

# Class\_10\_020924

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## Whats in the PDB?

Downloaded a CSV file from PDB Link: <http://rcsb.org/stats/summary>

```
pdbstats <- read.csv("pdb_stats.csv", row.names = 1)
head(pdbstats)
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	161,663	12,592	12,337	200	74	32
Protein/Oligosaccharide	9,348	2,167	34	8	2	0
Protein/NA	8,404	3,924	286	7	0	0
Nucleic acid (only)	2,758	125	1,477	14	3	1
Other	164	9	33	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
Total						
Protein (only)	186,898					
Protein/Oligosaccharide	11,559					
Protein/NA	12,621					
Nucleic acid (only)	4,378					
Other	206					
Oligosaccharide (only)	22					

```
as.numeric(pdbstats$X.ray)
```

Warning: NAs introduced by coercion

```
[1] NA NA NA NA 164 11
```

```
# we want to remove all commas in the data so that its not characters anymore:
```

```
#x <- "2,2222,22"
```

```
#gsub(",", "xoxoxoxoxox",x)
```

```
#gsub(",", "",x)
```

```
#as.numeric(gsub(",", "",x))
```

```
#now we want to sum everything for the purpose of this
```

```
comma_sum <- function(x) {  
  sum(as.numeric(gsub(",", "",x)))  
}
```

```
#go to code > Extract function > make your function
```

```
comma_sum(pdbstats$X.ray)
```

```
[1] 182348
```

```
comma_sum(pdbstats$Total)
```

```
[1] 215684
```

```
comma_sum(pdbstats["Protein (only)","Total"])
```

```
[1] 186898
```

```
#applying to the whole table
```

```
head(pdbstats)
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	161,663	12,592	12,337	200	74	32
Protein/Oligosaccharide	9,348	2,167	34	8	2	0
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Total						
Protein (only)	186,898					
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Protein/NA	12,621					
Nucleic acid (only)	4,378					
Other	206					
Oligosaccharide (only)	22					

```
apply(pdbstats, 2, comma_sum) / comma_sum(pdbstats$Total)
```

X.ray	EM	NMR	Multiple.methods
0.8454405519	0.0872433746	0.0657118748	0.0010663749
Neutron	Other	Total	
0.0003662766	0.0001715473	1.0000000000	

```
# I want to round it up now:
round(apply(pdbstats, 2, comma_sum) / comma_sum(pdbstats$Total)*100,2)
```

X.ray	EM	NMR	Multiple.methods
84.54	8.72	6.57	0.11
Neutron	Other	Total	
0.04	0.02	100.00	

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

ANSWER: X.ray: 84.54% EM: 8.72%

Q2: What proportion of structures in the PDB are protein?

```
#pdbstats$Total["Protein (only)", ]

#pdbstats[pdbstats$Total == "Protein (only)", "Total"]

# Assuming pdbstats is the data frame containing information about structures in the PDB
# Access the value for "Protein (only)" from the "Total" column
#protein_only_count <- pdbstats[pdbstats$Total == "Other", "Total"]
```

```

#protein_only_count

# Calculate the proportion of protein structures
#proportion_protein <- protein_only_count / sum((pdbstats$Total)

# Print the proportion
#print(proportion_protein)

Protein_proportion <- (comma_sum(pdbstats["Protein (only)","Total"])) / comma_sum(pdbstats$
Protein_proportion

```

[1] 86.65362

ANSWER: 86.65%

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

```

library(rentrez)

#search and retrieve HIV-1 proteases
search_result <- entrez_search(db = "structure", term = "HIV-1 protease", retmax = 10000)

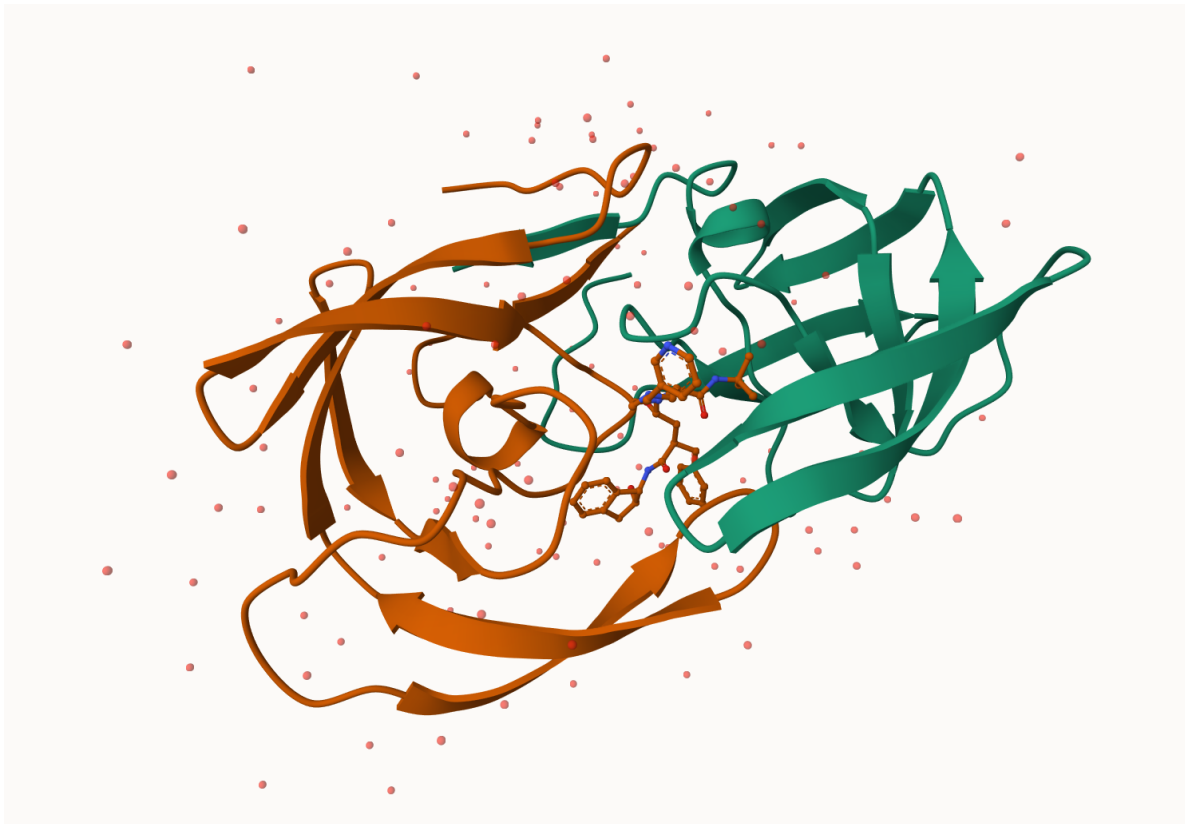
#Count
HIV1_Count <- search_result$count

# Print the count of HIV-1 protease structures in the PDB
HIV1_Count

```

[1] 1065

ANSWER=1065



## Working with structures in R

We will use the package `bio3d` for structural bioinformatics.

```
library(bio3d)  
  
hiv <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
hiv
```

Call: `read.pdb(file = "1hsg")`



Figure 1: A nice display showing the homodimeric inhibitor with the Asp25 highlighted

```

Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

```

```

Protein sequence:
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

```

```

+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call

```

```
head(hiv$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

```
aa123(pdbseq(hiv)[25])
```

```
[1] "ASP"
```

## Predicting functional motions of a single structure

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file  
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

```
Call: read.pdb(file = "6s36")
```

```
Total Models#: 1
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
```

Protein sequence:

```
MRILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV
DELVIALVKERIAQEDCRNGFLLDGFPRTPQADAMKEAGINVDYVLEFDVPDELIVDKI
VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQM TAPLIG
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

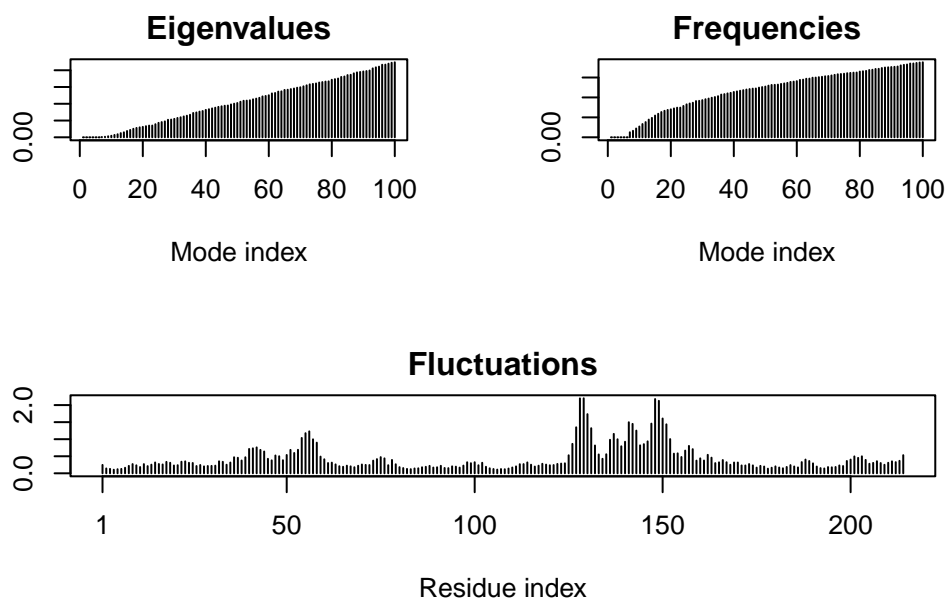
## Normal Mode Analysis (NMA) a tool to predict motions and large-scale structure changes

```
m <- nma(adk)
```

```
Building Hessian...      Done in 0.02 seconds.
Diagonalizing Hessian... Done in 0.462 seconds.
```



```
plot(m)
```



Let's make a movie (a.k.a "trajectory")

```
#To view a "movie" of these predicted motions we can generate a molecular "trajectory" with  
mktrj(m, file="adk_m7.pdb")
```

## Quick comparative analysis of structures

Workflow:

1-PDB seq is in adk 2-Get seq 3-BLAST against PDB 4-Download all the hits 5-Superpose all structures from the blast hit

```
s <- pdbseq(adk)  
blast <- blast.pdb(s)
```

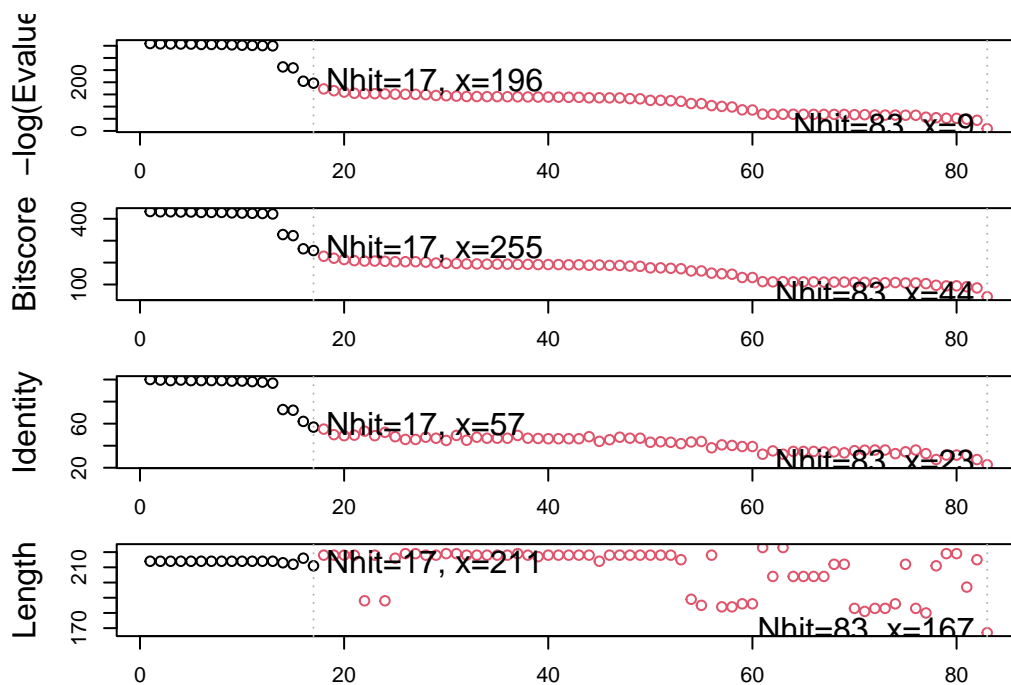
Searching ... please wait (updates every 5 seconds) RID = WMHWK27X01N

Reporting 83 hits

```
plot(blast)
```

```
* Possible cutoff values: 196 9  
    Yielding Nhits: 17 83
```

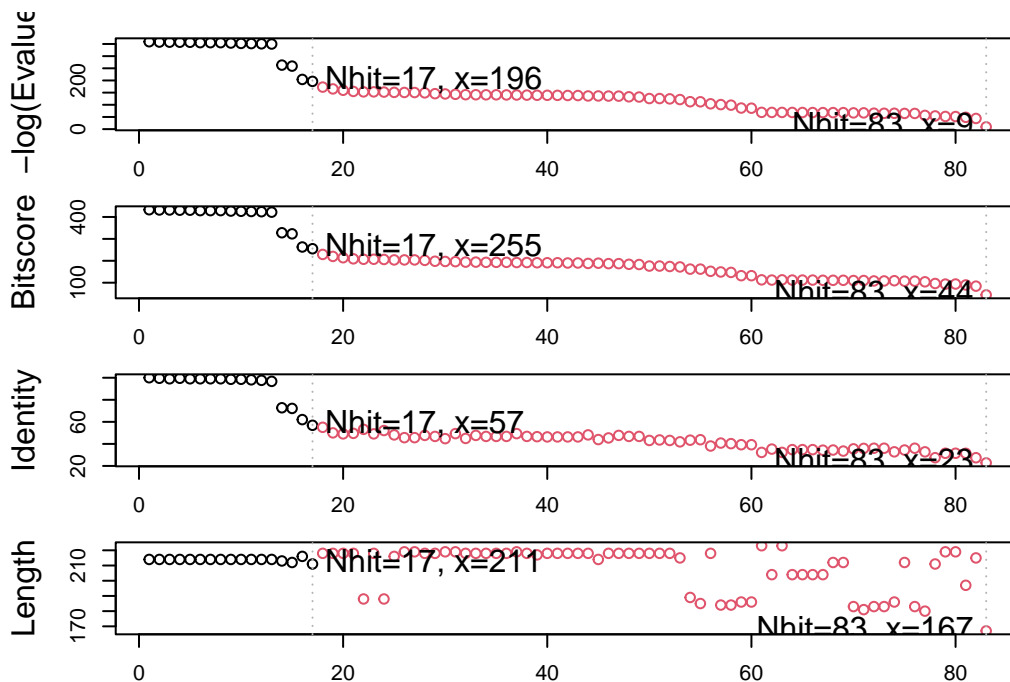
```
* Chosen cutoff value of: 196  
    Yielding Nhits: 17
```



```
hits <- plot(blast)
```

```
* Possible cutoff values: 196 9  
    Yielding Nhits: 17 83
```

```
* Chosen cutoff value of: 196  
    Yielding Nhits: 17
```



#this will give us all the accession numbers of the 17 hits that matched the protein sequence

```
hits$ pdb.id
```

```
[1] "6S36_A" "1AKE_A" "8BQF_A" "6RZE_A" "4X8M_A" "4X8H_A" "1E4V_A" "3HPR_A"
[9] "5EJE_A" "1E4Y_A" "3X2S_A" "6HAP_A" "6HAM_A" "4K46_A" "4NP6_A" "3GMT_A"
[17] "4PZL_A"
```

But, lets automate this process using a string of code:

```
# Download releated PDB files in a "pdbs" folder
files <- get.pdb(hits$ pdb.id, path="pdbs", split=TRUE, gzip=TRUE)
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6S36.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1AKE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/8BQF.pdb.gz exists. Skipping download
```

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/6RZE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/4X8M.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/4X8H.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/1E4V.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/3HPR.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/5EJE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/1E4Y.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/3X2S.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/6HAP.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/6HAM.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/4K46.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/4NP6.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/3GMT.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/4PZL.pdb.gz exists. Skipping download

	0%
====	6%
=====	12%
=====	18%
=====	24%
=====	29%
=====	35%
=====	41%
=====	47%
=====	53%
=====	59%
=====	65%
=====	71%
=====	76%
=====	82%
=====	88%
=====	94%
=====	100%

```
# Align related PDBs using MSA and putting structures on top of each other.
pdbbs <- pdbaln(files, fit = TRUE, exefile="msa")
```

Reading PDB files:

```
pdbbs/split_chain/6S36_A.pdb
pdbbs/split_chain/1AKE_A.pdb
pdbbs/split_chain/8BQF_A.pdb
pdbbs/split_chain/6RZE_A.pdb
pdbbs/split_chain/4X8M_A.pdb
pdbbs/split_chain/4X8H_A.pdb
pdbbs/split_chain/1E4V_A.pdb
pdbbs/split_chain/3HPR_A.pdb
pdbbs/split_chain/5EJE_A.pdb
pdbbs/split_chain/1E4Y_A.pdb
pdbbs/split_chain/3X2S_A.pdb
pdbbs/split_chain/6HAP_A.pdb
pdbbs/split_chain/6HAM_A.pdb
pdbbs/split_chain/4K46_A.pdb
pdbbs/split_chain/4NP6_A.pdb
pdbbs/split_chain/3GMT_A.pdb
pdbbs/split_chain/4PZL_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.... PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.... PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
....
```

Extracting sequences

```
pdb/seq: 1   name: pdbbs/split_chain/6S36_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 2   name: pdbbs/split_chain/1AKE_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 3   name: pdbbs/split_chain/8BQF_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 4   name: pdbbs/split_chain/6RZE_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 5   name: pdbbs/split_chain/4X8M_A.pdb
```

```

pdb/seq: 6   name: pdbc/split_chain/4X8H_A.pdb
pdb/seq: 7   name: pdbc/split_chain/1E4V_A.pdb
pdb/seq: 8   name: pdbc/split_chain/3HPR_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 9   name: pdbc/split_chain/5EJE_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 10  name: pdbc/split_chain/1E4Y_A.pdb
pdb/seq: 11  name: pdbc/split_chain/3X2S_A.pdb
pdb/seq: 12  name: pdbc/split_chain/6HAP_A.pdb
pdb/seq: 13  name: pdbc/split_chain/6HAM_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 14  name: pdbc/split_chain/4K46_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 15  name: pdbc/split_chain/4NP6_A.pdb
pdb/seq: 16  name: pdbc/split_chain/3GMT_A.pdb
pdb/seq: 17  name: pdbc/split_chain/4PZL_A.pdb

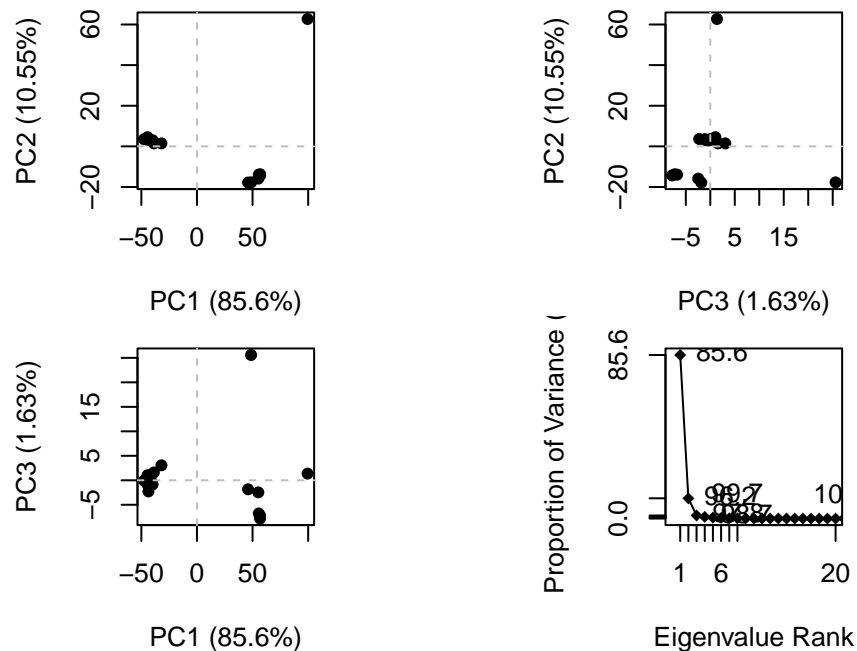
```

## PCA of structures

```

pc.xray <- pca(pdbc)
plot(pc.xray)

```

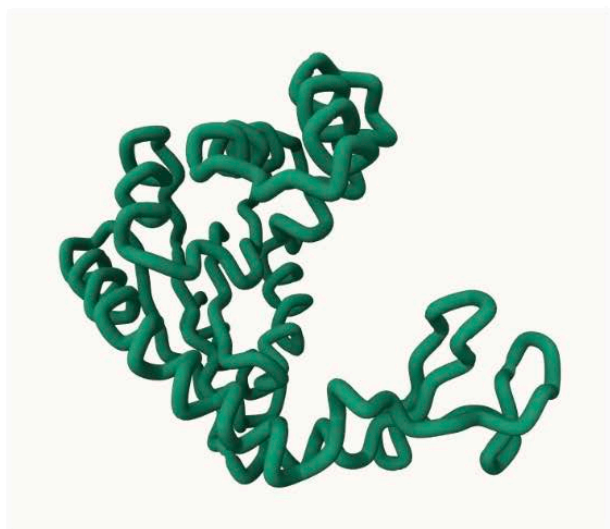


Let's make a trajectory:

```
mktrj(pc.xray, file="pca_movie.pdb")
```

Here is the final image:

![A overlapping figure of the ADK](ADK\_M7.PDB.png)



Note: For the last figure, I added it manually (inserted from screenshot) because NCBI blast was taking very long when rendering the file.