Species diversity and stand structure as drivers of canopy complexity in southern African woodlands

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Abstract

Atmospheric CO₂ enrichment and human-induced climate change are expected to drive woody encroachment and an increase in tree cover across African savannas, with consequences for ecosystem function, particularly related to carbon dynamics. The patch dynamics of savanna-woodland mosaics are complex however, as woody growth is mediated by seasonal fire that is itself driven by properties of the woody overtstorey. It is unclear how variation in tree species composition and stand structure in this ecosystem affects woody canopy structure, and how this might determine future vegetation dynamics. Here, I conducted a study of canopy structure in southern African savannas using terrestrial LiDAR, at sites in Bicuar National Park, Angola and Mtarure Forest Reserve, Tanzania, to explore relationships between tree species diversity, species composition, canopy structure, and the spatial distribution of trees. I found consistent weak positive effects of species diversity on plot scale canopy complexity. Species diversity caused an increase in canopy height, canopy closure, and within-canopy structural complexity. However, stochasticity in neighbourhood scale woody structure masked species diversity effects at small spatial scales. Finally I found that spatial clustering of trees in space led to a reduction in canopy closure, even within clustered areas, suggesting that disturbance by fire and herbivory not only reduce canopy cover at the landscape scale, but also reduce canopy cover at smaller spatial scales But how does disturbance lead to clustering? Could just say "suggesting"

20 1 Introduction

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Atmospheric CO₂ enrichment, coupled with climate change and changing disturbance regimes, 21 is expected to drive woody encroachment, i.e. proliferation of trees in previously non-wooded 22 areas, and increased growth of trees in currently wooded areas, across the savanna biome over 23 the coming century (Criado et al., 2020; Mitchard & Flintrop, 2013; Stevens et al., 2016). As 24 atmospheric CO₂ concentrations increase, C₃ trees are expected to gain a competitive edge over 25 C₄ grasses due to differences in photosynthetic pathway and carbon use efficiency (Buitenwerf 26 et al., 2012), with cascading effects on canopy cover, grass growth, and disturbance regime (Bond 27 & Midgley, 2012). If realised, woody encroachment and woody densification will have significant 28 effects on the global carbon cycle, as more CO₂ is stored in woody biomass, as well as myriad 29 other effects on ecosystem structure (Donohue et al., 2013). Indeed, tropical savannas have been 30 identified as the fastest increasing component of the terrestrial carbon sink (Sitch et al., 2015). 31 Previous studies however, have reported wide variation in rates of woody encroachment and 32 densification (Mitchard & Flintrop, 2013), particularly in disturbance-prone savannas such as 33 miombo woodlands in southern Africa (Axelsson & Hanan, 2018), and it is unclear how the 34 35 fertilisation effect of atmospheric CO₂ enrichment will interact with other ecosystem properties to alter vegetation structure (Körner, 2017; Reich et al., 2014). 36 Savanna vegetation is defined by the coexistence of trees and grasses (Scholes & Archer, 1997). 37 In the tropical mesic savannas of southern Africa, disturbance by fire and herbivory are the main 38 limitations on tree cover, preventing the competitive exclusion of shade-sensitive C₄ grasses

where climatic conditions would otherwise allow for closed canopy forest (Sankaran et al., 2005). C₄ grasses also provide the main fuel source for seasonal fires in these savannas (Frost, 1996),

producing a positive feedback where an increase in tree cover reduces grass fuel load, reducing fire frequency and intensity, increasing tree cover, and so on (Staver & Koerner, 2015). As such, even small perturbations in tree cover can lead to large changes in vegetation structure if critical thresholds of tree cover are crossed (Hirota et al., 2011). Previous research has sought to identify environmental factors which affect tree cover and its responses to atmospheric CO₂ enrichment, but few have considered the functional role of the existing tree community and its effect on ecosystem processes.

Canopy structure describes the spatial distribution and density of tree canopy foliage (Lowman & 49 Rinker, 2004). Canopy structural complexity, i.e. the spatial heterogeneity of foliage distribution within the canopy, has been linked to increased net ecosystem productivity (Baldocchi & Wilson, 51 2001; Chen et al., 2012; Gough et al., 2019; Hardiman et al., 2011; Law et al., 2001; Morin, 2015), 52 increased resilience of productivity (Pretzsch, 2014), reduced understorey light penetration (Fotis 53 et al., 2018; Scheuermann et al., 2018), and greater moderation of understorey micro-climate 54 (Wright et al., 2017). Furthermore, in temperate and boreal forests, functional differences among 55 coexisting tree species in their vertical and horizontal canopy occupation provides a link between 56 species diversity, canopy structural complexity and canopy density, with canopy complexity 57 constituting a mechanism for observed positive biodiversity-ecosystem function effects in wooded 58 ecosystems (Barry et al., 2019; Pretzsch, 2014). In tropical savannas, tree species diversity might 59 therefore influence ecosystem-level woody thickening in response to elevated atmospheric CO₂, 60 where competition effects in diverse tree communities are reducted due to niche separation, and 61 can more effectively increase foliage density and reduce understorey light penetration, excluding grass and thus reducing the probability of disturbance. 63

As well as the species diversity of trees, the spatial distribution and relative size of tree stems, 64 i.e. stand structure, is also expected to affect canopy structural complexity (Stark et al., 2015). Heterogeneity in stem size, whether a result of species diversity, disturbance history or some other 66 factor, is expected to increase canopy complexity and canopy density as individuals of different 67 sizes occupy different parts of the vertical canopy space (Panzou et al., 2020), and may differ in 68 light requirements (Charles-Dominique et al., 2018). Additionally, clustering of individuals in 69 space is expected to increase canopy structural heterogeneity across the wider savanna landscape, but ultimately decrease total foliage density due to an increase in competitive interactions (Dohn 71 et al., 2017). Clustering may occur as a result of disturbance history, facilitation effects among 72 individuals in stressful environments (Ratcliffe et al., 2017), or due to other limitations on 73 establishment arising from growth strategy (). More diverse communities may allow greater stem 74 density and greater foliage density, as differences in canopy occupancy among species reduce 75 negative effects competition among individuals on growth (Gough et al., 2019). 76

Functional differences among floristic types of savanna may also drive variation in canopy 77 complexity, irrespective of species diversity. Some savanna trees form denser canopies than 78 others, as a result of variation in leaf size and branch architecture. Previous studies have 79 compared the branch architecture of ex-Acacia (e.g. Senegalia and Vachellia spp.) and miombo 80 (e.g. Julberardia, Brachystegia, and Isoberlinia) archetypal tree species. While ex-Acacia species 81 tend to inhabit drier, heavily grazed areas, miombo species tend to inhabit dystrophic wetter 82 areas structured heavily by fire (Ribeiro et al., 2020). These studies have shown that ex-Acacia 83 species develop sparser canopies, cagey branch architecture, and wider spreading crowns, while 84 dominant Fabaceae species from the miombo develop thicker, taller canopies, and can grow to 85 large trees (Archibald & Bond, 2003; Mugasha et al., 2013; Privette et al., 2004). Similarly, 86 dominant miombo Fabaceae species from the Detarioideae subfamily have been shown to exhibit 87 wider crowns and grow taller than coexisting species from the Combretaceae family (). Shenkin 88 et al. (2020) showed that Fabaceae tree species from tropical forests exhibit wider and more 89 voluminous tree crowns than other common families of tropical trees. Under identical stem densities, miombo woodland species may therefore exclude grass more effectively than ex-Acacia

or Combretaceae species given these differences in growth form.

Canopy complexity is multi-dimensional and has previously been explained using a plethora 93 of simple metrics that originated in forest and community ecology (Kershaw et al., 2017). 94 Assessments of canopy complexity have most often modelled tree canopies as a series of ellipses 95 (2D), ellipsoids or cones (3D) based on field measurements with measuring tapes (Jucker et al., 96 2015), or used surrogate proxies for 3D canopy structure, due to its inherent complexity (Seidel et 97 al., 2011). Measurements of this kind are time consuming and yet remain an over-simplification of 98 canopy structure. Alternatively, canopy closure is often measured using indirect optical methods 99 which partition sky from canopy material, i.e. with hemispherical photography or the commonly 100 used LAI-2000, providing a 2D representation of the canopy but lacking information on vertical 101 canopy structure (Jonckheere et al., 2004). In recent years, particularly in temperate and boreal 102 forests, LiDAR (Light Detection And Ranging) has emerged as a suitable technology for rapidly 103 and precisely assessing canopy structure in 3D, conserving information on 3D structure of the 104 calibre that is required to understand it's complexities (Calders et al., 2020; Muir et al., 2018). 105 In tropical savannas, very few studies have used terrestrial LiDAR for vegetation analyses, and 106 in southern Africa all existing studies have been located at the Skukuza Flux Tower in Kruger 107 National Park, South Africa (Muumbe et al., 2021). Pioneering work describing the ecology of 108 southern African savannas placed large emphasis on canopy structural diversity as a mediator of 109 ecosystem function (Solbrig et al., 1996), but much of that understanding of savanna vegetation 110 structure was derived from traditional mensuration methods. Using terrestrial LiDAR to measure 111 canopy complexity in southern African savannas therefore offers a unique chance to validate 112 accepted theory and describe differences in ecosystem structure among savanna vegetation types 113 in finer detail than previously possible. 114

In this study I applied terrestrial LiDAR techniques to woodland-savanna mosaics at two 115 sites in southern Africa, with the aim of increasing understanding of how various measures of 116 tree canopy complexity relate to tree diversity and stand structure. I hypothesise that tree 117 neighbourhoods with greater tree species diversity, and greater heterogeneity in stem size may 118 allow greater canopy complexity and foliage density. Thus, more diverse savannas might exhibit 119 a higher potential woody biomass, and more effectively increase their growth under elevated 120 atmospheric CO₂, promoting woody thickening. I also consider the functional differences in 121 canopy architecture among tree communities and how this affects canopy closure and total 122 canopy occupancy. 123

¹²⁴ 2 Materials and methods

125 2.1 Study sites

Field measurements were conducted at two sites, Bicuar National Park, in southwest Angola 126 (S15.1°, E14.8°), and Mtarure Forest Reserve, in southeast Tanzania (S9.0°, E39.0°) (Figure 1) 127 At each site, 1 ha (100×100 m) plots were located in areas of savanna-woodland vegetation, 128 across a gradient of stem density and a range of savanna floristic archetypes. In Angola, 15 plots 129 were sampled, while in Tanzania, seven were sampled following the curtailment of fieldwork 130 due to COVID-19 travel restrictions. Fieldwork was conducted between February and April at 131 both sites, during the peak growth period of each site in order to capture the maximum foliage 132 volume in the canopy. 133

2.2 Field measurements

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Within each 1 ha plot, each woody stem ≥ 5 cm stem diameter was identified to species, the stem Diameter at Breast Height (DBH) was measured at 1.3 m above the ground, and the stem

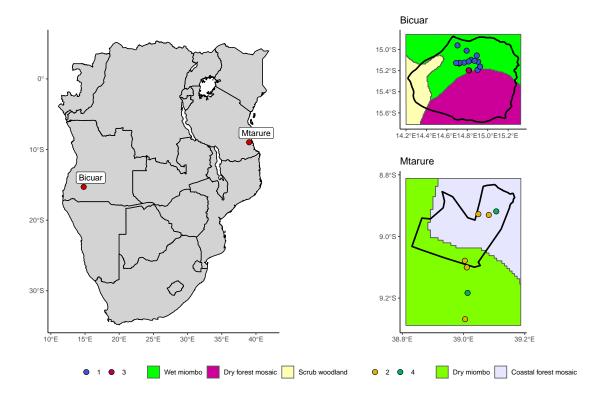


Figure 1: Location of study sites within southern Africa (left), and of 1 ha plots within each site (right). The black outlines in each site map denote the boundaries of protected areas which encompass the majority of study sites, Bicuar National Park in Angola (top), and Mtarure Forest Reserve in Tanzania (bottom). The background of each site map is a re-classified version of White's vegetation map (White, 1983). Points in site maps are shaded according to vegetation type identified by hierarchical clustering of tree genera abundances. Note that all maps are on different scales.

location within the plot was recorded using tape measures. Each 1 ha plot was sampled by nine 137 10 m diameter circular subplots arranged in a regular grid, with a 15 m buffer from the plot edge 138 and 35 m between subplots. For each subplot, the distance and direction from the subplot centre 139 of each stem >5 cm diameter with canopy material inside the subplot was recorded. Within each 140 subplot, a variable number of scans were recorded using a Leica HDS6100 phase-shift Terrestrial 141 Laser Scanner (TLS). The number and position of scans within a subplot was determined by the arrangement of canopy material in the subplot, to minimise shadows within the canopy of the 143 subplot, and to maximise canopy penetration. The number of scans per subplot ranged between 144 one and five across both sites. Extended field methods and data analysis methods are described 145 in Chapter 6. 146

147 2.3 Data analysis

\sim 2.3.1 TLS processing

Point clouds from scans in each subplot were registered and unified using Leica Cyclone (version 149 9.1), using five reflective cross targets visible to all scans as anchor points. Point clouds were 150 voxelised to cubic voxels of different sizes depending on the application of the data. Subplot 151 height profile estimation and gap fraction was conducted using 5 cm³ voxels, while whole plot 152 canopy rugosity was estimated using 50 cm³ voxels. Voxels were classified as 'filled' if they 153 intersected one or more points. Variation in voxel size reflects the spatial scale of each analysis, 154 and is bounded by the beam divergence of the scanner over longer distances (Cifuentes et al., 155 2014). Choosing voxels that are too small can result in pock-marked representations of surfaces 156 that are especially problematic when calculating larger scale canopy complexity metrics such as 157 canopy top roughness, while voxels that are too large can result in an over-estimation of plant 158 volume when estimating canopy foliage density at the subplot scale (Cifuentes et al., 2014; Seidel 159 et al., 2012). 160

The noise reduction algorithm from Rusu et al. (2008) was used to discard points based on 161 mean nearest neighbour distances, with a mean number of neighbours of eight, and a standard 162 deviation threshold of 1.96. This effectively removed 'ghost points' produced by partial beam 163 interceptions and also removed many erroneous returns caused by airborne dust particles, which was common at these study sites. Raw points clouds for each subplot had a mean of ~2.9e+08 165 points, $\sim 4.5e + 07$ points after voxelisation to 5 cm³, and $\sim 2.1e + 07$ points after noise reduction. 166 Ground points were classified using the Progressive Morphological Filter (PMF) from Zhang et al. (2003). Point cloud height was reclassified based on this revised ground layer by measuring 168 the vertical distance between the nearest ground point and each point. Points below 1.3 m 169 height above ground were discarded for calculations of foliage density, canopy cover, and canopy 170 complexity, as points below this threshold where often occupied by long grass. 171

2.3.2 Canopy complexity metrics

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Ray-tracing was used to estimate canopy closure in each subplot, i.e. the proportion of the 173 sky hemisphere occluded by plant material at the subplot centre from multiple TLS scans. 174 Hemispherical images were created using the POV-Ray ray-tracing software (Persistence of 175 Vision Pty. Ltd., 2004). Filled voxels were represented as black cubes filling the voxel volume, 176 with a white sky box and no light source. A 'camera' with a 180° fisheye lens was placed at the 177 subplot centre within POV-Ray, at a height of 1.8 m pointing directly upwards. The images produced by POV-Ray were analysed using Hemiphot (ter Steege, 2018) to estimate canopy 179 closure. Canopy closure estimates from the TLS were validated with hemispherical photographs 180 taken at the same location and processed using the same method in Hemiphot, and compared 181

using Pearson's correlation (r(195)=0.87, p<0.001). A plot level estimate of canopy closure was calculated as the mean of subplot canopy closure measurements.

Effective Number of Layers (ENL) was calculated according to Ehbrecht et al. (2016) to measure vertical variation in subplot foliage density. ENL is calculated as the exponential Shannon index (i.e. the Hill number of order q=1) of foliage density among 50 cm vertical layers within each subplot:

$$ENL = \exp\left(-\sum_{i=1}^{N} p_i \times \ln p_i\right)$$
 (1)

Where p_i is the proportion of filled voxels in the 50 cm layer i, and N is the total number of layers.

Total foliage density was calculated within each subplot as the area under the curve of the foliage height profile. Total foliage density was also calculated at the plot level as the sum of filled 50 cm³ voxels across the plot.

Plot level canopy surface models were extracted using the 99th percentile of canopy height in 10 cm² columns. A pit-filling algorithm provided by Khosravipour et al. (2014) was applied at 50 cm² resolution to reduce the effects of incomplete canopy penetration in dense canopies. Whole plot canopy complexity was measured by two metrics. Canopy top roughness was measured as the coefficient of variation of canopy height across the plot. Canopy rugosity was measured according to Hardiman et al. (2011), as the coefficient of variation of vertical and horizontal foliage density within 0.5 m³ cubic bins.

2.3.3 Stand structure and diversity

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For each subplot, an adapted version of the Iterative Hegyi index was used to estimate crowding, as an alternative to stem density which does not adequately capture crowding at small spatial scales when only a small number of trees are included in the sample (Hegyi, 1974). The coefficient of variation of stem diameter was calculated as a measure of the heterogeneity of tree size in the neighbourhood.

At the plot level, the regularity of species spatial distribution was estimated using the spatial mingling index (von Gadow & Hui, 2002), which scores each tree based on whether it shares 207 species identity with its nearest neighbours. The spatial regularity of trees was estimated using 208 the uniform angle index (winkelmass) (von Gadow & Hui, 2002), which scores each tree based 209 on the angles between nearest neighbours. Additionally, the degree of spatial clustering of trees 210 was measured using Voronoi tessellation, as the coefficient of variation of Voronoi cell areas (Ong 211 et al., 2012). Finally, plot level tree density was calculated to estimate crowding at the plot 212 scale. See Chapter 6 for more information on the behaviour of the spatial mingling index and 213 uniform angle index. 214

Species diversity at both the subplot and plot level was measured using the exponential Shannon index (i.e. the Hill number of order q=1), calculated using tree species abundance. At the subplot level trees were included if they had canopy material inside the 10 m diameter subplot, while at the plot level trees were included if the largest stem was inside the plot boundaries.

2.3.4 Statistical analysis

Non-metric Multi-dimensional Scaling (NMDS) was used to describe variation in species composition among plots, using genus-level basal area weighted abundance in each plot. Stems that could not be identified to genus were excluded from this analysis, which accounted for 0.2% of

Table 1: Description of the vegetation type clusters, identified using the Ward algorithm based on basal area weighted genus abundance. AGB = Above-Ground woody Biomass. Species richness, stem density and AGB are reported as the median among plots, with the interquartile range in parentheses.

Site	Cluster	N sites	Richness	Stem density (Stems ha ⁻¹)	AGB (t ha ⁻¹)
Bicuar	1	12	17(2)	642(194)	41(8.4)
Mtarure	2	5	23(4)	411(137)	72(11.9)
Bicuar	3	3	6(1)	196(55)	77(7.3)
Mtarure	4	2	12(2)	288(73)	9(0.2)

the total basal area recorded. Four distinct vegetation types, two from each site (Table 1), were identified using hirerarchical clustering of the four dominant NMDS ordination axes. Clusters were further described using Dufrêne-Legendre indicator species analysis and by ranking tree species according to abundance across all plots within each cluster.

Linear mixed effects models tested the effects of tree species diversity and stand structural diversity on subplot canopy complexity metrics. Mixed models used a nested random intercept structure to account for the sampling design of subplots within plots and plots within vegetation types. Separate models were fitted for each canopy complexity metric, resulting in four models at the subplot level. Effect sizes among fixed effects in maximal models were compared for each canopy complexity metric, using the 95% confidence interval of the effect size to ascertain whether a fixed effect was significant by whether the confidence interval overlapped zero (Nakagawa & Cuthill, 2007). AIC values and Akaike weights of models with different combinations of fixed effects were compared to determine which combination of diversity and structural metrics best explained variation in each canopy complexity metric.

Path analysis was used to test whether tree species diversity may influence canopy complexity indirectly through its effect on stand structure, using the piecewiseSEM R package (Lefcheck, 2016). The path analysis investigated the direct effect of plot species diversity on mean plot canopy closure, as well as the indirect effect of diversity on canopy closure via the coefficient of variation of basal area, with random intercept terms for each vegetation type. The ex-Acacia vegetation type was represented by only two plots and could not be included in this model due to lack of replication.

Statistical analysis of the determinants of plot level canopy complexity metrics were conducted using linear models. Again, these models excluded the ex-Acacia vegetation type due to lack of replication. As with the subplot linear mixed models, predictor variable effect sizes were used to assess predictor variable significance, and comparison of candidate models using AIC, Akaike weights, and model R² values was used to determine which combination of predictors best explained each canopy complexity metric.

$_{ ext{50}}$ 3 Results

3.1 Description of vegetation types

Indicator species analysis shows that the four identified vegetation types constitute common southern African savanna floristic archetypes (Table 2). Cluster 1, found in Bicuar National Park contains typical miombo species from the Detarioideae subfamily, such as *Julbernardia paniculata*. Cluster 1 is the most frequent vegetation type in this study, with 12 plots. Cluster 1 has the

Table 2: Floristic description of the vegetation type clusters. Dominant species are the most abundant individuals across all plots per cluster. Indicator species are derived from Dufrêne-Legendre indicator species analysis with the three highest indicator values.

Cluster	Dominant species	Indicator species	Indicator value
1	Julbernardia paniculata	Strychnos spinosa	0.83
	Burkea africana	Combretum collinum	0.74
	Combretum collinum	Julbernardia paniculata	0.70
2	Diplorhynchus condylocarpon	Pteleopsis myrtifolia	1.00
	Pseudolachnostylis maprouneifolia	Diplorhynchus condylocarpon	0.89
	Gymnosporia senegalensis	Pseudolachnostylis maprouneifolia	0.81
3	Baikiaea plurijuga	Baikiaea plurijuga	0.94
	Baphia massaiensis	Baphia massaiensis	0.83
	Philenoptera nelsii	Philenoptera nelsii	0.45
4	Combretum apiculatum	Vachellia nilotica	0.99
	Burkea africana	Combretum apiculatum	0.70
	Bauhinia petersiana	Senegalia polyacantha	0.62

highest stem density, but lower AGB than Clusters 2 or 3, which contain larger individuals with disproportionately higher biomass. Cluster 2, found in Mtarure Forest Reserve, is dominated by *Pteleopsis myrtifolia*, a common miombo species from the Combretaceae family. Indeed, Cluster 2 also contained other common miombo species shared with plots in Cluster 1, such as *Julbernardia globiflora* and *Pseudolachnostylis maprouneifolia*, but these clusters remain distinct due to biogeographic variation in endemic genera at the longitudinal extremes of the miombo ecoregion represented by the two sites in this study. Cluster 3 represents *Baikiaea* woodland, found on Kalahari sands in southern Angola. It is species poor and dominated by *Baikiaea plurijuga* which forms large spreading canopy trees with high AGB. Other shrubby species that coppice readily in response to disturbance by fire such as *Baphia massaiensis* are also common. Cluster 4, found in Mtarure is a type of ex-Acacia woodland, dominated by *Vachellia* and *Senegalia* spp. This vegetation type was not well represented in the study, with only two plots, precluding its use in some multi-level statistical analyses due to lack of replication. Cluster 4 had far lower AGB than the other clusters (Table 1).

Differences in canopy structure among the four vegetation types are evident through observation of canopy surface models for typical plots within each vegetation type (Figure 5), and by comparing canopy complexity metrics (Figure 6). Cluster 1 shows many overlapping crowns forming a nearly contiguous canopy surface, and the heighest plot foliage density of all clusters. Though most trees in Cluster 1 have smaller crowns than those in Cluster 2, which also forms a nearly contiguous canopy. The largest trees in Cluster 2 grow taller and have a wider spreading canopy than those in other vegetation types. Cluster 3 shows two distinct size classes of tree, the large *Baikiaea plurijuga* forming clear isolated canopies, and much smaller scattered shrubby individuals in the understorey. Cluster 4 shows many small shrubby individuals with irregular canopy shapes, but a greater total crown area coverage than Cluster 3.

3.2 Bivariate relationships

Bivariate plots and linear models show that subplot species diversity, measured as the truenumbers equivalent of the Shannon diversity index of the tree neighbourhood around each 10

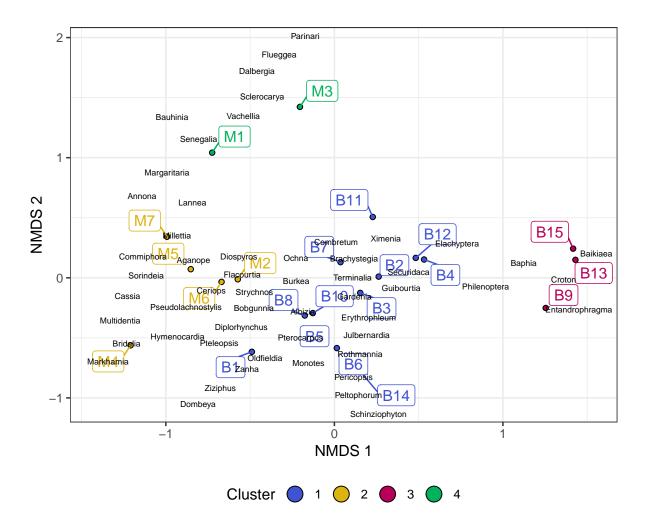


Figure 2: The first two axes of a Non-metric Multi-Dimensional Scaling (NMDS) analysis of tree genus diversity in each plot. Genus scores are labelled as black text, while plot scores are labelled as coloured points. Plots are shaded by vegetation types identified by hierarchical clustering: 1) B1-B8, B10-B12, B14, dominated by core miombo species such as *Julbernardia* spp., *Brachystegia* spp.; 2) M2, M5, M6, and M7, also dominated by core miombo genera with some genera not found in Bicuar National Park such as *Commiphora* and *Sorindeia*; 3) B9, B13 and B15, dominated by *Baikiaea plurijuga*; and 4) M1, M3, and M4, dominated by *Senegalia* spp., *Vachellia* spp., and *Combretum* spp.

m diameter subplot, appears to have weak positive effects on subplot canopy layer diversity, 283 canopy closure and foliage density (Figure 3, Table 3). The Hegyi crowding index had strong 284 positive effects on all canopy complexity metrics, as expected. The effect of Hegyi crowding on 285 subplot canopy complexity metrics was similar across all vegetation types (Table 6). Structural 286 diversity, measured as the coefficient of variation of subplot stem basal area had significant weak 287 positive effects on total canopy foliage, layer diversity, and canopy closure. 288

At the plot level, effects of species diversity and stand structure on canopy complexity were 289 similarly weak, but not significant except for the effect on canopy height, which explained more 290 variance in canopy height than tree density (Figure 4, Table 3). The effect of spatial regularity of 291 stems on canopy closure, measured by uniform angle index, was clearly negative, while the effect 292 of spatial clustering of stems, measured by the coefficient of variation of Voronoi cell area, was 293 negligible. Additionally, there was a non-significant negative effect of the coefficient of variation 294 of basal area on whole canopy rugosity. As expected, tree density had strong positive and 295 significant effects on foliage density and canopy closure, and negative effects on canopy roughness 296 and canopy rugosity. Cluster 4 represented an outlier in plot level bivariate relationships, with low canopy closure, low canopy height, low species diversity, and low variation in stem size. 298

3.3 Subplot mixed models 299

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Linear mixed effects models showed that species diversity of the subplot neighbourhood con-300 tributed to both layer diversity and canopy closure (Table 4), despite their low R² in bivariate linear models, and low effect sizes in maximal linear mixed models (Figure 7). As also seen in the subplot bivariate relationships Figure 3, the Hegyi crowding index had strong positive effects on all measured canopy complexity metrics, though these effects were non-significant for vegetation clusters 3 and 4. Heterogeneity of stem basal area had a significant positive effect on layer diversity and foliage density, but there was wide variation in vegetation type marginal effects for Clusters 3 and 4, due to low levels of replication. Cluster 3 had strong positive effects of species diversity on foliage uniformity and layer diversity. The random effects of vegetation type and plot identity described most of the variation in layer diversity and foliage density. Foliage uniformity was poorly explained by all combinations of fixed effects, with the best model only explaining 29%. All models were better than random effects only models according to AIC values.

3.4 Whole-plot multivariate linear models

While species diversity had varying effects on different plot level canopy complexity metrics, the 314 confidence intervals on these effect sizes were wide (Figure 8). Species diversity had a significant 315 positive effect on canopy height ($\beta=3\pm0.96$, p<0.05), a non-significant positive effect on canopy closure (β =0.07±0.085, p=0.41), but a negative effect on canopy surface roughness (β =-13±6.8, 317 p=0.09) and whole canopy rugosity (β =-111±71, p=0.15). Plot tree density had negligible 318 effects on canopy complexity, except for canopy rugosity (β =-61±42, p=0.17), in contrast to the 319 effect of Hegyi crowding on subplot canopy complexity. Spatially explicit measures of structural 320 diversity, measured by the uniform angle index and Voronoi cell area CV respectively, had 321 negligible effects on all canopy complexity metrics. One exception was the effect of uniform 322 angle index, i.e. the spatial clustering of stems, on canopy closure, which was clearly negative 323 $(\beta=-0.08\pm0.043, p=0.1)$, and the effect of Voronoi cell area on foliage density, which was positive 324 $(\beta = 6199 \pm 3312, p = 0.09).$ 325

Despite the weak effect sizes of species diverity on canopy complexity at the plot level, model 326 selection showed that canopy height and canopy roughness were better explained by models 327 which included species diversity (Table 5). Additionally, the best model for canopy height also 328

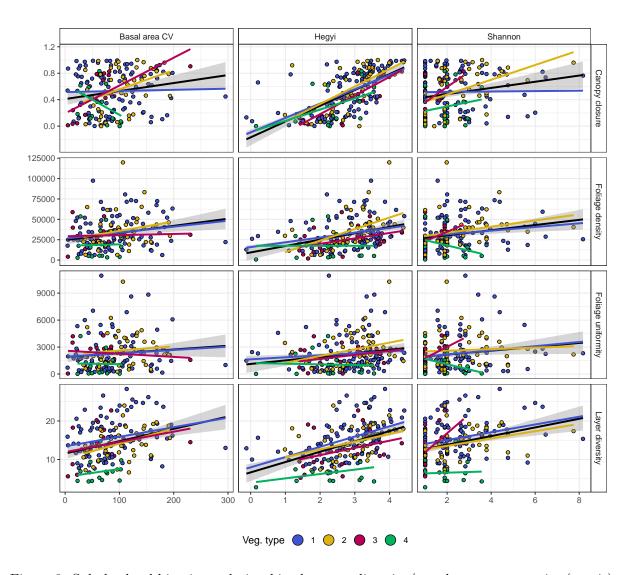


Figure 3: Subplot level bivariate relationships between diversity/stand structure metrics (x axis) and canopy complexity metrics (y axis). Points and linear model lines of best fit are coloured by vegetation type. Black lines of best fit are linear models including all plots, with a 95% confidence interval. See Table 6 for a comparison of linear model fits by vegetation type.

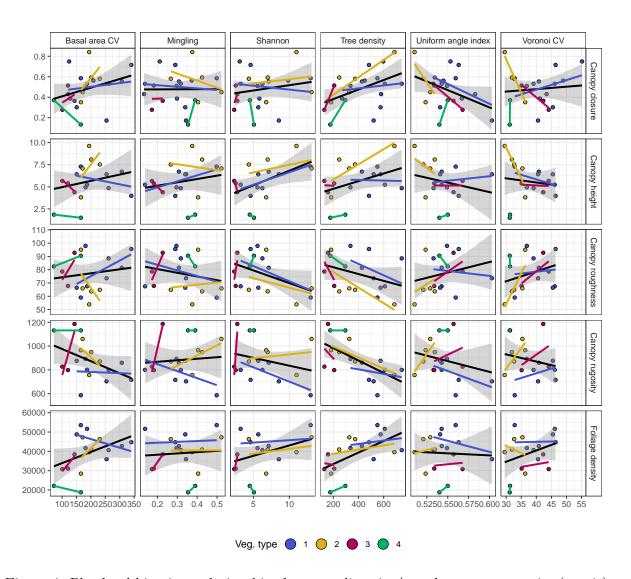


Figure 4: Plot level bivariate relationships between diversity/stand structure metrics (x axis) and canopy complexity metrics (y axis). Points and linear model lines of best fit are coloured by vegetation type. Black lines of best fit are linear models including all plots, with a 95% confidence interval. See Table 6 for a comparison of linear model fits by vegetation type.

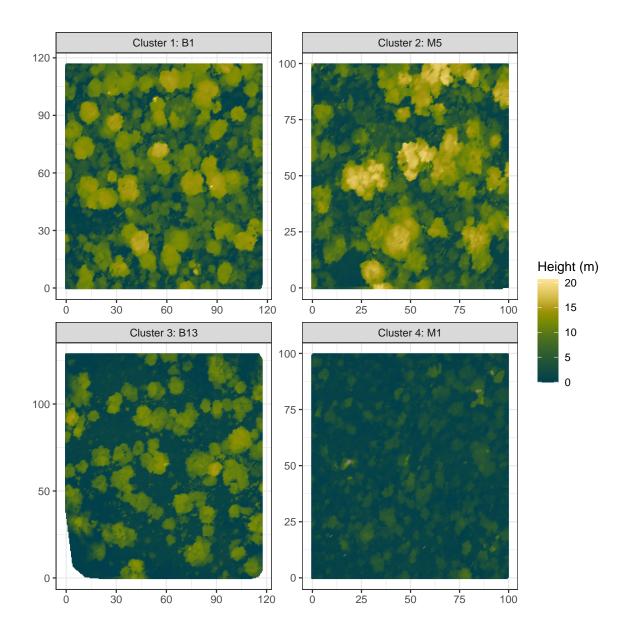


Figure 5: Representative canopy surface models for each vegetation type identified in the Non-metric Multi-dimensional Scaling (NMDS) clustering analysis. Panel titles show the plot name and the vegetation type.

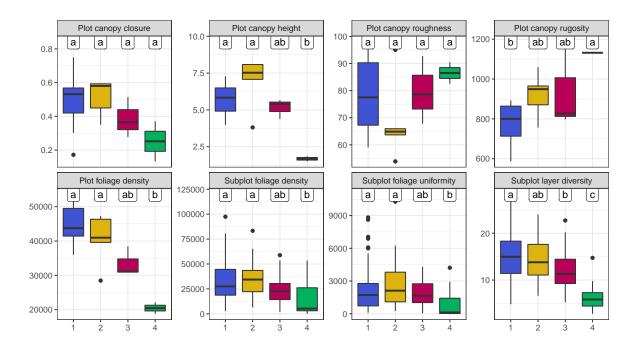


Figure 6: Box plots showing variation in canopy complexity metrics among the four vegetation types identified in the Non-metric Multi-dimensional Scaling (NMDS) clustering analysis. Thick lines show the median, boxes show the interquartile range (IQR), whiskers show $1.5 \times IQR$, and points show outliers beyond these limits. Labels above each box plot group vegetation types according to significant differences in pairwise Tukey's tests; vegetation types sharing a letter are not significantly different.

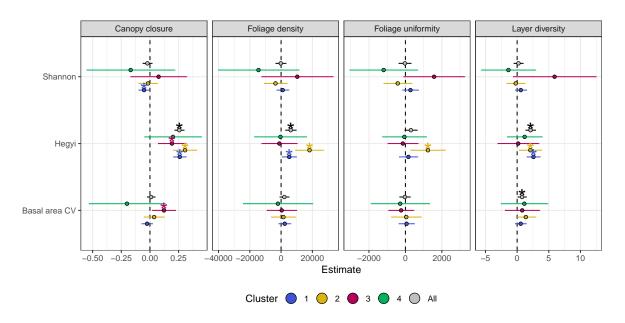


Figure 7: Standardised fixed effect slopes for each model of a canopy complexity metric. Slope estimates where the interval (± 1 standard error) does not overlap zero are considered to be significant effects, marked with asterisks. Points are coloured according to vegetation type.

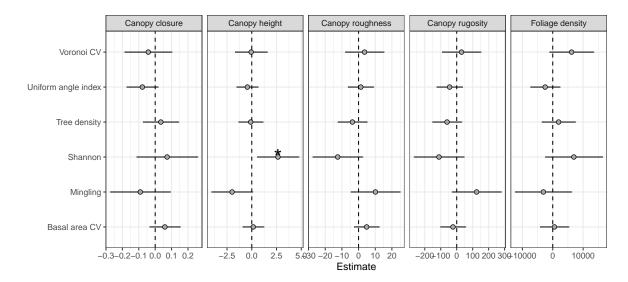


Figure 8: Standardised effect sizes for whole-plot canopy rugosity. Slope estimates where the interval (± 1 standard error) does not overlap zero are considered to be significant effects, marked with asterisks.

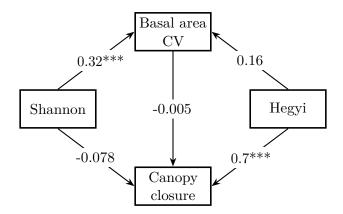


Figure 9: Directed Acyclic Graph showing standardised path coefficients of paths in the path analysis of the indirect effect of subplot species diversity (Shannon diversity index) on canopy closure via coefficient of variation of basal area. Asterisks define p-value thresholds: *<0.05, **<0.01, ***<0.001.

included spatial mingling of tree species. The model for canopy roughness was no better than a null model, and all models had non-significant p-values. Voronoi cell area CV was not included in any models.

3.5 Path analysis

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The path analysis investigating the indirect effect of subplot species diversity on canopy closure via the coefficient of variation of basal area showed that while species diversity had a strong positive significant effect on basal area variation, the effect of basal area variation on canopy closure remained negligible (Figure 9). The indirect effect of species diversity on canopy closure via basal area coefficient of variation was -0.0016, while the direct effect was -0.078. As in the bivariate relationships and plot level linear models, species diversity had a weak positive significant effect on canopy closure, while the major driver of canopy closure was the Hegyi crowding index.

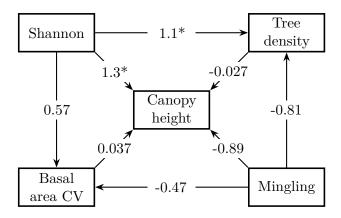


Figure 10: Directed Acyclic Graph showing standardised path coefficients of paths in the path analysis of the indirect effect of plot species diversity (Shannon diversity index) and species mingling on mean canopy height via stand structural metrics of coefficient of variation of basal area and tree density. Asterisks define p-value thresholds: *<0.05, **<0.01, ***<0.001.

3.6 Covariance of subplot and plot measures of canopy complexity

Plot and subplot canopy complexity metrics were highly correlated in many cases, with similar relationships among vegetation types (Figure 8). Most subplot and plot level canopy metrics covaried in a predictable manner. For example, increased canopy height led to an increase in canopy closure. Plot canopy height especially, tended to be strongly positively correlated with subplot canopy complexity metrics. Additionally, as canopy rugosity increased, many subplot canopy complexity and density metrics decreased. Subplot metrics varied greatly within plots, producing large uncertainty in plot level estimates of these metrics. All subplot level canopy complexity metrics positive correlated with each other (Figure 13). Plot level canopy complexity also generally correlated (Figure 12). Plot level measures of spatial heterogeneity in canopy structure, i.e. canopy surface roughness and canopy rugosity, were negatively correlated with measures of canopy density, i.e. foliage density, canopy closure, and canopy height. Measures of canopy spatial heterogeneity positively correlated with each other, as did measures of canopy density.

355 4 Discussion

This study investigated relationships between tree species diversity, stand structure, and several metrics of tree canopy complexity using terrestrial LiDAR in southern African savannas, with a view to improving understanding of the biotic drivers of variation in canopy complexity and vegetation dynamics. Species diversity appeared to generally have weak positive effects on canopy complexity metrics related to canopy density at both the subplot and plot scales. Plots with greater species diversity produced taller tree canopies, with greater canopy closure and foliage density.

Plot level canopy complexity metrics were generally better predicted by species diversity and stand structure than subplot metrics. While positive relationships between species diversity and subplot canopy complexity metrics were observed in the subplot bivariate models, subplot linear mixed effects models did not show strong species diversity effects. Additionally, none of the best models for subplot canopy complexity metrics included species diversity as a fixed effect. This finding suggests a large degree of stochastic variability in canopy complexity within plots, that masks species effects at smaller spatial scales. The prevalence of disturbance events such as fire and damage by elephants in southern African woodlands, as well as tree-fall, small-scale

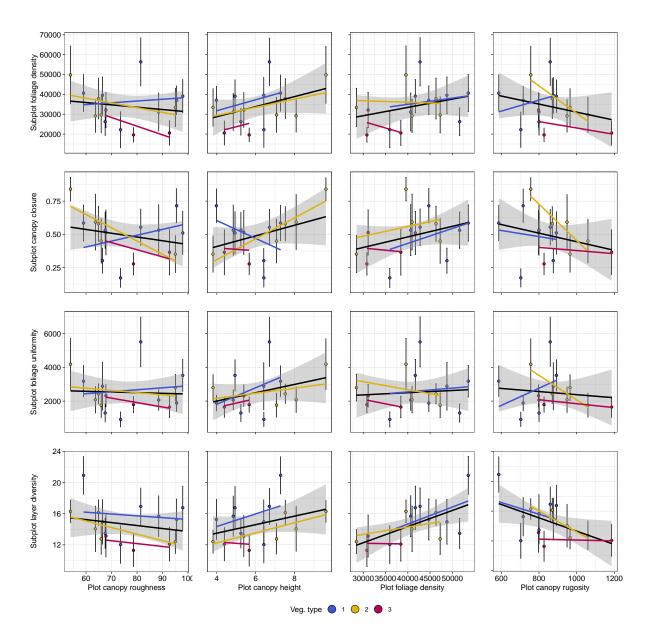


Figure 11: Bivariate plots comparing canopy structural metrics at the plot (x axis) and subplot scale (y axis). Each point represents the mean values of a single plot. Points and linear model fits are coloured according to vegetation type. The black linear model combines all vegetation types. Error bars on points are the standard deviation of mean subplot metrics across the plot. Note that because plot level canopy closure is calculated as the mean of subplot canopy closure, a comparison of subplot and plot canopy closure is not made in this figure.

variability in edaphic factors, and stochastic tree germination all contribute to heterogeneity in canopy complexity (). This study demonstrates the importance of large sample units and a high degree of replication when measuring canopy complexity, especially in disturbed systems, to effectively account for the inherent heterogeneity in the system ().

Spatial clustering of stems, measured using the uniform angle index, caused a clear decrease in 375 canopy closure, with similar behaviour across vegetation types. Uniform angle index was also 376 included in the best multivariate model predicting canopy closure. This finding is expected, 377 as spatial clustering results in reduced canopy cover in areas outside clusters, and a non-378 compensatory increase in canopy closure within clusters, due to competition among individuals 379 (). Clustering of trees in savannas can result from positive feedback effects from disturbance 380 by fire and herbivory (). This study suggests that as well as reducing canopy cover, i.e. the 381 ground area covered by tree canopies, disturbances in savannas may also serve to decrease canopy 382 closure, i.e. the visible sky proportion from both within canopy gaps and between canopy gaps, 383 indirectly by increasing spatial clustering of trees. The negative effects of spatial clustering 384 on canopy closure are expected to increase in species poor woodlands, due to a lack of niche complementarity among coexisting individuals (). 386

Stand structural diversity caused positive canopy complexity effects for within-canopy structural 387 metrics such as layer diversity and canopy surface roughness, but had negligible effects on canopy 388 density. This is in line with other studies in forest ecosystems, which report that variation in 389 tree size increases total canopy volume occupancy by increasing the number of canopy layers, 390 but does not necessarily result in a concomitant increase in canopy closure, as the resulting 391 canopies are often more sparse, especially for understorey individuals (). The path analysis 392 also supports this conclusion, where species diversity was found to cause an increase in stand 393 structural diversity, but this did not extend to an increase in canopy closure. 394

The effect of stand structure on canopy complexity in this system appears to be a result of demographic effects rather than variation in growth form as a function of species diversity. The path analysis testing the indirect effect of species diversity on canopy closure via stand structural diversity did not find a significant indirect effect of species diversity on canopy closure. While other studies in forests have found a species diversity effect on stand structural diversity (), it is suggested here that prevailing disturbance pressures mask any species diversity effect.

Effects of species composition, measured by vegetation type, on canopy complexity were small. 401 While vegetation types differed in mean values for stand structural and species diversity metrics, 402 variation in these metrics produced results of similar direction and magnitude among vegetation 403 types in most cases. Small sample sizes for Baikiaea and ex-Acacia vegetation however, led to 404 wide errors on most relationships especially at the plot level, such that it is impossible to draw 405 deeper conclusions about the behaviour of these vegetation types. Variation in mean values of 406 canopy complexity metrics among vegetation types is likely driven by species identity (), though 407 species composition itself is likely driven by environmental factors and disturbance regime (). 408

Thoughts on the suitability of different canopy complexity metrics in sparse and clustered savannas. Lots of stochstic variation in stand structure

411 Lots of disturbance is exogenous, i.e. not controlled by stand composition, lots of noise

In bivariate relationships tree density had a strong effect on canopy complexity metrics at the plot level, but did not feature prominently in the best quality mixed models for those metrics. Tree density covaried with basal area coefficient of variation, with denser plots also exhibiting greater variation in tree size. This is also reflected in the bivariate relationship between tree density and canopy height. Further path analysis showed that tree species diversity and spatial mingling of tree species increased tree density and variation in tree size, which caused an increase in canopy height.

Hegyi index is a key driver of canopy complexity at subplot, so what drives variation in the

- 420 Hegyi index. Discussion
- 421 All subplot canopy complexity metrics postiviely covaried. This indicates that for rapid assess-
- ments of canopy complexity, simple canopy closure measurements are sufficient to characterise
- canopy complexity, at the spatial scale of an individual tree neighbourhood. In closed-canopy
- forest ecosystems, measures of canopy spatial heterogeneity often correlate with canopy closure
- (). In the savannas studied here however, the sparser and discontinuous canopy results in a
- ⁴²⁶ negative relationship. MORE

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Table 6: Summary statistics of bivariate linear models comparing canopy complexity metrics with diversity and stand structural metrics. Slope refers to the slope of the predictor term in the model, \pm 1 standard error. T is the t-value of the slope of the predictor term in the model, Asterisks indicate the p-value of these terms (***<0.001, **<0.01, *<0.05).

Response	Predictor	Cluster	Slope	F	\mathbb{R}^2	Т
		1	$7.3e+01\pm3.7e+01$	4.0(2,97)	0.04	1.99*
Foliage density	Recel eres CV	2	$1.1e+02\pm7.9e+01$	2.1(2,38)	0.05	1.44
ronage density	Dasar area C v	3	$1.4e + 01 \pm 7.2e + 01$	0.0(2,14)	0.00	0.20
		4	$1.6e + 01 \pm 2.0e + 02$	-01±3.7e+01	0.08	
		1	$5.9e+03\pm2.1e+03$	8.2(2,102)	0.07	2.86**
Foliage density	Ности	2	$1.4e + 04 \pm 3.6e + 03$	15.2(2,40)	0.28	3.90***
ronage density	Hegyi	3	$6.6e + 03 \pm 3.0e + 03$	4.8(2,23)	0.17	2.18*
		4	$1.5e + 01 \pm 5.5e + 03$	0.0(2,13)	0.00	0.00
		1	$2.2e+03\pm1.3e+03$	2.8(2,102)	0.03	1.67
Foliago dongitu	Channon	2	$3.8e + 03 \pm 2.4e + 03$	2.6(2,39)	0.06	1.61
Foliage density	Shannon	3	$1.1e+04\pm6.5e+03$	3.1(2,20)	0.13	1.77
		4	$-6.5e + 03 \pm 6.5e + 03$	1.0(2,13)	0.07	-1.01
		1	$1.7e-04\pm6.0e-04$	0.1(2,97)	0.00	0.28
Conony aloguro	Dogal area CV	2	$2.9e-03\pm1.1e-03$	6.9(2,39)	0.15	2.62*
Canopy closure	Dasar area Cv	3	$4.2e-03\pm1.1e-03$	15.1(2,14)	0.52	3.89**
		4	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0.16	-1.50	
		1	$2.2\text{e-}01\pm2.8\text{e-}02$	62.3(2,102)	0.38	7.89***
Comony aloguno	TT .	2	$2.6e-01\pm5.1e-02$	27.0(2,41)	0.40	5.19***
Canopy closure	педуг	3	$2.8e-01\pm4.0e-02$	50.7(2,23)	0.69	7.12***
		4	$1.7e-01\pm 8.0e-02$	4.5(2,13)	0.26	2.12
C		1	$3.1e-03\pm2.2e-02$	0.0(2,102)	0.00	0.14
	Shannon	2	$1.1e-01\pm3.2e-02$	12.1(2,40)	0.23	3.48**
Canopy closure		3	$2.3e-01\pm1.4e-01$	2.9(2,20)	0.13	1.69
		4	$6.7 \text{e-} 02 \pm 1.1 \text{e-} 01$	0.4(2,13)	0.03	0.60
		1	$3.7e + 00 \pm 4.0e + 00$	0.9(2,97)	0.01	0.92
Foliago uniformity	Dogal area CV	2	$4.5e+00\pm7.4e+00$	0.4(2,38)	0.01	0.61
ronage unnormity	Basal area CV	3	$-3.5e+00\pm5.9e+00$	0.4(2,14)	0.02	-0.59
		4	1.4e+01±7.2e+01	0.00	-0.06	
		1	$2.2e + 02 \pm 2.3e + 02$	1.0(2,102)	0.01	0.98
Foliomo uniformitar	Hamri	2	$7.5e + 02 \pm 3.7e + 02$	4.0(2,40)	0.09	2.00
ronage unnormity	Basal area CV Hegyi Hegyi Shannon Basal area CV Hegyi Hegyi Basal area CV Hegyi Hegyi	3	$4.5e+02\pm2.6e+02$	2.9(2,23)	0.11	1.72
		4	$-7.5e + 01 \pm 4.0e + 02$	0.0(2,13)	0.00	-0.19
		1	$2.3e+02\pm1.4e+02$	2.6(2,102)	0.02	1.61
Foliago uniformity	Shannon	2	$8.6e + 01 \pm 2.2e + 02$	0.1(2,39)	0.00	0.38
ronage uniformity	Shannon	3	$1.3e + 03 \pm 5.1e + 02$	· / /	0.23	2.48*
		4	$-5.9e + 02 \pm 4.7e + 02$	1.6(2,13)	0.11	-1.27
		1	$2.5e-02\pm9.3e-03$	7.1(2,97)	0.07	2.66**
Layer diversity	Rasal area CV	2	$3.9e-02\pm1.4e-02$	8.0(2,38)	0.17	2.83**
Layer diversity	Dasai alea UV	3		` ' '		1.15
		4	$2.1e-02\pm3.1e-02$	0.5(2,12)	0.04	0.67
		1	$2.7e + 00 \pm 4.9e - 01$	$29.1(2,\overline{102})$	0.22	5.39***
Layer diversity	Hogwi	2	$2.0\mathrm{e}{+00}{\pm}7.5\mathrm{e}{-01}$	7.1(2,40)	0.15	2.66*
Layer diversity	11cgy1	3	$1.9e+00\pm1.0e+00$	3.6(2,23)	0.13	1.89
		4	$1.1e+00\pm 8.5e-01$	1.8(2,13)	0.12	1.33
		1	$1.0e + 00 \pm 3.4e - 01$	8.7(2,102)	0.08	2.95**
Layer diversity	Shannon			· · /		
Layer diversity	SHAIIHOH					

		2	$9.5e-01\pm4.3e-01$	4.8(2,39)	0.11	2.18*
		3	$4.9e + 00 \pm 1.8e + 00$	7.2(2,20)	0.11	2.68*
		4	$1.8e-01\pm1.1e+00$	0.0(2,13)	0.20	0.16
		1	$1.2e-01\pm6.9e-02$	2.9(2,6)	0.33	1.72
Canopy roughness	Basal area CV	2	$-3.2e-01\pm2.9e-01$	1.2(2,3)	0.29	-1.10
		3	$3.5e-01\pm4.7e-01$	0.6(2,1)	0.36	0.74
		4				
		1	$2.6e-01\pm1.2e+00$	0.0(2,6)	0.01	0.22
Canopy roughness	Voronoi CV	2	$4.6e + 00 \pm 1.9e + 00$	6.1(2,3)	0.67	2.48
Canopy roughness	voronor C v	3	$1.8e + 00 \pm 1.9e + 00$	1.0(2,1)	0.49	0.99
		4				
		1	$-4.2e+01\pm5.7e+01$	0.5(2,6)	0.08	-0.74
C 1	ъ. г. 1.	2	$1.6e + 01 \pm 9.7e + 01$	0.0(2,3)	0.01	0.17
Canopy roughness	Mingling	3	$3.5e + 02 \pm 2.5e + 02$	2.0(2,1)	0.67	1.42
		4		, ,		
		1	$-4.3e-02\pm4.5e-02$	0.9(2,6)	0.13	-0.96
		2	$-5.9e-02\pm3.1e-02$	3.6(2,3)	0.54	-1.89
Canopy roughness	Tree density	3	$-1.8e-01\pm2.6e-01$	0.5(2,1)	0.31	-0.68
		4		010(=,=)	0.02	0.00
		1	$-2.3e+00\pm1.7e+00$	1.7(2,6)	0.22	-1.32
	Shannon	2	$-1.4e + 00 \pm 2.4e + 00$	0.4(2,3)	0.22	-0.60
Canopy roughness		3	$3.4e + 0.0\pm 2.4e + 0.0$ $3.4e + 0.0\pm 4.7e + 0.0$	0.4(2,3) 0.5(2,1)	0.11 0.34	-0.00 0.72
		$\frac{3}{4}$	J.4c 01±4.7c 01	0.0(2,1)	0.54	0.12
			7.40+01+9.60+09	0.1(9.6)	0.01	-0.28
		1	$-7.4e + 01 \pm 2.6e + 02$	0.1(2,6)	0.01	-0.28 0.43
Canopy roughness	Uniform angle index	$\frac{2}{3}$	$4.1e+02\pm9.5e+02$ $4.4e+02\pm5.7e+02$	$0.2(2,3) \\ 0.6(2,1)$	0.00 0.37	$0.45 \\ 0.76$
		4	4.40+02±3.70+02	0.0(2,1)	0.57	0.70
			6.5-02.16.1-02	1 1(0 6)	0.16	1.07
		1	$-6.5e-03\pm6.1e-03$	1.1(2,6)	$0.16 \\ 0.28$	-1.07 1.08
Canopy height	Basal area CV	$\frac{2}{3}$	$4.3e-02\pm4.0e-02$ $-3.1e-02\pm8.7e-03$	$ 1.2(2,3) \\ 12.3(2,1) $	0.28 0.92	-3.51
Canopy neight		4	-3.1e-02±6.7e-03	12.3(2,1)	0.92	-5.51
			1.001.10.600	1 [(0,0)	0.00	1.01
		1	$-1.0e-01\pm8.6e-02$ $-7.0e-01\pm2.0e-01$	1.5(2.6)	0.20	-1.21
Canopy height	Voronoi CV	$\frac{2}{3}$	$-1.0e-01\pm 2.0e-01$ $-1.8e-02\pm 1.4e-01$	$ 12.7(2,3) \\ 0.0(2,1) $	$0.81 \\ 0.02$	$-3.57* \\ -0.13$
		3 4	-1.6e-02±1.4e-01	0.0(2,1)	0.02	-0.13
				0.7/2.3		
		1	$6.8e + 00 \pm 3.8e + 00$	3.2(2,6)	0.34	1.78
Canopy height	Mingling	2	$-3.3e+0.0\pm1.3e+0.1$	0.1(2,3)	0.02	-0.25
. U	5 0	3	$-2.3e+01\pm9.3e-01$	619.2(2,1)	1.00	-24.88*
		4				
		1	$-3.5e-04\pm3.8e-03$	0.0(2,6)	0.00	-0.09
Canopy height	Tree density	2	$8.6e-03\pm4.0e-03$	4.7(2,3)	0.61	2.16
оапору пеідпі	rice density	3	$-1.0e-03\pm1.7e-02$	0.0(2,1)	0.00	-0.06
		4				
		1	$2.8e-01\pm1.1e-01$	7.1(2,6)	0.54	2.66*
0 1 1 1	CI	2	$1.7e-01\pm3.3e-01$	0.3(2,3)	0.08	0.52
Canopy height	Shannon	3	$-3.0e+00\pm9.0e-01$	11.1(2,1)	0.92	-3.32
			• • • • • •	())		

		4				
		1	$1.0e+01\pm2.1e+01$	0.2(2,6)	0.04	0.49
Canopy height	Uniform angle index	2	$-7.2e + 01 \pm 1.3e + 02$	0.3(2,3)	0.09	-0.56
Canopy neight	Official angle fidex	3	$6.0e-02\pm3.9e+01$	0.0(2,1)	0.00	0.00
		4				
		1	$3.6e-04\pm6.9e-04$	0.3(2,10)	0.03	0.53
Conony alaguma	Basal area CV	2	$3.5e-03\pm3.5e-03$	1.0(2,3)	0.24	0.98
Canopy closure	Dasai area Cv	3	$1.9e-03\pm5.3e-03$	0.1(2,1)	0.11	0.35
		4				
		1	9.3e-03±8.2e-03	1.3(2,10)	0.11	1.13
C	Voronoi CV	2	$-6.6e-02\pm7.9e-03$	69.7(2,3)	0.96	-8.35**
Canopy closure	voronoi C v	3	$-2.5e-02\pm4.6e-03$	29.0(2,1)	0.97	-5.39
		4				
		1	$-1.6e-01\pm5.1e-01$	0.1(2,10)	0.01	-0.31
Conony alaguma	Min alin a	2	$-6.9e-01\pm1.1e+00$	0.4(2,3)	0.12	-0.63
Canopy closure	Mingling	3	$7.6e-02\pm4.1e+00$	0.0(2,1)	0.00	0.02
		4		, , ,		
		1	$1.4e-04\pm4.0e-04$	0.1(2,10)	0.01	0.36
C 1	m 1 :	2	$8.5e-04\pm2.4e-04$	12.2(2,3)	0.80	3.50*
Canopy closure	Tree density	3	$3.0e-03\pm4.3e-06$	499683.9(2,1)	1.00	706.88***
		4		, , ,		
		1	$-7.6e-03\pm1.7e-02$	0.2(2,10)	0.02	-0.45
C 1	CI	2	$8.5e-03\pm3.0e-02$	0.1(2,3)	0.03	0.28
Canopy closure	Shannon	3	$1.9e-01\pm5.2e-01$	0.1(2,1)	0.12	0.37
		4		())		
		1	$-3.9e + 00 \pm 2.3e + 00$	2.9(2,10)	0.23	-1.71
G 1	Uniform angle index	2	$-1.2e + 01 \pm 9.3e + 00$	1.7(2,3)	0.36	-1.30
Canopy closure		3	$-6.9e + 00 \pm 3.9e - 01$	306.2(2,1)	1.00	-17.50*
		4		())		
		1	$-4.5e + 01 \pm 2.9e + 01$	2.3(2,6)	0.28	-1.52
T-10	D 1 ~	2	$1.5e + 02 \pm 1.4e + 02$	1.1(2,3)	0.27	1.05
Foliage density	Basal area CV	3	$1.8e + 02 \pm 8.9e + 01$	4.2(2,1)	0.81	2.06
		4		())		
		1	$3.5e + 0.1 \pm 5.0e + 0.2$	0.0(2,6)	0.00	0.07
T 1	17 . 017	$\overline{2}$	$-7.7e + 02 \pm 1.5e + 03$	0.3(2,3)	0.08	-0.51
Foliage density	Voronoi CV	3	$2.7e + 02 \pm 8.7e + 02$	0.1(2,1)	0.09	0.31
		4		())		
		1	$4.5e + 03 \pm 2.5e + 04$	0.0(2,6)	0.01	0.18
D.1	3.61 11	$\overline{2}$	$8.0e + 02 \pm 4.7e + 04$	0.0(2,3)	0.00	0.02
Foliage density	Mingling	3	$1.5e + 05 \pm 2.0e + 04$	54.1(2,1)	0.98	7.35
		4		()-/		
		1	$8.8e + 00 \pm 2.0e + 01$	0.2(2,6)	0.03	0.45
		2	$1.1e+01\pm2.1e+01$	0.3(2,3)	0.08	0.51
Foliage density	Tree density	3	$-1.3e+01\pm2.1e+01$	0.0(2,1)	0.01	-0.12
		4	, 011.10 02	(=,±)		J <u>-</u>
		1	$2.5e + 02 \pm 8.1e + 02$	0.1(2,6)	0.02	0.31
		1	∠.9e+U2±0.1e+U2	0.1(2,0)	0.02	0.91
Foliage density	Shannon					

		2 3 4	$5.0e+02\pm1.2e+03$ $1.8e+04\pm9.1e+03$	0.2(2,3) $3.9(2,1)$	0.05 0.80	0.42 1.98
Foliage density	Uniform angle index	1 2 3 4	$-1.1e+05\pm1.0e+05$ $1.2e+05\pm4.7e+05$ $4.3e+04\pm2.5e+05$	$ \begin{array}{c} 1.3(2,6) \\ 0.1(2,3) \\ 0.0(2,1) \end{array} $	0.18 0.02 0.03	-1.15 0.25 0.18
Canopy rugosity	Basal area CV	1 2 3 4	$-1.0e-01\pm6.1e-01$ $-2.2e+00\pm2.2e+00$ $8.7e+00\pm5.4e+00$	$0.0(2,6) \\ 1.1(2,3) \\ 2.6(2,1)$	0.00 0.26 0.73	-0.17 -1.03 1.62
Canopy rugosity	Voronoi CV	1 2 3 4	$7.9e+00\pm8.2e+00$ $3.5e+01\pm1.3e+01$ $1.8e+01\pm4.2e+01$	0.9(2,6) 6.8(2,3) 0.2(2,1)	0.13 0.69 0.15	0.96 2.61 0.42
Canopy rugosity	Mingling	1 2 3 4	$-5.9e+02\pm3.6e+02$ $8.5e+02\pm5.2e+02$ $7.2e+03\pm1.7e+03$	2.7(2,6) 2.7(2,3) 17.6(2,1)	0.31 0.47 0.95	-1.63 1.63 4.19
Canopy rugosity	Tree density	1 2 3 4	$-1.9e-01\pm3.4e-01$ $-4.6e-01\pm2.1e-01$ $-1.2e+00\pm5.4e+00$	$0.3(2,6) \\ 4.9(2,3) \\ 0.0(2,1)$	0.05 0.62 0.05	-0.56 -2.22 -0.22
Canopy rugosity	Shannon	1 2 3 4	$-2.4e+01\pm1.0e+01$ $6.4e+00\pm1.8e+01$ $8.5e+02\pm5.4e+02$	5.3(2,6) 0.1(2,3) 2.5(2,1)	0.47 0.04 0.71	-2.31 0.35 1.57
Canopy rugosity	Uniform angle index	1 2 3 4	$-2.6e+03\pm1.6e+03$ $1.0e+04\pm4.1e+03$ $3.4e+03\pm1.2e+04$	2.5(2,6) 6.1(2,3) 0.1(2,1)	0.30 0.67 0.07	-1.58 2.47 0.28

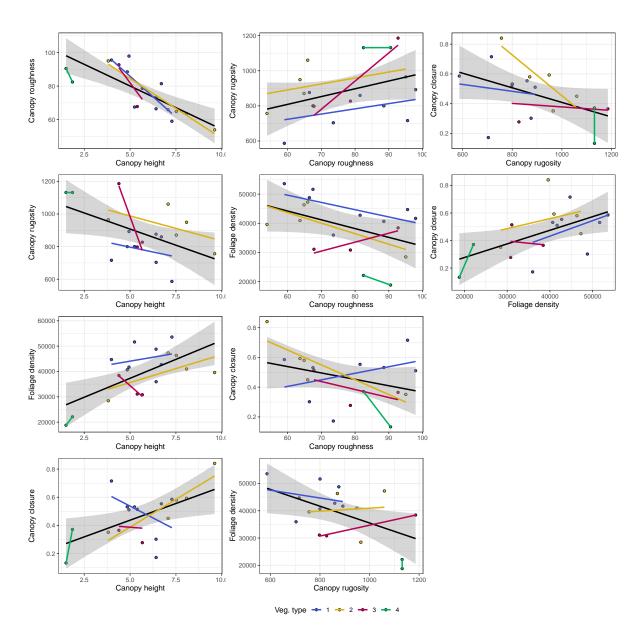


Figure 12: Bivariate scatter plots of plot level canopy complexity metrics.

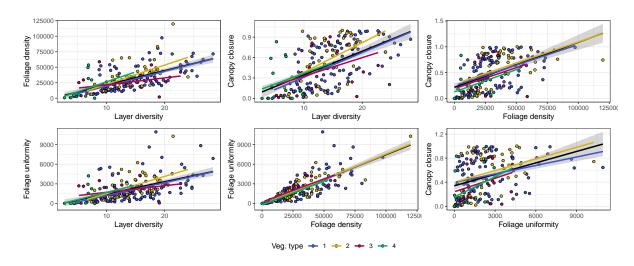


Figure 13: Bivariate scatter plots of subplot level canopy complexity metrics.

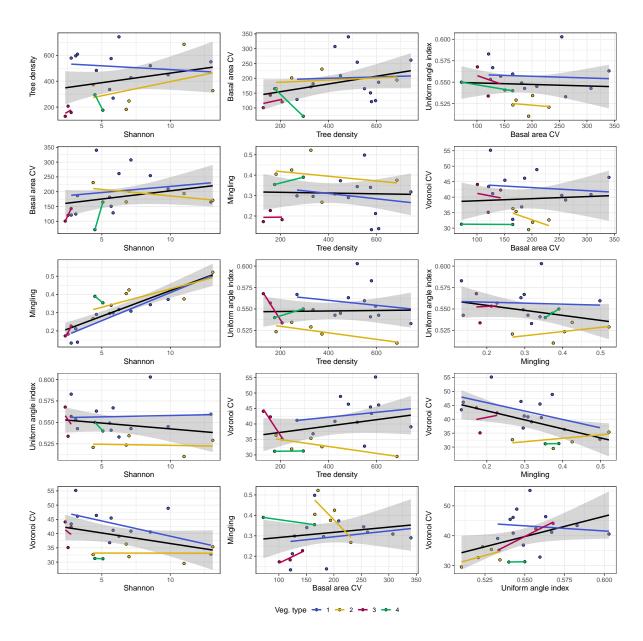


Figure 14: Bivariate scatter plots of plot level diversity and stand structural metrics.

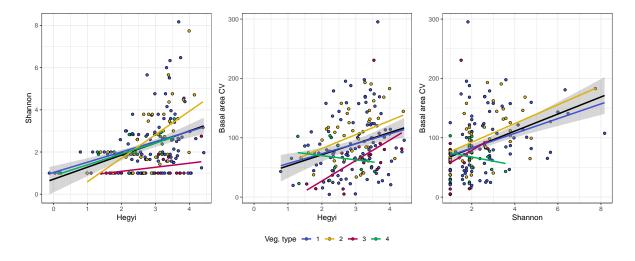


Figure 15: Bivariate scatter plots of subplot level diversity and stand structural metrics.

Table 3: Summary statistics of bivariate linear models comparing canopy complexity metrics with diversity and stand structural metrics across all vegetation types. Slope refers to the slope of the predictor term in the model, \pm 1 standard error. T is the t-value of the slope of the predictor term in the model, Asterisks indicate the p-value of these terms (***<0.001, **<0.01, *<0.05).

Response	Predictor	Slope	F	\mathbb{R}^2	$\overline{\mathrm{T}}$
	Basal area CV	$8.7e + 01 \pm 3.0e + 01$	8.6(2,167)	0.05	2.93**
Foliage density	Hegyi	$7.8e + 03 \pm 1.6e + 03$	25.5(2,184)	0.12	5.05***
	Shannon	$3.2e+03\pm1.1e+03$	8.9(2,180)	0.05	2.98**
	Basal area CV	$1.2e-03\pm4.8e-04$	6.3(2,168)	0.04	2.52*
Canopy closure	Hegyi	$2.4e-01\pm2.1e-02$	132.8(2,185)	0.42	11.52***
	Shannon	$4.7e-02\pm1.7e-02$	7.3(2,181)	0.04	2.70**
	Basal area CV	$4.1e+00\pm3.0e+00$	1.9(2,167)	0.01	1.37
Foliage uniformity	Hegyi	$4.0e + 02 \pm 1.6e + 02$	6.2(2,184)	0.03	2.49*
	Shannon	$2.2e + 02 \pm 1.1e + 02$	4.1(2,180)	0.02	2.04*
	Basal area CV	$3.2 \text{e-} 02 \pm 7.6 \text{e-} 03$	17.6(2,167)	0.10	4.20***
Layer diversity	Hegyi	$2.7e+00\pm3.9e-01$	46.8(2,184)	0.20	6.84***
	Shannon	$1.1e+00\pm2.7e-01$	16.8(2,180)	0.09	4.10***
	Basal area CV	$3.0 \text{e-} 02 \pm 5.0 \text{e-} 02$	0.4(2,16)	0.02	0.60
	Voronoi CV	$7.5e-01\pm 5.9e-01$	1.6(2,16)	0.09	1.26
Canopy roughness	Mingling	$-2.8e+01\pm3.3e+01$	0.7(2,16)	0.04	-0.86
Canopy roughness	Tree density	$-2.6e-02\pm1.7e-02$	2.3(2,16)	0.12	-1.51
	Shannon	$-1.9e + 00 \pm 9.5e - 01$	4.0(2,16)	0.20	-2.01
	Uniform angle index	$1.6e + 02 \pm 1.6e + 02$	1.0(2,16)	0.06	0.98
	Basal area CV	$7.1e-03\pm7.3e-03$	0.9(2,16)	0.06	0.97
	Voronoi CV	$-4.7e-02\pm9.1e-02$	0.3(2,16)	0.02	-0.52
Canopy height	Mingling	$3.8e+00\pm4.8e+00$	0.6(2,16)	0.04	0.79
Canopy neight	Tree density	$4.3e-03\pm2.5e-03$	3.1(2,16)	0.16	1.76
	Shannon	$3.3e-01\pm1.3e-01$	6.0(2,16)	0.27	2.45*
	Uniform angle index	$-2.2e+01\pm2.4e+01$	0.8(2,16)	0.05	-0.90
	Basal area CV	$8.5 \text{e-}04 \pm 5.7 \text{e-}04$	2.2(2,20)	0.10	1.50
	Voronoi CV	$2.4e-03\pm5.8e-03$	0.2(2,20)	0.01	0.41
Canopy closure	Mingling	$7.2e-03\pm3.7e-01$	0.0(2,20)	0.00	0.02
Canopy closure	Tree density	$4.7e-04\pm1.9e-04$	6.3(2,20)	0.24	2.50*
	Shannon	$1.0e-02\pm1.2e-02$	0.7(2,20)	0.04	0.86
	Uniform angle index	$-3.4e + 00 \pm 1.7e + 00$	3.9(2,20)	0.16	-1.98
	Basal area CV	$5.8e + 01 \pm 3.2e + 01$	3.3(2,16)	0.17	1.80
	Voronoi CV	$5.8e + 02 \pm 4.1e + 02$	2.1(2,16)	0.11	1.43
Foliage density	Mingling	$6.6e + 03 \pm 2.3e + 04$	0.1(2,16)	0.01	0.29
romage demoney	Tree density	$3.0e+01\pm1.0e+01$	8.6(2,16)	0.35	2.93**
	Shannon	$1.1e + 03 \pm 6.9e + 02$	2.5(2,16)	0.13	1.57
	Uniform angle index	$-2.1e+04\pm1.1e+05$	0.0(2,16)	0.00	-0.18
	Basal area CV	$-1.0e + 00 \pm 5.3e - 01$	3.7(2,16)	0.19	-1.92
	Voronoi CV	$-6.0e + 00 \pm 7.0e + 00$	0.7(2,16)	0.04	-0.86
Canopy rugosity	Mingling	$1.3e + 02 \pm 3.8e + 02$	0.1(2,16)	0.01	0.33
camply ragonity	Tree density	$-5.2e-01\pm1.7e-01$	10.0(2,16)	0.38	-3.16**
	Shannon	$-1.3e+01\pm1.2e+01$	1.2(2,16)	0.07	-1.11
	Uniform angle index	$-1.8e + 03 \pm 1.9e + 03$	0.9(2,16)	0.06	-0.97

Table 4: Explanatory variables included in the best model for each subplot canopy structure variable. ΔAIC shows the difference in model AIC value compared to a null model which included only the random effects of vegetation type and plot. Positive ΔAIC values >2 indicate that the model is of better quality than the null model. R^2_c is the R^2 of the best model, while R^2_m is the R^2 of the model fixed effects only.

Response	Hegyi	Shannon	Basal area CV	$\Delta { m AIC}$	${ m R^2_{\ c}}$	R^2_{m}
Layer diversity	✓	✓	√	37.0	0.50	0.17
Foliage density	\checkmark		\checkmark	47.6	0.27	0.09
Foliage uniformity	\checkmark			13.1	0.28	0.02
Canopy closure	\checkmark	\checkmark		101.9	0.60	0.46

Table 5: Explanatory variables included in the best linear model for each plot-level canopy complexity metric. ΔAIC shows the difference in model AIC value compared to a null model. Positive ΔAIC values >2 indicate that the model is of better quality than the null model.

Response	Shannon	Tree density	Basal area CV	Mingling	Uniform angle index	Voronoi CV	$\Delta { m AIC}$	\mathbb{R}^2	Prob.
Foliage density		✓					3.8	0.58	0.08
Canopy closure			\checkmark		\checkmark		1.9	0.54	0.13
Canopy height	\checkmark			\checkmark			2.1	0.54	0.12
Canopy roughness	\checkmark						0.3	0.50	0.19
Canopy rugosity		\checkmark					4.0	0.59	0.08