Species diversity and stand structure as drivers of canopy complexity in southern African woodlands

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1 Abstract

Atmospheric CO₂ enrichment and human-induced climate change are expected to drive woody encroachment and an increase in tree cover across African savannas, with consequences for ecosystem function, particularly related to carbon dynamics. The patch dynamics of savanna-woodland mosaics are complex however, as woody growth is mediated by seasonal fire that is itself driven by properties of the woody overtstorey. It is unclear how variation in tree species composition and stand structure in this ecosystem affects woody canopy complexity, and how this might determine future vegetation dynamics. Here, I conducted a study of canopy structure in southern African savannas using terrestrial LiDAR, at sites in Bicuar National Park, Angola and Mtarure Forest Reserve, Tanzania, to explore relationships between tree species diversity, species composition, the spatial distribution of trees, variation in tree size and canopy complexity. I found consistent weak positive effects of species diversity on plot scale canopy complexity metrics related to canopy density, but a negative effect on metrics related to the spatial heterogeneity of the distribution of canopy material. Species diversity caused an increase in canopy height, canopy closure, and within-canopy structural complexity. However, stochasticity in neighbourhood scale woody structure masked species diversity effects at small spatial scales. Finally I found that spatial clustering of trees in space led to a reduction in canopy closure, even within clustered areas, suggesting that disturbance by fire and herbivory not only reduce canopy cover at the landscape scale, but also reduce canopy cover at smaller spatial scales But how does disturbance lead to clustering? Could just say "suggesting"

$_{\scriptscriptstyle 2}$ 1 Introduction

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Atmospheric CO₂ enrichment, coupled with climate change and changing disturbance regimes, is expected to drive woody encroachment, i.e. proliferation of trees in previously non-wooded 24 areas, and increased growth of trees in currently wooded areas, across the savanna biome over 25 the coming century (Criado et al., 2020; Mitchard & Flintrop, 2013; Stevens et al., 2016). As 26 atmospheric CO₂ concentrations increase, C₃ trees are expected to gain a competitive edge over 27 C₄ grasses due to differences in photosynthetic pathway and carbon use efficiency (Buitenwerf 28 et al., 2012), with cascading effects on canopy cover, grass growth, and disturbance regime (Bond 29 & Midgley, 2012). If realised, woody encroachment and woody densification will have significant effects on the global carbon cycle, as more CO₂ is stored in woody biomass, as well as myriad 31 other effects on ecosystem structure (Donohue et al., 2013). Indeed, tropical savannas have been 32 identified as the fastest increasing component of the terrestrial carbon sink (Sitch et al., 2015). 33 Previous studies however, have reported wide variation in rates of woody encroachment and 34 densification (Mitchard & Flintrop, 2013), particularly in disturbance-prone savannas such as 35 miombo woodlands in southern Africa (Axelsson & Hanan, 2018), and it is unclear how the 36 fertilisation effect of atmospheric CO₂ enrichment will interact with other ecosystem properties 37 to alter vegetation structure (Körner, 2017; Reich et al., 2014). 38

- Savanna vegetation is defined by the coexistence of trees and grasses (Scholes & Archer, 1997).
- 40 In the tropical mesic savannas of southern Africa, disturbance by fire and herbivory are the main
- limitations on tree cover, preventing the competitive exclusion of shade-sensitive C₄ grasses

where climatic conditions would otherwise allow for closed canopy forest (Sankaran et al., 2005). C₄ grasses also provide the main fuel source for seasonal fires in these savannas (Frost, 1996), 43 producing a positive feedback where an increase in tree cover reduces grass fuel load, reducing 44 fire frequency and intensity, increasing tree cover, and so on (Staver & Koerner, 2015). As such, 45 even small perturbations in tree cover can lead to large changes in vegetation structure if critical 46 thresholds of tree cover are crossed (Hirota et al., 2011). Previous research has sought to identify 47 environmental factors which affect tree cover and its responses to atmospheric CO₂ enrichment, 48 but few have considered the functional role of the existing tree community and its effect on 49 ecosystem processes. 50

Canopy structure describes the spatial distribution and density of tree canopy foliage (Lowman & 51 Rinker, 2004). Canopy structural complexity, i.e. the spatial heterogeneity of foliage distribution 52 within the canopy, has been linked to increased net ecosystem productivity (Baldocchi & Wilson, 53 2001; Chen et al., 2012; Gough et al., 2019; Hardiman et al., 2011; Law et al., 2001; Morin, 2015), 54 increased resilience of productivity (Pretzsch, 2014), reduced understorey light penetration (Fotis 55 et al., 2018; Scheuermann et al., 2018), and greater moderation of understorey micro-climate 56 (Wright et al., 2017). Furthermore, in temperate and boreal forests, functional differences among 57 coexisting tree species in their vertical and horizontal canopy occupation provides a link between 58 species diversity, canopy structural complexity and canopy density, with canopy complexity 59 constituting a mechanism for observed positive biodiversity-ecosystem function effects in wooded 60 ecosystems (Barry et al., 2019; Pretzsch, 2014). In tropical savannas, tree species diversity might 61 therefore influence ecosystem-level woody thickening in response to elevated atmospheric CO₂, where competition effects in diverse tree communities are reducted due to niche separation, and 63 can more effectively increase foliage density and reduce understorey light penetration, excluding 64 grass and thus reducing the probability of disturbance. 65

As well as the species diversity of trees, the spatial distribution and relative size of tree stems, i.e. stand structure, is also expected to affect canopy structural complexity (Stark et al., 2015). 67 Heterogeneity in stem size, whether a result of species diversity, disturbance history or some other 68 factor, is expected to increase canopy complexity and canopy density as individuals of different 69 sizes occupy different parts of the vertical canopy space (Panzou et al., 2020), and may differ in light requirements (Charles-Dominique et al., 2018). Additionally, clustering of individuals in 71 space is expected to increase canopy structural heterogeneity across the wider savanna landscape, 72 but ultimately decrease total foliage density due to an increase in competitive interactions (Dohn 73 et al., 2017). Clustering may occur as a result of disturbance history, facilitation effects among 74 individuals in stressful environments (Ratcliffe et al., 2017), or due to other limitations on 75 establishment arising from growth strategy (). More diverse communities may allow greater stem 76 density and greater foliage density, as differences in canopy occupancy among species reduce 77 negative effects competition among individuals on growth (Gough et al., 2019). 78

Functional differences among floristic types of savanna may also drive variation in canopy 79 complexity, irrespective of species diversity. Some savanna trees form denser canopies than 80 others, as a result of variation in leaf size and branch architecture. Previous studies have 81 compared the branch architecture of ex-Acacia (e.g. Senegalia and Vachellia spp.) and miombo 82 (e.g. Julberardia, Brachystegia, and Isoberlinia) archetypal tree species. While ex-Acacia species 83 tend to inhabit drier, heavily grazed areas, miombo species tend to inhabit dystrophic wetter 84 areas structured heavily by fire (Ribeiro et al., 2020). These studies have shown that ex-Acacia 85 species develop sparser canopies, cagey branch architecture, and wider spreading crowns, while 86 dominant Fabaceae species from the miombo develop thicker, taller canopies, and can grow to 87 large trees (Archibald & Bond, 2003; Mugasha et al., 2013; Privette et al., 2004). Similarly, 88 dominant miombo Fabaceae species from the Detarioideae subfamily have been shown to exhibit 89 wider crowns and grow taller than coexisting species from the Combretaceae family (). Shenkin et al. (2020) showed that Fabaceae tree species from tropical forests exhibit wider and more

voluminous tree crowns than other common families of tropical trees. Under identical stem densities, miombo woodland species may therefore exclude grass more effectively than ex-Acacia or Combretaceae species given these differences in growth form.

Canopy complexity is multi-dimensional and has previously been explained using a plethora 95 of simple metrics that originated in forest and community ecology (Kershaw et al., 2017). 96 Assessments of canopy complexity have most often modelled tree canopies as a series of ellipses 97 (2D), ellipsoids or cones (3D) based on field measurements with measuring tapes (Jucker et al., 98 2015), or used surrogate proxies for 3D canopy structure, due to its inherent complexity (Seidel et 99 al., 2011). Measurements of this kind are time consuming and yet remain an over-simplification of 100 canopy structure. Alternatively, canopy closure is often measured using indirect optical methods 101 which partition sky from canopy material, i.e. with hemispherical photography or the commonly 102 used LAI-2000, providing a 2D representation of the canopy but lacking information on vertical 103 canopy structure (Jonckheere et al., 2004). In recent years, particularly in temperate and boreal 104 forests, LiDAR (Light Detection And Ranging) has emerged as a suitable technology for rapidly 105 and precisely assessing canopy structure in 3D, conserving information on 3D structure of the 106 calibre that is required to understand it's complexities (Calders et al., 2020; Muir et al., 2018). 107 In tropical savannas, very few studies have used terrestrial LiDAR for vegetation analyses, and 108 in southern Africa all existing studies have been located at the Skukuza Flux Tower in Kruger 109 National Park, South Africa (Muumbe et al., 2021). Pioneering work describing the ecology of 110 southern African savannas placed large emphasis on canopy structural diversity as a mediator of 111 ecosystem function (Solbrig et al., 1996), but much of that understanding of savanna vegetation 112 structure was derived from traditional mensuration methods. Using terrestrial LiDAR to measure 113 canopy complexity in southern African savannas therefore offers a unique chance to validate 114 accepted theory and describe differences in ecosystem structure among savanna vegetation types 115 in finer detail than previously possible. 116

In this study I applied terrestrial LiDAR techniques to woodland-savanna mosaics at two sites in southern Africa, with the aim of increasing understanding of how various measures of tree canopy complexity relate to tree diversity and stand structure. I hypothesise that tree neighbourhoods with greater tree species diversity, greater heterogeneity in stem size, and greater heterogeneity in stem location, allow greater canopy complexity and foliage density. Thus, more diverse savannas might exhibit a higher potential woody biomass, and more effectively increase their growth under elevated atmospheric CO₂, promoting woody thickening. I also consider the functional differences in canopy architecture among tree communities and how this affects canopy closure and total canopy occupancy.

¹²⁶ 2 Materials and methods

2.1 Study sites

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Field measurements were conducted at two sites, Bicuar National Park, in southwest Angola (S15.1°, E14.8°), and Mtarure Forest Reserve, in southeast Tanzania (S9.0°, E39.0°) (Figure 1).

At each site, 1 ha (100×100 m) plots were located in areas of savanna-woodland vegetation, across a gradient of stem density and a range of savanna floristic archetypes. In Angola, 15 plots were sampled, while in Tanzania, seven were sampled following the curtailment of fieldwork due to COVID-19 travel restrictions. Fieldwork was conducted between February and April at both sites, during the peak growth period of each site in order to capture the maximum foliage volume in the canopy.

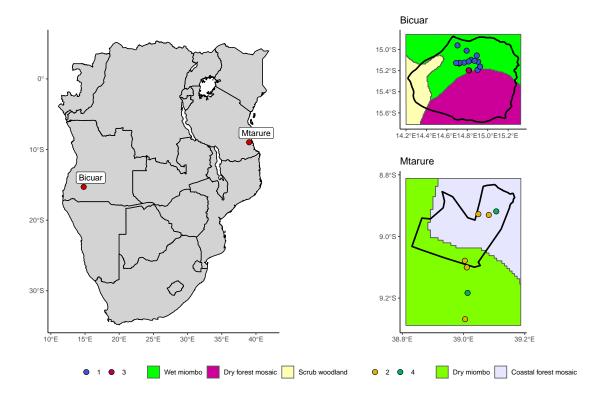


Figure 1: Location of study sites within southern Africa (left), and of 1 ha plots within each site (right). The black outlines in each site map denote the boundaries of protected areas which encompass the majority of study sites, Bicuar National Park in Angola (top), and Mtarure Forest Reserve in Tanzania (bottom). The background of each site map is a re-classified version of White's vegetation map (White, 1983). Points in site maps are shaded according to vegetation type identified by hierarchical clustering of tree genera abundances. Note that all maps are on different scales.

136 2.2 Field measurements

Within each 1 ha plot, each woody stem ≥ 5 cm stem diameter was identified to species, the 137 stem Diameter at Breast Height (DBH) was measured at 1.3 m above the ground, and the stem 138 location within the plot was recorded using tape measures. Each 1 ha plot was sampled by nine 139 10 m diameter circular subplots arranged in a regular grid, with a 15 m buffer from the plot edge 140 and 35 m between subplots. For each subplot, the distance and direction from the subplot centre of each stem >5 cm diameter with canopy material inside the subplot was recorded. Within each 142 subplot, a variable number of scans were recorded using a Leica HDS6100 phase-shift Terrestrial 143 Laser Scanner (TLS). The number and position of scans within a subplot was determined by the 144 arrangement of canopy material in the subplot, to minimise shadows within the canopy of the 145 subplot, and to maximise canopy penetration. The number of scans per subplot ranged between 146 one and five across both sites. Extended field methods and data analysis methods are described in Chapter 6. 148

149 2.3 Data analysis

150 2.3.1 TLS processing

Point clouds from scans in each subplot were registered and unified using Leica Cyclone (version 151 9.1), using five reflective cross targets visible to all scans as anchor points. Point clouds were 152 voxelised to cubic voxels of different sizes depending on the application of the data. Subplot 153 height profile estimation and gap fraction was conducted using 5 cm³ voxels, while whole plot 154 canopy rugosity was estimated using 50 cm³ voxels. Voxels were classified as 'filled' if they 155 intersected one or more points. Variation in voxel size reflects the spatial scale of each analysis, 156 and is bounded by the beam divergence of the scanner over longer distances (Cifuentes et al., 157 2014). Choosing voxels that are too small can result in pock-marked representations of surfaces 158 that are especially problematic when calculating larger scale canopy complexity metrics such as 159 canopy top roughness, while voxels that are too large can result in an over-estimation of plant volume when estimating canopy foliage density at the subplot scale (Cifuentes et al., 2014; Seidel 161 et al., 2012). 162

The noise reduction algorithm from Rusu et al. (2008) was used to discard points based on 163 mean nearest neighbour distances, with a mean number of neighbours of eight, and a standard 164 deviation threshold of 1.96. This effectively removed 'ghost points' produced by partial beam 165 interceptions and also removed many erroneous returns caused by airborne dust particles, which 166 was common at these study sites. Raw points clouds for each subplot had a mean of ~2.9e+08 167 points, ~4.5e+07 points after voxelisation to 5 cm³, and ~2.1e+07 points after noise reduction. 168 Ground points were classified using the Progressive Morphological Filter (PMF) from Zhang 169 et al. (2003). Point cloud height was reclassified based on this revised ground layer by measuring 170 the vertical distance between the nearest ground point and each point. Points below 1.3 m 171 height above ground were discarded for calculations of foliage density, canopy cover, and canopy 172 complexity, as points below this threshold where often occupied by long grass. 173

2.3.2 Canopy complexity metrics

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Ray-tracing was used to estimate canopy closure in each subplot, i.e. the proportion of the sky hemisphere occluded by plant material at the subplot centre from multiple TLS scans. Hemispherical images were created using the POV-Ray ray-tracing software (Persistence of Vision Pty. Ltd., 2004). Filled voxels were represented as black cubes filling the voxel volume, with a white sky box and no light source. A 'camera' with a 180° fisheye lens was placed at the

subplot centre within POV-Ray, at a height of 1.8 m pointing directly upwards. The images produced by POV-Ray were analysed using Hemiphot (ter Steege, 2018) to estimate canopy closure. Canopy closure estimates from the TLS were validated with hemispherical photographs taken at the same location and processed using the same method in Hemiphot, and compared using Pearson's correlation (r(195)=0.87, p<0.001). A plot level estimate of canopy closure was calculated as the mean of subplot canopy closure measurements. See Chapter 6 for expanded methods and explanation of the behaviour of the different canopy complexity metrics.

Effective Number of Layers (ENL) was calculated according to Ehbrecht et al. (2016) to measure 187 vertical variation in subplot foliage density. ENL is calculated as the exponential Shannon index 188 (i.e. the Hill number of order q=1) of foliage density among 50 cm vertical layers within each 189 subplot: 190

$$ENL = \exp\left(-\sum_{i=1}^{N} p_i \times \ln p_i\right) \tag{1}$$

Where p_i is the proportion of filled voxels in the 50 cm layer i, and N is the total number of 191 layers. ENL increases with canopy height and thus number of layers, and also with variation in 192 foliage density among those layers, but not with increased total foliage density. 193

Total foliage density was calculated within each subplot as the area under the curve of the foliage 194 height profile. Total foliage density was also calculated at the plot level as the sum of filled 50 195 cm³ voxels across the plot. Vertical variation in subplot foliage density was calculated by fitting 196 a linear model to the cumulative foliage density profile, then calculating the sum of squared 197 residuals of that model. 198

Plot level canopy surface models were extracted using the 99th percentile of canopy height in 10 cm² columns. A pit-filling algorithm provided by Khosravipour et al. (2014) was applied at 50 ${
m cm}^2$ resolution to reduce the effects of incomplete canopy penetration in dense canopies. Whole plot canopy complexity was measured by three metrics. Canopy top roughness was measured as the coefficient of variation (CV) of canopy height across the plot. Canopy rugosity was measured according to Hardiman et al. (2011), as the CV of vertical and horizontal foliage density within 0.5 m³ cubic bins. Finally, canopy height was calculated as the mean of the canopy surface roughness model across the plot.

2.3.3 Stand structure and diversity

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For each subplot, an adapted version of the Iterative Hegyi index was used to estimate crowding, as an alternative to stem density which does not adequately capture crowding at small spatial scales when only a small number of trees are included in the sample (Hegyi, 1974). The CV of 210 stem diameter was calculated as a measure of the heterogeneity of tree size in the neighbourhood. At the plot level, the regularity of species spatial distribution was estimated using the spatial mingling index (von Gadow & Hui, 2002), which scores each tree based on whether it shares species identity with its nearest neighbours. The spatial regularity of trees was estimated using the uniform angle index (winkelmass) (von Gadow & Hui, 2002), which scores each tree based on the angles between nearest neighbours. Additionally, the degree of spatial clustering of trees was measured using Voronoi tessellation, as the CV of Voronoi cell areas (Ong et al., 2012). Finally, plot level tree density was calculated to estimate crowding at the plot scale. See Chapter 6 for 218 more information on the behaviour of the spatial mingling index and uniform angle index. 219

Species diversity at both the subplot and plot level was measured using the exponential Shannon 220 index (i.e. the Hill number of order q=1), calculated using tree species abundance (Jost, 221 2006). At the subplot level trees were included if they had canopy material inside the 10 m

diameter subplot, while at the plot level trees were included if the largest stem was inside the plot boundaries.

225 2.3.4 Statistical analysis

Non-metric Multi-dimensional Scaling (NMDS) was used to describe variation in species composition among plots, using genus-level basal area weighted abundance in each plot. Stems that
could not be identified to genus were excluded from this analysis, which accounted for 0.2% of
the total basal area recorded. Four distinct vegetation types, two from each site (Table 1), were
identified using hirerarchical clustering of the four dominant NMDS ordination axes. Clusters
were further described using Dufrêne-Legendre indicator species analysis and by ranking tree
species according to abundance across all plots within each cluster.

Linear mixed effects models tested the effects of tree species diversity and stand structural 233 diversity on subplot canopy complexity metrics. Mixed models used a nested random intercept 234 structure to account for the sampling design of subplots within plots and plots within vegetation 235 types. Separate models were fitted for each canopy complexity metric, resulting in four models 236 at the subplot level. Effect sizes among fixed effects in maximal models were compared for each 237 canopy complexity metric, using the 95% confidence interval of the effect size to ascertain whether 238 a fixed effect was significant by whether the confidence interval overlapped zero (Nakagawa & 239 Cuthill, 2007). AIC values and Akaike weights of models with different combinations of fixed 240 effects were compared to determine which combination of diversity and structural metrics best 241 explained variation in each canopy complexity metric. 242

Path analysis was used to test whether tree species diversity influences canopy complexity 243 indirectly through its effect on stand structure, using the piecewiseSEM R package (Lefcheck, 244 2016). Two path analyses were conducted, one at the plot level and one at the subplot level. 245 Subplot path analysis investigated the direct effect of species diversity on canopy closure, as well as the indirect effect of diversity on canopy closure via the CV of basal area, with random 247 intercept terms for each vegetation type. The ex-Acacia vegetation type was represented by only 248 two plots and could not be included in this model due to lack of replication. Plot level path 249 analysis investigated the direct effects of species diversity and spatial mingling of species on 250 mean canopy height, as well as the indirect effects of these metrics on canopy height via tree density and basal area CV. Again, ex-Acacia plots were excluded from this path analysis. 252

Statistical analysis of the determinants of plot level canopy complexity metrics were conducted using linear models. Again, these models excluded the ex-Acacia vegetation type due to lack of replication. As with the subplot linear mixed models, predictor variable effect sizes were used to assess predictor variable significance, and comparison of candidate models using AIC, Akaike weights, and model R² values was used to determine which combination of predictors best explained each canopy complexity metric.

259 3 Results

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3.1 Description of vegetation types

Indicator species analysis shows that the four identified vegetation types constitute common southern African savanna floristic archetypes (Table 2). Cluster 1, found in Bicuar National Park contains typical miombo species from the Detarioideae subfamily, such as *Julbernardia paniculata*. Cluster 1 is the most frequent vegetation type in this study, with 12 plots. Cluster 1 has the highest stem density, but lower AGB than Clusters 2 or 3, which contain larger individuals with disproportionately higher biomass. Cluster 2, found in Mtarure Forest Reserve, is dominated

Table 1: Description of the vegetation type clusters, identified using the Ward algorithm based on basal area weighted genus abundance. AGB = Above-Ground woody Biomass. Species richness, stem density and AGB are reported as the median among plots, with the interquartile range in parentheses.

Site	Cluster	N sites	Richness	Stem density (Stems ha ⁻¹)	AGB (t ha ⁻¹)
Bicuar	1	12	17(2)	642(194)	41(8.4)
Mtarure	2	5	23(4)	411(137)	72(11.9)
Bicuar	3	3	6(1)	196(55)	77(7.3)
Mtarure	4	2	12(2)	288(73)	9(0.2)

Table 2: Floristic description of the vegetation type clusters. Dominant species are the most abundant individuals across all plots per cluster. Indicator species are derived from Dufrêne-Legendre indicator species analysis with the three highest indicator values.

Cluster	Dominant species	Indicator species	Indicator value
1	Julbernardia paniculata	Strychnos spinosa	0.83
	Burkea africana	Combretum collinum	0.74
	Combretum collinum	Julbernardia paniculata	0.70
2	Diplorhynchus condylocarpon	Pteleopsis myrtifolia	1.00
	Pseudolachnostylis maprouneifolia	Diplorhynchus condylocarpon	0.89
	Gymnosporia senegalensis	Pseudolachnostylis maprouneifolia	0.81
3	Baikiaea plurijuga	Baikiaea plurijuga	0.94
	Baphia massaiensis	Baphia massaiensis	0.83
	Philenoptera nelsii	Philenoptera nelsii	0.45
4	Combretum apiculatum	Vachellia nilotica	0.99
	Burkea africana	Combretum apiculatum	0.70
	Bauhinia petersiana	Senegalia polyacantha	0.62

by Pteleopsis myrtifolia, a common miombo species from the Combretaceae family. Indeed, 267 Cluster 2 also contained other common miombo species shared with plots in Cluster 1, such as 268 Julbernardia globiflora and Pseudolachnostylis maprouneifolia, but these clusters remain distinct 269 due to biogeographic variation in endemic genera at the longitudinal extremes of the miombo 270 ecoregion represented by the two sites in this study. Cluster 3 represents Baikiaea woodland, 271 found on Kalahari sands in southern Angola. It is species poor and dominated by Baikiaea 272 plurijuga which forms large spreading canopy trees with high AGB. Other shrubby species that 273 coppice readily in response to disturbance by fire such as Baphia massaiensis are also common. 274 Cluster 4, found in Mtarure is a type of ex-Acacia woodland, dominated by Vachellia and 275 Senegalia spp. This vegetation type was not well represented in the study, with only two plots, 276 precluding its use in some multi-level statistical analyses due to lack of replication. Cluster 4 277 had far lower AGB than the other clusters (Table 1).

Differences in canopy structure among the four vegetation types are evident through observation 279 of canopy surface models for typical plots within each vegetation type (Figure 5), and by 280 comparing canopy complexity metrics (Figure 6). Cluster 1 shows many overlapping crowns 281 forming a nearly contiguous canopy surface, and the heighest plot foliage density of all clusters. 282 Though most trees in Cluster 1 have smaller crowns than those in Cluster 2, which also forms a 283 nearly contiguous canopy. The largest trees in Cluster 2 grow taller and have a wider spreading 284 canopy than those in other vegetation types. Cluster 3 shows two distinct size classes of tree, 285 the large Baikiaea plurijuqa forming clear isolated canopies, and much smaller scattered shrubby 286 individuals in the understorey. Cluster 4 shows many small shrubby individuals with irregular 287 canopy shapes, but a greater total crown area coverage than Cluster 3. 288

3.2 Bivariate relationships 289

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Bivariate plots and linear models show that subplot species diversity, measured as the true-290 numbers equivalent of the Shannon diversity index of the tree neighbourhood around each 10 291 m diameter subplot, appears to have weak positive effects on subplot canopy layer diversity, 292 canopy closure and foliage density (Figure 3, Table 3). The Hegyi crowding index had strong 293 positive effects on all canopy complexity metrics, as expected. The effect of Hegyi crowding on subplot canopy complexity metrics was similar across all vegetation types (Table 6). Structural 295 diversity, measured as the CV of subplot stem basal area had significant weak positive effects on 296 total canopy foliage, layer diversity, and canopy closure. 297

At the plot level, effects of species diversity and stand structure on canopy complexity were similarly weak, but not significant except for the effect on canopy height, which explained more variance in canopy height than tree density (Figure 4, Table 3). The effect of spatial regularity of 300 stems on canopy closure, measured by uniform angle index, was clearly negative, while the effect of spatial clustering of stems, measured by Voronoi cell area CV, was negligible. Additionally, there was a non-significant negative effect of basal area CV on whole canopy rugosity. As expected, tree density had strong positive and significant effects on foliage density and canopy closure, and negative effects on canopy roughness and canopy rugosity. Cluster 4 represented an outlier in plot level bivariate relationships, with low canopy closure, low canopy height, low species diversity, and low variation in stem size.

3.3 Subplot mixed models

Linear mixed effects models showed that species diversity of the subplot neighbourhood contributed to both layer diversity and canopy closure (Table 4), despite their low R² in bivariate 310 linear models, and low effect sizes in maximal linear mixed models (Figure 7). As also seen 311 in the subplot bivariate relationships Figure 3, the Hegyi crowding index had strong positive 312

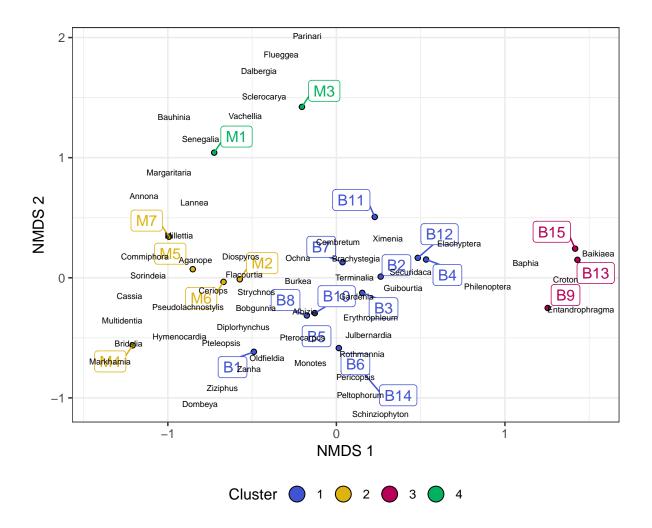


Figure 2: The first two axes of a Non-metric Multi-Dimensional Scaling (NMDS) analysis of tree genus diversity in each plot. Genus scores are labelled as black text, while plot scores are labelled as coloured points. Plots are shaded by vegetation types identified by hierarchical clustering: 1) B1-B8, B10-B12, B14, dominated by core miombo species such as *Julbernardia* spp., *Brachystegia* spp.; 2) M2, M5, M6, and M7, also dominated by core miombo genera with some genera not found in Bicuar National Park such as *Commiphora* and *Sorindeia*; 3) B9, B13 and B15, dominated by *Baikiaea plurijuga*; and 4) M1, M3, and M4, dominated by *Senegalia* spp., *Vachellia* spp., and *Combretum* spp.

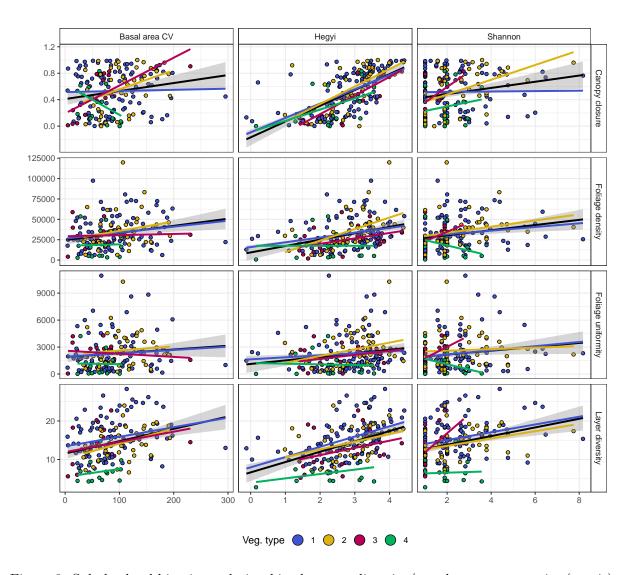


Figure 3: Subplot level bivariate relationships between diversity/stand structure metrics (x axis) and canopy complexity metrics (y axis). Points and linear model lines of best fit are coloured by vegetation type. Black lines of best fit are linear models including all plots, with a 95% confidence interval. See Table 6 for a comparison of linear model fits by vegetation type.

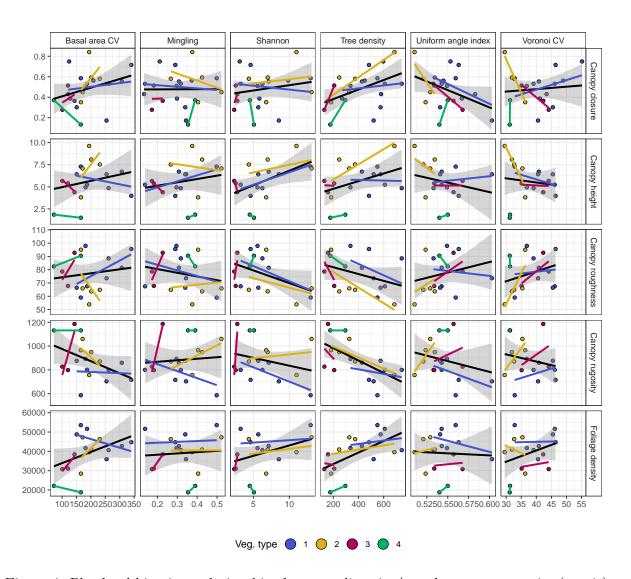


Figure 4: Plot level bivariate relationships between diversity/stand structure metrics (x axis) and canopy complexity metrics (y axis). Points and linear model lines of best fit are coloured by vegetation type. Black lines of best fit are linear models including all plots, with a 95% confidence interval. See Table 6 for a comparison of linear model fits by vegetation type.

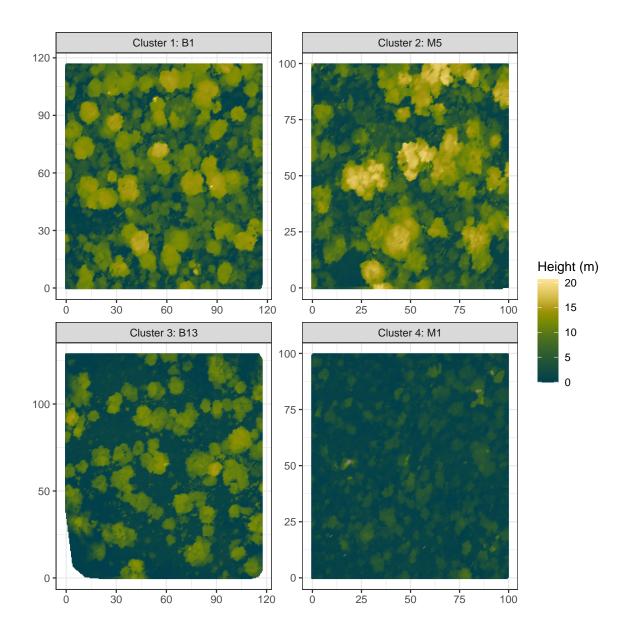


Figure 5: Representative canopy surface models for each vegetation type identified in the Non-metric Multi-dimensional Scaling (NMDS) clustering analysis. Panel titles show the plot name and the vegetation type.

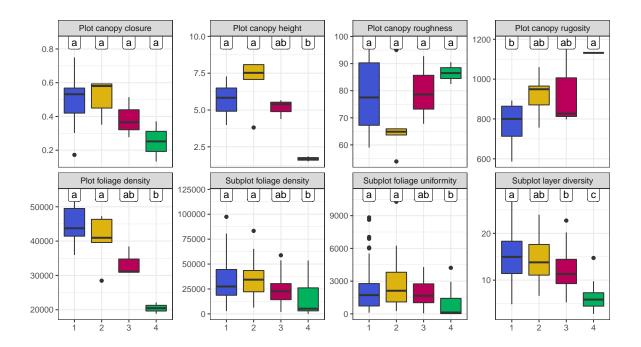


Figure 6: Box plots showing variation in canopy complexity metrics among the four vegetation types identified in the Non-metric Multi-dimensional Scaling (NMDS) clustering analysis. Thick lines show the median, boxes show the interquartile range (IQR), whiskers show $1.5 \times IQR$, and points show outliers beyond these limits. Labels above each box plot group vegetation types according to significant differences in pairwise Tukey's tests; vegetation types sharing a letter are not significantly different.

effects on all measured canopy complexity metrics, though these effects were non-significant for vegetation clusters 3 and 4. Heterogeneity of stem basal area had a significant positive effect on layer diversity and foliage density, but there was wide variation in vegetation type marginal effects for Clusters 3 and 4, due to low levels of replication. Cluster 3 had strong positive effects of species diversity on foliage uniformity and layer diversity. The random effects of vegetation type and plot identity described most of the variation in layer diversity and foliage density. Foliage uniformity was poorly explained by all combinations of fixed effects, with the best model only explaining 29%. All models were better than random effects only models according to AIC values.

3.4 Whole-plot multivariate linear models

While species diversity had varying effects on different plot level canopy complexity metrics, the confidence intervals on these effect sizes were wide (Figure 8). Species diversity had a significant positive effect on canopy height (β =3±0.96, p<0.05), a non-significant positive effect on canopy closure (β =0.07±0.085, p=0.41), but a negative effect on canopy surface roughness (β =-13±6.8, p=0.09) and whole canopy rugosity (β =-111±71, p=0.15). Spatial mingling of tree species had a positive effect on canopy surface roughness and canopy rugosity, but a strong negative effect on canopy height. Plot tree density had negligible effects on canopy complexity, except for canopy rugosity (β =-61±42, p=0.17), in contrast to the effect of Hegyi crowding on subplot canopy complexity. Spatially explicit measures of structural diversity, measured by the uniform angle index, Voronoi cell area CV, and basal area CV, had smaller effects on canopy complexity than species diversity, which were generally insignificant. One exception was the effect of uniform angle index, i.e. the spatial clustering of stems, on canopy closure, which was clearly negative,

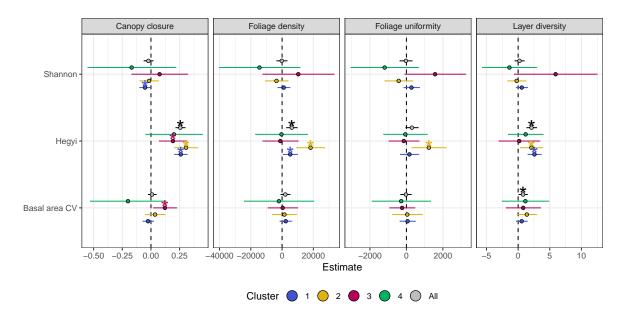


Figure 7: Standardised fixed effect slopes for each model of a canopy complexity metric. Slope estimates where the interval (± 1 standard error) does not overlap zero are considered to be significant effects, marked with asterisks. Points are coloured according to vegetation type.

though still insignificant (β =-0.08±0.043, p=0.1), the effect of Voronoi cell area CV on foliage density, which was positive (β =6199±3312, p=0.09), and the effect of basal area CV on canopy closure, which was positive (β =0.06±0.042, p=0.19).

Despite the weak effect sizes of species diverity on canopy complexity at the plot level, model selection showed that foliage density, canopy height and canopy roughness were better explained by models which included species diversity (Table 5). Additionally, the best models for canopy height and canopy roughness also included spatial mingling of tree species. The model for canopy roughness was only maginally better than a null model and the model did not have a significant p-value.

344 3.5 Path analysis

The subplot level path analysis investigating the indirect effect of subplot species diversity on canopy closure via the basal area CV showed that while species diversity had a strong positive significant effect on basal area variation, the effect of basal area variation on canopy closure remained negligible (Figure 9). The indirect effect of species diversity on canopy closure via basal area CV was -0.0016, while the direct effect was -0.078. The R² of this model was 0.47. As in the bivariate relationships and plot level linear models, species diversity had a weak positive significant effect on canopy closure, while the major driver of canopy closure was the Hegyi crowding index.

The plot level path analysis, which tested the effects of species diversity and species mingling on canopy height, showed that the main effect of species diversity on canopy height was direct (1.3*), while the indirect effects via basal area CV (0.0210), and tree density (-0.0294), remained small and insignificant. Shannon diversity had a strong positive effect on tree density. Species mingling had a moderately strong negative but insignificant direct effect on canopy height, as in the linear mixed models.

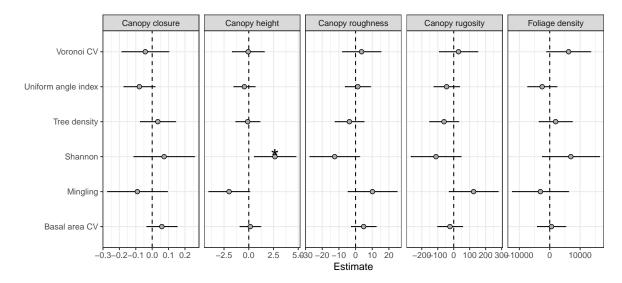


Figure 8: Standardised effect sizes for whole-plot canopy rugosity. Slope estimates where the interval (± 1 standard error) does not overlap zero are considered to be significant effects, marked with asterisks.

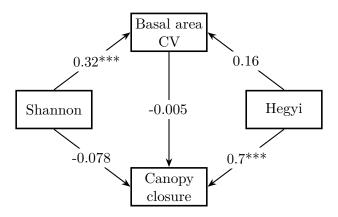


Figure 9: Directed Acyclic Graph showing standardised path coefficients of paths in the path analysis of the indirect effect of subplot species diversity (Shannon diversity index) on canopy closure via basal area CV. Asterisks define p-value thresholds: *<0.05, **<0.01, ***<0.001.

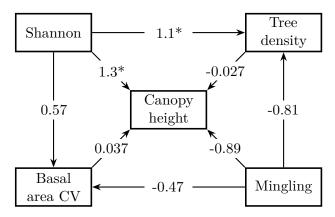


Figure 10: Directed Acyclic Graph showing standardised path coefficients of paths in the path analysis of the indirect effect of plot species diversity (Shannon diversity index) and species mingling on mean canopy height via stand structural metrics of basal area CV and tree density. Asterisks define p-value thresholds: *<0.05, **<0.01, ***<0.001.

3.6 Covariance of subplot and plot measures of canopy complexity

Plot and subplot canopy complexity metrics were highly correlated in many cases, with similar relationships among vegetation types (Figure 8). Most subplot and plot level canopy metrics covaried in a predictable manner. For example, increased canopy height led to an increase in canopy closure. Plot canopy height especially, tended to be strongly positively correlated with subplot canopy complexity metrics. Additionally, as canopy rugosity increased, many subplot canopy complexity and density metrics decreased. Subplot metrics varied greatly within plots, producing large uncertainty in plot level estimates of these metrics. All subplot level canopy complexity metrics positive correlated with each other (Figure 13). Plot level canopy complexity also generally correlated (Figure 12). Plot level measures of spatial heterogeneity in canopy structure, i.e. canopy surface roughness and canopy rugosity, were negatively correlated with measures of canopy density, i.e. foliage density, canopy closure, and canopy height. Measures of canopy spatial heterogeneity positively correlated with each other, as did measures of canopy density.

4 Discussion

This study investigated relationships between tree species diversity, stand structure, and several metrics of tree canopy complexity using terrestrial LiDAR in southern African savannas, with a view to improving understanding of the biotic drivers of variation in canopy complexity and vegetation dynamics. Species diversity appeared to generally have weak positive effects on canopy complexity metrics related to canopy density at both the subplot and plot scales. Plots with greater species diversity produced taller tree canopies, with greater canopy closure and foliage density. Species diversity generally had negative effects on canopy complexity metrics related to the spatial heterogeneity of foliage distribution. Species diversity caused a decrease in canopy surface roughness and canopy rugosity. The study did not, however, find support for the hypothesis that increased heterogeneity in tree stem size causes an increase in canopy complexity, and only partial support for the hypothesis that greater heterogeneity in stem location causes increased canopy complexity.

The result that species diversity increases canopy density suggests that woody encroachment may occur more readily in more diverse stands. Bivariate relationships show that some of this observed species diversity effect may be driven by vegetation type, and the variation in species diversity among vegetation types. The linear mixed model framework, which accounted for differences among vegetation types, still shows weak species diversity effects even after vegetation type is controlled for, strengthening the validity of the result. Species diversity is generally greater where resource availability is higher (), and is often reduced by human disturbances such as timber extraction (). Thus, in southern African savannas, it might be expected that woody encroachment may occur in areas of higher rainfall. The conflation of disturbance by fire and herbivory and rainfall complicates this prediction however. As rainfall increases in mesic savannas, so does fire frequency and intensity, limiting woody encroachment ().

Some aspects of stand structural diversity, such as variation in Voronoi cell area produced by spatial clustering of stems, are both the results of disturbance regime, and at the same time were found in this study to affect canopy complexity, which is expected to affect disturbance regime through its effect on understorey light environment (). Canopy complexity may therefore constitute a key mechanism by which the positive feedback between disturbance and understorey fuel generation is moderated. I suggest that future studies investigate more deeply the role of canopy complexity on understorey grassy fuel generation, and how this is affected by tree species composition and diversity.

Path analysis showed that indirect effects of species diversity on canopy complexity via stand

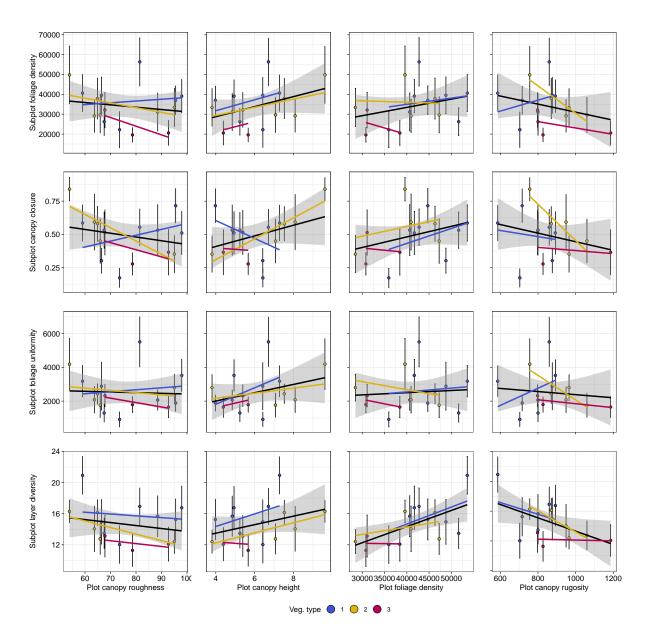


Figure 11: Bivariate plots comparing canopy structural metrics at the plot (x axis) and subplot scale (y axis). Each point represents the mean values of a single plot. Points and linear model fits are coloured according to vegetation type. The black linear model combines all vegetation types. Error bars on points are the standard deviation of mean subplot metrics across the plot. Note that because plot level canopy closure is calculated as the mean of subplot canopy closure, a comparison of subplot and plot canopy closure is not made in this figure.

structure were negligible, due to the lack of an effect of stand structure on canopy complexity at 406 both the subplot and plot scale. This finding suggests that the effects of species diversity on 407 canopy complexity are due to differences among species in their canopy dimensions and growth 408 strategy, rather than simply covarying with heterogeneity of tree stem size, and stregenthens 409 support for the hypothesis that species diversity causes increased canopy complexity. Previous 410 studies in temperate and boreal forests have suggested that increasing stand structural diversity 411 through active management and thinning can offset productivity losses caused by reduced species 412 diversity (Levick et al., 2009), but this study suggests that in southern African savannas at least, 413 this is not the case. Shannon diversity did however, have strong positive effects on basal area 414 CV and tree density. This can be interpreted as a niche complementarity effect, where diverse 415 savannas are able to support a greater density of trees by reducing the effects of intraspecific 416 competition and promoting canopy occupation in different vertical layers. Although basal area CV was included in best models for subplot layer diversity and foliage density, subplot level 418 path analysis suggests that covariance between basal area CV and species diversity, and the 419 strong direct effect of species diversity on canopy complexity, means that any observed effect of 420 basal area CV on subplot canopy complexity is actually due to species diversity rather than 421 basal area CV itself. 422

The standardised effect sizes of species diversity on canopy complexity metrics were generally greater at the plot level than at the subplot level. While positive and significant relationships between species diversity and subplot canopy complexity metrics were observed in the subplot bivariate models, subplot linear mixed effects models did not show strong species diversity effects, and were instead dominated by the effect of crowding. This finding suggests a large degree of stochastic variability in canopy complexity within plots, that masks species effects at smaller spatial scales. The prevalence of disturbance events such as fire and damage by elephants in southern African woodlands, as well as tree-fall, small-scale variability in edaphic factors, and stochastic tree germination all contribute to heterogeneity in canopy complexity (). While disturbances are controlled to some extent by stand structure and composition, due to the stochastic nature of disturbance events, a snapshot study such as this cannot capture the average disturbance regime and there is therefore a great deal of noise in measurements of canopy complexity and stand structure. The contrast in strength of species diversity effects at the subplot and plot level demonstrates the importance of large sample units, a high degree of spatial replication, and ideally a longer time scale when measuring canopy complexity, especially in disturbed systems, to effectively account for the inherent heterogeneity in the system ().

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In bivariate relationships tree density had a strong effect on canopy complexity metrics at the plot level, but did not feature prominently in the best quality mixed models for those metrics. Tree density covaried with basal area CV, with denser plots also exhibiting greater variation in tree size. This is also reflected in the bivariate relationship between tree density and canopy height. Further path analysis showed that tree species diversity allowed greater tree density and variation in tree size, but this did not translate into a positive effect of tree density or basal area CV on canopy height.

While species diversity caused increases in canopy complexity metrics related to canopy density, it caused a decrease in those related to the spatial heterogeneity of canopy foliage, particularly at the plot scale.

This study has raised concerns about the suitability of some canopy complexity metrics for characterising sparse savanna canopies. Metrics related to canopy foliage heterogeneity (i.e. canopy roughness and canopy rugosity), were highly sensitive to variation in canopy density.

Despite tree density having strong positive observed effects on canopy density, and negative effects on canopy heterogeneity in bivariate relationships, tree density only appeared in the best quality mixed model for canopy rugosity, and the effect sizes for tree density in maximal mixed models were small. Tree density was shown to covary with shannon diversity, basal area

CV and Voronoi cell area CV. This covariance may have led to the effect dropping out in the best models. Despite many explanatory variables showing positive relationships with canopy complexity metrics in bivariate plots, in the model selection many dropped out. This is because many explanatory variables covaried in the analysis.

Spatial regularity of stems, measured using the uniform angle index, caused a clear decrease 460 in canopy closure, with similar behaviour across vegetation types. Uniform angle index was 461 also included in the best multivariate model predicting canopy closure. Concurrently, spatial 462 clustering of stems, measured by Voronoi cell area CV was included in the best model for 463 foliage density but had a positive effect on this canopy complexity metric. This finding is 464 expected, as spatial clustering results in reduced canopy cover in areas outside clusters, and 465 a non-compensatory increase in canopy closure within clusters, due to competition among 466 individuals (). In contrast, changes in spatial regularity of trees do not imply changes in the 467 distance of stems, only their layout, but does spatial regularity does covary with spatial clustering 468 Clustering of trees in savannas can result from positive feedback effects from disturbance by fire 469 and herbivory ().

Stand structural diversity caused positive canopy complexity effects for within-canopy structural 471 metrics such as layer diversity and canopy surface roughness, but had negligible effects on canopy 472 density. This is in line with other studies in forest ecosystems, which report that variation in 473 tree size increases total canopy volume occupancy by increasing the number of canopy layers, 474 but does not necessarily result in a concomitant increase in canopy closure, as the resulting 475 canopies are often more sparse, especially for understorey individuals (). The path analysis 476 also supports this conclusion, where species diversity was found to cause an increase in stand 477 structural diversity, but this did not extend to an increase in canopy closure. 478

The effect of stand structure on canopy complexity in this system appears to be a result of demographic effects rather than variation in growth form as a function of species diversity. The path analysis testing the indirect effect of species diversity on canopy closure via stand structural diversity did not find a significant indirect effect of species diversity on canopy closure. While other studies in forests have found a species diversity effect on stand structural diversity (), it is suggested here that prevailing disturbance pressures mask any species diversity effect.

Canopy complexity metrics differed among vegetation types, but significant differences among vegetation types only occurred in a few cases. Canopy density and total foliage volume was lowest in ex-Acacia plots, as expected, while the highest canopy density occurred in miombo plots in clusters 1 and 2. Miombo woodlands frequently have contiguous canopies with overlapping individual tree canopies (), while ex-Acacia savannas show greater negative density dependence of individuals, forming patchy canopies with simpler vertical profiles ().

While vegetation types differed in mean values for stand structural and species diversity metrics, variation in these metrics produced results of similar direction and magnitude among vegetation types in most cases. Small sample sizes for *Baikiaea* and ex-Acacia vegetation however, led to wide errors on most relationships especially at the plot level, such that it is impossible to draw deeper conclusions about the behaviour of these vegetation types. Variation in mean values of canopy complexity metrics among vegetation types is likely driven by species identity (), though species composition itself is driven by environmental factors and disturbance regime ().

At the subplot level the Hegyi crowding index was a key driver of canopy complexity. Crowding of stems at the subplot level correlates with both tree density and spatial clustering of stems at the plot level. Hegyi crowding is expected to positively correlate with resource availability, and negatively correlate with disturbance which causes tree mortality and reduces tree growth.

All subplot canopy complexity metrics positively covaried. For rapid assessments of canopy complexity, simple canopy closure measurements are sufficient to characterise canopy complexity, at the spatial scale of an individual tree neighbourhood. In closed-canopy forest ecosystems,

measures of canopy spatial heterogeneity often correlate with canopy closure (). In the savannas studied here however, the sparser and discontinuous canopy results in a negative relationship. MORE

508 5 Conclusion

References

- Archibald, S. & W. J. Bond (2003). 'Growing tall vs growing wide: tree architecture and allometry of Acacia karroo in forest, savanna, and arid environments'. In: *Oikos* 102.1, pp. 3–14. DOI: 10.1034/j.1600-0706.2003.12181.x.
- Axelsson, C. R. & N. P. Hanan (2018). 'Rates of woody encroachment in African savannas reflect water constraints and fire disturbance'. In: *Journal of Biogeography* 45.6, pp. 1209–1218. DOI: 10.1111/jbi.13221.
- Baldocchi, D. D. & K. B. Wilson (2001). 'Modeling CO2 and water vapor exchange of a temperate
 broadleaved forest across hourly to decadal time scales'. In: *Ecological Modelling* 142.1-2,
 pp. 155–184. DOI: 10.1016/s0304-3800(01)00287-3.
- Barry, K. E., L. Mommer, J. van Ruijven, C. Wirth, A. J. Wright, Y. Bai, J. Connolly, G. B. D. Deyn, H. de Kroon, F. Isbell et al. (2019). 'The Future of Complementarity: Disentangling Causes from Consequences'. In: *Trends in Ecology & Evolution* 34.2, pp. 167–180. DOI: 10.1016/j.tree.2018.10.013.
- Bond, W. J. & G. F. Midgley (2012). 'Carbon dioxide and the uneasy interactions of trees and savannah grasses'. In: *Philosophical Transactions of the Royal Society B: Biological Sciences* 367.1588, pp. 601–612. DOI: 10.1098/rstb.2011.0182.
- Buitenwerf, R., W. J. Bond, N. Stevens & W. S. W. Trollope (2012). 'Increased tree densities in South African savannas: >50 years of data suggests CO₂ as a driver'. In: Global Change Biology 18.2, pp. 675–684. DOI: 10.1111/j.1365-2486.2011.02561.x.
- Calders, K., J. Adams, J. Armston, H. Bartholomeus, S. Bauwens, L. P. Bentley, J. Chave, F. M. Danson, M. Demol, M. Disney et al. (2020). 'Terrestrial laser scanning in forest ecology: Expanding the horizon'. In: *Remote Sensing of Environment* 251, p. 112102. DOI: 10.1016/j.rse.2020.112102.
- Charles-Dominique, T., G. F. Midgley, K. W. Tomlinson & W. J. Bond (2018). 'Steal the
 light: shade vs fire adapted vegetation in forest-savanna mosaics'. In: New Phytologist 218.4,
 pp. 1419–1429. DOI: 10.1111/nph.15117.
- Chen, J. M., G. Mo, J. Pisek, J. Liu, F. Deng, M. Ishizawa & D. Chan (2012). 'Effects of foliage clumping on the estimation of global terrestrial gross primary productivity'. In: Global Biogeochemical Cycles 26, GB1019. DOI: 10.1029/2010gb003996.
- Cifuentes, R., D. V. der Zande, J. Farifteh, C. Salas & P. Coppin (2014). 'Effects of voxel size
 and sampling setup on the estimation of forest canopy gap fraction from terrestrial laser
 scanning data'. In: Agricultural and Forest Meteorology 194, pp. 230–240. DOI: 10.1016/j.
 agrformet.2014.04.013.
- Criado, M. G., I. H. Myers-Smith, A. D. Bjorkman, C. E. R. Lehmann & N. Stevens (2020).
 'Woody plant encroachment intensifies under climate change across tundra and savanna biomes'.
 In: Global Ecology and Biogeography 29.5, pp. 925–943. DOI: 10.1111/geb.13072.
- Dohn, J., D. J. Augustine, N. P. Hanan, J. Ratnam & M. Sankaran (2017). 'Spatial vegetation
 patterns and neighborhood competition among woody plants in an East African savanna'. In:
 Ecology 98.2, pp. 478–488. DOI: 10.1002/ecy.1659.
- Donohue, R. J., M. L. Roderick, T. R. McVicar & G. D. Farquhar (2013). 'Impact of CO₂
 fertilization on maximum foliage cover across the globe's warm, arid environments'. In:
 Geophysical Research Letters 40.12, pp. 3031–3035. DOI: 10.1002/grl.50563.

- Ehbrecht, M., P. Schall, J. Juchheim, C. Ammer & D. Seidel (2016). 'Effective number of layers:

 A new measure for quantifying three-dimensional stand structure based on sampling with
 terrestrial LiDAR'. In: Forest Ecology and Management 380, pp. 212–223. DOI: 10.1016/j.
 foreco.2016.09.003.
- Fotis, A. T., T. H. Morin, R. T. Fahey, B. S. Hardiman, G. Bohrer & P. S. Curtis (2018).

 'Forest structure in space and time: Biotic and abiotic determinants of canopy complexity and
 their effects on net primary productivity'. In: Agricultural and Forest Meteorology 250-251,
 pp. 181–191. DOI: 10.1016/j.agrformet.2017.12.251.
- Frost, P. (1996). 'The ecology of miombo woodlands'. In: The miombo in transition: woodlands
 and welfare in Africa. Ed. by B. Campbell. Bogor, Indonesia: Center for International Forestry
 Research, pp. 11–55.
- Gough, C. M., J. W. Atkins, R. T. Fahey & B. S. Hardiman (2019). 'High rates of primary production in structurally complex forests'. In: *Ecology* 100.10. DOI: 10.1002/ecy.2864.
- Hardiman, B. S., G. Bohrer, C. M. Gough, C. S. Vogel & P. S. Curtis (2011). 'The role of canopy
 structural complexity in wood net primary production of a maturing northern deciduous
 forest'. In: *Ecology* 92.9, pp. 1818–1827. DOI: 10.1890/10-2192.1.
- Hegyi, F. (1974). 'A simulation model for managing jack-pine stands'. In: Royal College of
 Forestry, editor. Stockholm, Sweden: Royal College of Forestry, pp. 74–90.
- Hirota, M., M. Holmgren, E. H. Van Nes & M. Scheffer (2011). 'Global resilience of tropical forest and savanna to critical transitions'. In: *Science* 334, pp. 232–235. DOI: 10.1126/science. 1210657.
- Jonckheere, I., S. Fleck, K. Nackaerts, B. Muys, P. Coppin, M. Weiss & F. Baret (2004). 'Review of methods for in situ leaf area index determination'. In: *Agricultural and Forest Meteorology* 121.1-2, pp. 19–35. DOI: 10.1016/j.agrformet.2003.08.027.
- Jost, L. (2006). 'Entropy and diversity'. In: *Oikos* 113.2, pp. 363–375. doi: 10.1111/j.2006. 0030-1299.14714.x.
- Jucker, T., O. Bouriaud & D. A. Coomes (2015). 'Crown plasticity enables trees to optimize canopy packing in mixed-species forests'. In: Functional Ecology 29.8, pp. 1078–1086. DOI: 10.1111/1365-2435.12428.
- Kershaw, J. A., M. J. Ducey, T. W. Beers & B. Husch (2017). Forest Mensuration. Chichester,
 UK: John Wiley & Sons. ISBN: 9781118902035.
- Khosravipour, A., A. K. Skidmore, M. Isenburg, T. Wang & Y. A. Hussin (2014). 'Generating Pit-free Canopy Height Models from Airborne LiDAR'. In: *Photogrammetric Engineering & Remote Sensing* 80.9, pp. 863–872. DOI: 10.14358/pers.80.9.863.
- Körner, C. (2017). 'A matter of tree longevity'. In: *Science* 355.6321, pp. 130–131. DOI: 10.1126/ssr science.aal2449.
- Law, B. E., A. Cescatti & D. D. Baldocchi (2001). 'Leaf area distribution and radiative transfer in open-canopy forests: implications for mass and energy exchange'. In: *Tree Physiology* 21.12-13, pp. 777-787. DOI: 10.1093/treephys/21.12-13.777.
- Lefcheck, J. S. (2016). 'piecewiseSEM: Piecewise structural equation modeling in R for ecology, evolution, and systematics'. In: *Methods in Ecology and Evolution* 7.5, pp. 573–579. DOI: 10.1111/2041-210X.12512.
- Levick, S. R., G. P. Asner, T. Kennedy-Bowdoin & D. E. Knapp (2009). 'The relative influence of fire and herbivory on savanna three-dimensional vegetation structure'. In: *Biological Conservation* 142.8, pp. 1693–1700. DOI: 10.1016/j.biocon.2009.03.004.
- Lowman, M. D. & H. B. Rinker (2004). Forest Canopies. Physiological Ecology. Burlington MA, USA: Elsevier Science. ISBN: 9780080491349.
- Mitchard, E. T. A. & C. M. Flintrop (2013). 'Woody encroachment and forest degradation in sub-Saharan Africa's woodlands and savannas 1982-2006'. In: *Philosophical Transactions of the* Royal Society B: Biological Sciences 368.1625, p. 20120406. DOI: 10.1098/rstb.2012.0406.

- Morin, X. (2015). 'Species richness promotes canopy packing: a promising step towards a better understanding of the mechanisms driving the diversity effects on forest functioning'. In:

 Functional Ecology 29.8, pp. 993–994. DOI: 10.1111/1365-2435.12473.
- Mugasha, W. A., O. M. Bollandsås & T. Eid (2013). 'Relationships between diameter and height
 of trees in natural tropical forest in Tanzania'. In: Southern Forests: a Journal of Forest
 Science 75.4, pp. 221–237. DOI: 10.2989/20702620.2013.824672.
- Muir, J., S. Phinn, T. Eyre & P. Scarth (2018). 'Measuring plot scale woodland structure using
 terrestrial laser scanning'. In: Remote Sensing in Ecology and Conservation 4.4, pp. 320–338.
 DOI: 10.1002/rse2.82.
- Muumbe, T. P., J. Baade, J. Singh, C. Schmullius & C. Thau (2021). 'Terrestrial Laser Scanning
 for Vegetation Analyses with a Special Focus on Savannas'. In: Remote Sensing 13.3, p. 507.
 DOI: 10.3390/rs13030507.
- Nakagawa, S. & I. C. Cuthill (2007). 'Effect size, confidence interval and statistical significance: a practical guide for biologists'. In: *Biological Reviews* 82.4, pp. 591–605. DOI: 10.1111/j.1469-185x.2007.00027.x.
- Ong, M. S., Y. C. Kuang & M. P.-L. Ooi (2012). 'Statistical measures of two dimensional point set uniformity'. In: *Computational Statistics & Data Analysis* 56.6, pp. 2159–2181. DOI: 10.1016/j.csda.2011.12.005.
- Panzou, G. J. L., A. Fayolle, T. Jucker, O. L. Phillips, S. Bohlman, L. F. Banin, S. L. Lewis,
 K. Affum-Baffoe, L. F. Alves, C. Antin et al. (2020). 'Pantropical variability in tree crown
 allometry'. In: Global Ecology and Biogeography 30.2, pp. 459–475. DOI: 10.1111/geb.13231.
- Persistence of Vision Pty. Ltd. (2004). Persistence of Vision Raytracer (Version 3.7). [Computer software].
- Pretzsch, H. (2014). 'Canopy space filling and tree crown morphology in mixed-species stands compared with monocultures'. In: Forest Ecology and Management 327, pp. 251–264. DOI: http://dx.doi.org/10.1016/j.foreco.2014.04.027.
- Privette, J., Y. Tian, G. Roberts, R. Scholes, Y. Wang, K. Caylor, P. Frost & M. Mukelabai (2004).
 'Vegetation structure characteristics and relationships of Kalahari woodlands and savannas'.
 In: Global Change Biology 10.3, pp. 281–291. DOI: 10.1111/j.1365-2486.2004.00740.x.
- Ratcliffe, S., C. Wirth, T. Jucker, F. van der Plas, M. Scherer-Lorenzen, K. Verheyen, E. Allan, R. Benavides, H. Bruelheide, B. Ohse et al. (2017). 'Biodiversity and ecosystem functioning relations in European forests depend on environmental context'. In: *Ecology Letters* 20, pp. 1414–1426. DOI: http://dx.doi.org/10.1111/ele.12849.
- Reich, P. B., S. E. Hobbie & T. D. Lee (2014). 'Plant growth enhancement by elevated CO2 eliminated by joint water and nitrogen limitation'. In: *Nature Geoscience* 7.12, pp. 920–924.

 DOI: 10.1038/ngeo2284.
- Ribeiro, N. S., P. L. Silva de Miranda & J. Timberlake (2020). 'Biogeography and Ecology of Miombo Woodlands'. In: *Miombo Woodlands in a Changing Environment: Securing the Resilience and Sustainability of People and Woodlands*. Ed. by N. S. Ribeiro, Y. Katerere, P. W. Chirwa & I. M. Grundy. Springer International Publishing, pp. 9–53. DOI: 10.1007/978-3-030-50104-4_2.
- Rusu, R. B., Z. C. Marton, N. Blodow, M. Dolha & M. Beetz (2008). 'Towards 3D Point cloud based object maps for household environments'. In: *Robotics and Autonomous Systems* 56.11, pp. 927–941. DOI: 10.1016/j.robot.2008.08.005.
- Sankaran, M., N. P. Hanan, R. J. Scholes, J. Ratnam, D. J. Augustine, B. S. Cade, J. Gignoux,
 S. I. Higgins, X. Le Roux, F. Ludwig et al. (2005). 'Determinants of woody cover in African savannas'. In: *Nature* 438.8, pp. 846–849. DOI: http://dx.doi.org/10.1038/nature04070.
- Scheuermann, C. M., L. E. Nave, R. T. Fahey, K. J. Nadelhoffer & C. M. Gough (2018). 'Effects
 of canopy structure and species diversity on primary production in upper Great Lakes forests'.
 In: Oecologia 188.2, pp. 405–415. DOI: 10.1007/s00442-018-4236-x.

- Scholes, R. J. & S. R. Archer (1997). 'Tree grass interactions in savannas'. In: Annual Review of
 Ecology and Systematics.
- Seidel, D., S. Fleck & C. Leuschner (2012). 'Analyzing forest canopies with ground-based
 laser scanning: A comparison with hemispherical photography'. In: Agricultural and Forest
 Meteorology 154-155, pp. 1–8. DOI: 10.1016/j.agrformet.2011.10.006.
- Seidel, D., S. Fleck, C. Leuschner & T. Hammett (2011). 'Review of ground-based methods to measure the distribution of biomass in forest canopies'. In: *Annals of Forest Science* 68.2, pp. 225–244. DOI: 10.1007/s13595-011-0040-z.
- Shenkin, A., L. P. Bentley, I. Oliveras, N. Salinas, S. Adu-Bredu, B. H. Marimon-Junior, B. S.
 Marimon, T. Peprah, E. L. Choque, L. T. Rodriguez et al. (2020). 'The Influence of Ecosystem and Phylogeny on Tropical Tree Crown Size and Shape'. In: Frontiers in Forests and Global Change 3. DOI: 10.3389/ffgc.2020.501757. URL: https://doi.org/10.3389%2Fffgc.2020.501757.
- Sitch, S., P. Friedlingstein, N. Gruber, S. D. Jones, G. Murray-Tortarolo, A. Ahlström, S. C.
 Doney, H. Graven, C. Heinze, C. Huntingford et al. (2015). 'Recent trends and drivers of
 regional sources and sinks of carbon dioxide'. In: *Biogeosciences* 12.3, pp. 653–679. DOI:
 10.5194/bg-12-653-2015.
- Solbrig, O. T., E. Medina & J. F. Silva (1996). Biodiversity and Savanna Ecosystem Processes.
 Berlin, Germany: Springer-Verlag.
- Stark, S. C., B. J. Enquist, S. R. Saleska, V. Leitold, J. Schietti, M. Longo, L. F. Alves, P. B.
 Camargo & R. C. Oliveira (2015). 'Linking canopy leaf area and light environments with tree
 size distributions to explain Amazon forest demography'. In: *Ecology Letters* 18.7, pp. 636–645.
 DOI: 10.1111/ele.12440.
- Staver, A. C. & S. E. Koerner (2015). 'Top-down and bottom-up interactions determine tree and herbaceous layer dynamics in savanna grasslands'. In: *Trophic Ecology: Bottom-up and Top-Down Interactions Across Aquatic and Terrestrial Systems*. Ed. by K. J. La Pierre & T. C. Hanley. Cambridge, United Kingdom: Cambridge University Press, pp. 86–106.
- Stevens, N., C. E. R. Lehmann, B. P. Murphy & G. Durigan (2016). 'Savanna woody encroachment is widespread across three continents'. In: *Global Change Biology* 23.1, pp. 235–244. DOI: 10.1111/gcb.13409.
- ter Steege, H. (2018). Hemiphot.R: Free R scripts to analyse hemispherical photographs for canopy openness, leaf area index and photosynthetic active radiation under forest canopies. Unpublished report. Leiden, The Netherlands: Naturalis Biodiversity Center. URL: https://github.com/Naturalis/Hemiphot.
- von Gadow, K. & G. Hui (2002). Characterising forest spatial structure and diversity. Ed. by
 L. Bjoerk. Lund, Sweden, pp. 20–30.
- White, F. (1983). The Vegetation of Africa: A descriptive memoir to accompany the UN ESCO/AETFAT/UNSO vegetation map of Africa. Paris, France: UNESCO. DOI: 10.2307/
 2260340.
- Wright, A. J., W. D. A. Wardle, W. R. Callaway & A. Gaxiola (2017). 'The overlooked role of facilitation in biodiversity experiments'. In: Trends in Ecology & Evolution 32, pp. 383–390.
 DOI: 10.1016/j.tree.2017.02.011.
- Zhang, K., S.-C. Chen, D. Whitman, M.-L. Shyu, J. Yan & C. Zhang (2003). 'A progressive morphological filter for removing nonground measurements from airborne LIDAR data'. In:
 IEEE Transactions on Geoscience and Remote Sensing 41.4, pp. 872–882. DOI: 10.1109/tgrs.
 2003.810682.

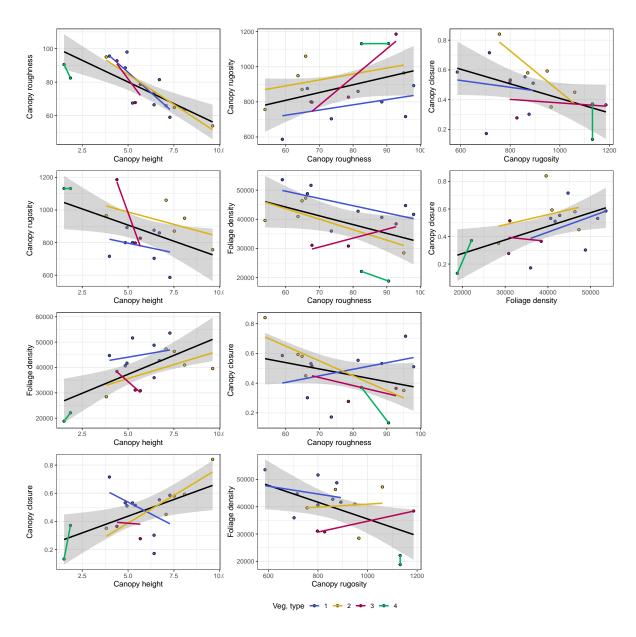


Figure 12: Bivariate scatter plots of plot level canopy complexity metrics.

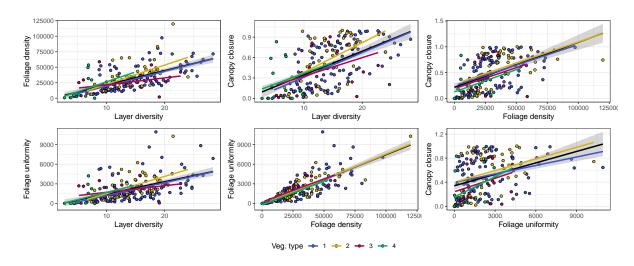


Figure 13: Bivariate scatter plots of subplot level canopy complexity metrics.

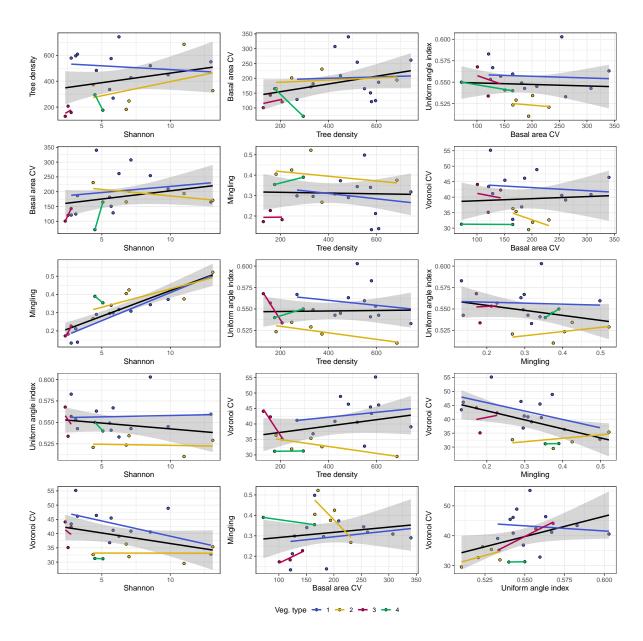


Figure 14: Bivariate scatter plots of plot level diversity and stand structural metrics.

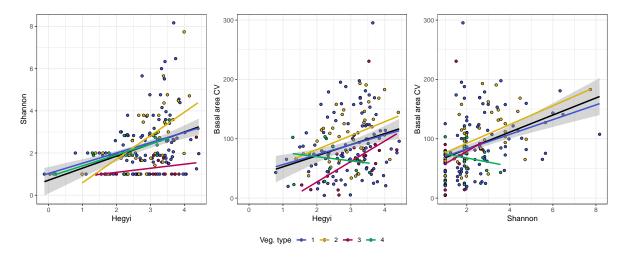


Figure 15: Bivariate scatter plots of subplot level diversity and stand structural metrics.

Table 6: Summary statistics of bivariate linear models comparing canopy complexity metrics with diversity and stand structural metrics, grouped by vegetation type. Note that models plot level canopy complexity metrics could not be fitted for Cluster 4, as this cluster only contained two plots. Slope refers to the slope of the predictor term in the model, \pm 1 standard error. T is the t-value of the slope of the predictor term in the model, Asterisks indicate the p-value of these terms (***<0.001, **<0.01, *<0.05).

Foliage density	Response	Predictor	Cluster	Slope	F	\mathbb{R}^2	T
Foliage density Basal area CV				$7.3e + 01 \pm 3.7e + 01$, , ,		1.99*
Foliage density Hegyi	Foliage density	Rasal area CV		$1.1e + 02 \pm 7.9e + 01$	2.1(2,38)	0.05	1.44
Foliage density Hegyi	ronage density	Dasar area C v	3	$1.4e + 01 \pm 7.2e + 01$	0.0(2,14)	0.00	0.20
Foliage density Hegyi 2 1.4e+04±3.6e+03 15.2(2,40) 0.28 3.90*** 3 6.6e+03±3.0e+03 4.8(2,23) 0.17 2.18** 4 1.5e+01±5.5e+03 0.0(2,13) 0.00 0.00 Foliage density Shannon 1 2.2e+03±1.3e+03 2.8(2,102) 0.03 1.67 4 2.2e+03±1.3e+03 2.6(2,39) 0.06 1.61 3 1.1e+04±6.5e+03 3.1(2,20) 0.13 1.77 4 -6.5e+03±6.5e+03 1.0(2,13) 0.07 -1.01 Canopy closure Basal area CV 2 2.9e-03±1.1e-03 6.9(2,39) 0.15 2.62** 3 4.2e+03±1.1e-03 15.1(2,14) 0.52 3.89** 4 -4.6e+03±3.0e+03 2.2(2,12) 0.16 -1.50 Canopy closure Hegyi 2 2.6e+01±2.8e+02 62.3(2,102) 0.38 7.89*** 4 1.7e+01±8.0e+02 4.5(2,13) 0.26 2.12 Canopy closure Basal area CV 3 2.8e+01±4.0e+02 50.7(2,23) 0.69 7.12*** Canopy closure Foliage uniformity Basal area CV 3 2.3e+01±1.e01 0.4(2,13) 0.03 0.60 Foliage uniformity Basal area CV 3 2.5e+00±7.9e+00 0.4(2,14) 0.02 0.92 4 6.7e+02±1.1e-01 0.4(2,13) 0.03 0.60 Foliage uniformity Hegyi 1 2.2e+02±3.e+02 0.0(2,102) 0.01 0.92 4 0.9a-01±1.5e+01 0.0(2,12) 0.00 0.10.61 4 0.9a-01±1.5e+01 0.0(2,12) 0.00 0.06 Foliage uniformity Hegyi 1 2.2e+02±3.e+02 0.0(2,102) 0.01 0.98 Foliage uniformity Hegyi 2 7.5e+01±4.0e+02 0.9(2,37) 0.11 1.72 Foliage uniformity Shannon 1 2.2e+02±3.e+02 0.0(2,13) 0.00 0.09 Foliage uniformity Hegyi 1 2.2e+02±3.e+02 0.0(2,13) 0.00 0.09 Foliage uniformity Hegyi 1 2.3e+00±4.0e+00 0.4(2,14) 0.02 0.05 4 0.9a-01±1.5e+01 0.0(2,12) 0.00 0.06 Foliage uniformity Shannon 2 2.8e+00±7.4e+00 0.4(2,38) 0.01 0.61 4 0.9a-01±1.5e+01 0.0(2,12) 0.00 0.06 Foliage uniformity Hegyi 2 7.5e+01±4.0e+02 0.0(2,13) 0.00 0.09 Foliage uniformity Shannon 3 1.3e+03±5.1e+02 0.0(2,13) 0.00 0.01 Foliage uniformity Shannon 3 1.3e+03±5.1e+02 0.0(2,13) 0.00 0.01 Foliage uniformity Shannon 3 1.3e+03±5.1e+02 0.0(2,13) 0.00 0.03 8			4	$1.6e + 01 \pm 2.0e + 02$	0.0(2,12)	0.00	0.08
Foliage density Hegyi 3 6.6e+03±3.0e+03 4.8(2,23) 0.17 2.18* 4 1.5e+01±5.5e+03 0.0(2,13) 0.00 0.00 Foliage density Shannon 1 2.2e+03±1.3e+03 2.8(2,102) 0.03 1.67 2 3.8e+03±2.4e+03 2.6(2,39) 0.06 1.61 3 1.1e+04±6.5e+03 3.1(2,20) 0.13 1.77 4 -6.5e+03±6.5e+03 1.0(2,13) 0.07 -1.01 Canopy closure Basal area CV 2 2.9e+03±1.1e+03 6.9(2,39) 0.15 2.62* 3 4.2e+03±1.1e+03 6.9(2,39) 0.15 2.62* 4 -4.6e+03±3.0e+03 2.2(2,12) 0.16 -1.50 Canopy closure Hegyi 1 2.2e+01±2.8e+02 62.3(2,102) 0.38 7.89*** 2 2.6e+01±5.1e+02 27.0(2,41) 0.40 5.19*** 3 2.8e+01±4.0e+02 50.7(2,23) 0.69 7.12*** 4 1.7e+01±8.0e+02 4.5(2,13) 0.26 2.12 Canopy closure Foliage uniformity Basal area CV 2 1.1e+01±3.2e+02 12.1(2,40) 0.23 3.48** Foliage uniformity Hegyi 2 4.5e+00±7.4e+00 0.4(2,13) 0.03 0.60 Foliage uniformity Hegyi 2 7.5e+02±3.7e+00 0.4(2,14) 0.02 -0.59 4 9.3e+01±1.5e+01 0.0(2,12) 0.00 -0.06 Foliage uniformity Hegyi 2 7.5e+02±3.7e+00 0.4(2,14) 0.02 -0.59 4 9.3e+01±1.5e+01 0.0(2,12) 0.00 -0.06 Foliage uniformity Hegyi 2 7.5e+02±3.7e+02 0.0(2,102) 0.01 0.98 Foliage uniformity Hegyi 3 4.5e+02±2.6e+02 0.9(2,23) 0.01 1.7e 4 -7.5e+01±4.0e+02 0.0(2,13) 0.00 -0.06 Foliage uniformity Shannon 3 4.8e+02±2.6e+02 0.9(2,23) 0.11 1.7e 4 -7.5e+01±4.0e+02 0.0(2,13) 0.00 -0.09 Foliage uniformity Shannon 3 4.8e+02±2.6e+02 0.9(2,23) 0.11 1.7e 4 -7.5e+01±4.0e+02 0.0(2,13) 0.00 -0.09 Foliage uniformity Shannon 3 4.5e+02±2.6e+02 0.9(2,23) 0.11 1.7e 5 4 -7.5e+01±4.0e+02 0.0(2,13) 0.00 -0.09 Foliage uniformity Shannon 3 4.5e+02±2.6e+02 0.1(2,39) 0.00 0.38 3 1.3e+03±2.1e+02 0.1(2,39) 0.00 0.38 3 1.3e+03±2.1e+02 0.1(2,39) 0.00 0.38 4 0.5e+02±4.7e+02 0.1(2,39) 0.00 0.38 5 0.6e+01±2.2e+02 0.1(2,39) 0.00 0.38 5 0.3e+02±4.7e+02 0.1(2,39) 0.00 0.38 5 0.3e+02±4.7e+02 0.1(2,39) 0.00 0.38 5 0.3e+02±4.7e+02 0.1(2,39) 0.00 0.38 6 0.17 0.2e6** 6 0.2e02±1.4e+02 0.0(2,33) 0.11 1.7e 7 0.2e6** 6 0.2e02±1.4e+02 0.0(2,33) 0.11 1.7e 7 0.2e6** 8 0.2e02±1.4e+02 0.0(2,33) 0.11 1.7e 8 0.2e02±1.4e+02 0.0(2,33) 0.11 1.7e 8 0.2e02±1.4e+02 0.0(2,33) 0.11 1.7e 8 0.2e02±1.4e+02 0.0(2,33) 0.11 1			1	$5.9e + 03 \pm 2.1e + 03$	8.2(2,102)	0.07	
Solution	Foliago donsity	Hogwi		$1.4e + 04 \pm 3.6e + 03$	15.2(2,40)	0.28	3.90***
Foliage density Shannon $ \begin{array}{c} 1 & 2.2 e + 0.3 \pm 1.3 e + 0.3 & 2.8 (2,102) & 0.03 & 1.67 \\ 2 & 3.8 e + 0.3 \pm 2.4 e + 0.3 & 2.6 (2,39) & 0.06 & 1.61 \\ 3 & 1.1 e + 0.4 \pm 6.5 e + 0.3 & 3.1 (2,20) & 0.13 & 1.77 \\ 4 & -6.5 e + 0.3 \pm 6.5 e + 0.3 & 1.0 (2,13) & 0.07 & -1.01 \\ \end{array} $ $ \begin{array}{c} \text{Canopy closure} \\ \text{Basal area CV} \\ \end{array} \begin{array}{c} \text{Basal area CV} \\ \text{2} \\ 2.9 e - 0.3 \pm 1.1 e - 0.3 & 6.9 (2.39) & 0.15 & 2.62 * \\ 3 & 4.2 e - 0.3 \pm 1.1 e - 0.3 & 6.9 (2.39) & 0.15 & 2.62 * \\ 3 & 4.2 e - 0.3 \pm 1.1 e - 0.3 & 6.9 (2.39) & 0.15 & 2.62 * \\ 3 & 4.2 e - 0.3 \pm 1.1 e - 0.3 & 15.1 (2,14) & 0.52 & 3.89 * * \\ 4 & -4.6 e - 0.3 \pm 3.0 e - 0.3 & 2.2 (2,12) & 0.16 & -1.50 \\ \end{array} \\ \text{Canopy closure} \\ \text{Canopy closure} \\ \text{Canopy closure} \\ \text{Basal area CV} \\ \text{Basal area CV} \\ \text{2} \\ 2.2 e - 0.0 \pm 1.2 e - 0.0 & 2.7 (0.241) & 0.40 & 5.19 * * * \\ 2.2 e - 0.0 \pm 1.2 e - 0.0 & 2.7 (0.241) & 0.40 & 5.19 * * * \\ 4 & 1.7 e - 0.1 \pm 8.0 e - 0.2 & 5.0 7 (2.23) & 0.69 & 7.12 * * * \\ 4 & 1.7 e - 0.1 \pm 8.0 e - 0.2 & 4.5 (2,13) & 0.26 & 2.12 \\ \end{array} \\ \text{Canopy closure} \\ \text{Canopy closure} \\ \text{Shannon} \\ \text{3} \\ \text{3} \\ 1.3 e - 0.3 \pm 2.2 e - 0.2 & 0.0 (2,102) & 0.00 & 0.14 \\ 2 & 1.1 e - 0.1 \pm 3.2 e - 0.2 & 12.1 (2,40) & 0.23 & 3.48 * * \\ 3 & 2.3 e - 0.1 \pm 1.4 e - 01 & 2.9 (2.20) & 0.13 & 1.69 \\ 4 & 6.7 e - 0.2 \pm 1.1 e - 01 & 0.4 (2.13) & 0.03 & 0.60 \\ \end{array} \\ \text{Foliage uniformity} \\ \text{Basal area CV} \\ \text{2} \\ \text{3} \\ \text{3} \\ \text{3} \\ \text{3} \\ \text{5} \\ \text{6} \\ \text{0} \\ \text{1} \\ \text{2} \\ \text{2} \\ \text{7} \\ \text{5} \\ \text{6} \\ \text{0} \\ \text{1} \\ \text{2} \\ \text{2} \\ \text{3} \\ \text{5} \\ \text{6} \\ \text{0} \\ \text{1} \\ \text{2} \\ \text{2} \\ \text{3} \\ \text{3} \\ \text{3} \\ \text{4} \\ \text{5} \\ \text{9} \\ \text{2} \\ \text{2} \\ \text{3} \\ \text{3} \\ \text{4} \\ \text{5} \\ \text{9} \\ \text{2} \\ \text{2} \\ \text{3} \\ \text{2} \\ \text{3} \\ \text{3} \\ \text{4} \\ \text{5} \\ \text{9} \\ \text{2} \\ \text{2} \\ \text{3} \\ \text{2} \\ \text{3} \\ \text{3} \\ \text{4} \\ \text{5} \\ \text{9} \\ \text{2} \\ \text{2} \\ \text{3} \\ \text{3} \\ \text{3} \\ \text{3} \\ \text{4} \\ \text{5} \\ \text{9} \\ \text{2} \\ \text{3} \\ \text{3} \\ \text{4} \\ \text{5} \\ \text{9} \\ \text{2} \\ \text{3} \\ \text{3} \\ \text{3} \\ \text{3} \\ \text{3} \\ \text{4} \\ \text{5} \\ \text{9} \\ \text{2} \\ \text{3} \\ \text{3} \\ \text{3} \\ \text{4} \\ $	ronage density	Hegyi	3	$6.6e + 03 \pm 3.0e + 03$	4.8(2,23)	0.17	2.18*
$ \begin{array}{c} \text{Foliage density} \\ \text{Foliage density} \\ \text{Foliage density} \\ \text{Foliage density} \\ \text{Poliage uniformity} \\ Poliage $			4	$1.5e + 01 \pm 5.5e + 03$	0.0(2,13)	0.00	0.00
Foliage density Foliage density Foliage density Shannon $\begin{array}{c} 3 & 1.1e+04\pm6.5e+03 & 3.1(2,20) & 0.13 & 1.77 \\ 4 & -6.5e+03\pm6.5e+03 & 1.0(2,13) & 0.07 & -1.01 \\ \hline \\ A & -6.5e+03\pm6.5e+03 & 1.0(2,13) & 0.07 & -1.01 \\ \hline \\ Canopy closure \\ Canopy closure \\ \end{array} \begin{array}{c} 1 & 1.7e-04\pm6.0e-04 & 0.1(2,97) & 0.00 & 0.28 \\ 2 & 2.9e-03\pm1.1e-03 & 6.9(2,39) & 0.15 & 2.62* \\ 3 & 4.2e-03\pm1.1e-03 & 15.1(2,14) & 0.52 & 3.89*** \\ 4 & -4.6e-03\pm3.0e-03 & 2.2(2,12) & 0.16 & -1.50 \\ \hline \\ Canopy closure \\ \end{array} \begin{array}{c} 1 & 2.2e-01\pm2.8e-02 & 62.3(2,102) & 0.38 & 7.89*** \\ 2 & 2.6e-01\pm5.1e-02 & 27.0(2,41) & 0.40 & 5.19**** \\ 3 & 2.8e-01\pm4.0e-02 & 50.7(2,23) & 0.69 & 7.12*** \\ 4 & 1.7e-01\pm8.0e-02 & 4.5(2,13) & 0.26 & 2.12 \\ \hline \\ Canopy closure \\ \end{array} \begin{array}{c} 1 & 3.1e-03\pm2.2e-02 & 0.0(2,102) & 0.00 & 0.14 \\ 3 & 2.3e-01\pm1.4e-01 & 2.9(2,20) & 0.13 & 1.69 \\ 4 & 6.7e-02\pm1.1e-01 & 0.4(2,13) & 0.03 & 0.60 \\ \hline \\ Foliage uniformity \\ \end{array} \begin{array}{c} 1 & 3.7e+00\pm4.0e+00 & 0.9(2,97) & 0.01 & 0.92 \\ 2 & 4.5e+00\pm7.4e+00 & 0.4(2,13) & 0.03 & 0.60 \\ 3 & -3.5e+00\pm5.9e+00 & 0.4(2,14) & 0.02 & -0.59 \\ 4 & -9.3e-01\pm1.5e+01 & 0.0(2,12) & 0.00 & -0.06 \\ \hline \\ Foliage uniformity \\ \end{array} \begin{array}{c} 1 & 2.2e+02\pm2.3e+02 & 1.0(2,102) & 0.01 & 0.98 \\ -9.3e-01\pm1.5e+01 & 0.0(2,12) & 0.00 & -0.06 \\ \hline \\ \hline \\ Foliage uniformity \\ \end{array} \begin{array}{c} 1 & 2.2e+02\pm2.3e+02 & 1.0(2,102) & 0.01 & 0.98 \\ -9.3e-01\pm1.5e+01 & 0.0(2,12) & 0.00 & -0.06 \\ \hline \\ \hline \\ Foliage uniformity \\ \end{array} \begin{array}{c} 1 & 2.3e+02\pm1.4e+02 & 2.9(2,23) & 0.11 & 1.72 \\ -9.3e-01\pm1.5e+01 & 0.0(2,13) & 0.00 & -0.19 \\ \hline \\ \hline \\ Foliage uniformity \\ \end{array} \begin{array}{c} 1 & 2.3e+02\pm1.4e+02 & 2.6(2,102) & 0.02 & 1.61 \\ -9.3e-01\pm1.5e+01 & 0.0(2,13) & 0.00 & -0.19 \\ \hline \\ \hline \\ Foliage uniformity \\ \end{array} \begin{array}{c} 1 & 2.5e+02\pm3.6e+02 & 2.9(2,23) & 0.11 & 1.72 \\ -7.5e+01\pm4.0e+02 & 0.1(2,39) & 0.00 & 0.38 \\ -7.5e+01\pm4.0e+02 & 0.1(2,39) & 0.00 & 0.38 \\ -7.5e+02\pm4.7e+02 & 1.6(2,13) & 0.11 & -1.27 \\ \hline \end{array} \begin{array}{c} 1 & 2.5e+02\pm3.6e+02 & 2.9(2,23) & 0.11 & 1.72 \\ -7.5e+02\pm4.7e+02 & 1.6(2,13) & 0.11 & -1.27 \\ \hline \end{array} \begin{array}{c} 1 & 2.5e+02\pm9.3e-03 & 7.1(2,97) & 0.07 & 2.66e** \\ -7.5e+02\pm4.7e+02 & 1.6(2,13) & 0.11 & -1.27 \\ \hline \end{array} $			1	$2.2e+03\pm1.3e+03$	2.8(2,102)	0.03	1.67
Canopy closure Basal area CV $ \begin{array}{c} 3 & 1.1e+04\pm 6.5e+03 & 3.1(2,20) & 0.13 & 1.77 \\ 4 & -6.5e+03\pm 6.5e+03 & 1.0(2,13) & 0.07 & -1.01 \\ 1 & 1.7e-04\pm 6.0e+04 & 0.1(2,97) & 0.00 & 0.28 \\ 2 & 2.9e+03\pm 1.1e+03 & 6.9(2,39) & 0.15 & 2.6e^* \\ 3 & 4.2e+03\pm 1.1e+03 & 15.1(2,14) & 0.52 & 3.89^{***} \\ 4 & -4.6e+03\pm 3.0e+03 & 2.2(2,12) & 0.16 & -1.50 \\ 2 & 2.6e+01\pm 5.1e+02 & 27.0(2,41) & 0.40 & 5.19^{****} \\ 2 & 2.6e+01\pm 5.1e+02 & 27.0(2,41) & 0.40 & 5.19^{****} \\ 3 & 2.8e+01\pm 4.0e+02 & 50.7(2,23) & 0.69 & 7.12^{****} \\ 4 & 1.7e+01\pm 8.0e+02 & 4.5(2,13) & 0.26 & 2.12 \\ 2 & 1.1e+01\pm 3.2e+02 & 0.0(2,102) & 0.00 & 0.14 \\ 3 & 2.3e+01\pm 1.4e+01 & 2.9(2,20) & 0.13 & 1.69 \\ 4 & 6.7e+02\pm 1.1e+01 & 0.4(2,13) & 0.03 & 0.60 \\ 4 & 6.7e+02\pm 1.1e+01 & 0.4(2,13) & 0.03 & 0.60 \\ 4 & 6.7e+02\pm 1.1e+01 & 0.4(2,13) & 0.03 & 0.60 \\ 4 & 9.3e+01\pm 1.5e+01 & 0.0(2,12) & 0.00 & -0.59 \\ 4 & 9.3e+01\pm 1.5e+01 & 0.0(2,12) & 0.00 & -0.06 \\ 5 & 4 & 9.3e+01\pm 1.5e+01 & 0.0(2,12) & 0.00 & -0.06 \\ 6 & 9.3e+01\pm 1.5e+01 & 0.0(2,12) & 0.00 & -0.06 \\ 7 & 5e+02\pm 3.7e+02 & 4.0(2,40) & 0.09 & 2.00 \\ 8 & 4.5e+02\pm 2.6e+02 & 2.9(2,23) & 0.11 & 1.72 \\ 4 & -7.5e+01\pm 4.0e+02 & 0.0(2,13) & 0.00 & -0.19 \\ 7 & 5e+02\pm 3.7e+02 & 4.0(2,40) & 0.09 & 2.00 \\ 8 & 4.5e+02\pm 2.6e+02 & 2.9(2,23) & 0.11 & 1.72 \\ 4 & -7.5e+01\pm 4.0e+02 & 0.0(2,13) & 0.00 & -0.19 \\ 7 & 5e+02\pm 4.7e+02 & 0.1(2,39) & 0.00 & 0.38 \\ 8 & 1.3e+03\pm 5.1e+02 & 6.1(2,20) & 0.23 & 2.48^* \\ 4 & -5.9e+02\pm 4.7e+02 & 1.6(2,13) & 0.11 & -1.27 \\ 8 & 4.5e+02\pm 2.6e+02 & 2.1(2,30) & 0.03 & 3.48^{**} \\ 4 & -5.9e+02\pm 4.7e+02 & 1.6(2,13) & 0.11 & -1.27 \\ 8 & 5e+02\pm 3.6e+03 & 7.1(2,97) & 0.07 & 2.66^{**} \\ 4 & -5.9e+02\pm 4.7e+02 & 1.6(2,13) & 0.11 & -1.27 \\ 8 & 5e+02\pm 3.6e+03 & 7.1(2,97) & 0.07 & 2.66^{**} \\ 4 & -5.9e+02\pm 4.7e+02 & 1.6(2,13) & 0.11 & -1.27 \\ 8 & 5e+02\pm 3.6e+03 & 7.1(2,97) & 0.07 & 2.66^{**} \\ 4 & -5.9e+02\pm 4.7e+02 & 8.0(2,38) & 0.17 & 2.83^{**} \\ 8 & 5e+02\pm 3.6e+03 & 8.0(2,38) & 0.17 & 2.83^{**} \\ 8 & 5e+02\pm 3.6e+03 & 8.0(2,38) & 0.17 & 2.83^{**} \\ 8 & 5e+02\pm 3.6e+03 & 8.0(2,38) & 0.17 & 2.83^{**} \\ 8 & 5e+02\pm 4.7e$	Foliago dongitu	Channon	2	$3.8e + 03 \pm 2.4e + 03$	2.6(2,39)	0.06	1.61
Canopy closure Basal area CV $\begin{bmatrix} 1 & 1.7e-04\pm6.0e-04 & 0.1(2,97) & 0.00 & 0.28 \\ 2 & 2.9e-03\pm1.1e-03 & 6.9(2,39) & 0.15 & 2.62* \\ 3 & 4.2e-03\pm1.1e-03 & 15.1(2,14) & 0.52 & 3.89** \\ 4 & -4.6e-03\pm3.0e-03 & 2.2(2,12) & 0.16 & -1.50 \\ \end{bmatrix}$ Canopy closure Hegyi $\begin{bmatrix} 1 & 2.2e-01\pm2.8e-02 & 62.3(2,102) & 0.38 & 7.89*** \\ 2 & 2.6e-01\pm5.1e-02 & 27.0(2,41) & 0.40 & 5.19*** \\ 3 & 2.8e-01\pm4.0e-02 & 50.7(2,23) & 0.69 & 7.12*** \\ 4 & 1.7e-01\pm8.0e-02 & 4.5(2,13) & 0.26 & 2.12 \\ \end{bmatrix}$ Canopy closure Shannon $\begin{bmatrix} 1 & 3.1e-03\pm2.2e-02 & 0.0(2,102) & 0.00 & 0.14 \\ 2 & 1.1e-01\pm3.2e-02 & 0.0(2,102) & 0.00 & 0.14 \\ 4 & 6.7e-02\pm1.1e-01 & 0.4(2,13) & 0.03 & 3.48** \\ 4 & 6.7e-02\pm1.1e-01 & 0.4(2,13) & 0.03 & 0.60 \\ \end{bmatrix}$ Foliage uniformity Basal area CV $\begin{bmatrix} 1 & 3.7e+00\pm4.0e+00 & 0.9(2,97) & 0.01 & 0.92 \\ 2 & 4.5e+00\pm7.4e+00 & 0.4(2,38) & 0.01 & 0.61 \\ 3 & -3.5e+00\pm5.9e+00 & 0.4(2,14) & 0.02 & -0.59 \\ 4 & -9.3e-01\pm1.5e+01 & 0.0(2,12) & 0.00 & -0.06 \\ \end{bmatrix}$ Foliage uniformity Hegyi $\begin{bmatrix} 1 & 2.2e+02\pm2.3e+02 & 1.0(2,102) & 0.01 & 0.98 \\ 2 & 7.5e+02\pm3.7e+02 & 4.0(2,40) & 0.09 & 2.00 \\ 3 & 4.5e+02\pm2.6e+02 & 2.9(2,23) & 0.11 & 1.72 \\ 4 & -7.5e+01\pm4.0e+02 & 0.0(2,13) & 0.00 & -0.19 \\ \end{bmatrix}$ Foliage uniformity Shannon $\begin{bmatrix} 1 & 2.3e+02\pm1.4e+02 & 2.6(2,102) & 0.02 & 1.61 \\ 2 & 8.6e+01\pm2.2e+02 & 0.1(2,39) & 0.00 & 0.38 \\ 3 & 1.3e+03\pm5.1e+02 & 6.1(2,20) & 0.23 & 2.48* \\ 4 & -5.9e+02\pm4.7e+02 & 1.6(2,13) & 0.11 & -1.27 \\ \end{bmatrix}$ Lavar diversity Basal area CV $\begin{bmatrix} 1 & 2.5e-02\pm9.3e-03 & 7.1(2,97) & 0.07 & 2.66e** \\ 2 & 3.9e-02\pm1.4e-02 & 8.0(2,38) & 0.17 & 2.83** \\ \end{bmatrix}$	ronage density	Shannon	3	$1.1e+04\pm6.5e+03$	3.1(2,20)	0.13	1.77
$ \begin{array}{c} \text{Canopy closure} \\ \text{Canopy closure} \\ \text{Basal area CV} \\ \end{array}{c} \begin{array}{c} 2 \\ 3 \\ 4.2 \text{e} - 0.3 \pm 1.1 \text{e} - 0.3 \\ 4.2 \text{e} - 0.3 \pm 1.1 \text{e} - 0.3 \\ 4.2 \text{e} - 0.3 \pm 1.1 \text{e} - 0.3 \\ 4.2 \text{e} - 0.0 \pm 1.1 \text{e} - 0.3 \\ 4.2 \text{e} - 0.0 \pm 1.1 \text{e} - 0.3 \\ 4.2 \text{e} - 0.0 \pm 1.1 \text{e} - 0.3 \\ 2.2 \text{e} - 0.0 \pm 1.1 \text{e} - 0.3 \\ 2.2 \text{e} - 0.0 \pm 1.1 \text{e} - 0.3 \\ 2.2 \text{e} - 0.0 \pm 1.1 \text{e} - 0.2 \\ 2.0 \text{e} - 0.0 \text{e} - 0.0 \\ 2.0 \text{e} - 0.0 \\ 2.0 \text{e} - 0.0 \text{e} - 0.2 \\ 2.0 \text{e} - 0.0 \text{e} - 0.2 \\ 2.0 \text{e}$			4	$-6.5e + 03 \pm 6.5e + 03$	1.0(2,13)	0.07	-1.01
Canopy closure Basal area CV $\frac{3}{4} + \frac{4.2e-03\pm 1.1e-03}{-4.6e-03\pm 3.0e-03} = \frac{15.1(2,14)}{2.2(2,12)} = 0.16 -1.50$ Canopy closure Hegyi $\frac{1}{2} = \frac{2.2e-01\pm 2.8e-02}{2.6e-01\pm 5.1e-02} = \frac{62.3(2,102)}{2.7.0(2,41)} = 0.40 -5.19^{****}$ Canopy closure Hegyi $\frac{1}{2} = \frac{2.2e-01\pm 2.8e-02}{4.1.7e-01\pm 8.0e-02} = \frac{62.3(2,102)}{2.7.0(2,41)} = 0.40 -5.19^{****}$ Canopy closure Shannon $\frac{1}{2} = \frac{3.1e-03\pm 2.2e-02}{4.5(2,13)} = 0.00 -0.00 -0.14$ Canopy closure Shannon $\frac{1}{2} = \frac{3.1e-03\pm 2.2e-02}{1.1e-01\pm 3.2e-02} = \frac{10.0(2,102)}{1.2(40)} = 0.00 -0.13 -0.69$ Foliage uniformity Basal area CV $\frac{1}{2} = \frac{3.7e+00\pm 4.0e+00}{4.5e+00\pm 7.4e+00} = 0.9(2,20) -0.01 -0.92$ Foliage uniformity Hegyi $\frac{1}{2} = \frac{2.2e+02\pm 2.3e+02}{4.5e+00\pm 7.4e+00} = 0.4(2,14) -0.02 -0.59$ Foliage uniformity Hegyi $\frac{1}{2} = \frac{2.2e+02\pm 2.3e+02}{4.5e+00\pm 2.2e+02} = \frac{1.0(2,102)}{2.0(2,102)} = 0.01 -0.98$ Foliage uniformity Hegyi $\frac{1}{2} = \frac{2.2e+02\pm 2.3e+02}{2.9(2,23)} = \frac{1.0(2,102)}{0.01} = 0.01$ Foliage uniformity Shannon $\frac{1}{2} = \frac{2.3e+02\pm 3.7e+02}{4.9e+02\pm 2.6e+02} = \frac{2.9(2,23)}{2.9(2,23)} = 0.11 -1.72$ Foliage uniformity Shannon $\frac{1}{2} = \frac{2.3e+02\pm 1.4e+02}{4.9e+02\pm 4.6e+02} = \frac{2.6(2,102)}{0.0(2,13)} = 0.00 -0.19$ Foliage uniformity Shannon $\frac{1}{2} = \frac{2.3e+02\pm 1.4e+02}{4.9e+02\pm 4.7e+02} = \frac{2.6(2,102)}{0.0(2,13)} = 0.00 -0.19$			1	$1.7e-04\pm6.0e-04$	0.1(2,97)	0.00	0.28
Canopy closure Hegyi $3 $	Conony aloguro	Pagal area CV	2	$2.9 \text{e-} 03 \pm 1.1 \text{e-} 03$	6.9(2,39)	0.15	2.62*
Canopy closure Hegyi $ \begin{array}{c} 1 \\ 2 \\ 2 \\ 2.6e-01\pm 5.1e-02 \\ 3 \\ 2.8e-01\pm 4.0e-02 \\ 4.5(2,13) \\ 2.6e-01\pm 5.1e-02 \\ 50.7(2,23) \\ 4.5(2,13) \\ 2.6e-01\pm 8.0e-02 \\ 2.6e-01\pm 8.0e-02 \\ 4.5(2,13) \\ 2.6e-01\pm 8.0e-02 \\ 2.6e-01\pm 8.0e-02 \\ 4.5(2,13) \\ 2.6e-01\pm 8.0e-02 \\ 2.6e-01\pm 8.$	Canopy closure	Dasar area Cv	3	$4.2 \text{e-} 03 \pm 1.1 \text{e-} 03$	15.1(2,14)	0.52	3.89**
Canopy closure Hegyi $ \begin{array}{c} 2 & 2.6e - 01 \pm 5.1e - 02 \\ 3 & 2.8e - 01 \pm 4.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 4 & 1.7e - 01 \pm 8.0e - 02 \\ 1 & 1.1e - 01 \pm 3.2e - 02 \\ 1$			4	$-4.6e-03\pm3.0e-03$	2.2(2,12)	0.16	-1.50
Canopy closure Hegyl 3 2.8e-01±4.0e-02 50.7(2,23) 0.69 7.12*** 4 1.7e-01±8.0e-02 4.5(2,13) 0.26 2.12 1 3.1e-03±2.2e-02 0.0(2,102) 0.00 0.14 2 1.1e-01±3.2e-02 12.1(2,40) 0.23 3.48** 3 2.3e-01±1.4e-01 2.9(2,20) 0.13 1.69 4 6.7e-02±1.1e-01 0.4(2,13) 0.03 0.60 Foliage uniformity Basal area CV 1 3 -3.5e+00±4.0e+00 0.9(2,97) 0.01 0.92 2 4.5e+00±7.4e+00 0.4(2,38) 0.01 0.61 3 -3.5e+00±5.9e+00 0.4(2,14) 0.02 -0.59 4 -9.3e-01±1.5e+01 0.0(2,12) 0.00 -0.06 Foliage uniformity Hegyl 1 2.2e+02±2.3e+02 1.0(2,102) 0.01 0.98 2 7.5e+02±3.7e+02 4.0(2,40) 0.09 2.00 3 4.5e+02±2.6e+02 2.9(2,23) 0.11 1.72 4 -7.5e+01±4.0e+02 0.0(2,13) 0.00 -0.19 Foliage uniformity Shannon 1 2.3e+02±1.4e+02 2.6(2,102) 0.02 1.61 2 8.6e+01±2.2e+02 0.1(2,39) 0.00 0.38 3 1.3e+03±5.1e+02 6.1(2,20) 0.23 2.48* 4 -5.9e+02±4.7e+02 1.6(2,13) 0.11 -1.27 Layer diversity Basal area CV 2 3.9e-02±1.4e-02 8.0(2,38) 0.17 2.83**	Canopy closure	Hegyi	1	$2.2e-01\pm2.8e-02$	62.3(2,102)	0.38	7.89***
Canopy closure Shannon $ \begin{array}{c} 3 & 2.8e-01\pm 4.0e-02 & 50.7(2,23) & 0.69 & 7.12^{-12-12-12-12-12-12-12-12-12-12-12-12-12-$			2	$2.6e-01\pm5.1e-02$	27.0(2,41)	0.40	5.19***
Canopy closure Shannon			3	$2.8e-01\pm4.0e-02$	50.7(2,23)	0.69	7.12***
Canopy closure Shannon $ \begin{array}{c} 2 & 1.1e \cdot 01\pm 3.2e \cdot 02 & 12.1(2,40) & 0.23 & 3.48^{**} \\ 3 & 2.3e \cdot 01\pm 1.4e \cdot 01 & 2.9(2,20) & 0.13 & 1.69 \\ 4 & 6.7e \cdot 02\pm 1.1e \cdot 01 & 0.4(2,13) & 0.03 & 0.60 \\ \end{array} $ Foliage uniformity $ \begin{array}{c} 1 & 3.7e + 00\pm 4.0e + 00 & 0.9(2,97) & 0.01 & 0.92 \\ 2 & 4.5e + 00\pm 7.4e + 00 & 0.4(2,38) & 0.01 & 0.61 \\ 3 & -3.5e + 00\pm 5.9e + 00 & 0.4(2,14) & 0.02 & -0.59 \\ 4 & -9.3e \cdot 01\pm 1.5e + 01 & 0.0(2,12) & 0.00 & -0.06 \\ \end{array} $ Foliage uniformity $ \begin{array}{c} 1 & 2.2e + 02\pm 2.3e + 02 & 1.0(2,102) & 0.01 & 0.98 \\ 2 & 7.5e + 02\pm 3.7e + 02 & 4.0(2,40) & 0.09 & 2.00 \\ 3 & 4.5e + 02\pm 2.6e + 02 & 2.9(2,23) & 0.11 & 1.72 \\ 4 & -7.5e + 01\pm 4.0e + 02 & 0.0(2,13) & 0.00 & -0.19 \\ \end{array} $ Foliage uniformity $ \begin{array}{c} 1 & 2.3e + 02\pm 1.4e + 02 & 2.6(2,102) & 0.02 & 1.61 \\ 2 & 8.6e + 01\pm 2.2e + 02 & 0.1(2,39) & 0.00 & 0.38 \\ 3 & 1.3e + 03\pm 5.1e + 02 & 6.1(2,20) & 0.23 & 2.48^* \\ 4 & -5.9e + 02\pm 4.7e + 02 & 1.6(2,13) & 0.11 & -1.27 \\ \end{array} $ Layer diversity $ \begin{array}{c} 1 & 2.5e \cdot 02\pm 9.3e \cdot 03 & 7.1(2,97) & 0.07 & 2.66^{**} \\ 2 & 3.9e \cdot 02\pm 1.4e \cdot 02 & 8.0(2,38) & 0.17 & 2.83^{**} \\ \end{array} $			4	$1.7e-01\pm 8.0e-02$	4.5(2,13)	0.26	2.12
Canopy closure Shannon 3 2.3e-01 \pm 1.4e-01 2.9(2,20) 0.13 1.69 4 6.7e-02 \pm 1.1e-01 0.4(2,13) 0.03 0.60 Foliage uniformity Basal area CV $ \begin{array}{c} 1 & 3.7e+00\pm4.0e+00 & 0.9(2,97) & 0.01 & 0.92 \\ 2 & 4.5e+00\pm7.4e+00 & 0.4(2,38) & 0.01 & 0.61 \\ 3 & -3.5e+00\pm5.9e+00 & 0.4(2,14) & 0.02 & -0.59 \\ 4 & -9.3e-01\pm1.5e+01 & 0.0(2,12) & 0.00 & -0.06 \\ \end{array} $ Foliage uniformity Hegyi $ \begin{array}{c} 1 & 2.2e+02\pm2.3e+02 & 1.0(2,102) & 0.01 & 0.98 \\ 2 & 7.5e+02\pm3.7e+02 & 4.0(2,40) & 0.09 & 2.00 \\ 3 & 4.5e+02\pm2.6e+02 & 2.9(2,23) & 0.11 & 1.72 \\ 4 & -7.5e+01\pm4.0e+02 & 0.0(2,13) & 0.00 & -0.19 \\ \end{array} $ Foliage uniformity Shannon $ \begin{array}{c} 1 & 2.3e+02\pm1.4e+02 & 2.6(2,102) & 0.02 & 1.61 \\ 2 & 8.6e+01\pm2.2e+02 & 0.1(2,39) & 0.00 & 0.38 \\ 3 & 1.3e+03\pm5.1e+02 & 6.1(2,20) & 0.23 & 2.48* \\ 4 & -5.9e+02\pm4.7e+02 & 1.6(2,13) & 0.11 & -1.27 \\ \end{array} $ Layer diversity Basal area CV $ \begin{array}{c} 1 & 2.5e-02\pm9.3e-03 & 7.1(2,97) & 0.07 & 2.66** \\ 2 & 3.9e-02\pm1.4e-02 & 8.0(2,38) & 0.17 & 2.83** \\ \end{array} $			1	$3.1e-03\pm2.2e-02$	0.0(2,102)	0.00	0.14
Foliage uniformity Basal area CV $ \begin{array}{c} 3 & 2.3e\text{-}01\pm1.4e\text{-}01 & 2.9(2,20) & 0.13 & 1.69 \\ 4 & 6.7e\text{-}02\pm1.1e\text{-}01 & 0.4(2,13) & 0.03 & 0.60 \\ \\ 1 & 3.7e+00\pm4.0e+00 & 0.9(2,97) & 0.01 & 0.92 \\ 2 & 4.5e+00\pm7.4e+00 & 0.4(2,38) & 0.01 & 0.61 \\ 3 & -3.5e+00\pm5.9e+00 & 0.4(2,14) & 0.02 & -0.59 \\ 4 & -9.3e\text{-}01\pm1.5e+01 & 0.0(2,12) & 0.00 & -0.06 \\ \\ Foliage uniformity & Hegyi & 1 & 2.2e+02\pm2.3e+02 & 1.0(2,102) & 0.01 & 0.98 \\ 2 & 7.5e+02\pm3.7e+02 & 4.0(2,40) & 0.09 & 2.00 \\ 3 & 4.5e+02\pm2.6e+02 & 2.9(2,23) & 0.11 & 1.72 \\ 4 & -7.5e+01\pm4.0e+02 & 0.0(2,13) & 0.00 & -0.19 \\ \\ Foliage uniformity & Shannon & 1 & 2.3e+02\pm1.4e+02 & 2.6(2,102) & 0.02 & 1.61 \\ 2 & 8.6e+01\pm2.2e+02 & 0.1(2,39) & 0.00 & 0.38 \\ 3 & 1.3e+03\pm5.1e+02 & 6.1(2,20) & 0.23 & 2.48* \\ 4 & -5.9e+02\pm4.7e+02 & 1.6(2,13) & 0.11 & -1.27 \\ \\ 1 & 2.5e-02\pm9.3e-03 & 7.1(2,97) & 0.07 & 2.66** \\ 2 & 3.9e-02\pm1.4e-02 & 8.0(2,38) & 0.17 & 2.83** \\ \end{array} $	Canony aloguro	Channon	2	$1.1e-01\pm3.2e-02$	12.1(2,40)	0.23	3.48**
Foliage uniformity Basal area CV	Canopy closure	Shannon	3	$2.3e-01\pm1.4e-01$	2.9(2,20)	0.13	1.69
Foliage uniformity Basal area CV $ \begin{array}{c} 2 \\ 3 \\ -3.5 \\ e+00 \\ \pm 5.9 \\ e+00 \\ \end{array} \begin{array}{c} 0.4(2,38) \\ 0.01 \\ 0.61 \\ 0.02 \\ -0.59 \\ 0.00 \\ -0.06 \\ \end{array} $ Foliage uniformity Hegyi $ \begin{array}{c} 2 \\ 3 \\ -3.5 \\ e+00 \\ \pm 2.3 \\ e+02 \\ \pm 2.3 \\ e+02 \\ \pm 3.7 \\ e+02 \\ \pm 3.7 \\ e+02 \\ \pm 2.6 \\ e+02 \\ \pm 2.6 \\ e+02 \\ \pm 2.9 \\ (2,33) \\ 0.11 \\ 1.72 \\ 4 \\ -7.5 \\ e+01 \\ \pm 4.0 \\ e+02 \\ 0.0 \\ (2,13) \\ 0.00 \\ -0.19 \\ \end{array} $ Foliage uniformity Shannon $ \begin{array}{c} 1 \\ 2.3 \\ e+02 \\ \pm 1.4 \\ e+02 \\ 2.6 \\ (2,102) \\ 0.02 \\ 1.61 \\ 2.3 \\ e+01 \\ \pm 2.2 \\ e+02 \\ 2.6 \\ (2,102) \\ 0.02 \\ 0.03 \\ 2.48 \\ 4 \\ -5.9 \\ e+02 \\ \pm 4.7 \\ e+02 \\ 1.6 \\ (2,13) \\ 0.11 \\ -1.27 \\ \end{array} $ Lever diversity Basal area CV $ \begin{array}{c} 2 \\ 3.9 \\ e+02 \\ \pm 1.4 \\ e+02 \\ 2.5 \\ e+02 \\ \pm 4.7 \\ e+02 \\ 1.6 \\ (2,13) \\ 0.11 \\ -1.27 \\ \end{array} $			4	$6.7 \text{e-} 02 \pm 1.1 \text{e-} 01$	0.4(2,13)	0.03	0.60
Foliage uniformity Basal area CV $ \begin{array}{c} 3 & -3.5 \text{e} + 00 \pm 5.9 \text{e} + 00 \\ 4 & -9.3 \text{e} - 01 \pm 1.5 \text{e} + 01 \\ \end{array} \begin{array}{c} 0.0(2,12) & 0.02 & -0.59 \\ 0.00(2,12) & 0.00 & -0.06 \\ \end{array} \\ \text{Foliage uniformity Hegyi} \\ \end{array} \begin{array}{c} 1 & 2.2 \text{e} + 02 \pm 2.3 \text{e} + 02 \\ 2 & 7.5 \text{e} + 02 \pm 3.7 \text{e} + 02 \\ 3 & 4.5 \text{e} + 02 \pm 2.6 \text{e} + 02 \\ 4 & -7.5 \text{e} + 01 \pm 4.0 \text{e} + 02 \\ 4 & -7.5 \text{e} + 01 \pm 4.0 \text{e} + 02 \\ 4 & -7.5 \text{e} + 01 \pm 4.0 \text{e} + 02 \\ 4 & -7.5 \text{e} + 01 \pm 2.2 \text{e} + 02 \\ 4 & -8.6 \text{e} + 01$			1	$3.7e + 00 \pm 4.0e + 00$	0.9(2,97)	0.01	0.92
Foliage uniformity Hegyi $ \begin{array}{c} 3 \\ 4 \\ -9.3 \text{e} - 01 \pm 1.5 \text{e} + 00 \\ 2 \\ 7.5 \text{e} + 02 \pm 2.3 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 02 \pm 3.7 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 4.0 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 4.0 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 4.0 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 2.2 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 4.0 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 4.0 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 2.2 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 2.2 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 2.2 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 2.2 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 2.2 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 2.2 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 2.2 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 01 \pm 2.2 \text{e} + 02 \\ 4 \\ -7.5 \text{e} + 02 \pm 4.7 \text{e} + 02 \\ 4 \\ -7.5 e$	Foliago uniformity	Bagal area CV		$4.5e+00\pm7.4e+00$	0.4(2,38)	0.01	0.61
Foliage uniformity Hegyi $ \begin{array}{c} 1 \\ 2.2e+02\pm2.3e+02 \\ 2 \\ 7.5e+02\pm3.7e+02 \\ 3 \\ 4.5e+02\pm2.6e+02 \\ 4 \\ -7.5e+01\pm4.0e+02 \\ 2 \\ 0.0(2,13) \\ 0.00 \\ -0.19 \\ \end{array} $ Foliage uniformity Shannon $ \begin{array}{c} 1 \\ 2.3e+02\pm1.4e+02 \\ 2.6(2,102) \\ 2.6(2,102) \\ 3.9e+02\pm1.4e+02 \\ 2.6(2,102) \\ 0.02 \\ 0.12,39 \\ 0.00 \\ 0.38 \\ 0.11 \\ 0.38 \\ 0.48^* \\ 1.3e+03\pm5.1e+02 \\ 1.6(2,13) \\ 0.11 \\ 0.12 \\ 0.23 \\ 0.23 \\ 0.248^* \\ 1.25e+02\pm4.7e+02 \\ 1.27 \\ 0.27 \\ 0.27 \\ 0.27 \\ 0.28 \\ 0.28 \\ 0.238 \\ 0.17 \\ 0.28 \\ 0.28 \\ 0.28 \\ 0.28 \\ 0.28 \\ 0.28 \\ 0.28 \\ 0.28 \\ 0.27 \\ 0.07 \\ 0.07 \\ 0.28 \\ 0.28 \\ 0.28 \\ 0.17 \\ 0.28 \\ 0.2$	ronage unnormity	Dasar area C v	3	$-3.5e+00\pm5.9e+00$	0.4(2,14)	0.02	-0.59
Foliage uniformity Hegyi $ \begin{array}{c ccccccccccccccccccccccccccccccccccc$			4	$-9.3e-01\pm1.5e+01$	0.0(2,12)	0.00	-0.06
Foliage uniformity Hegyl $ \begin{array}{ccccccccccccccccccccccccccccccccccc$			1	$2.2e + 02 \pm 2.3e + 02$	1.0(2,102)	0.01	0.98
Foliage uniformity Shannon $ \begin{array}{ccccccccccccccccccccccccccccccccccc$	Foliago uniformity	Ности	2	$7.5e + 02 \pm 3.7e + 02$	4.0(2,40)	0.09	2.00
Foliage uniformity Shannon $ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	ronage unnormity	педуг	3	$4.5e+02\pm2.6e+02$	2.9(2,23)	0.11	1.72
Foliage uniformity Shannon $ \begin{array}{ccccccccccccccccccccccccccccccccccc$			4	$-7.5e + 01 \pm 4.0e + 02$	0.0(2,13)	0.00	-0.19
Foliage uniformity Snannon $ \begin{array}{ccccccccccccccccccccccccccccccccccc$			1	$2.3e + 02 \pm 1.4e + 02$	2.6(2,102)	0.02	1.61
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Foliago uniformit	Channon	2	$8.6e + 01 \pm 2.2e + 02$	0.1(2,39)	0.00	0.38
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	ronage unnormity	SHAIIIOH	3	$1.3e+03\pm5.1e+02$	6.1(2,20)	0.23	2.48*
Layer diversity Basal area CV $2 3.9e-02\pm1.4e-02 8.0(2,38) 0.17 2.83**$			4	$-5.9e + 02 \pm 4.7e + 02$	1.6(2,13)	0.11	-1.27
Laver diversity Basal area Liv			1	$2.5e-02\pm9.3e-03$	7.1(2,97)	0.07	2.66**
Tayer diversity Basar area 0.7 3 $2.7e-02\pm2.3e-02$ $1.3(2,14)$ 0.09 1.15	Lavor diversity	Regal eres CV	2	$3.9 \text{e-} 02 \pm 1.4 \text{e-} 02$	8.0(2,38)	0.17	2.83**
	Layer diversity	Dasai aiea ∪ V	3	$2.7e-02\pm2.3e-02$	1.3(2,14)	0.09	1.15

		4	$2.1e-02\pm3.1e-02$	0.5(2,12)	0.04	0.67
		1	$2.7e + 00 \pm 4.9e - 01$	29.1(2,102)	0.22	5.39**
T 1	тт •	2	$2.0e+00\pm7.5e-01$	7.1(2,40)	0.15	2.66*
Layer diversity	Hegyi	3	$1.9e + 00 \pm 1.0e + 00$	3.6(2,23)	0.13	1.89
		4	$1.1\mathrm{e}{+00}{\pm}8.5\mathrm{e}{-01}$	1.8(2,13)	0.12	1.33
		1	$1.0e+00\pm3.4e-01$	8.7(2,102)	0.08	2.95**
Layer diversity	Shannon	2	$9.5e-01\pm4.3e-01$	4.8(2,39)	0.11	2.18*
Layer diversity	Silainion	3	$4.9e+00\pm1.8e+00$	7.2(2,20)	0.26	2.68*
		4	$1.8e-01\pm1.1e+00$	0.0(2,13)	0.00	0.16
		1	$1.2 \text{e-} 01 \pm 6.9 \text{e-} 02$	2.9(2,6)	0.33	1.72
Canopy roughness	Basal area CV		$-3.2e-01\pm2.9e-01$	1.2(2,3)	0.29	-1.10
Canopy roughness	Dasar area C v		$3.5e-01\pm4.7e-01$	0.6(2,1)	0.36	0.74
		4				
		1	$2.6e-01\pm1.2e+00$	0.0(2,6)	0.01	0.22
Canopy roughness	Voronoi CV		$4.6e + 00 \pm 1.9e + 00$	6.1(2,3)	0.67	2.48
Canopy roughness	voronor C v		$1.8e + 00 \pm 1.9e + 00$	1.0(2,1)	0.49	0.99
		4				
		1	$-4.2e+01\pm5.7e+01$	0.5(2,6)	0.08	-0.74
Canopy roughness	Mingling	2	$1.6e + 01 \pm 9.7e + 01$	0.0(2,3)	0.01	0.17
Canopy roughness	Willighing	3	$3.5e + 02 \pm 2.5e + 02$	2.0(2,1)	0.67	1.42
		4				
		1	$-4.3e-02\pm4.5e-02$	0.9(2,6)	0.13	-0.96
Canopy roughness	Tron dongity	2	$-5.9e-02\pm3.1e-02$	3.6(2,3)	0.54	-1.89
Canopy roughness	Tree density	3	$-1.8e-01\pm2.6e-01$	0.5(2,1)	0.31	-0.68
		4				
		1	$-2.3e+00\pm1.7e+00$	1.7(2,6)	0.22	-1.32
Canopy roughness	Channon	2	$-1.4e + 00 \pm 2.4e + 00$	0.4(2,3)	0.11	-0.60
Canopy roughness	Shannon	3	$3.4e + 01 \pm 4.7e + 01$	0.5(2,1)	0.34	0.72
		4				
		1	$-7.4e + 01 \pm 2.6e + 02$	0.1(2,6)	0.01	-0.28
Can any mangh naga	Uniform andle index	2	$4.1e+02\pm9.5e+02$	0.2(2,3)	0.06	0.43
Canopy roughness	Official angle fildex	3	$4.4e + 02 \pm 5.7e + 02$	0.6(2,1)	0.37	0.76
		ea CV 2 3 4 CV 3 4 1 2 3 4 1 2 3 4 1 2 3 4 1 2 3 4 1 2 3 4 CV 2 3 4 CV 3 4 1 2 3 4 1 2 3 4 CV 3 4 1 2 3 4 1 2 3 4 1 2 3 4 1 2 3 4				
		1	$-6.5 \text{e-} 03 \pm 6.1 \text{e-} 03$	1.1(2,6)	0.16	-1.07
Canopy height	Basal area CV	2	$4.3e-02\pm4.0e-02$	1.2(2,3)	0.28	1.08
Canopy neight	Dasar area C v	3	$-3.1e-02\pm8.7e-03$	12.3(2,1)	0.92	-3.51
		4				
		1	-1.0e-01±8.6e-02	1.5(2,6)	0.20	-1.21
Canopy height	Voronoi CV	2	$-7.0e-01\pm2.0e-01$	12.7(2,3)	0.81	-3.57*
оапору пегди	AOTOHOL (A	3	$-1.8e-02\pm1.4e-01$	0.0(2,1)	0.02	-0.13
		4		·		
		1	$6.8e + 00 \pm 3.8e + 00$	3.2(2,6)	0.34	1.78
Canony baisht	Mingling	2	$-3.3e+00\pm1.3e+01$	0.1(2,3)	0.02	-0.25
Canopy height	Mingling	3	$-2.3e+01\pm9.3e-01$	619.2(2,1)	1.00	-24.88*
		4		. ,		
		1	$-3.5e-04\pm3.8e-03$	0.0(2,6)	0.00	-0.09
Canopy height	Tree density			,		
Canopy neight	Tree delisity					

		2	$8.6e-03\pm4.0e-03$	4.7(2,3)	0.61	2.16
		3	$-1.0e-03\pm1.7e-02$	0.0(2,1)	0.00	-0.06
		4		() /		
		1	2.8e-01±1.1e-01	7.1(2,6)	0.54	2.66*
		2	$1.7e-01\pm3.3e-01$	0.3(2,3)	0.04	0.52
Canopy height	Shannon	$\frac{2}{3}$	$-3.0e + 00 \pm 9.0e - 01$	11.1(2,1)	0.08 0.92	-3.32
		$\frac{3}{4}$	-3.0e+00±3.0e-01	11.1(2,1)	0.92	-5.52
				2 2 (2 2)		
		1	$1.0e+01\pm2.1e+01$	0.2(2,6)	0.04	0.49
Canopy height	Uniform angle index	2	$-7.2e + 01 \pm 1.3e + 02$	0.3(2,3)	0.09	-0.56
10 0	O	3	$6.0e-02\pm3.9e+01$	0.0(2,1)	0.00	0.00
		4				
		1	$3.6e-04\pm6.9e-04$	0.3(2,10)	0.03	0.53
Canopy closure	Basal area CV	2	$3.5e-03\pm3.5e-03$	1.0(2,3)	0.24	0.98
Canopy closure	Dasar area CV	3	$1.9e-03\pm5.3e-03$	0.1(2,1)	0.11	0.35
		4				
		1	9.3e-03±8.2e-03	1.3(2,10)	0.11	1.13
		2	$-6.6e-02\pm7.9e-03$	69.7(2,3)	0.96	-8.35**
Canopy closure	Voronoi CV	3	$-2.5e-02\pm4.6e-03$	29.0(2,1)	0.97	-5.39
		4		() /		
		1	-1.6e-01±5.1e-01	0.1(2.10)	0.01	-0.31
Canopy closure		1	$-6.9e-01\pm1.1e+00$	$0.1(2,10) \\ 0.4(2,3)$	0.01 0.12	-0.63
	Mingling	$\frac{2}{3}$	$7.6e-02\pm4.1e+00$	0.4(2,3) 0.0(2,1)	0.12 0.00	-0.03 0.02
		4	7.0e-02±4.1e+00	0.0(2,1)	0.00	0.02
		1	$1.4e-04\pm4.0e-04$	0.1(2,10)	0.01	0.36
Canopy closure	Tree density	2	$8.5e-04\pm2.4e-04$	12.2(2,3)	0.80	3.50*
10	· · · · · · · · · · · · · · · · · · ·	3	$3.0e-03\pm4.3e-06$	499683.9(2,1)	1.00	706.88***
		4				
		1	$-7.6e-03\pm1.7e-02$	0.2(2,10)	0.02	-0.45
Canopy closure	Shannon	2	$8.5 \text{e-} 03 \pm 3.0 \text{e-} 02$	0.1(2,3)	0.03	0.28
Canopy closure	Shaimon	3	$1.9e-01\pm5.2e-01$	0.1(2,1)	0.12	0.37
		4				
		1	$-3.9e + 00 \pm 2.3e + 00$	2.9(2,10)	0.23	-1.71
		$\overline{2}$	$-1.2e + 01 \pm 9.3e + 00$	1.7(2,3)	0.36	-1.30
Canopy closure	Uniform angle index	3	$-6.9e + 00 \pm 3.9e - 01$	306.2(2,1)	1.00	-17.50*
		4		() /		
		1	$-4.5e + 01 \pm 2.9e + 01$	2.3(2,6)	0.28	-1.52
		$\frac{1}{2}$	$1.5e + 02 \pm 1.4e + 02$	1.1(2,3)	0.28 0.27	-1.52 1.05
Foliage density	Basal area CV	$\frac{2}{3}$	$1.8e + 02 \pm 1.4e + 02$ $1.8e + 02 \pm 8.9e + 01$	4.2(2,1)	0.21 0.81	$\frac{1.05}{2.06}$
		$\frac{3}{4}$	1.06+02±0.96+01	4.2(2,1)	0.01	2.00
				0.2/2.3		
		1	$3.5e + 01 \pm 5.0e + 02$	0.0(2,6)	0.00	0.07
Foliage density	Voronoi CV	2	$-7.7e + 02 \pm 1.5e + 03$	0.3(2,3)	0.08	-0.51
G ,		3	$2.7e + 02 \pm 8.7e + 02$	0.1(2,1)	0.09	0.31
		4				
		1	$4.5e + 03 \pm 2.5e + 04$	0.0(2,6)	0.01	0.18
Foliama dan -:t	Mingling	2	$8.0e + 02 \pm 4.7e + 04$	0.0(2,3)	0.00	0.02
Foliage density	Mingling	3	$1.5e + 05 \pm 2.0e + 04$	54.1(2,1)	0.98	7.35

		4				
		1	$8.8e + 00 \pm 2.0e + 01$	0.2(2,6)	0.03	0.45
Foliomo domaites	The density	2	$1.1e+01\pm2.1e+01$	0.3(2,3)	0.08	0.51
Foliage density	Tree density	3	$-1.3e + 01 \pm 1.1e + 02$	0.0(2,1)	0.01	-0.12
		4				
		1	$2.5e + 02 \pm 8.1e + 02$	0.1(2,6)	0.02	0.31
T) 1: 1 ::	CI	2	$5.0e + 02 \pm 1.2e + 03$	0.2(2,3)	0.05	0.42
Foliage density	Shannon	3	$1.8e + 04 \pm 9.1e + 03$	3.9(2,1)	0.80	1.98
		4		, ,		
		1	$-1.1e + 05 \pm 1.0e + 05$	1.3(2,6)	0.18	-1.15
T 1: 1 :	TT : C 1 : 1	2	$1.2e + 05 \pm 4.7e + 05$	0.1(2,3)	0.02	0.25
Foliage density	Uniform angle index	3	$4.3e+04\pm2.5e+05$	0.0(2,1)	0.03	0.18
		4		, ,		
		1	-1.0e-01±6.1e-01	0.0(2,6)	0.00	-0.17
Canana magaita	Dagal area CV	2	$-2.2e+00\pm2.2e+00$	1.1(2,3)	0.26	-1.03
Canopy rugosity	Basal area CV	3	$8.7e + 00 \pm 5.4e + 00$	2.6(2,1)	0.73	1.62
		4				
		1	$7.9e + 00 \pm 8.2e + 00$	0.9(2,6)	0.13	0.96
	Voronoi CV	2	$3.5e+01\pm1.3e+01$	6.8(2,3)	0.69	2.61
Canopy rugosity	VOLOTIOL C A	3	$1.8e + 01 \pm 4.2e + 01$	0.2(2,1)	0.15	0.42
		4				
		1	$-5.9e + 02 \pm 3.6e + 02$	2.7(2,6)	0.31	-1.63
Canopy rugosity	Mingling	2	$8.5e + 02 \pm 5.2e + 02$	2.7(2,3)	0.47	1.63
Canopy rugosity	Mingling	3	$7.2e + 03 \pm 1.7e + 03$	17.6(2,1)	0.95	4.19
		4				
		1	$-1.9e-01\pm3.4e-01$	0.3(2,6)	0.05	-0.56
Canopy rugosity	Tree density	2	$-4.6e-01\pm2.1e-01$	4.9(2,3)	0.62	-2.22
Canopy rugosity	Tree density	3	$-1.2e + 00 \pm 5.4e + 00$	0.0(2,1)	0.05	-0.22
		4				
		1	$-2.4e+01\pm1.0e+01$	5.3(2,6)	0.47	-2.31
Canopy rugosity	Shannon	2	$6.4e + 00 \pm 1.8e + 01$	0.1(2,3)	0.04	0.35
Canopy rugosity	Shannon	3	$8.5e + 02 \pm 5.4e + 02$	2.5(2,1)	0.71	1.57
		4				
		1	$-2.6e + 03 \pm 1.6e + 03$	2.5(2,6)	0.30	-1.58
Canopy rugosity	Uniform angle index	2	$1.0e+04\pm4.1e+03$	6.1(2,3)	0.67	2.47
Canopy rugosity	omnorm angle muex	3	$3.4e + 03 \pm 1.2e + 04$	0.1(2,1)	0.07	0.28
		4				

Table 3: Summary statistics of bivariate linear models comparing canopy complexity metrics with diversity and stand structural metrics across all vegetation types. Slope refers to the slope of the predictor term in the model, \pm 1 standard error. T is the t-value of the slope of the predictor term in the model, Asterisks indicate the p-value of these terms (***<0.001, **<0.01, *<0.05).

Response	Predictor	Slope	F	\mathbb{R}^2	Т
	Basal area CV	$8.7e + 01 \pm 3.0e + 01$	8.6(2,167)	0.05	2.93**
Foliage density	Hegyi	$7.8e + 03 \pm 1.6e + 03$	25.5(2,184)	0.12	5.05***
	Shannon	$3.2e+03\pm1.1e+03$	8.9(2,180)	0.05	2.98**
	Basal area CV	$1.2e-03\pm4.8e-04$	6.3(2,168)	0.04	2.52*
Canopy closure	Hegyi	$2.4e-01\pm2.1e-02$	132.8(2,185)	0.42	11.52***
	Shannon	$4.7e-02\pm1.7e-02$	7.3(2,181)	0.04	2.70**
	Basal area CV	$4.1e+00\pm3.0e+00$	1.9(2,167)	0.01	1.37
Foliage uniformity	Hegyi	$4.0e+02\pm1.6e+02$	6.2(2,184)	0.03	2.49*
	Shannon	$2.2e+02\pm1.1e+02$	4.1(2,180)	0.02	2.04*
	Basal area CV	$3.2 \text{e-} 02 \pm 7.6 \text{e-} 03$	17.6(2,167)	0.10	4.20***
Layer diversity	Hegyi	$2.7e+00\pm3.9e-01$	46.8(2,184)	0.20	6.84***
	Shannon	$1.1e+00\pm2.7e-01$	16.8(2,180)	0.09	4.10***
	Basal area CV	$3.0e-02\pm5.0e-02$	0.4(2,16)	0.02	0.60
	Voronoi CV	$7.5 \text{e-}01 \pm 5.9 \text{e-}01$	1.6(2,16)	0.09	1.26
Con ones nou alon ogg	Mingling	$-2.8e+01\pm3.3e+01$	0.7(2,16)	0.04	-0.86
Canopy roughness	Tree density	$-2.6e-02\pm1.7e-02$	2.3(2,16)	0.12	-1.51
	Shannon	$-1.9e + 00 \pm 9.5e - 01$	4.0(2,16)	0.20	-2.01
	Uniform angle index	$1.6e + 02 \pm 1.6e + 02$	1.0(2,16)	0.06	0.98
Community in the	Basal area CV	$7.1 \text{e-} 03 \pm 7.3 \text{e-} 03$	0.9(2,16)	0.06	0.97
	Voronoi CV	$-4.7e-02\pm9.1e-02$	0.3(2,16)	0.02	-0.52
	Mingling	$3.8e+00\pm4.8e+00$	0.6(2,16)	0.04	0.79
Canopy height	Tree density	$4.3e-03\pm2.5e-03$	3.1(2,16)	0.16	1.76
	Shannon	$3.3e-01\pm1.3e-01$	6.0(2,16)	0.27	2.45*
	Uniform angle index	$-2.2e+01\pm2.4e+01$	0.8(2,16)	0.05	-0.90
	Basal area CV	$8.5 \text{e-}04 \pm 5.7 \text{e-}04$	2.2(2,20)	0.10	1.50
	Voronoi CV	$2.4e-03\pm5.8e-03$	0.2(2,20)	0.01	0.41
Canopy closure	Mingling	$7.2e-03\pm3.7e-01$	0.0(2,20)	0.00	0.02
Canopy closure	Tree density	$4.7e-04\pm1.9e-04$	6.3(2,20)	0.24	2.50*
	Shannon	$1.0e-02\pm1.2e-02$	0.7(2,20)	0.04	0.86
	Uniform angle index	$-3.4e + 00 \pm 1.7e + 00$	3.9(2,20)	0.16	-1.98
	Basal area CV	$5.8e + 01 \pm 3.2e + 01$	3.3(2,16)	0.17	1.80
	Voronoi CV	$5.8e + 02 \pm 4.1e + 02$	2.1(2,16)	0.11	1.43
Foliage density	Mingling	$6.6e + 03 \pm 2.3e + 04$	0.1(2,16)	0.01	0.29
Tollage delibity	Tree density	$3.0e+01\pm1.0e+01$	8.6(2,16)	0.35	2.93**
	Shannon	$1.1e + 03 \pm 6.9e + 02$	2.5(2,16)	0.13	1.57
	Uniform angle index	$-2.1e+04\pm1.1e+05$	0.0(2,16)	0.00	-0.18
	Basal area CV	$-1.0e + 00 \pm 5.3e - 01$	3.7(2,16)	0.19	-1.92
	Voronoi CV	$-6.0e + 00 \pm 7.0e + 00$	0.7(2,16)	0.04	-0.86
Canopy rugosity	Mingling	$1.3e + 02 \pm 3.8e + 02$	0.1(2,16)	0.01	0.33
Carropy ragosity	Tree density	$-5.2e-01\pm1.7e-01$	10.0(2,16)	0.38	-3.16**
	Shannon	$-1.3e+01\pm1.2e+01$	1.2(2,16)	0.07	-1.11
	Uniform angle index	$-1.8e + 03 \pm 1.9e + 03$	0.9(2,16)	0.06	-0.97

Table 4: Explanatory variables included in the best model for each subplot canopy structure variable. ΔAIC shows the difference in model AIC value compared to a null model which included only the random effects of vegetation type and plot. Positive ΔAIC values >2 indicate that the model is of better quality than the null model. R^2_c is the R^2 of the best model, while R^2_m is the R^2 of the model fixed effects only.

Response	Hegyi	Shannon	Basal area CV	$\Delta { m AIC}$	${ m R^2_{\ c}}$	R^2_{m}
Layer diversity	✓	✓	√	37.0	0.50	0.17
Foliage density	\checkmark		\checkmark	47.6	0.27	0.09
Foliage uniformity	\checkmark			13.1	0.28	0.02
Canopy closure	\checkmark	\checkmark		101.9	0.60	0.46

Table 5: Explanatory variables included in the best linear model for each plot-level canopy complexity metric. ΔAIC shows the difference in model AIC value compared to a null model. Positive ΔAIC values >2 indicate that the model is of better quality than the null model.

Response	Shannon	Tree density	Basal area CV	Mingling	Uniform angle index	Voronoi CV	$\Delta { m AIC}$	\mathbb{R}^2	Prob.
Foliage density	✓					√	5.8	0.42	< 0.05
Canopy closure			\checkmark		\checkmark		5.8	0.42	< 0.05
Canopy height	\checkmark			\checkmark			8.2	0.49	< 0.01
Canopy roughness	\checkmark			\checkmark			2.5	0.30	0.07
Canopy rugosity		\checkmark			\checkmark		6.9	0.45	< 0.05