# Species diversity and stand structure as drivers of canopy complexity in southern African woodlands

John L. Godlee

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1 Abstract

Atmospheric CO<sub>2</sub> enrichment and human-induced climate change are expected to drive woody encroachment and an increase in tree cover across African savannas, with consequences for ecosystem function, particularly related to carbon dynamics. The patch dynamics of savanna-woodland mosaics are complex however, as woody growth is mediated by seasonal fire that is itself driven by properties of the woody overtstorey. It is unclear how variation in tree species composition and stand structure in this ecosystem affects woody canopy complexity, and how this might determine future vegetation dynamics. Here, I conducted a study of canopy structure in southern African savannas using terrestrial LiDAR, at sites in Bicuar National Park, Angola and Mtarure Forest Reserve, Tanzania, to explore relationships between tree species diversity, species composition, the spatial distribution of trees, variation in tree size and canopy complexity. I found consistent weak positive effects of species diversity on plot scale canopy complexity metrics related to canopy density, but a negative effect on metrics related to the spatial heterogeneity of the distribution of canopy material. Species diversity caused an increase in canopy height, canopy closure, and within-canopy structural complexity. However, stochasticity in neighbourhood scale woody structure masked species diversity effects at small spatial scales. Finally I found that spatial clustering of trees in space led to a reduction in canopy closure, even within clustered areas, suggesting that disturbance by fire and herbivory not only reduce canopy cover at the landscape scale, but also reduce canopy cover at smaller spatial scales But how does disturbance lead to clustering? Could just say "suggesting"

## $_{\scriptscriptstyle 2}$ 1 Introduction

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Atmospheric CO<sub>2</sub> enrichment, coupled with climate change and changing disturbance regimes, is expected to drive woody encroachment, i.e. proliferation of trees in previously non-wooded 24 areas, and increased growth of trees in currently wooded areas, across the savanna biome over 25 the coming century (Criado et al., 2020; Mitchard & Flintrop, 2013; Stevens et al., 2016). As 26 atmospheric CO<sub>2</sub> concentrations increase, C<sub>3</sub> trees are expected to gain a competitive edge over 27 C<sub>4</sub> grasses due to differences in photosynthetic pathway and carbon use efficiency (Buitenwerf 28 et al., 2012), with cascading effects on canopy cover, grass growth, and disturbance regime (Bond 29 & Midgley, 2012). If realised, woody encroachment and woody densification will have significant effects on the global carbon cycle, as more CO<sub>2</sub> is stored in woody biomass, as well as myriad 31 other effects on ecosystem structure (Donohue et al., 2013). Indeed, tropical savannas have been 32 identified as the fastest increasing component of the terrestrial carbon sink (Sitch et al., 2015). 33 Previous studies however, have reported wide variation in rates of woody encroachment and 34 densification (Mitchard & Flintrop, 2013), particularly in disturbance-prone savannas such as 35 miombo woodlands in southern Africa (Axelsson & Hanan, 2018), and it is unclear how the 36 fertilisation effect of atmospheric CO<sub>2</sub> enrichment will interact with other ecosystem properties 37 to alter vegetation structure (Körner, 2017; Reich et al., 2014). 38

- Savanna vegetation is defined by the coexistence of trees and grasses (Scholes & Archer, 1997).
- 40 In the tropical mesic savannas of southern Africa, disturbance by fire and herbivory are the main
- limitations on tree cover, preventing the competitive exclusion of shade-sensitive C<sub>4</sub> grasses

where climatic conditions would otherwise allow for closed canopy forest (Sankaran et al., 2005). C<sub>4</sub> grasses also provide the main fuel source for seasonal fires in these savannas (Frost, 1996), 43 producing a positive feedback where an increase in tree cover reduces grass fuel load, reducing 44 fire frequency and intensity, increasing tree cover, and so on (Staver & Koerner, 2015). As such, 45 even small perturbations in tree cover can lead to large changes in vegetation structure if critical 46 thresholds of tree cover are crossed (Hirota et al., 2011). Previous research has sought to identify 47 environmental factors which affect tree cover and its responses to atmospheric CO<sub>2</sub> enrichment, 48 but few have considered the functional role of the existing tree community and its effect on 49 ecosystem processes. 50

Canopy structure describes the spatial distribution and density of tree canopy foliage (Lowman & 51 Rinker, 2004). Canopy structural complexity, i.e. the spatial heterogeneity of foliage distribution 52 within the canopy, has been linked to increased net ecosystem productivity (Baldocchi & Wilson, 53 2001; Chen et al., 2012; Gough et al., 2019; Hardiman et al., 2011; Law et al., 2001; Morin, 2015), 54 increased resilience of productivity (Pretzsch, 2014), reduced understorey light penetration (Fotis 55 et al., 2018; Scheuermann et al., 2018), and greater moderation of understorey micro-climate 56 (Wright et al., 2017). Furthermore, in temperate and boreal forests, functional differences among 57 coexisting tree species in their vertical and horizontal canopy occupation provides a link between 58 species diversity, canopy structural complexity and canopy density, with canopy complexity 59 constituting a mechanism for observed positive biodiversity-ecosystem function effects in wooded 60 ecosystems (Barry et al., 2019; Pretzsch, 2014). In tropical savannas, tree species diversity might 61 therefore influence ecosystem-level woody thickening in response to elevated atmospheric CO<sub>2</sub>, where competition effects in diverse tree communities are reducted due to niche separation, and 63 can more effectively increase foliage density and reduce understorey light penetration, excluding 64 grass and thus reducing the probability of disturbance. 65

As well as the species diversity of trees, the spatial distribution and relative size of tree stems, i.e. stand structure, is also expected to affect canopy structural complexity (Stark et al., 2015). 67 Heterogeneity in stem size, whether a result of species diversity, disturbance history or some other 68 factor, is expected to increase canopy complexity and canopy density as individuals of different 69 sizes occupy different parts of the vertical canopy space (Panzou et al., 2020), and may differ in light requirements (Charles-Dominique et al., 2018). Additionally, clustering of individuals in 71 space is expected to increase canopy structural heterogeneity across the wider savanna landscape, 72 but ultimately decrease total foliage density due to an increase in competitive interactions (Dohn 73 et al., 2017). Clustering may occur as a result of disturbance history, facilitation effects among 74 individuals in stressful environments (Ratcliffe et al., 2017), or due to other limitations on 75 establishment arising from growth strategy (). More diverse communities may allow greater stem 76 density and greater foliage density, as differences in canopy occupancy among species reduce 77 negative effects competition among individuals on growth (Gough et al., 2019). 78

Functional differences among floristic types of savanna may also drive variation in canopy 79 complexity, irrespective of species diversity. Some savanna trees form denser canopies than 80 others, as a result of variation in leaf size and branch architecture. Previous studies have 81 compared the branch architecture of ex-Acacia (e.g. Senegalia and Vachellia spp.) and miombo 82 (e.g. Julberardia, Brachystegia, and Isoberlinia) archetypal tree species. While ex-Acacia species 83 tend to inhabit drier, heavily grazed areas, miombo species tend to inhabit dystrophic wetter 84 areas structured heavily by fire (Ribeiro et al., 2020). These studies have shown that ex-Acacia 85 species develop sparser canopies, cagey branch architecture, and wider spreading crowns, while 86 dominant Fabaceae species from the miombo develop thicker, taller canopies, and can grow to 87 large trees (Archibald & Bond, 2003; Mugasha et al., 2013; Privette et al., 2004). Similarly, 88 dominant miombo Fabaceae species from the Detarioideae subfamily have been shown to exhibit 89 wider crowns and grow taller than coexisting species from the Combretaceae family (). Shenkin et al. (2020) showed that Fabaceae tree species from tropical forests exhibit wider and more

voluminous tree crowns than other common families of tropical trees. Under identical stem densities, miombo woodland species may therefore exclude grass more effectively than ex-Acacia or Combretaceae species given these differences in growth form.

Canopy complexity is multi-dimensional and has previously been explained using a plethora 95 of simple metrics that originated in forest and community ecology (Kershaw et al., 2017). 96 Assessments of canopy complexity have most often modelled tree canopies as a series of ellipses 97 (2D), ellipsoids or cones (3D) based on field measurements with measuring tapes (Jucker et al., 98 2015), or used surrogate proxies for 3D canopy structure, due to its inherent complexity (Seidel et 99 al., 2011). Measurements of this kind are time consuming and yet remain an over-simplification of 100 canopy structure. Alternatively, canopy closure is often measured using indirect optical methods 101 which partition sky from canopy material, i.e. with hemispherical photography or the commonly 102 used LAI-2000, providing a 2D representation of the canopy but lacking information on vertical 103 canopy structure (Jonckheere et al., 2004). In recent years, particularly in temperate and boreal 104 forests, LiDAR (Light Detection And Ranging) has emerged as a suitable technology for rapidly 105 and precisely assessing canopy structure in 3D, conserving information on 3D structure of the 106 calibre that is required to understand it's complexities (Calders et al., 2020; Muir et al., 2018). 107 In tropical savannas, very few studies have used terrestrial LiDAR for vegetation analyses, and 108 in southern Africa all existing studies have been located at the Skukuza Flux Tower in Kruger 109 National Park, South Africa (Muumbe et al., 2021). Pioneering work describing the ecology of 110 southern African savannas placed large emphasis on canopy structural diversity as a mediator of 111 ecosystem function (Solbrig et al., 1996), but much of that understanding of savanna vegetation 112 structure was derived from traditional mensuration methods. Using terrestrial LiDAR to measure 113 canopy complexity in southern African savannas therefore offers a unique chance to validate 114 accepted theory and describe differences in ecosystem structure among savanna vegetation types 115 in finer detail than previously possible. 116 In this study I applied terrestrial LiDAR techniques to woodland-savanna mosaics at two sites in 117

In this study I applied terrestrial LiDAR techniques to woodland-savanna mosaics at two sites in southern Africa, with the aim of increasing understanding of how various measures of tree canopy complexity relate to tree diversity and stand structure. I hypothesise that tree neighbourhoods with greater tree species diversity, and greater heterogeneity in stem size allow greater canopy complexity and foliage density. Thus, more diverse savannas might exhibit a higher potential woody biomass, and more effectively increase their growth under elevated atmospheric CO<sub>2</sub>, promoting woody thickening. I also consider the functional differences in canopy architecture among tree communities and how this affects canopy closure and total canopy occupancy.

## <sup>125</sup> 2 Materials and methods

#### 126 2.1 Study sites

Field measurements were conducted at two sites, Bicuar National Park, in southwest Angola (S15.1°, E14.8°), and Mtarure Forest Reserve, in southeast Tanzania (S9.0°, E39.0°) (Figure 1). At each site, 1 ha (100×100 m) plots were located in areas of savanna-woodland vegetation, across a gradient of stem density and a range of savanna floristic archetypes. In Angola, 15 plots were sampled, while in Tanzania, seven were sampled following the curtailment of fieldwork due to COVID-19 travel restrictions. Fieldwork was conducted between February and April at both sites, during the peak growth period of each site in order to capture the maximum foliage volume in the canopy.

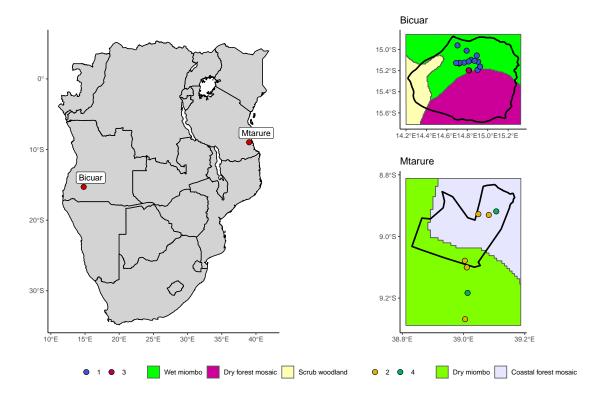


Figure 1: Location of study sites within southern Africa (left), and of 1 ha plots within each site (right). The black outlines in each site map denote the boundaries of protected areas which encompass the majority of study sites, Bicuar National Park in Angola (top), and Mtarure Forest Reserve in Tanzania (bottom). The background of each site map is a re-classified version of White's vegetation map (White, 1983). Points in site maps are shaded according to vegetation type identified by hierarchical clustering of tree genera abundances. Note that all maps are on different scales.

#### 135 2.2 Field measurements

Within each 1 ha plot, each woody stem  $\geq 5$  cm stem diameter was identified to species, the 136 stem Diameter at Breast Height (DBH) was measured at 1.3 m above the ground, and the stem 137 location within the plot was recorded using tape measures. Each 1 ha plot was sampled by nine 138 10 m diameter circular subplots arranged in a regular grid, with a 15 m buffer from the plot edge 139 and 35 m between subplots. For each subplot, the distance and direction from the subplot centre of each stem >5 cm diameter with canopy material inside the subplot was recorded. Within each 141 subplot, a variable number of scans were recorded using a Leica HDS6100 phase-shift Terrestrial 142 Laser Scanner (TLS). The number and position of scans within a subplot was determined by the 143 arrangement of canopy material in the subplot, to minimise shadows within the canopy of the 144 subplot, and to maximise canopy penetration. The number of scans per subplot ranged between 145 one and five across both sites. Extended field methods and data analysis methods are described in Chapter 6. 147

#### 148 2.3 Data analysis

## 149 2.3.1 TLS processing

Point clouds from scans in each subplot were registered and unified using Leica Cyclone (version 150 9.1), using five reflective cross targets visible to all scans as anchor points. Point clouds were 151 voxelised to cubic voxels of different sizes depending on the application of the data. Subplot 152 height profile estimation and gap fraction was conducted using 5 cm<sup>3</sup> voxels, while whole plot 153 canopy rugosity was estimated using 50 cm<sup>3</sup> voxels. Voxels were classified as 'filled' if they 154 intersected one or more points. Variation in voxel size reflects the spatial scale of each analysis, 155 and is bounded by the beam divergence of the scanner over longer distances (Cifuentes et al., 156 2014). Choosing voxels that are too small can result in pock-marked representations of surfaces 157 that are especially problematic when calculating larger scale canopy complexity metrics such as 158 canopy top roughness, while voxels that are too large can result in an over-estimation of plant 159 volume when estimating canopy foliage density at the subplot scale (Cifuentes et al., 2014; Seidel 160 et al., 2012). 161

The noise reduction algorithm from Rusu et al. (2008) was used to discard points based on mean nearest neighbour distances, with a mean number of neighbours of eight, and a standard 163 deviation threshold of 1.96. This effectively removed 'ghost points' produced by partial beam 164 interceptions and also removed many erroneous returns caused by airborne dust particles, which 165 was common at these study sites. Raw points clouds for each subplot had a mean of ~2.9e+08 166 points, ~4.5e+07 points after voxelisation to 5 cm<sup>3</sup>, and ~2.1e+07 points after noise reduction. 167 Ground points were classified using the Progressive Morphological Filter (PMF) from Zhang 168 et al. (2003). Point cloud height was reclassified based on this revised ground layer by measuring 169 the vertical distance between the nearest ground point and each point. Points below 1.3 m 170 height above ground were discarded for calculations of foliage density, canopy cover, and canopy 171 complexity, as points below this threshold where often occupied by long grass. 172

### 2.3.2 Canopy complexity metrics

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Ray-tracing was used to estimate canopy closure in each subplot, i.e. the proportion of the sky hemisphere occluded by plant material at the subplot centre from multiple TLS scans. Hemispherical images were created using the POV-Ray ray-tracing software (Persistence of Vision Pty. Ltd., 2004). Filled voxels were represented as black cubes filling the voxel volume, with a white sky box and no light source. A 'camera' with a 180° fisheye lens was placed at the

subplot centre within POV-Ray, at a height of 1.8 m pointing directly upwards. The images produced by POV-Ray were analysed using Hemiphot (ter Steege, 2018) to estimate canopy closure. Canopy closure estimates from the TLS were validated with hemispherical photographs taken at the same location and processed using the same method in Hemiphot, and compared using Pearson's correlation (r(195)=0.87, p<0.001). A plot level estimate of canopy closure was calculated as the mean of subplot canopy closure measurements. See Chapter 6 for expanded methods and explanation of the behaviour of the different canopy complexity metrics.

Effective Number of Layers (ENL) was calculated according to Ehbrecht et al. (2016) to measure vertical variation in subplot foliage density. ENL is calculated as the exponential Shannon index (i.e. the Hill number of order q=1) of foliage density among 50 cm vertical layers within each subplot:

$$ENL = \exp\left(-\sum_{i=1}^{N} p_i \times \ln p_i\right) \tag{1}$$

Where  $p_i$  is the proportion of filled voxels in the 50 cm layer i, and N is the total number of layers. ENL increases with canopy height and thus number of layers, and also with variation in foliage density among those layers, but not with increased total foliage density.

Total foliage density was calculated within each subplot as the area under the curve of the foliage height profile. Total foliage density was also calculated at the plot level as the sum of filled 50 cm<sup>3</sup> voxels across the plot. Vertical variation in foliage density was calculated by fitting a linear model to the cumulative foliage density profile, then calculating the sum of squared residuals of that model.

Plot level canopy surface models were extracted using the 99th percentile of canopy height in 10 cm<sup>2</sup> columns. A pit-filling algorithm provided by Khosravipour et al. (2014) was applied at 50 cm<sup>2</sup> resolution to reduce the effects of incomplete canopy penetration in dense canopies. Whole plot canopy complexity was measured by two metrics. Canopy top roughness was measured as the coefficient of variation (CV) of canopy height across the plot. Canopy rugosity was measured according to Hardiman et al. (2011), as the CV of vertical and horizontal foliage density within 0.5 m<sup>3</sup> cubic bins.

#### 2.3.3 Stand structure and diversity

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For each subplot, an adapted version of the Iterative Hegyi index was used to estimate crowding, as an alternative to stem density which does not adequately capture crowding at small spatial scales when only a small number of trees are included in the sample (Hegyi, 1974). The CV of 208 stem diameter was calculated as a measure of the heterogeneity of tree size in the neighbourhood. 209 At the plot level, the regularity of species spatial distribution was estimated using the spatial mingling index (von Gadow & Hui, 2002), which scores each tree based on whether it shares 211 species identity with its nearest neighbours. The spatial regularity of trees was estimated using 212 the uniform angle index (winkelmass) (von Gadow & Hui, 2002), which scores each tree based on 213 the angles between nearest neighbours. Additionally, the degree of spatial clustering of trees was 214 measured using Voronoi tessellation, as the CV of Voronoi cell areas (Ong et al., 2012). Finally, plot level tree density was calculated to estimate crowding at the plot scale. See Chapter 6 for 216 more information on the behaviour of the spatial mingling index and uniform angle index. 217 Species diversity at both the subplot and plot level was measured using the exponential Shannon index (i.e. the Hill number of order q=1), calculated using tree species abundance (**Jost2006**). 219

At the subplot level trees were included if they had canopy material inside the 10 m diameter

subplot, while at the plot level trees were included if the largest stem was inside the plot

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#### 2.3.4 Statistical analysis

Non-metric Multi-dimensional Scaling (NMDS) was used to describe variation in species composition among plots, using genus-level basal area weighted abundance in each plot. Stems that
could not be identified to genus were excluded from this analysis, which accounted for 0.2% of
the total basal area recorded. Four distinct vegetation types, two from each site (Table 1), were
identified using hirerarchical clustering of the four dominant NMDS ordination axes. Clusters
were further described using Dufrêne-Legendre indicator species analysis and by ranking tree
species according to abundance across all plots within each cluster.

Linear mixed effects models tested the effects of tree species diversity and stand structural 231 diversity on subplot canopy complexity metrics. Mixed models used a nested random intercept 232 structure to account for the sampling design of subplots within plots and plots within vegetation 233 types. Separate models were fitted for each canopy complexity metric, resulting in four models 234 at the subplot level. Effect sizes among fixed effects in maximal models were compared for each 235 canopy complexity metric, using the 95% confidence interval of the effect size to ascertain whether a fixed effect was significant by whether the confidence interval overlapped zero (Nakagawa & 237 Cuthill, 2007). AIC values and Akaike weights of models with different combinations of fixed 238 effects were compared to determine which combination of diversity and structural metrics best 239 explained variation in each canopy complexity metric. 240

Path analysis was used to test whether tree species diversity influences canopy complexity 241 indirectly through its effect on stand structure, using the piecewiseSEM R package (Lefcheck, 242 2016). Two path analyses were conducted, one at the plot level and one at the subplot level. 243 Subplot path analysis investigated the direct effect of species diversity on canopy closure, as 244 well as the indirect effect of diversity on canopy closure via the CV of basal area, with random intercept terms for each vegetation type. The ex-Acacia vegetation type was represented by only 246 two plots and could not be included in this model due to lack of replication. Plot level path 247 analysis investigated the direct effects of species diversity and spatial mingling of species on 248 mean canopy height, as well as the indirect effects of these metrics on canopy height via tree 249 density and basal area CV. Again, ex-Acacia plots were excluded from this path analysis.

Statistical analysis of the determinants of plot level canopy complexity metrics were conducted using linear models. Again, these models excluded the ex-Acacia vegetation type due to lack of replication. As with the subplot linear mixed models, predictor variable effect sizes were used to assess predictor variable significance, and comparison of candidate models using AIC, Akaike weights, and model R<sup>2</sup> values was used to determine which combination of predictors best explained each canopy complexity metric.

## 257 3 Results

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#### 3.1 Description of vegetation types

Indicator species analysis shows that the four identified vegetation types constitute common southern African savanna floristic archetypes (Table 2). Cluster 1, found in Bicuar National Park contains typical miombo species from the Detarioideae subfamily, such as *Julbernardia paniculata*. Cluster 1 is the most frequent vegetation type in this study, with 12 plots. Cluster 1 has the highest stem density, but lower AGB than Clusters 2 or 3, which contain larger individuals with disproportionately higher biomass. Cluster 2, found in Mtarure Forest Reserve, is dominated by *Pteleopsis myrtifolia*, a common miombo species from the Combretaceae family. Indeed,

Table 1: Description of the vegetation type clusters, identified using the Ward algorithm based on basal area weighted genus abundance. AGB = Above-Ground woody Biomass. Species richness, stem density and AGB are reported as the median among plots, with the interquartile range in parentheses.

Site	Cluster	N sites	Richness	Stem density (Stems ha <sup>-1</sup> )	AGB (t ha <sup>-1</sup> )
Bicuar	1	12	17(2)	642(194)	41( 8.4)
Mtarure	2	5	23(4)	411(137)	72(11.9)
Bicuar	3	3	6(1)	196(55)	77(7.3)
Mtarure	4	2	12(2)	288(73)	9(0.2)

Table 2: Floristic description of the vegetation type clusters. Dominant species are the most abundant individuals across all plots per cluster. Indicator species are derived from Dufrêne-Legendre indicator species analysis with the three highest indicator values.

Cluster	Dominant species	Indicator species	Indicator value
1	Julbernardia paniculata	Strychnos spinosa	0.83
	Burkea africana	Combretum collinum	0.74
	Combretum collinum	Julbernardia paniculata	0.70
2	Diplorhynchus condylocarpon	Pteleopsis myrtifolia	1.00
	Pseudolachnostylis maprouneifolia	Diplorhynchus condylocarpon	0.89
	Gymnosporia senegalensis	Pseudolachnostylis maprouneifolia	0.81
3	Baikiaea plurijuga	Baikiaea plurijuga	0.94
	Baphia massaiensis	Baphia massaiensis	0.83
	Philenoptera nelsii	Philenoptera nelsii	0.45
4	Combretum apiculatum	Vachellia nilotica	0.99
	Burkea africana	Combretum apiculatum	0.70
	Bauhinia petersiana	Senegalia polyacantha	0.62

Cluster 2 also contained other common miombo species shared with plots in Cluster 1, such as 266 Julbernardia globiflora and Pseudolachnostylis maprouneifolia, but these clusters remain distinct 267 due to biogeographic variation in endemic genera at the longitudinal extremes of the miombo 268 ecoregion represented by the two sites in this study. Cluster 3 represents Baikiaea woodland. 269 found on Kalahari sands in southern Angola. It is species poor and dominated by Baikiaea 270 plurijuga which forms large spreading canopy trees with high AGB. Other shrubby species that 271 coppice readily in response to disturbance by fire such as Baphia massaiensis are also common. 272 Cluster 4, found in Mtarure is a type of ex-Acacia woodland, dominated by Vachellia and 273 Senegalia spp. This vegetation type was not well represented in the study, with only two plots, 274 precluding its use in some multi-level statistical analyses due to lack of replication. Cluster 4 275 had far lower AGB than the other clusters (Table 1). 276

Differences in canopy structure among the four vegetation types are evident through observation 277 of canopy surface models for typical plots within each vegetation type (Figure 5), and by 278 comparing canopy complexity metrics (Figure 6). Cluster 1 shows many overlapping crowns 279 forming a nearly contiguous canopy surface, and the heighest plot foliage density of all clusters. Though most trees in Cluster 1 have smaller crowns than those in Cluster 2, which also forms a 281 nearly contiguous canopy. The largest trees in Cluster 2 grow taller and have a wider spreading 282 canopy than those in other vegetation types. Cluster 3 shows two distinct size classes of tree, 283 the large Baikiaea plurijuqa forming clear isolated canopies, and much smaller scattered shrubby 284 individuals in the understorey. Cluster 4 shows many small shrubby individuals with irregular 285 canopy shapes, but a greater total crown area coverage than Cluster 3. 286

## 3.2 Bivariate relationships

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Bivariate plots and linear models show that subplot species diversity, measured as the truenumbers equivalent of the Shannon diversity index of the tree neighbourhood around each 10 m diameter subplot, appears to have weak positive effects on subplot canopy layer diversity, canopy closure and foliage density (Figure 3, Table 3). The Hegyi crowding index had strong positive effects on all canopy complexity metrics, as expected. The effect of Hegyi crowding on subplot canopy complexity metrics was similar across all vegetation types (Table 6). Structural diversity, measured as the CV of subplot stem basal area had significant weak positive effects on total canopy foliage, layer diversity, and canopy closure.

At the plot level, effects of species diversity and stand structure on canopy complexity were similarly weak, but not significant except for the effect on canopy height, which explained more variance in canopy height than tree density (Figure 4, Table 3). The effect of spatial regularity of stems on canopy closure, measured by uniform angle index, was clearly negative, while the effect of spatial clustering of stems, measured by Voronoi cell area CV, was negligible. Additionally, there was a non-significant negative effect of basal area CV on whole canopy rugosity. As expected, tree density had strong positive and significant effects on foliage density and canopy closure, and negative effects on canopy roughness and canopy rugosity. Cluster 4 represented an outlier in plot level bivariate relationships, with low canopy closure, low canopy height, low species diversity, and low variation in stem size.

## 306 3.3 Subplot mixed models

Linear mixed effects models showed that species diversity of the subplot neighbourhood contributed to both layer diversity and canopy closure (Table 4), despite their low R<sup>2</sup> in bivariate linear models, and low effect sizes in maximal linear mixed models (Figure 7). As also seen in the subplot bivariate relationships Figure 3, the Hegyi crowding index had strong positive effects on all measured canopy complexity metrics, though these effects were non-significant for

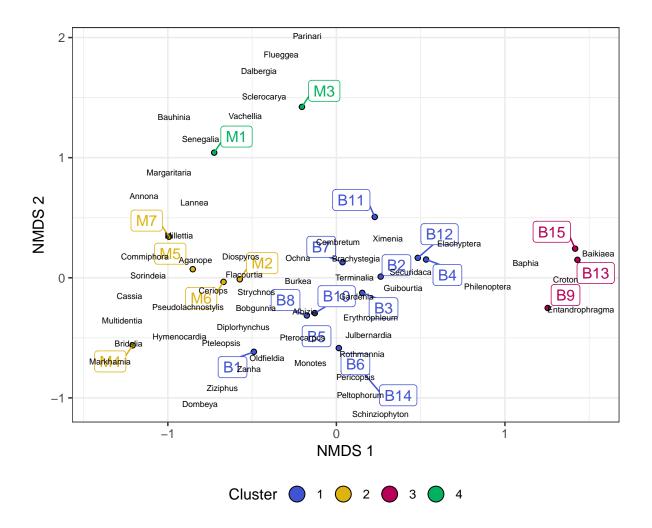


Figure 2: The first two axes of a Non-metric Multi-Dimensional Scaling (NMDS) analysis of tree genus diversity in each plot. Genus scores are labelled as black text, while plot scores are labelled as coloured points. Plots are shaded by vegetation types identified by hierarchical clustering: 1) B1-B8, B10-B12, B14, dominated by core miombo species such as *Julbernardia* spp., *Brachystegia* spp.; 2) M2, M5, M6, and M7, also dominated by core miombo genera with some genera not found in Bicuar National Park such as *Commiphora* and *Sorindeia*; 3) B9, B13 and B15, dominated by *Baikiaea plurijuga*; and 4) M1, M3, and M4, dominated by *Senegalia* spp., *Vachellia* spp., and *Combretum* spp.

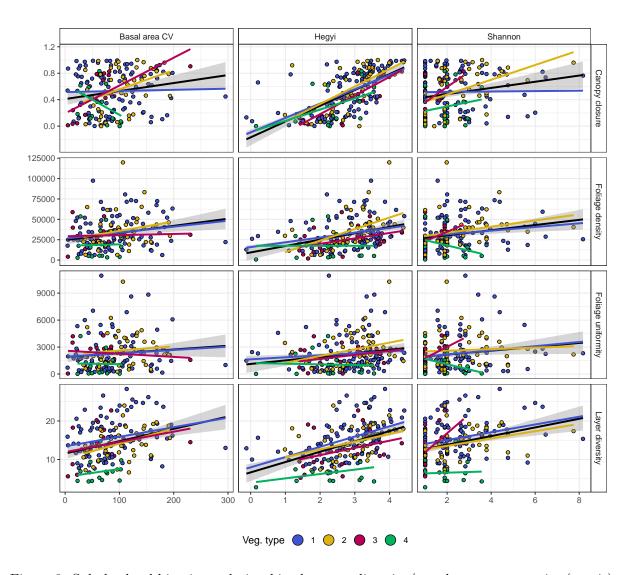


Figure 3: Subplot level bivariate relationships between diversity/stand structure metrics (x axis) and canopy complexity metrics (y axis). Points and linear model lines of best fit are coloured by vegetation type. Black lines of best fit are linear models including all plots, with a 95% confidence interval. See Table 6 for a comparison of linear model fits by vegetation type.

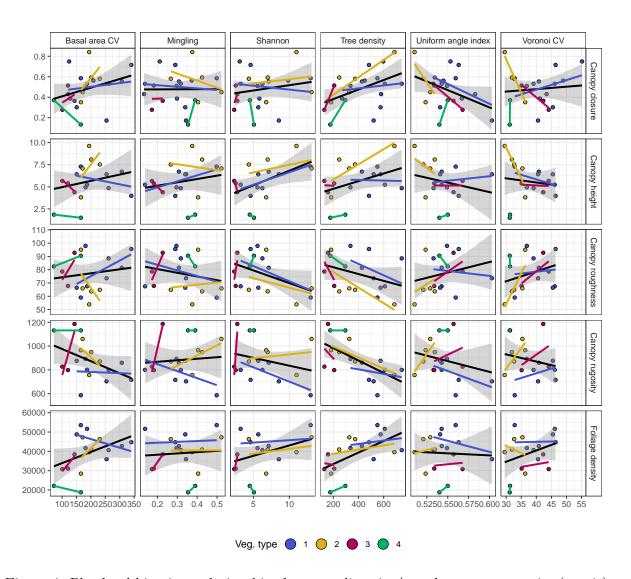


Figure 4: Plot level bivariate relationships between diversity/stand structure metrics (x axis) and canopy complexity metrics (y axis). Points and linear model lines of best fit are coloured by vegetation type. Black lines of best fit are linear models including all plots, with a 95% confidence interval. See Table 6 for a comparison of linear model fits by vegetation type.

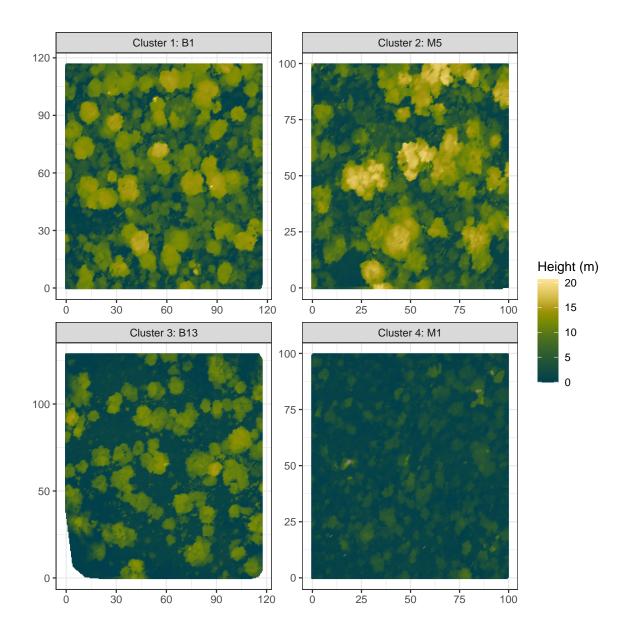


Figure 5: Representative canopy surface models for each vegetation type identified in the Non-metric Multi-dimensional Scaling (NMDS) clustering analysis. Panel titles show the plot name and the vegetation type.

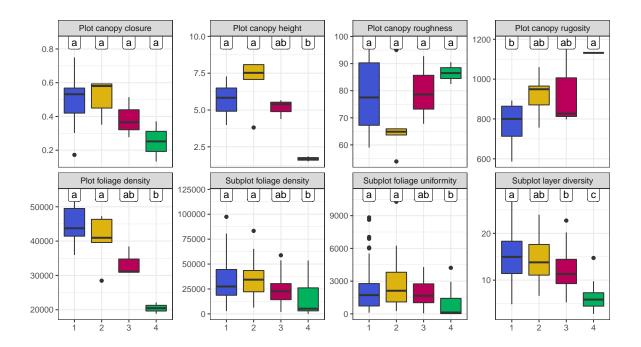


Figure 6: Box plots showing variation in canopy complexity metrics among the four vegetation types identified in the Non-metric Multi-dimensional Scaling (NMDS) clustering analysis. Thick lines show the median, boxes show the interquartile range (IQR), whiskers show  $1.5 \times IQR$ , and points show outliers beyond these limits. Labels above each box plot group vegetation types according to significant differences in pairwise Tukey's tests; vegetation types sharing a letter are not significantly different.

vegetation clusters 3 and 4. Heterogeneity of stem basal area had a significant positive effect on layer diversity and foliage density, but there was wide variation in vegetation type marginal effects for Clusters 3 and 4, due to low levels of replication. Cluster 3 had strong positive effects of species diversity on foliage uniformity and layer diversity. The random effects of vegetation type and plot identity described most of the variation in layer diversity and foliage density. Foliage uniformity was poorly explained by all combinations of fixed effects, with the best model only explaining 29%. All models were better than random effects only models according to AIC values.

#### 3.4 Whole-plot multivariate linear models

While species diversity had varying effects on different plot level canopy complexity metrics, the confidence intervals on these effect sizes were wide (Figure 8). Species diversity had a significant positive effect on canopy height ( $\beta$ =3±0.96, p<0.05), a non-significant positive effect on canopy closure ( $\beta$ =0.07±0.085, p=0.41), but a negative effect on canopy surface roughness ( $\beta$ =-13±6.8, p=0.09) and whole canopy rugosity ( $\beta$ =-111±71, p=0.15). Spatial mingling of tree species had a positive effect on canopy surface roughness and canopy rugosity, but a strong negative effect on canopy height. Plot tree density had negligible effects on canopy complexity, except for canopy rugosity ( $\beta$ =-61±42, p=0.17), in contrast to the effect of Hegyi crowding on subplot canopy complexity. Spatially explicit measures of structural diversity, measured by the uniform angle index, Voronoi cell area CV, and basal area CV, had smaller effects on canopy complexity than species diversity, which were generally insignificant. One exception was the effect of uniform angle index, i.e. the spatial clustering of stems, on canopy closure, which was clearly negative, though still insignificant ( $\beta$ =-0.08±0.043, p=0.1), the effect of Voronoi cell area CV on foliage

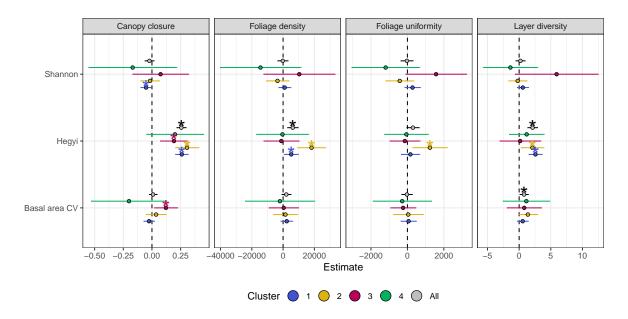


Figure 7: Standardised fixed effect slopes for each model of a canopy complexity metric. Slope estimates where the interval ( $\pm 1$  standard error) does not overlap zero are considered to be significant effects, marked with asterisks. Points are coloured according to vegetation type.

density, which was positive ( $\beta$ =6199±3312, p=0.09), and the effect of basal area CV on canopy closure, which was positive ( $\beta$ =0.06±0.042, p=0.19).

Despite the weak effect sizes of species diverity on canopy complexity at the plot level, model selection showed that foliage density, canopy height and canopy roughness were better explained by models which included species diversity (Table 5). Additionally, the best models for canopy height and canopy roughness also included spatial mingling of tree species. The model for canopy roughness was only maginally better than a null model and the model did not have a significant p-value.

#### 342 3.5 Path analysis

The subplot level path analysis investigating the indirect effect of subplot species diversity on canopy closure via the basal area CV showed that while species diversity had a strong positive significant effect on basal area variation, the effect of basal area variation on canopy closure remained negligible (Figure 9). The indirect effect of species diversity on canopy closure via basal area CV was -0.0016, while the direct effect was -0.078. The R<sup>2</sup> of this model was 0.47. As in the bivariate relationships and plot level linear models, species diversity had a weak positive significant effect on canopy closure, while the major driver of canopy closure was the Hegyi crowding index.

The plot level path analysis, which tested the effects of species diversity and species mingling on canopy height, showed that the main effect of species diversity on canopy height was direct (1.3\*), while the indirect effects via basal area CV (0.0210), and tree density (-0.0294), remained small and insignificant. Shannon diversity had a strong positive effect on tree density. Species mingling had a moderately strong negative but insignificant direct effect on canopy height, as in the linear mixed models.

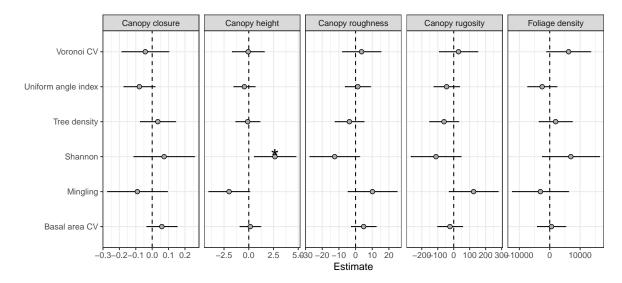


Figure 8: Standardised effect sizes for whole-plot canopy rugosity. Slope estimates where the interval ( $\pm 1$  standard error) does not overlap zero are considered to be significant effects, marked with asterisks.

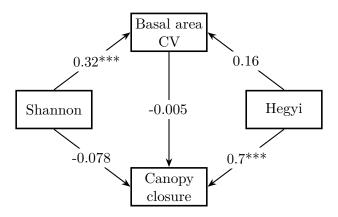


Figure 9: Directed Acyclic Graph showing standardised path coefficients of paths in the path analysis of the indirect effect of subplot species diversity (Shannon diversity index) on canopy closure via basal area CV. Asterisks define p-value thresholds: \*<0.05, \*\*<0.01, \*\*\*<0.001.

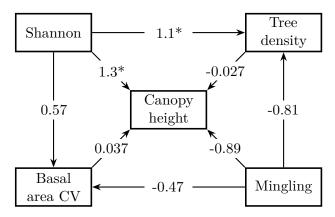


Figure 10: Directed Acyclic Graph showing standardised path coefficients of paths in the path analysis of the indirect effect of plot species diversity (Shannon diversity index) and species mingling on mean canopy height via stand structural metrics of basal area CV and tree density. Asterisks define p-value thresholds: \*<0.05, \*\*<0.01, \*\*\*<0.001.

## 3.6 Covariance of subplot and plot measures of canopy complexity

Plot and subplot canopy complexity metrics were highly correlated in many cases, with similar 358 relationships among vegetation types (Figure 8). Most subplot and plot level canopy metrics 359 covaried in a predictable manner. For example, increased canopy height led to an increase in 360 canopy closure. Plot canopy height especially, tended to be strongly positively correlated with 361 subplot canopy complexity metrics. Additionally, as canopy rugosity increased, many subplot 362 canopy complexity and density metrics decreased. Subplot metrics varied greatly within plots, 363 producing large uncertainty in plot level estimates of these metrics. All subplot level canopy 364 complexity metrics positive correlated with each other (Figure 13). Plot level canopy complexity 365 also generally correlated (Figure 12). Plot level measures of spatial heterogeneity in canopy 366 structure, i.e. canopy surface roughness and canopy rugosity, were negatively correlated with 367 measures of canopy density, i.e. foliage density, canopy closure, and canopy height. Measures of canopy spatial heterogeneity positively correlated with each other, as did measures of canopy 369 density. 370

# 4 Discussion

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This study investigated relationships between tree species diversity, stand structure, and several metrics of tree canopy complexity using terrestrial LiDAR in southern African savannas, with a view to improving understanding of the biotic drivers of variation in canopy complexity and vegetation dynamics. Species diversity appeared to generally have weak positive effects on canopy complexity metrics related to canopy density at both the subplot and plot scales. Plots with greater species diversity produced taller tree canopies, with greater canopy closure and foliage density. Species diversity generally had negative effects on canopy complexity metrics related to the spatial heterogeneity of foliage distribution. Species diversity caused a decrease in canopy surface roughness and canopy rugosity. The study did not, however, find support for the hypothesis that increased heterogeneity in tree stem size causes an increase in canopy complexity.

Path analysis showed that indirect effects of species diversity on canopy complexity via stand 383 structure were small, due to the lack of an effect of stand structure on canopy complexity at both 384 the subplot and plot scale. However, tree species diversity did positively influence structural 385 diversity and tree density. This finding suggests that the effects of species diversity on canopy 386 complexity are due to differences among species in their canopy dimensions and growth strategy, 387 rather than simply covarying with heterogeneity of tree stem size. Previous studies in temperate 388 and boreal forests have suggested that increasing stand structural diversity through active 389 management and thinning can offset productivity losses caused by reduced species diversity 390 (Levick et al., 2009), but this study suggests that in southern African savannas at least, this is 391 not the case. 392

Shannon diversity did have strong positive effects on basal area CV and tree density. This is interpreted as a niche complementarity effect, where diverse savannas are able to support a greater density of trees by reducing the effects of intraspecific competition and

Although basal area CV was included in best models for subplot layer diversity and foliage density, subplot level path analysis suggests that covariance between basal area CV and species diversity, and the strong direct effect of species diversity on canopy complexity, means that any observed effect of basal area CV on subplot canopy complexity is actually due to species diversity rather than basal area CV itself.

The standardised effect sizes of species diversity on canopy complexity metrics were generally greater at the plot level than at the subplot level. While positive and significant relationships

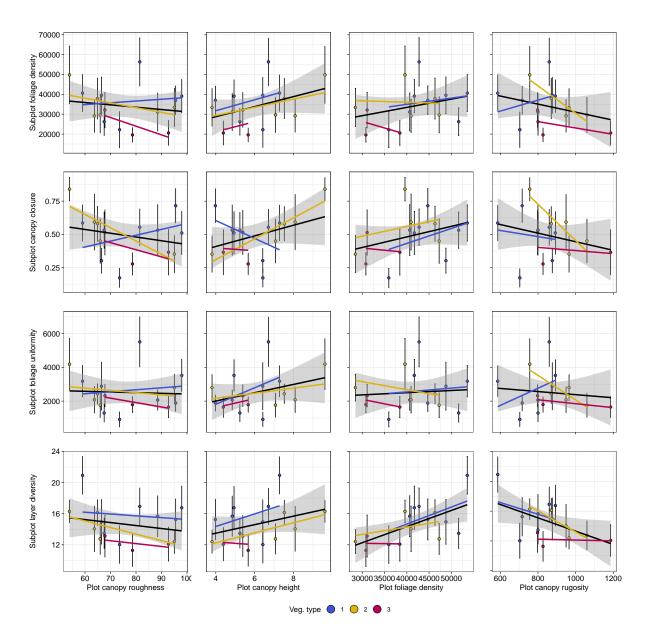


Figure 11: Bivariate plots comparing canopy structural metrics at the plot (x axis) and subplot scale (y axis). Each point represents the mean values of a single plot. Points and linear model fits are coloured according to vegetation type. The black linear model combines all vegetation types. Error bars on points are the standard deviation of mean subplot metrics across the plot. Note that because plot level canopy closure is calculated as the mean of subplot canopy closure, a comparison of subplot and plot canopy closure is not made in this figure.

between species diversity and subplot canopy complexity metrics were observed in the subplot 403 bivariate models, subplot linear mixed effects models did not show strong species diversity 404 effects, and were instead dominated by the effect of crowding. This finding suggests a large 405 degree of stochastic variability in canopy complexity within plots, that masks species effects 406 at smaller spatial scales. The prevalence of disturbance events such as fire and damage by 407 elephants in southern African woodlands, as well as tree-fall, small-scale variability in edaphic 408 factors, and stochastic tree germination all contribute to heterogeneity in canopy complexity 409 (). While disturbances are controlled to some extent by stand structure and composition, due 410 to the stochastic nature of disturbance events, a snapshot study such as this cannot capture 411 the average disturbance regime and there is therefore a great deal of noise in measurements of 412 canopy complexity and stand structure. The contrast in strength of species diversity effects at 413 the subplot and plot level demonstrates the importance of large sample units, a high degree of spatial replication, and ideally a longer time scale when measuring canopy complexity, especially 415 in disturbed systems, to effectively account for the inherent heterogeneity in the system (). 416

In bivariate relationships tree density had a strong effect on canopy complexity metrics at the plot level, but did not feature prominently in the best quality mixed models for those metrics. Tree density covaried with basal area CV, with denser plots also exhibiting greater variation in tree size. This is also reflected in the bivariate relationship between tree density and canopy height. Further path analysis showed that tree species diversity allowed greater tree density and variation in tree size, but this did not translate into a positive effect of tree density or basal area CV on canopy height.

Despite tree density having strong positive observed effects on canopy density, and negative 424 effects on canopy heterogeneity in bivariate relationships, tree density only appeared in the 425 best quality mixed model for canopy rugosity, and the effect sizes for tree density in maximal 426 mixed models were small. Tree density was shown to covary with shannon diversity, basal area 427 CV and Voronoi cell area CV. This covariance may have led to the effect dropping out in the 428 best models. Despite many explanatory variables showing positive relationships with canopy 429 complexity metrics in bivariate plots, in the model selection many dropped out. This is because 430 many explanatory variables covaried in the analysis. 431

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Spatial regularity of stems, measured using the uniform angle index, caused a clear decrease in canopy closure, with similar behaviour across vegetation types. Uniform angle index was also included in the best multivariate model predicting canopy closure. Concurrently, spatial clustering of stems, measured by Voronoi cell area CV was included in the best model for foliage density but had a positive effect on this canopy complexity metric. This finding is expected, as spatial clustering results in reduced canopy cover in areas outside clusters, and a non-compensatory increase in canopy closure within clusters, due to competition among individuals (). In contrast, changes in spatial regularity of trees do not imply changes in the distance of stems, only their layout, but does spatial regularity does covary with spatial clustering. Clustering of trees in savannas can result from positive feedback effects from disturbance by fire and herbivory ().

Stand structural diversity caused positive canopy complexity effects for within-canopy structural metrics such as layer diversity and canopy surface roughness, but had negligible effects on canopy density. This is in line with other studies in forest ecosystems, which report that variation in tree size increases total canopy volume occupancy by increasing the number of canopy layers, but does not necessarily result in a concomitant increase in canopy closure, as the resulting canopies are often more sparse, especially for understorey individuals (). The path analysis also supports this conclusion, where species diversity was found to cause an increase in stand structural diversity, but this did not extend to an increase in canopy closure.

The effect of stand structure on canopy complexity in this system appears to be a result of demographic effects rather than variation in growth form as a function of species diversity. The

path analysis testing the indirect effect of species diversity on canopy closure via stand structural diversity did not find a significant indirect effect of species diversity on canopy closure. While other studies in forests have found a species diversity effect on stand structural diversity (), it is suggested here that prevailing disturbance pressures mask any species diversity effect.

Canopy complexity metrics differed among vegetation types, but significant differences among vegetation types only occurred in a few cases. Canopy density and total foliage volume was lowest in ex-Acacia plots, as expected, while the highest canopy density occurred in miombo plots in clusters 1 and 2. Miombo woodlands frequently contiguous canopies with overlapping individual tree canopies (), while ex-Acacia savannas show greater negative density dependence of individuals, forming patchy canopies with simpler vertical profiles ().

While vegetation types differed in mean values for stand structural and species diversity metrics, variation in these metrics produced results of similar direction and magnitude among vegetation types in most cases. Small sample sizes for *Baikiaea* and ex-Acacia vegetation however, led to wide errors on most relationships especially at the plot level, such that it is impossible to draw deeper conclusions about the behaviour of these vegetation types. Variation in mean values of canopy complexity metrics among vegetation types is likely driven by species identity (), though species composition itself is driven by environmental factors and disturbance regime ().

At the subplot level the Hegyi crowding index was a key driver of canopy complexity. Crowding
of stems at the subplot level correlates with both tree density and spatial clustering of stems at
the plot level. Values of the Hegyi crowding index are expected to increase under greater species
richness due to niche complementarity of canopy layer occupancy.

All subplot canopy complexity metrics positively covaried. For rapid assessments of canopy complexity, simple canopy closure measurements are sufficient to characterise canopy complexity, at the spatial scale of an individual tree neighbourhood. In closed-canopy forest ecosystems, measures of canopy spatial heterogeneity often correlate with canopy closure (). In the savannas studied here however, the sparser and discontinuous canopy results in a negative relationship.

479 **MORE** 

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Table 6: Summary statistics of bivariate linear models comparing canopy complexity metrics with diversity and stand structural metrics, grouped by vegetation type. Note that models plot level canopy complexity metrics could not be fitted for Cluster 4, as this cluster only contained two plots. Slope refers to the slope of the predictor term in the model,  $\pm$  1 standard error. T is the t-value of the slope of the predictor term in the model, Asterisks indicate the p-value of these terms (\*\*\*<0.001, \*\*<0.01, \*<0.05).

Response	Predictor	Cluster	Slope	F	$\mathbb{R}^2$	Τ
		1	$7.3e + 01 \pm 3.7e + 01$	4.0(2,97)	0.04	1.99*
Foliomo donaites	Basal area CV	2	$1.1e+02\pm7.9e+01$	2.1(2,38)	0.05	1.44
Foliage density	Dasai area Uv	3	$1.4e + 01 \pm 7.2e + 01$	0.0(2,14)	0.00	0.20
		4	$1.6e + 01 \pm 2.0e + 02$	0.0(2,12)	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0.08
		1	$5.9e + 03 \pm 2.1e + 03$	8.2(2,102)	0.07	2.86**
Foliage density	Hegyi	2	$1.4e + 04 \pm 3.6e + 03$	15.2(2,40)	0.28	3.90***
ronage density	negyi	3	$6.6e + 03 \pm 3.0e + 03$	4.8(2,23)	0.17	2.18*
		4	$1.5e + 01 \pm 5.5e + 03$	0.0(2,13)	0.00	0.00
		1	$2.2e+03\pm1.3e+03$	2.8(2,102)	0.03	1.67
Foliago dongity	Shannon	2	$3.8e + 03 \pm 2.4e + 03$	2.6(2,39)	0.06	1.61
Foliage density		3	$1.1e+04\pm6.5e+03$	3.1(2,20)	0.13	1.77
		4	$-6.5e + 03 \pm 6.5e + 03$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0.07	-1.01
Canopy closure		1	$1.7e-04\pm6.0e-04$		0.00	
	Basal area CV	2	$2.9e-03\pm1.1e-03$		0.15	2.62*
Canopy closure	Dasar area C v	3	$4.2e-03\pm1.1e-03$	15.1(2,14)	0.52	3.89**
		4	$-4.6e-03\pm3.0e-03$	3.7e+01       4.0(2,97)       0.04         7.9e+01       2.1(2,38)       0.05         7.2e+01       0.0(2,14)       0.00         2.0e+02       0.0(2,12)       0.00         2.1e+03       8.2(2,102)       0.07         3.6e+03       15.2(2,40)       0.28         3.0e+03       4.8(2,23)       0.17         5.5e+03       0.0(2,13)       0.00         1.3e+03       2.8(2,102)       0.03         2.4e+03       2.6(2,39)       0.06         6.5e+03       3.1(2,20)       0.13         6.5e+03       1.0(2,13)       0.07         6.0e-04       0.1(2,97)       0.00         1.1e-03       6.9(2,39)       0.15         1.1e-03       15.1(2,14)       0.52         3.0e-03       2.2(2,12)       0.16         2.8e-02       62.3(2,102)       0.38         5.1e-02       27.0(2,41)       0.40         4.0e-02       50.7(2,23)       0.69         8.0e-02       4.5(2,13)       0.26         2.2e-02       0.0(2,102)       0.00         3.2e-02       12.1(2,40)       0.23	-1.50	
		1	$2.2 \text{e-} 01 \pm 2.8 \text{e-} 02$	62.3(2,102)	0.38	7.89***
Canopy closure	Hegyi	2	$2.6e-01\pm5.1e-02$	27.0(2,41)	0.40	5.19***
Canopy closure	Hegyi	3	$2.8e-01\pm4.0e-02$	50.7(2,23)	0.69	7.12***
		4	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0.26	2.12	
			$3.1 \text{e-} 03 \pm 2.2 \text{e-} 02$	, , ,		
Canopy closure	Shannon		$1.1e-01\pm3.2e-02$	12.1(2,40)	0.23	3.48**
Сапору стояще	Mannon	3	$2.3e-01\pm1.4e-01$	2.9(2,20)	0.13	1.69

		4	$6.7e-02\pm1.1e-01$	0.4(2,13)	0.03	0.60
				. , ,	0.03	0.92
		$\frac{1}{2}$	$3.7e+00\pm4.0e+00$ $4.5e+00\pm7.4e+00$	0.9(2,97)	0.01	0.92 $0.61$
Foliage uniformity	Basal area CV	$\frac{2}{3}$		0.4(2,38)	0.01 $0.02$	-0.59
			$-3.5e+00\pm5.9e+00$	0.4(2,14)		
		4	$-9.3e-01\pm1.5e+01$	0.0(2,12)	0.00	-0.06
		1	$2.2e+02\pm2.3e+02$	1.0(2,102)	0.01	0.98
Foliage uniformity	Hegyi	2	$7.5e + 02 \pm 3.7e + 02$	4.0(2,40)	0.09	2.00
ronage annormity	1108,11	3	$4.5e + 02 \pm 2.6e + 02$	2.9(2,23)	0.11	1.72
		4	$-7.5e + 01 \pm 4.0e + 02$	0.0(2,13)	0.00	-0.19
		1	$2.3e+02\pm1.4e+02$	2.6(2,102)	0.02	1.61
Foliage uniformity	Channon	2	$8.6e + 01 \pm 2.2e + 02$	0.1(2,39)	0.00	0.38
ronage uniformity	Shaimon	3	$1.3e+03\pm5.1e+02$	6.1(2,20)	0.23	2.48*
		4	$-5.9e + 02 \pm 4.7e + 02$	1.6(2,13)	0.11	-1.27
		1	$2.5 \text{e-} 02 \pm 9.3 \text{e-} 03$	7.1(2,97)	0.07	2.66**
T 1: '4	D 1 CV	2	$3.9e-02\pm1.4e-02$	8.0(2,38)	0.17	2.83**
Layer diversity	Basal area CV	3	$2.7e-02\pm2.3e-02$	1.3(2,14)	0.09	1.15
		4	$2.1e-02\pm3.1e-02$	0.5(2,12)	0.04	0.67
		1	$2.7e + 00 \pm 4.9e - 01$	29.1(2,102)	0.22	5.39***
T 1	TT .	2	$2.0e + 00 \pm 7.5e - 01$	7.1(2,40)	0.15	2.66*
Layer diversity	Hegyi	3	$1.9e + 00 \pm 1.0e + 00$	3.6(2,23)	0.13	1.89
		4	$1.1e+00\pm 8.5e-01$	1.8(2,13)	0.12	1.33
		1	$1.0e + 00 \pm 3.4e - 01$	8.7(2,102)	0.08	2.95**
		$\frac{1}{2}$	$9.5e-01\pm4.3e-01$	4.8(2,39)	0.11	2.18*
Layer diversity	Shannon	3	$4.9e + 00 \pm 1.8e + 00$	7.2(2,20)	0.26	2.68*
		4	$1.8e-01\pm1.1e+00$	0.0(2,13)	0.00	0.16
		1	$1.2e-01\pm6.9e-02$	2.9(2,6)	0.33	1.72
		2	$-3.2e-01\pm2.9e-01$	1.2(2,3)	0.29	-1.10
Canopy roughness	Basal area CV	3	$3.5e-01\pm4.7e-01$	0.6(2,1)	0.36	0.74
		4	0.00 01±1.70 01	0.0(2,1)	0.00	0.11
		1	$2.6e-01\pm1.2e+00$	0.0(2,6)	0.01	0.22
		$\frac{1}{2}$	$4.6e + 00 \pm 1.9e + 00$	6.1(2,3)	0.67	2.48
Canopy roughness	Voronoi CV	3	$1.8e + 00 \pm 1.9e + 00$	1.0(2,1)	0.49	0.99
		4	1.00   00±1.00   00	1.0(2,1)	0.40	0.00
		1	$-4.2e+01\pm5.7e+01$	0.5(2,6)	0.08	-0.74
		$\frac{1}{2}$	$1.6e + 01 \pm 9.7e + 01$	$0.0(2,0) \\ 0.0(2,3)$	0.08	-0.74 $0.17$
Canopy roughness	Mingling	$\frac{2}{3}$	$3.5e+02\pm2.5e+02$	2.0(2,1)	0.67	1.42
		4	3.9e+02±2.9e+02	2.0(2,1)	0.07	1.42
			4.20.00   4.5-00	0.0(9.6)	0.19	0.06
		1	$-4.3e-02\pm4.5e-02$ $-5.9e-02\pm3.1e-02$	0.9(2,6) $3.6(2,3)$	$0.13 \\ 0.54$	-0.96 $-1.89$
Canopy roughness	Tree density	$\frac{2}{3}$	$-5.9e-02\pm3.1e-02$ $-1.8e-01\pm2.6e-01$	* ' '	0.54 $0.31$	-1.89 $-0.68$
		3 4	-1.0e-U1±2.0e-U1	0.5(2,1)	0.51	-0.08
			0.9-100117 100	1 7(0.0)	0.00	1 20
		1	$-2.3e+0.0\pm1.7e+0.0$	1.7(2.6)	0.22	-1.32
Canopy roughness	Shannon	2	$-1.4e + 00 \pm 2.4e + 00$	0.4(2,3)	0.11	-0.60
_		$\frac{3}{4}$	$3.4e + 01 \pm 4.7e + 01$	0.5(2,1)	0.34	0.72
		4				
		1	$-7.4e + 01 \pm 2.6e + 02$	0.1(2,6)	0.01	-0.28

Canopy roughness Uniform angle index

		2	$4.1e+02\pm9.5e+02$	0.2(2,3)	0.06	0.43
		3	$4.4e + 02 \pm 5.7e + 02$	0.6(2,1)	0.37	0.76
		4				
		1	$-6.5e-03\pm6.1e-03$	1.1(2,6)	0.16	-1.07
Canopy height	Basal area CV	2	$4.3e-02\pm4.0e-02$	1.2(2,3)	0.28	1.08
Canopy neight	Dasar area e v	3	$-3.1e-02\pm8.7e-03$	12.3(2,1)	0.92	-3.51
		4				
		1	$-1.0e-01\pm8.6e-02$	1.5(2,6)	0.20	-1.21
Canopy height	Voronoi CV	2	$-7.0e-01\pm2.0e-01$	12.7(2,3)	0.81	-3.57*
Canopy neight	voronoi C v	3	$-1.8e-02\pm1.4e-01$	0.0(2,1)	0.02	-0.13
		4	$\begin{array}{cccccccccccccccccccccccccccccccccccc$			
		1	$6.8e + 00 \pm 3.8e + 00$	3.2(2,6)	0.34	1.78
Canopy height	Mingling	2	$-3.3e+00\pm1.3e+01$	0.1(2,3)	0.02	-0.25
Canopy neight	Willighing	3	$-2.3e+01\pm9.3e-01$	619.2(2,1)	1.00	-24.88*
		4				
		1	$-3.5e-04\pm3.8e-03$	0.0(2,6)	0.00	-0.09
Conony bainly	Tron donait	2	$8.6e-03\pm4.0e-03$	4.7(2,3)	0.61	2.16
Canopy height	Tree density	3	$-1.0e-03\pm1.7e-02$	0.0(2,1)	0.00	-0.06
		4				
		1	2.8e-01±1.1e-01	7.1(2,6)	0.54	2.66*
C 1:14	CI	2	$1.7e-01\pm3.3e-01$	0.3(2,3)	0.08	0.52
Canopy height	Shannon	3	$-3.0e + 00 \pm 9.0e - 01$	11.1(2,1)	0.92	-3.32
		4		, , ,		
		1	$1.0e + 01 \pm 2.1e + 01$	0.2(2,6)	0.04	0.49
C 1 1 1 1	TT • C 1 • 1	2		0.3(2,3)	0.09	-0.56
Canopy height	Uniform angle index	3	$6.0e-02\pm3.9e+01$	0.0(2,1)	0.00	0.00
		4		<b>,</b> , ,		
		1	$3.6e-04\pm6.9e-04$	0.3(2,10)	0.03	0.53
C 1	D 1 CW	2	$3.5e-03\pm3.5e-03$	1.0(2,3)	0.24	0.98
Canopy closure	Basal area CV	3	$1.9e-03\pm5.3e-03$	0.1(2,1)	0.11	0.35
		4		, ,		
		1	9.3e-03±8.2e-03	1.3(2,10)	0.11	1.13
C 1		2	$-6.6e-02\pm7.9e-03$	69.7(2,3)	0.96	-8.35**
Canopy closure	Voronoi CV	3	$-2.5e-02\pm4.6e-03$	29.0(2,1)	0.97	-5.39
		4				
		1	-1.6e-01±5.1e-01	0.1(2,10)	0.01	-0.31
C 1	3.f. 1.	$\overline{2}$	$-6.9e-01\pm1.1e+00$	0.4(2,3)	0.12	-0.63
Canopy closure	Mingling	3	$7.6e-02\pm4.1e+00$	0.0(2,1)	0.00	0.02
		4		· · · /		
		1	1.4e-04±4.0e-04	0.1(2,10)	0.01	0.36
C 1	m 1 **	$\overline{2}$	$8.5e-04\pm2.4e-04$	12.2(2,3)	0.80	3.50*
Canopy closure	Tree density	3	$3.0e-03\pm4.3e-06$	499683.9(2,1)	1.00	706.88***
		4		· · · /		
		1	-7.6e-03±1.7e-02	0.2(2,10)	0.02	-0.45
	C)	2	$8.5e-03\pm3.0e-02$	0.1(2,3)	0.03	0.28
Canopy closure	Shannon	3	$1.9e-01\pm5.2e-01$	0.1(2,1)	0.12	0.37
		-	<del> </del>	- (-,-)		

		4				
		1	$-3.9e+00\pm2.3e+00$	2.9(2,10)	0.23	-1.71
Canany alaguna	Uniform angle index	2	$-1.2e + 01 \pm 9.3e + 00$	1.7(2,3)	0.36	-1.30
Canopy closure	Omform angle index	3	$-6.9e + 00 \pm 3.9e - 01$	306.2(2,1)	1.00	-17.50*
		4		, ,		
		1	$-4.5e + 01 \pm 2.9e + 01$	2.3(2,6)	0.28	-1.52
77.14	D 1 CT1	2	$1.5e + 02 \pm 1.4e + 02$	1.1(2,3)	0.27	1.05
Foliage density	Basal area CV	3	$1.8e + 02 \pm 8.9e + 01$	4.2(2,1)	0.81	2.06
		4	1.00   02 100   01	112(2,1)	0.01	
		1	$3.5e + 0.1 \pm 5.0e + 0.02$	0.0(2,6)	0.00	0.07
		2	$-7.7e + 02 \pm 1.5e + 03$	0.3(2,3)	0.08	-0.51
Foliage density	Voronoi CV	3	$2.7e + 02 \pm 8.7e + 02$	0.1(2,1)	0.09	0.31
		4	2	011( <del>=</del> ,1)	0.00	0.01
		1	$4.5e + 03 \pm 2.5e + 04$	0.0(2,6)	0.01	0.18
		2	$8.0e + 0.02 \pm 4.7e + 0.04$	0.0(2,3)	0.00	0.02
Foliage density	Mingling	3	$1.5e + 0.00 \pm $	54.1(2,1)	0.98	7.35
		4	1.00   00 ± 2.00   04	04.1(2,1)	0.50	1.00
		1	$8.8e + 00 \pm 2.0e + 01$	0.2(2,6)	0.03	0.45
		2	$1.1e+01\pm2.1e+01$	0.2(2,3) $0.3(2,3)$	0.08	0.51
Foliage density	Tree density	3	$-1.3e+0.1\pm1.1e+0.2$	0.0(2,3) $0.0(2,1)$	0.01	-0.12
		$\frac{3}{4}$	-1.90   01±1.10   02	0.0(2,1)	0.01	0.12
		1	$2.5e + 02 \pm 8.1e + 02$	0.1(2,6)	0.02	0.31
		2	$5.0e + 02 \pm 0.1e + 02$ $5.0e + 02 \pm 1.2e + 03$	0.2(2,3)	0.02	0.42
Foliage density	Shannon	3	$1.8e + 04 \pm 9.1e + 03$	3.9(2,1)	0.80	1.98
		4	1.06+04±9.16+03	3.9(2,1)	0.00	1.90
		1	$-1.1e + 05 \pm 1.0e + 05$	1.3(2,6)	0.18	-1.15
		2	$1.2e + 05 \pm 4.7e + 05$	0.1(2,3)	0.02	0.25
Foliage density	Uniform angle index	3	$4.3e + 04 \pm 2.5e + 05$	0.1(2,3) $0.0(2,1)$	0.03	0.18
		$\frac{3}{4}$	4.90   0412.90   00	0.0(2,1)	0.05	0.10
		1	-1.0e-01±6.1e-01	0.0(2,6)	0.00	-0.17
		2	$-2.2e+00\pm2.2e+00$	1.1(2,3)	0.26	-1.03
Canopy rugosity	Basal area CV	3	$8.7e + 00 \pm 5.4e + 00$	2.6(2,1)	0.73	1.62
		$\frac{3}{4}$	0.7e+00±3.4e+00	2.0(2,1)	0.15	1.02
		1	$7.9e + 00 \pm 8.2e + 00$	0.9(2,6)	0.13	0.96
		2	$3.5e+00\pm0.2e+00$ $3.5e+01\pm1.3e+01$	6.8(2,3)	0.69	2.61
Canopy rugosity	Voronoi CV	3	$1.8e + 01 \pm 4.2e + 01$	0.2(2,1)	0.05	0.42
		$\frac{3}{4}$	1.06+01±4.26+01	0.2(2,1)	0.10	0.42
		1	$-5.9e + 02 \pm 3.6e + 02$	2.7(2,6)	0.31	-1.63
		2	$8.5e + 02 \pm 5.2e + 02$	2.7(2,3) $2.7(2,3)$	0.31 $0.47$	1.63
Canopy rugosity	Mingling	$\frac{2}{3}$	$7.2e+03\pm1.7e+03$	17.6(2,1)	0.47 $0.95$	4.19
		3 4	1.26+00±1.76+00	11.0(4,1)	0.30	4.13
		1	-1.9e-01±3.4e-01	0.3(2,6)	0.05	-0.56
		2	$-4.6e-01\pm 3.4e-01$	4.9(2,3)	0.62	-2.22
Canopy rugosity	Tree density	3	$-1.2e + 00 \pm 5.4e + 00$	0.0(2,1)	0.02	-2.22 $-0.22$
			-1.46〒UU上り.46 <b>〒</b> UU	0.0(2,1)	0.00	-0.22
		4	0.4 +04+4.0 +04	F 0/0 3)	0.17	0.01
		1	$-2.4e + 01 \pm 1.0e + 01$	5.3(2,6)	0.47	-2.31
Canopy rugosity	Shannon					

		2 3 4	$6.4e+00\pm1.8e+01$ $8.5e+02\pm5.4e+02$	$0.1(2,3) \\ 2.5(2,1)$	0.04 0.71	0.35 1.57
Canopy rugosity	Uniform angle index	1 2 3 4	$-2.6e+03\pm1.6e+03$ $1.0e+04\pm4.1e+03$ $3.4e+03\pm1.2e+04$	2.5(2,6) 6.1(2,3) 0.1(2,1)	0.30 0.67 0.07	-1.58 2.47 0.28

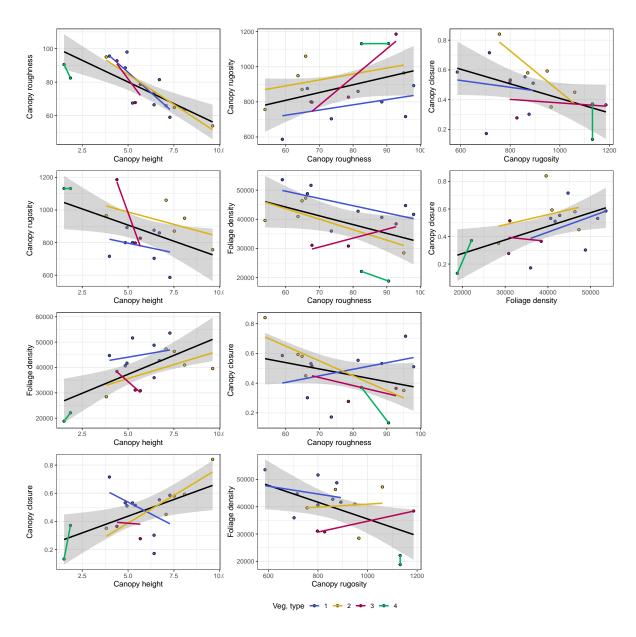


Figure 12: Bivariate scatter plots of plot level canopy complexity metrics.

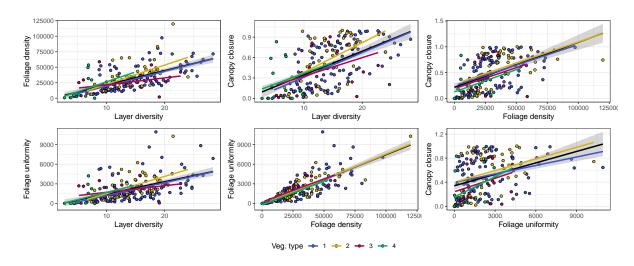


Figure 13: Bivariate scatter plots of subplot level canopy complexity metrics.

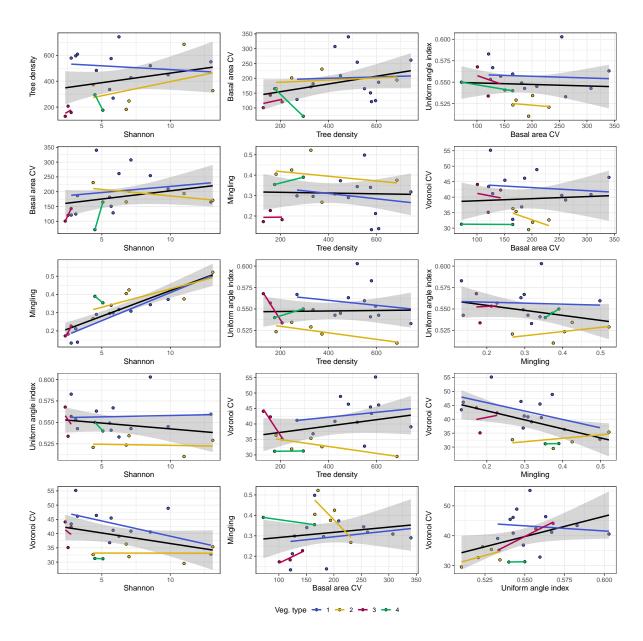


Figure 14: Bivariate scatter plots of plot level diversity and stand structural metrics.

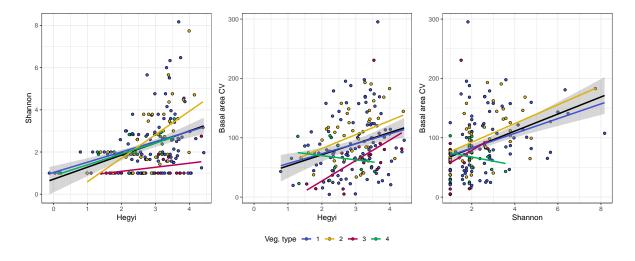


Figure 15: Bivariate scatter plots of subplot level diversity and stand structural metrics.

Table 3: Summary statistics of bivariate linear models comparing canopy complexity metrics with diversity and stand structural metrics across all vegetation types. Slope refers to the slope of the predictor term in the model,  $\pm$  1 standard error. T is the t-value of the slope of the predictor term in the model, Asterisks indicate the p-value of these terms (\*\*\*<0.001, \*\*<0.01, \*<0.05).

Response	Predictor	Slope	F	$\mathbb{R}^2$	Т
	Basal area CV	$8.7e + 01 \pm 3.0e + 01$	8.6(2,167)	0.05	2.93**
Foliage density	Hegyi	$7.8e + 03 \pm 1.6e + 03$	25.5(2,184)	0.12	5.05***
	Shannon	$3.2e+03\pm1.1e+03$	8.9(2,180)	0.05	2.98**
	Basal area CV	$1.2e-03\pm4.8e-04$	6.3(2,168)	0.04	2.52*
Canopy closure	Hegyi	$2.4e-01\pm2.1e-02$	132.8(2,185)	0.42	11.52***
	Shannon	$4.7e-02\pm1.7e-02$	7.3(2,181)	0.04	2.70**
	Basal area CV	$4.1e+00\pm3.0e+00$	1.9(2,167)	0.01	1.37
Foliage uniformity	Hegyi	$4.0e+02\pm1.6e+02$	6.2(2,184)	0.03	2.49*
	Shannon	$2.2e+02\pm1.1e+02$	4.1(2,180)	0.02	2.04*
	Basal area CV	$3.2 \text{e-} 02 \pm 7.6 \text{e-} 03$	17.6(2,167)	0.10	4.20***
Layer diversity	Hegyi	$2.7e+00\pm3.9e-01$	46.8(2,184)	0.20	6.84***
	Shannon	$1.1e+00\pm2.7e-01$	16.8(2,180)	0.09	4.10***
	Basal area CV	$3.0e-02\pm5.0e-02$	0.4(2,16)	0.02	0.60
	Voronoi CV	$7.5 \text{e-}01 \pm 5.9 \text{e-}01$	1.6(2,16)	0.09	1.26
Con ones nou alon ogg	Mingling	$-2.8e+01\pm3.3e+01$	0.7(2,16)	0.04	-0.86
Canopy roughness	Tree density	$-2.6e-02\pm1.7e-02$	2.3(2,16)	0.12	-1.51
	Shannon	$-1.9e + 00 \pm 9.5e - 01$	4.0(2,16)	0.20	-2.01
	Uniform angle index	$1.6e + 02 \pm 1.6e + 02$	1.0(2,16)	0.06	0.98
	Basal area CV	$7.1 \text{e-} 03 \pm 7.3 \text{e-} 03$	0.9(2,16)	0.06	0.97
	Voronoi CV	$-4.7e-02\pm9.1e-02$	0.3(2,16)	0.02	-0.52
Canopy height	Mingling	$3.8e+00\pm4.8e+00$	0.6(2,16)	0.04	0.79
Canopy neight	Tree density	$4.3e-03\pm2.5e-03$	3.1(2,16)	0.16	1.76
	Shannon	$3.3e-01\pm1.3e-01$	6.0(2,16)	0.27	2.45*
	Uniform angle index	$-2.2e+01\pm2.4e+01$	0.8(2,16)	0.05	-0.90
	Basal area CV	$8.5 \text{e-}04 \pm 5.7 \text{e-}04$	2.2(2,20)	0.10	1.50
	Voronoi CV	$2.4e-03\pm5.8e-03$	0.2(2,20)	0.01	0.41
Canopy closure	Mingling	$7.2e-03\pm3.7e-01$	0.0(2,20)	0.00	0.02
Canopy closure	Tree density	$4.7e-04\pm1.9e-04$	6.3(2,20)	0.24	2.50*
	Shannon	$1.0e-02\pm1.2e-02$	0.7(2,20)	0.04	0.86
	Uniform angle index	$-3.4e + 00 \pm 1.7e + 00$	3.9(2,20)	0.16	-1.98
	Basal area CV	$5.8e + 01 \pm 3.2e + 01$	3.3(2,16)	0.17	1.80
	Voronoi CV	$5.8e + 02 \pm 4.1e + 02$	2.1(2,16)	0.11	1.43
Foliage density	Mingling	$6.6e + 03 \pm 2.3e + 04$	0.1(2,16)	0.01	0.29
Tollage delibity	Tree density	$3.0e+01\pm1.0e+01$	8.6(2,16)	0.35	2.93**
	Shannon	$1.1e + 03 \pm 6.9e + 02$	2.5(2,16)	0.13	1.57
	Uniform angle index	$-2.1e+04\pm1.1e+05$	0.0(2,16)	0.00	-0.18
	Basal area CV	$-1.0e + 00 \pm 5.3e - 01$	3.7(2,16)	0.19	-1.92
	Voronoi CV	$-6.0e + 00 \pm 7.0e + 00$	0.7(2,16)	0.04	-0.86
Canopy rugosity	Mingling	$1.3e + 02 \pm 3.8e + 02$	0.1(2,16)	0.01	0.33
Carropy ragosity	Tree density	$-5.2e-01\pm1.7e-01$	10.0(2,16)	0.38	-3.16**
	Shannon	$-1.3e+01\pm1.2e+01$	1.2(2,16)	0.07	-1.11
	Uniform angle index	$-1.8e + 03 \pm 1.9e + 03$	0.9(2,16)	0.06	-0.97

Table 4: Explanatory variables included in the best model for each subplot canopy structure variable.  $\Delta AIC$  shows the difference in model AIC value compared to a null model which included only the random effects of vegetation type and plot. Positive  $\Delta AIC$  values >2 indicate that the model is of better quality than the null model.  $R^2_c$  is the  $R^2$  of the best model, while  $R^2_m$  is the  $R^2$  of the model fixed effects only.

Response	Hegyi	Shannon	Basal area CV	$\Delta { m AIC}$	${ m R^2_{ c}}$	$R^2_{m}$
Layer diversity	✓	✓	<b>√</b>	37.0	0.50	0.17
Foliage density	$\checkmark$		$\checkmark$	47.6	0.27	0.09
Foliage uniformity	$\checkmark$			13.1	0.28	0.02
Canopy closure	$\checkmark$	$\checkmark$		101.9	0.60	0.46

Table 5: Explanatory variables included in the best linear model for each plot-level canopy complexity metric.  $\Delta AIC$  shows the difference in model AIC value compared to a null model. Positive  $\Delta AIC$  values >2 indicate that the model is of better quality than the null model.

Response	Shannon	Tree density	Basal area CV	Mingling	Uniform angle index	Voronoi CV	$\Delta { m AIC}$	$\mathbb{R}^2$	Prob.
Foliage density	<b>√</b>					<b>√</b>	5.8	0.42	< 0.05
Canopy closure			$\checkmark$		$\checkmark$		5.8	0.42	< 0.05
Canopy height	$\checkmark$			$\checkmark$			8.2	0.49	< 0.01
Canopy roughness	$\checkmark$			$\checkmark$			2.5	0.30	0.07
Canopy rugosity		$\checkmark$			$\checkmark$		6.9	0.45	< 0.05