Dynamics of an Endangered Frog Population Recovering from Bd-Driven Decline

4 Abstract

Mountain yellow-legged frogs ($Rana\ sierrae\$ and $R.\ muscosa$) have disappeared from >90% of their historic

6 range due to Batrachochytrium dendrobatidis (Bd) infection, yet a handful of populations now persist and

recover in the pathogen's presence. We studied one such recovering population at Mossy Pond, Tahoe

National Forest, for five summers (2014–2018) using robust-design capture–mark–recapture methods and

⁹ a Bayesian Jolly–Seber model. From 3,631 captures of 657 uniquely marked adults, estimated abundance

varied from 117 to 245 individuals, while 629 new adults recruited over the study. Monthly survival was

strongly seasonal—mean 0.97 in winter versus 0.80 in summer—with male survival exceeding female survival

by 2-7% in each interval. Survey-specific detection probability averaged 0.45. Bd was present in 67% of

skin-swab samples and infection loads remained two-three orders of magnitude below lethal thresholds.

14 Recruitment and abundance declined sharply after severe winters, yet overwinter adult survival showed no

5 snow effect. These findings confirm functional Bd resistance, reveal contrasting seasonal pressures that

shape population dynamics, and identify this robust northern Sierra Nevada population as a prime donor

17 for translocation-based recovery efforts.

18 Introduction

19 Amphibians continue to experience global declines, with infectious diseases recognized as primary drivers of

biodiversity loss (Luedtke et al. 2023). Chytridiomycosis, caused by the fungal pathogen Batrachochytrium

dendrobatidis (Bd), has driven declines or extinctions of approximately 200 amphibian species globally (Sker-

ratt et al. 2007). Amphibian responses to Bd infection vary dramatically; some populations show minimal

impact, others rapidly go extinct, and a rare few exhibit persistent recovery following severe declines (Lips 2016). The mechanisms supporting persistence or recovery remain poorly understood but likely involve interactions among ecological, evolutionary, immunological, and demographic factors (Scheele et al. 2017). The mountain yellow-legged (MYL) frog species complex (Rana sierrae and R. muscosa), endemic to California's Sierra Nevada and Transverse mountain ranges, provides a particularly well-documented case of amphibian decline and recovery from Bd infection. Starting in the 1970s, Bd spread through the Sierra Nevada, causing widespread and rapid extirpations of MYL frog populations (Vredenburg et al. 2010, 2019). By the early 2000s, over 90% of their historical range had been lost, due to combined impacts from Bd and habitat degradation caused by introduced trout (Knapp and Matthews 2000). Despite these losses, a few populations survived initial Bd outbreaks, eventually exhibiting increased abundance and apparent disease resistance (Knapp et al. 2016). These dynamics exemplify patterns of evolutionary rescue (Carlson, Cunningham, and Westley 2014), in which populations rapidly acquire adaptive genetic variation that restores demographic viability under persistent threats (Knapp et al. 2024). While broad-scale studies have described these declines and recoveries, detailed demographic analyses at the individual population level remain scarce, primarily due to logistical challenges. Most fine-scale demographic research has focused on southern populations, particularly within Yosemite National Park (Fellers et al. 2013; Maxwell B. Joseph and Knapp 2018; Knapp et al. 2024). In contrast, the population dynamics of frogs in the northern Sierra Nevada—where habitats are more heterogeneous and populations are smaller and lower in elevation—remain understudied (MYLF ITT 2018) (Brown et al. 2019). To address this gap, we conducted a five-year capture-mark-recapture (CMR) study of a naturally recovering, Bd-persistent R. sierrae population in Tahoe National Forest. Our objectives were to estimate key demographic parameters—abundance, survival, recruitment, and detection probability—to provide insights into the ecological processes and factors that contribute to population viability under endemic Bd infection. We also examined the role of climate variability in shaping these demographic parameters, as climate change is increasingly recognized as a primary driver of amphibian declines worldwide, comparable in impact to disease and habitat loss (Luedtke et al. 2023). Understanding how climate influences abundance, survival, and recruitment is particularly important for high-elevation amphibian populations living with endemic Bd

- 50 infection.
- 51 Understanding demographic patterns in recovering MYL frog populations has significant conservation im-
- 52 plications. Recent research demonstrates these populations can serve as effective founders in reintroduction
- ₅₃ programs, establishing self-sustaining populations when translocated to suitable but currently unoccupied
- habitats (Knapp et al. 2024). Leveraging evolved disease resistance from naturally recovering populations
- offers a promising conservation strategy. Our study provides critical demographic data to support these
- 56 conservation efforts, particularly for the less-studied northern Sierra Nevada populations.

57 Methods

58 Study area and species

- 59 The Mossy Pond study area is a one square mile section of the Tahoe National Forest, north of Highway
- 80 in Nevada County, California. It is characterized by lakes, ponds, and streams set on granite benches,
- ₆₁ ranging from 6,400 feet to 7,100 feet in elevation. The study area includes the namesake Mossy Pond (6
- 62 hectare surface area, maximum depth 2.5 meters), its seasonally flowing outlet stream, and 12 ephemeral
- 63 ponds. The absence of deep-water habitat is unusual for areas supporting MYL frog populations, given their
- reliance on water bodies deeper than 4 meters for overwinter survival (Bradford 1983; Knapp et al. 2003).
- 65 R. sierrae is a federally endangered, mid-sized ranid frog that occupies montane waterbodies in California's
- 66 Sierra Nevada (Fish and Wildlife Service 2014). The US Geological Survey first detected R. sierrae in the
- 67 Mossy Pond area in 1998, and the California Department of Fish and Wildlife (CDFW) has been monitoring
- the population since 2001.

69 Bd status

- To assess the Mossy Pond R. sierrae population's Bd status, we collected epithelial swabs from 24 adult
- ₇₁ frogs over three years (2008, 2010, and 2021). Partner scientists quantified the amount of Bd DNA on each
- swab using real-time quantitative polymerase chain reaction (qPCR) analysis (Boyle et al. 2007). We report
- infection loads as the number of ITS1 (Internal Transcribed Spacer 1) gene copies detected (Longo et al.

⁷⁴ 2013; Maxwell B. Joseph and Knapp 2018). Quantifying Bd loads rather than simply testing for Bd presence ⁷⁵ allowed us to compare our study population with other known recovering populations. Partner scientists ⁷⁶ provided *R. sierrae* Bd load data from three known naturally recovering populations in Yosemite National ⁷⁷ Park (Knapp et al. 2024).

78 Frog population surveys

We studied R. sierrae with capture-mark-recapture (CMR) methods, which are widely used for estimating wildlife population parameters such as abundance, survival, and recruitment (Williams, Nichols, and Conroy 2001). We conducted CMR sampling according to Pollock's robust design (Pollock 1982), which involves sampling at two temporal scales: primary periods (between which the population is open to demographic change) and secondary periods, during which the population is assumed closed. Each summer from 2014 to 2018, we visited the study site 3 or 4 times (primary periods) for three consecutive days of surveys (secondary periods), for a total of 47 individual surveys. Surveys occurred between 0800 and 1800, when R. sierrae are most active. We systematically searched all water bodies in the study area, capturing frogs by hand or dip net for immediate processing. We first scanned each captured frog with a BioMark 601 passive integrated transponder (PIT) tag reader (BioMark, Boise, Idaho, USA) to detect if the frog was marked (i.e., a recapture). We then used calipers to measure the snout to urostyle (SUL) length of each captured frog, and released frogs <40 mm SUL (which are considered subadults) without further processing. Frogs larger than 40mm (adults) required further processing. New adult captures were PIT-tagged dorsally following McAllister et al. (2004). Next, we recorded the frog's sex and weighed it inside a tared plastic bag to the nearest 0.1 g using a Pesola spring scale. Finally, we collected a GPS point (estimated precision error ~3 m) for each capture using a handheld GPS unit. We only collected length and weight data during the first capture event within a three-day primary period. Afterward, if crews captured the same individual on a subsequent day during the same primary period, crews only recorded PIT tag, sex, and location data. Over the 5-year period of our study, we recorded 3,631 captures of 657 individual frogs.

99 Statistical Analysis

We estimated survival, detection, recruitment, and adult population size for the Mossy Pond *R. sierrae*population using a site-specific open-population Jolly–Seber CMR model in a Bayesian framework with R
statistical software (R Core Team, n.d.). Our implementation builds on the mrmr R package (Maxwell B.
Joseph 2019) (https://snarl1.github.io/mrmr/index.html), which tracks a superpopulation of M individuals
through not-recruited, alive, or dead states and models detection and demographic processes across primary
and secondary periods (Knapp et al. 2024). We assessed convergence with trace-plots and Gelman–Rubin
(R) diagnostics and compared model alternatives using leave-one-out cross-validation.

In our model—represents the probability that an individual alive at primary period t survives through to t + 1. Because the interval length Δ (in months) varies, we standardized each—to a per-month rate by taking the Δ th root of every posterior draw, producing monthly survival estimates (with 95% credible bounds) that are directly comparable across short summer and long overwinter intervals.

To test for sex differences in survival, we added a fixed "male" coefficient to the logit-scale survival predictor alongside the baseline intercept and period-specific random effects. Pseudo-individuals were assigned sex by sampling from the observed sex ratio, assuming equal detectability. For each draw and period we computed the female logit (intercept + period effect), added the male coefficient to get the male logit, transformed both via the logistic function, and recorded their difference. We summarize those interval-specific male—female differences by posterior mean, standard deviation, and 95 % credible interval.

To capture unmeasured heterogeneity in detection across survey occasions, we introduced survey-specific random effects on logit-scale detection. This parallels the hierarchical structure used for survival and recruitment in the original mrmr model and acknowledges that factors like temperature, effort, observer, and water clarity can affect detectability even when fixed covariate data are incomplete.

As a final analytical step, we used the posterior samples from our fitted CMR model to test for climate—
demography links. For each posterior draw, we calculated Pearson correlation coefficients between winter
severity (percent of average snow-water equivalent remaining on 1 April) and each demographic parameter—
monthly survival, recruitment, and adult abundance—thereby generating full posterior distributions of those

correlations. To capture potential delayed effects given the frog's multi-year tadpole overwintering, we repeated these analyses using winter severity at lag 0, 1, 2, and 3 years as well as a 3-year rolling average.

This approach quantifies both the strength and uncertainty of climate influences on population dynamics while preserving the hierarchical uncertainty structure of our Bayesian framework.

29 Environmental Covariates

We defined winter severity as the percentage of historical average snow-water equivalent (SWE) remaining
on April 1 each year (100% indicating average severity). We obtained SWE data from ten regional snowsurvey stations (4.5–19.5 km from Mossy Pond) to capture the spatial variability in snowpack present in
topographically complex mountainous environments (Revuelto et al. 2014). Annual SWE values (2011–
2018) were interpolated across the study area using inverse-distance weighting in ArcGIS Pro 3.4.3. From
the resulting continuous SWE raster, we extracted the mean percent SWE within our one-square-mile study
boundary. All raw SWE measurements were obtained from the California Data Exchange Center (CDEC;
http://cdec.water.ca.gov).

138 Results

139 Bd status

16 out of 24 samples tested positive for Bd, with infection intensities ranging from 16 to 43,318 ITS1 gene copies. These Bd loads are well below the ~600,000 gene copy threshold associated with severe chytridiomycosis and increased MYL frog mortality (Vredenburg et al. 2010; Longo et al. 2013). The infection intensities observed in our study population are typical of those measured in other known naturally recovering *R. sierrae* populations [Figure 1; Knapp et al. (2024)].

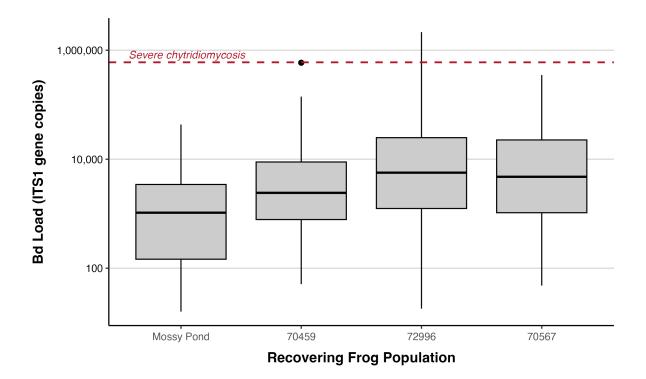


Figure 1: Figure 1. Batrachochytrium dendrobatidis (Bd) infection loads across naturally recovering mountain yellow-legged frog populations. Box plots show the distribution of Bd infection intensity (ITS1 gene copies) for Mossy Pond and three recovering Rana sierrae populations in Yosemite National Park (identified by ID numbers 70459, 72996, and 70567). The dashed red line indicates the approximate threshold (~600,000 gene copies) associated with severe chytridiomycosis and increased mortality in mountain yellow-legged frogs. Note the logarithmic scale on the y-axis. All recovering populations maintain Bd loads well below the mortality threshold, with Mossy Pond showing infection intensities comparable to those observed in the Yosemite populations.

Abundance, Survival, Recruitment, and Detection

146 Abundance

We calculated the abundance of the Mossy Pond *R. sierrae* population for each of the 16 primary survey periods of the study. The lowest abundance was observed in September 2018 (117 individuals, 95% credible interval (95% CrI): 103-132) and a maximum of 245 individuals (95% CrI: 237-256) was observed during the fifth primary period, in July 2015. Across all primary periods, the mean abundance of the population was 168 individuals.

2 Recruitment

A total of 629 frogs recruited into the adult population over the course of the study. Annual recruitment ranged from 60 individuals (95% CrI: 45-78) in 2018 to 166 individuals (95% CrI: 143-188) in 2015.

Survival

Monthly survival during the active (summer) season was markedly lower and more variable than during overwintering. Across 11 summer intervals, mean monthly survival was 0.803 (range: 0.673–0.904), whereas across 4 overwinter intervals, mean monthly survival was 0.966 (range: 0.945–0.974).

In every interval, mean male survival exceeded mean female survival, with differences ranging from 0.015 (95% CrI 0.004-0.029) between the first and second primary periods to 0.069 between the sixth and seventh primary periods (Figure 2). The 95% credible interval for each period lay entirely above zero, indicating consistently higher male survival across both summer and overwinter intervals. These results suggest a modest but statistically robust male survival advantage throughout the study.

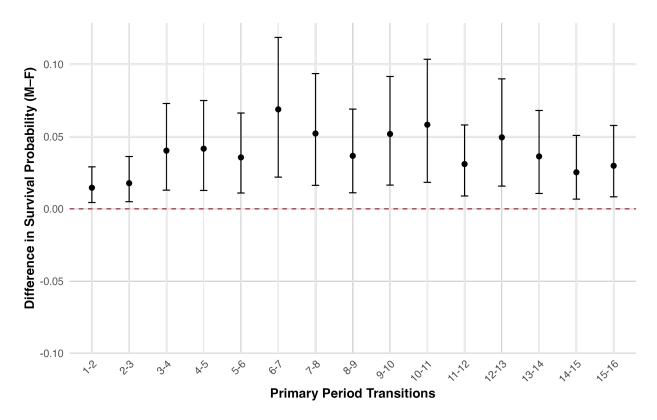


Figure 2: Figure 2. Sex-specific survival differences in the Mossy Pond Rana sierrae population, 2014-2018. Points represent the mean difference in monthly survival probability between males and females (M-F) across 15 primary period transitions. Vertical lines show 95% credible intervals. All intervals are entirely above zero, indicating consistently higher male survival across both summer and overwinter intervals throughout the study period. Overwinter periods occur during the 3-4, 6-7, 9-10, and 12-13 interval transitions.

164 Detection

For each survey (secondary period; n=47) of the study, we calculated the probability that an individual frog that is alive and available for detection is actually detected. Detection probability was lowest (median 0.26, 95% CrI: 0.2-0.31) during the sixteenth survey (09 September, 2015) and highest (median 0.61, 95% CrI: 0.54-0.67) during the forty-third survey (07 August, 2018). The average detection probability over 47 individual surveys was 0.45.

Winter severity effects on population dynamics

Our study period coincided with high variation in winter severity at the Mossy Pond study area, with April 1
SWE ranging from 3.5% of historical average in 2015 to 150% in 2017. Our analyses reveal a strong negative

correlation between winter severity lagged by one year and both the number of frogs recruited into the adult population (median correlation coefficient: -0.92; 95% CrI: -0.79 to -0.98; Figure 3) and the abundance of the population (median correlation coefficient: -0.84, 95% CrI: -0.8 to -0.89). Winter severity showed no notable relationship with overwinter survival probabilities.

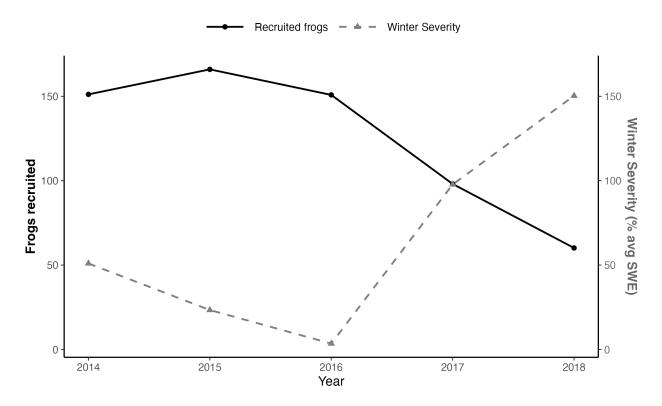


Figure 3: Figure 3: Relationship between adult Rana sierrae recruitment and prior year winter severity at Mossy Pond, 2014-2018. The solid line represents the number of frogs recruited into the adult population each year (left y-axis), while the dashed line shows winter severity as percent of average snowwater equivalent (SWE) measured on April 1 of the prior year (right y-axis). Note the inverse relationship, with high recruitment following mild winters (low SWE) and reduced recruitment following severe winters (high SWE).

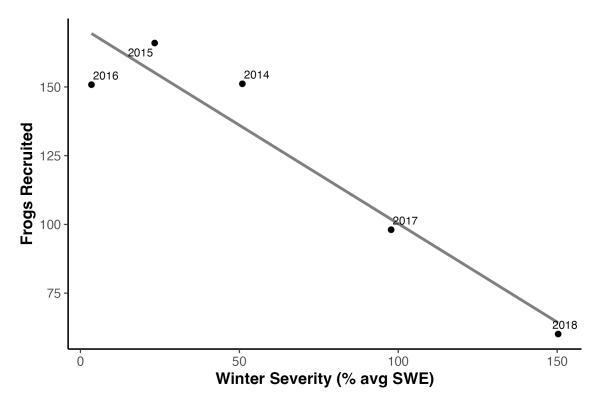


Figure 4: Figure 4. Negative correlation between adult Rana sierrae recruitment and prior year winter severity at Mossy Pond. Scatterplot showing the strong negative relationship (r = -0.92, 95% credible interval: -0.79 to -0.98) between annual adult frog recruitment and the prior year's winter severity (percent of average snow-water equivalent on April 1). Each point is labeled with the recruitment year (2014-2018). The gray line represents the linear regression of the posterior median number of frogs recruited against winter severity, illustrating the strong negative correlation between these variables. Years with mild prior winters (left side of x-axis) yielded substantially higher recruitment compared to years following severe winters (right side of x-axis).

177 Discussion

178 Bd infection dynamics and evidence for resistance

The Mossy Pond frogs carried Bd at moderate prevalence (67%) but at infection intensities that were two
to three orders of magnitude lower than levels associated with increased mortality in MYL frogs. Such low
loads mirror those reported for other naturally recovering populations in Yosemite National Park and stand
in stark contrast to the uniformly high loads (>600,000 ITS1 copies) and near-certain mortality that typify
collapsing populations (Vredenburg et al. 2010). Coupled with the population's stable abundance over the
study period, these patterns are consistent with the idea that some degree of Bd resistance has arisen in this
population. While such dynamics could reflect an evolutionary rescue process in which genotypes conferring

lower susceptibility have increased in frequency, additional genetic and experimental evidence would be required to confirm that mechanism. This expands the geographic scope of Bd-resistant R. sierrae beyond the southern Sierra Nevada, indicating that resistance is not confined to a single environmental setting or genetic lineage.

190 Winter severity as a driver of recruitment and abundance

Our Bayesian correlation analysis revealed a strong negative association between winter severity in the previous year and both adult recruitment and population size. Mild, low-snow winters may enhance juvenile overwinter survival and accelerate growth such that more animals reach the 40 mm adult threshold before the next breeding season. This mechanism has been invoked for long-lived amphibians in other montane systems (McCaffery and Maxell 2010) and is consistent with the lag-1 pattern we observe. The absence of any clear winter-severity signal in overwinter adult survival suggests that once frogs surpass a critical body size, they can tolerate a broad range of cold-season conditions.

198 Seasonal survival patterns

Monthly survival was consistently high during the four overwinter intervals (= 0.97) but dropped and became more variable in summer (= 0.80). Elevated summer mortality is probably multifactorial: frogs are active and conspicuous, water levels drop, and thermal and desiccation stress peak—all of which increase predation risk and physiological costs. Conversely, winter conditions at Mossy Pond may be unusually benign. Although the study area lacks the deep-water refuges (>4m) traditionally thought essential, our capture records show adults concentrated in the rocky outlet stream and mud-bottomed main pond—habitats that can stay unfrozen and oxygenated thanks to possible groundwater inputs and insulating effects.

The pronounced male survival advantage across all intervals ($\Delta = 0.02$ –0.07) contrasts with Fellers et

The pronounced male survival advantage across all intervals ($\Delta = 0.02$ –0.07) contrasts with Fellers et al. (2013), who reported nearly identical annual survival of male and female R. sierrae at Summit Meadow, with females sometimes faring slightly better. Our pattern instead mirrors Rocky Mountain tailed frogs (Ascaphus montanus), where males have shown modestly higher survival (Honeycutt et al. 2019).

Several non-exclusive factors could explain why females at Mossy Pond survived slightly less well. Most

obviously, the energetic burden of reproduction falls almost entirely on females: producing and depositing large gelatinous egg masses—often while bearing the added weight of one or more males in amplexus—depletes energy reserves and can delay post-breeding foraging. Post-oviposition condition declines have been linked to reduced annual survival in other anurans [e.g., common toads; Madsen and Loman (2010)]. Additionally, movement studies of A. montanus show that females travel farther than males to locate suitable oviposition sites, increasing exposure to predators and desiccating microhabitats (Honeycutt et al. 2019). Together, these reproductive and behavioural costs may magnify female summer mortality in the Mossy Pond R. sierrae population, driving the male-biased survival pattern we observed.

219 Climate change context

Across the western United States, April-1 SWE has declined by about 18% on average since 1955, with particularly steep losses in northern California (US EPA 2016). Regional climate projections point to continued
warming, more precipitation falling as rain rather than snow, and further reductions in spring snowpack
over the coming decades (Gottlieb and Mankin 2024). In the short term, such trends could benefit Mossy
Pond frogs by sustaining the sequence of "light" winters that boost recruitment. Over longer horizons,
however, chronic snowpack loss and more frequent summer droughts are likely to lower water levels, shorten
hydroperiods, and shrink the already shallow overwintering habitats available in the basin. The drop in
visual-encounter counts recorded after the 2018–2021 drought hints that these longer-term drying forces
may eventually override the positive recruitment signal.

Implications for conservation management

Our results place the Mossy Pond frogs squarely within the "resistant-donor" framework that Knapp et al.

(2024) used to successfully facilitate landscape-scale *R. sierrae* recovery in Yosemite National Park. In that

15-year experiment, reintroductions sourced from naturally recovering, Bd-resistant *R. sierrae* populations

established self-sustaining colonies at 75% of recipient sites despite the pathogen's continued presence, and

half of those new populations were projected to remain viable over 50 years. The Mossy Pond population's

large size, stable demographics, and demonstrated resistance make it a valuable additional conservation

resource, especially for northern Sierra reintroductions where local adaptation may confer an advantage for long-term population viability.

References

- Boyle, A. D. Hyatt D. G., V. Olsen, D. B. Boyle, L. Berger, D. Obendorf, A. Dalton, K. Kriger, et al.
- 2007. "Diagnostic Assays and Sampling Protocols for the Detection of Batrachochytrium Dendrobatidis."
- Diseases of Aquatic Organisms 73 (3): 175–92. https://doi.org/10.3354/dao073175.
- ²⁴² Bradford, David F. 1983. "Winterkill, Oxygen Relations, and Energy Metabolism of a Submerged Dormant
- Amphibian, Rana Muscosa." Ecology 64 (5): 1171–83. https://doi.org/10.2307/1937827.
- Brown, Cathy, Lucas R. Wilkinson, Kathryn K. Wilkinson, Tate Tunstall, Ryan Foote, Brian D. Todd, and
- Vance T. Vredenburg. 2019. "Demography, Habitat, and Movements of the Sierra Nevada Yellow-Legged
- Frog (Rana Sierrae) in Streams." Copeia 107 (4): 661. https://doi.org/10.1643/ce-19-196.
- ²⁴⁷ Carlson, Stephanie M., Curry J. Cunningham, and Peter A. H. Westley. 2014. "Evolutionary Rescue in a
- Changing World." Trends in Ecology & Evolution 29 (9): 521-30. https://doi.org/10.1016/j.tree.2014.
- 06.005.
- ²⁵⁰ Fellers, Gary M., Patrick M. Kleeman, David A. W. Miller, Brian J. Halstead, and William A. Link. 2013.
- ²⁵¹ "Population Size, Survival, Growth, and Movements of Rana Sierrae." Herpetologica 69 (2): 147–62.
- https://doi.org/10.1655/herpetologica-d-12-00045.
- Fish, U. S., and Wildlife Service. 2014. "Endangered and Threatened Wildlife and Plants; Endangered
- Species Status for Sierra Nevada Yellow-Legged Frog and Northern Distinct Population Segment of the
- Mountain Yellow-Legged Frog, and Threatened Species Status for Yosemite Toad."
- 256 Gottlieb, Alexander R., and Justin S. Mankin. 2024. "Evidence of Human Influence on Northern Hemisphere
- Snow Loss." Nature 625 (7994): 293–300. https://doi.org/10.1038/s41586-023-06794-v.
- ²⁵⁸ Honeycutt, R. Ken, Justin M. Garwood, Winsor H. Lowe, and Blake R. Hossack. 2019. "Spatial Capture—
- recapture Reveals Age- and Sex-Specific Survival and Movement in Stream Amphibians." Oecologia 190
- 260 (4): 821–33. https://doi.org/10.1007/s00442-019-04464-3.
- Joseph, Maxwell B. 2019. "mrmr: mark recapture miscellany in R." https://snarl1.github.io/mrmr/index.

- html.
- ²⁶³ Joseph, Maxwell B., and Roland A. Knapp. 2018. "Disease and Climate Effects on Individuals Drive
- Post-Reintroduction Population Dynamics of an Endangered Amphibian." Ecosphere 9 (11). https:
- //doi.org/10.1002/ecs2.2499.
- ²⁶⁶ Knapp, Roland A., Gary M. Fellers, Patrick M. Kleeman, David A. W. Miller, Vance T. Vredenburg, Erica
- Bree Rosenblum, and Cheryl J. Briggs. 2016. "Large-Scale Recovery of an Endangered Amphibian
- Despite Ongoing Exposure to Multiple Stressors." Proceedings of the National Academy of Sciences 113
- 269 (42): 11889–94. https://doi.org/10.1073/pnas.1600983113.
- 270 Knapp, Roland A., and Kathleen R. Matthews. 2000. "Non-Native Fish Introductions and the Decline of
- the Mountain Yellow-Legged Frog from Within Protected Areas." Conservation Biology 14 (2): 428–38.
- https://doi.org/10.1046/j.1523-1739.2000.99099.x.
- ²⁷³ Knapp, Roland A., Kathleen R. Matthews, Haiganoush K. Preisler, and Robert Jellison. 2003. "DEVEL-
- OPING PROBABILISTIC MODELS TO PREDICT AMPHIBIAN SITE OCCUPANCY IN A PATCHY
- LANDSCAPE." Ecological Applications 13 (4): 1069–82. https://doi.org/10.1890/1051-0761(2003)13%
- 5B1069:dpmtpa%5D2.0.co;2.
- Knapp, Roland A., Mark Q. Wilber, Maxwell B. Joseph, Thomas C. Smith, and Robert L. Grasso. 2024.
- "Reintroduction of Resistant Frogs Facilitates Landscape-Scale Recovery in the Presence of a Lethal
- Fungal Disease." Nature Communications 15 (1). https://doi.org/10.1038/s41467-024-53608-4.
- Lips, Karen R. 2016. "Overview of Chytrid Emergence and Impacts on Amphibians." Philosophical Trans-
- actions of the Royal Society B: Biological Sciences 371 (1709): 20150465. https://doi.org/10.1098/rstb.
- 2015.0465.
- Longo, Ana V., David Rodriguez, Domingos da Silva Leite, Luís Felipe Toledo, Cinthya Mendoza Almer-
- alla, Patricia A. Burrowes, and Kelly R. Zamudio. 2013. "ITS1 Copy Number Varies Among Ba-
- trachochytrium Dendrobatidis Strains: Implications for qPCR Estimates of Infection Intensity from
- Field-Collected Amphibian Skin Swabs." Edited by Tom Coenye. PLoS ONE 8 (3): e59499. https:
- //doi.org/10.1371/journal.pone.0059499.
- Luedtke, Jennifer A., Janice Chanson, Kelsey Neam, Louise Hobin, Adriano O. Maciel, Alessandro Catenazzi,

- Amaël Borzée, et al. 2023. "Ongoing Declines for the World's Amphibians in the Face of Emerging
- 290 Threats." Nature 622 (7982): 308–14. https://doi.org/10.1038/s41586-023-06578-4.
- ²⁹¹ Madsen, Thomas, and Jon Loman. 2010. "Sex Ratio of Breeding Common Toads (Bufo Bufo) Influ-
- ence of Survival and Skipped Breeding." Amphibia-Reptilia 31 (4): 509–24. https://doi.org/10.1163/
- 293 017353710x524705.
- ²⁹⁴ McAllister, Kelly R., James W. Watson, Ken Risenhoover, and Tim McBride. 2004. "MARKING AND
- RADIOTELEMETRY OF OREGON SPOTTED FROGS (RANA PRETIOSA)." Edited by M. J. Adams.
- Northwestern Naturalist 85 (1): 20–25. https://doi.org/10.1898/1051-1733(2004)085%3C0020:maroos%
- 3E2.0.co;2.
- ²⁹⁸ McCaffery, Rebecca M., and Bryce A. Maxell. 2010. "Decreased Winter Severity Increases Viability of
- a Montane Frog Population." Proceedings of the National Academy of Sciences 107 (19): 8644–49.
- 300 https://doi.org/10.1073/pnas.0912945107.
- Pollock, Kenneth H. 1982. "A Capture-Recapture Design Robust to Unequal Probability of Capture." The
- Journal of Wildlife Management 46 (3): 752. https://doi.org/10.2307/3808568.
- R Core Team. n.d. R: A Language and Environment for Statistical Computing. Vienna, Austria: R
- Foundation for Statistical Computing. https://www.R-project.org/.
- Revuelto, J., J. I. López-Moreno, C. Azorin-Molina, and S. M. Vicente-Serrano. 2014. "Topographic Control
- of Snowpack Distribution in a Small Catchment in the Central Spanish Pyrenees: Intra- and Inter-Annual
- persistence." The Cryosphere 8 (5): 1989–2006. https://doi.org/10.5194/tc-8-1989-2014.
- 308 Scheele, Ben C., Lee F. Skerratt, Laura F. Grogan, David A. Hunter, Nick Clemann, Michael McFadden,
- David Newell, et al. 2017. "After the Epidemic: Ongoing Declines, Stabilizations and Recoveries in
- Amphibians Afflicted by Chytridiomycosis." Biological Conservation 206 (February): 37–46. https:
- //doi.org/10.1016/j.biocon.2016.12.010.
- 312 Skerratt, Lee Francis, Lee Berger, Richard Speare, Scott Cashins, Keith Raymond McDonald, Andrea Dawn
- Phillott, Harry Bryan Hines, and Nicole Kenyon. 2007. "Spread of Chytridiomycosis Has Caused the
- Rapid Global Decline and Extinction of Frogs." EcoHealth 4 (2). https://doi.org/10.1007/s10393-007-
- 0093-5.

- US EPA, OAR. 2016. "Climate Change Indicators: Snowpack." https://www.epa.gov/climate-indicators/
- climate-change-indicators-snowpack.
- Vredenburg, Vance T., Roland A. Knapp, Tate S. Tunstall, and Cheryl J. Briggs. 2010. "Dynamics of an
- Emerging Disease Drive Large-Scale Amphibian Population Extinctions." Proceedings of the National
- 320 Academy of Sciences 107 (21): 9689–94. https://doi.org/10.1073/pnas.0914111107.
- ³²¹ Vredenburg, Vance T., Samuel V. G. McNally, Hasan Sulaeman, Helen M. Butler, Tiffany Yap, Michelle S.
- Koo, Dirk S. Schmeller, et al. 2019. "Pathogen Invasion History Elucidates Contemporary Host Pathogen
- Dynamics." Edited by Jake Kerby. PLOS ONE 14 (9): e0219981. https://doi.org/10.1371/journal.pone.
- 0219981.
- ³²⁵ Williams, B. K., J. D. Nichols, and M. J. Conroy. 2001. Analysis and Management of Animal Populations.
- Academic Press, San Diego, CA, USA.