangles. The rest of the datasets were located correctly, and their positions are shown as red rectangles. For the heat-treated sample, all the datasets that were included in the overview image were correctly located. However, the areas covered by two of the datasets A and B were found to not be included in the overview image.

The accuracy of the matching for the different datasets are shown in Fig. 4.5 for the non-heated sample, and in Fig. 4.6 for the heated values. The horizontal axis shows the different datasets while the vertical axis is the matching value in percent, where 100% is defined to be the matching value of the dataset covering the biggest area in each respective sample. The matching of this dataset is assumed to be the most trustworthy due to there being fewer potential locations that resemble the actual location, as there are several distinct features. For the smaller datasets, or more accurately, the datasets with smaller survey images, there might be several different locations in the overview image that all give a fairly good match.

In both figures, the red star is the value of the dataset used as reference, the red circles are the correctly matched datasets while the blue squares are the datasets that resulted in a wrong location. The non-heated sample (Fig. 4.5) shows high values for all the correctly located datasets, and significantly lower values for the wrongly located ones. In addition, it must be

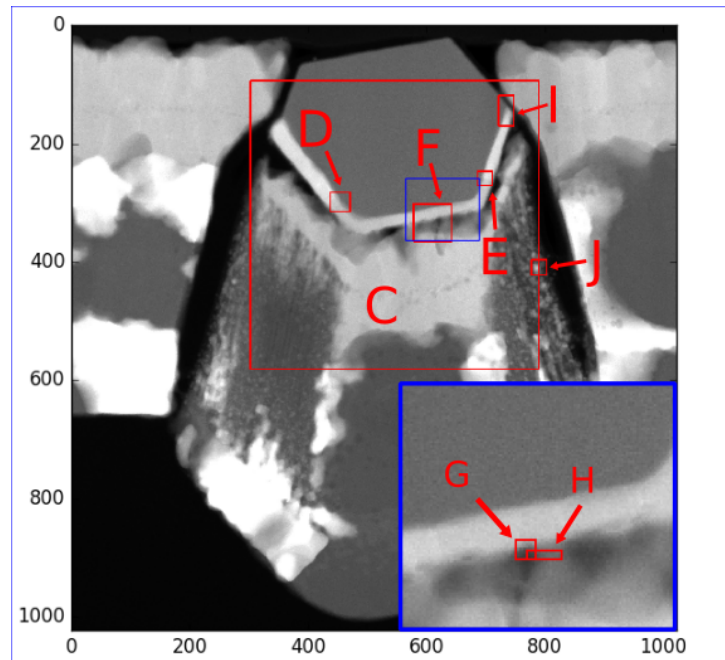


Figure 4.4

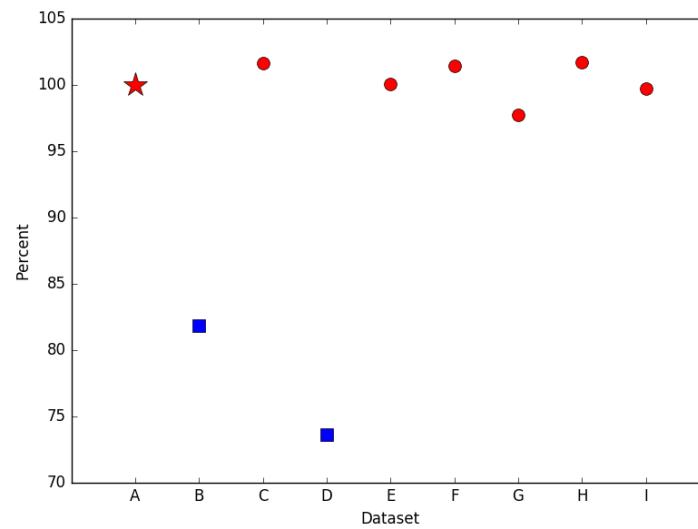


Figure 4.5

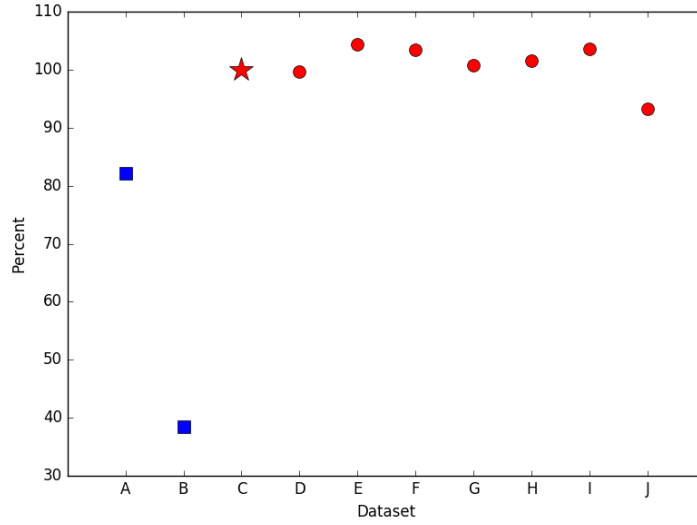


Figure 4.6

noted that the exact location of dataset F was impossible to verify visually due to the survey image not being large enough to distinguish specific features. In the heated sample (Fig. 4.6), all the red correctly located datasets have high values while the datasets that were not present in the reference image have distinguishably lower values.

## 4.2 Determination of $\zeta$ -values

The calculated  $\zeta$ -values for all the elements present in the sample are presented in Table 4.1, along with which dataset was used to calculate them. These values are calculated from the unheated sample. The elements

Table 4.1: ...

Dataset	Element	$\zeta$
Ga	A	608
Ga	C*	582
As	A	706
As	A*	689
Ge	D	741
Ge	B	732
Pd	E	1248
Pd	B	1318
Au	C	3473
Au	B	3450



# Appendix A

## Acronyms

**FTA** Fault tree analysis

**MTTF** Mean time to failure

**RAMS** Reliability, availability, maintainability, and safety

# Appendix B

## Additional Information

This is an example of an Appendix. You can write an Appendix in the same way as a chapter, with sections, subsections, and so on.

### B.1 Introduction

#### B.1.1 More Details

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