# Navigating the Molecular Maze: A Python-Powered Approach to Virtual Drug Screening

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#### Abstract

The COVID-19 pandemic has highlighted the power of using computational methods for virtual drug screening. However, the molecular search space is enormous and protein docking methods are still computationally intractable without access to the world's largest supercomputers. Instead, researchers are using AI methods to help guide docking campaigns. In such approaches, a lightweight surrogate model is trained and then used to identify promising candidates for screening. We present ParslDock, a Python-based pipeline using the Parsl parallel programming library and the K-Nearest Neighbors machine learning model to screen a huge molecular space of molecules against arbitrary receptors. We achieved a 38X speedup with ParslDock compared to a brute-force docking approach.

## 1 Introduction

Protein docking is a key computational method used in molecular biology to predict the structure of protein complexes formed when two or more proteins interact. This technique is vital in understanding biological processes and designing therapeutic drugs. Essentially, protein docking involves simulating the process by which proteins fit together or 'dock' to form a stable complex. This is akin to finding the correct way two puzzle pieces fit together among myriad possibilities.

Docking scoring functions estimate the binding free energy or affinity between a ligand and a protein. The lower the score, the more energetically favorable the binding interaction. The docking process is driven by several factors, including shape complementarity, electrostatic attractions, and hydrophobic interactions. As proteins are highly flexible and complex molecules, the problem of predicting their interactions is computationally demanding.

A typical docking computation can take over 10 minutes on 1-core; a typical workload involves multiple protein receptors (e.g., we found studies with 15

receptors) with millions of possible ligands to dock to (e.g., we found studies with 13M ligands), yielding a total compute complexity for a brute force approach to be over 32M CPU-hours (3710 years in a serial computation on a single core). [Clyde et al.(2021)]

## 2 Problem Statement

ParslDock aims to identify optimal ligands from a large dataset of potential molecules by efficiently combining simulation (docking) and ML algorithms on HPC resources. The goal is to reduce the computational complexity of the brute force docking application through ML methods. Our simulation runs the Docking application which aims to predict the optimal binding conformation of a protein receptor and ligand using a binding affinity scoring function. The main challenge is to identify a ML model that is sufficiently accurate to enable the identification of the best ligands from a relatively small subset of ligand dockings.

# 3 Proposed Solution

We propose ParslDock, an ML-based system that enables scalable and flexible screening of molecules. We assume that users provide a target receptor (in PDBQT format) and a list of candidate molecules (as SMILES strings). We aim for ParslDock to efficiently use allocated resources (e.g., from parallel or distributed systems) combining ML models and Monte Carlo Simulations.

#### 3.1 Software

We used AutoDock Vina (a leading docking implementation) that performs docking and utilizes a scoring function and gradient-based optimization algorithm. [Eberhardt et al.(2021)]

We used the Parsl parallel programming library to execute parts of the application in parallel. The power of Parsl lies in its ability to manage and coordinate the execution of tasks across a range of computing resources, from multicore workstations to HPC systems. Parsl's programming model relies on defining apps (to represent functions that can execute concurrently). Apps return 'futures', which are essentially placeholders for results that are computed asynchronously. Parsl automates the management of these futures, effectively handling task dependencies, synchronization, and data movement. Parsl allows users to write code that is agnostic of the underlying computing infrastructure, hence making scripts portable and scalable. [Babuji et al.(2019)]

#### 3.2 Machine Learning Model

We used a K-Nearest Neighbor (KNN) model. [Cover and Hart(1967)] Each molecule (a SMILES string) is converted into a Morgan Fingerprint that is

represented as a bit-vector.

The SMILES "CCN(CCCC(C)NC1=C2C=CC (=CC2=NC=C1)Cl)CCO" represents the Hydroxychloroquine molecule. This SMILES string is converted into the bit-vector "111001001111010111

## 3.3 ParslDock Pipeline

The ParslDock pipeline (Figure 1) is composed of 7 stages:

- 1. Dataset 4M: Input data is a target receptor (PDBQT) and a random list of 100K molecules (SMILES)
- 2. Data Format: Data preparation for AutoDock Vina docking; SMILES string to PDB file to PDBQT file
- 3. Docking: Executes Monte Carlo simulations between a protein receptor PDBQT file and a ligand PDBQT file, yielding a binding-affinity score output
- 4. Morgan Fingerprints: Generated 128-bit vector with a depth of 8 from a SMILES string [Pattanaik and Coley(2020)]
- 5. Machine Learning: Morgan Fingerprints and docking scores are paired as the input to KNN model to identify correlations between fingerprints and docking scores
- 6. Dataset Top-4K: The top 4K ligands identified based on the predicted docking scores (lowest binding-affinity scores) of all 4M molecules
- 7. Docking: Runs Monte Carlo simulations on the Top-4K subset of data to obtain a final list of top molecules ordered by docking scores

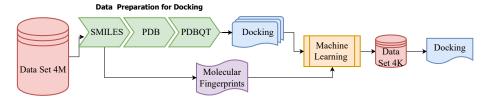


Figure 1: ParslDock pipeline

Most of these steps can be run concurrently across a set of input molecules. We implement each step as a Parsl app and establish dependencies between each

step. This allows us to scale out execution of each step across input data. All our code and datasets can be found on Github. [Raicu(2023)]

# 4 Software Stack, Testbeds, and Dataset

#### 4.1 Software Stack

The Programming Tools we used involved Python 3.8.3 to implement the computational pipeline. We used Parsl 1.3.0.dev0 to parallelize various stages of the computational pipeline. We made extensive use of Jupyter Notebook 6.5.4 in the development of ParslDock.

The main docking library we used was AutoDock Vina 1.2.3 which utilizes a scoring function and gradient-based optimization algorithm. We used the Visual Molecular Dynamics 1.9.3 application to visualize and analyze molecular simulations. Finally, we used Py3Dmol 2.0.3 to enable interactive 3D molecular visualizations. The machine learning library that implemented the KNN was Scikit-learn; we made use of other popular scientific and data analysis libraries in Python, such as NumPy, Pandas, and Matplotlib.

#### 4.2 Testbeds

The Hardware we use includes two testbeds to evaluate ParslDock. We conduct experiments on two testbeds: 8c-laptop and 192c-server. The laptop was a MacBook Pro from 2019 with an 8-core Intel Core i9 CPU, 2.4GHz, 64GB DDR4, 8TB NVMe, running MacOS 12.6.3. The server had 8x 24-core Intel Xeon Sky Lake CPUs (for a total of 192-cores), 2.1GHz, 786GB DDR4, 16TB SSD, running Ubuntu Linux 22.04.

#### 4.3 Datset

The 0.9 GB Figshare Dataset we used contained 8 million ligands stored as SMILES strings.

## 5 Evaluation

#### 5.1 Docking Scores

We used 4 million ligands from the 8 million ligand dataset and performed docking against the 1IEP receptor protein. This resulted in a new dataset that included 4M scores that are represented in the Figure 2. The scores follow a normal distribution and range from -187.3 to 46.65, with a median of -8.654. Our goal was to identify the top 0.1% of the ligands based on the docking score (green line at -34.32). All the ligand docking scores less than or equal to -34.32 were labeled as "best" while all docking scores higher than -34.32 were labeled as "worst".

#### 5.2 KNN Model

Once we obtained the docking scores for the entire dataset, we selected 100K random ligands and used the docking scores to build a KNN model as seen in Figure 3. We first explored use of the entire 100K random samples, which yielded surprisingly high accuracy results in the 99%+ range. Upon close inspection, we found that our dataset had a minority class (100 samples) and a majority class (99,900 samples). KNN are well known to perform poorly for unbalanced datasets. We set out to balance it by pulling a random sample of size 100 from the majority class of 99,900 samples.

Our final dataset we used to build the KNN was only 200 samples large, representing two classes that were of equal size. We tried larger sample sizes, but we did not see a significantly improved accuracy on the KNN testing data. We found that a relatively small number of molecules are needed to build the KNN while achieving good accuracy, so the  $O(n^2 * d)$  time and space complexity of KNN is tractable for modest values of n and d.

We balanced the dataset by choosing 100 of each. Larger samples did not significantly improve accuracy. We explored different values of dataset sizes (from 10K to 10M samples), and determined that 100K offers good enough accuracy of 87%. The number of neighbors we settled on for the KNN was 5. Different values of K did not yield significantly and consistently better results.

## 5.3 Molecular Fingerprints

We conducted a hyperparameter search of the "depth" (radius around atoms) and "size" (number of bits) of fingerprint. Figure 4 shows the accuracy of the KNN model on a training/testing dataset partitioned in 70% training and 30% testing. The best configuration (yellow) is when size is 128 and depth is 8, achieving an accuracy of 87%. Note that sub-optimal parameters for the Morgan Fingerprint can easily yield under 50% accuracy. One critical variable in building the machine learning model was the input data: a Morgan Fingerprint

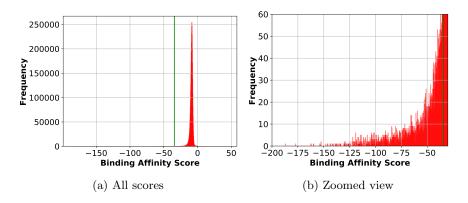


Figure 2: Binding affinity score for docking of 4M molecules.

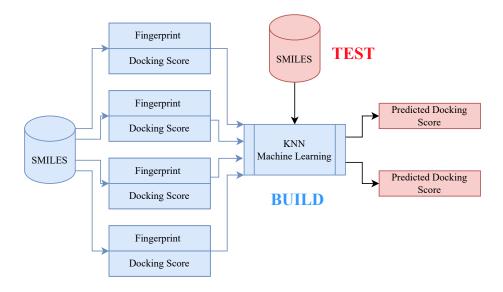


Figure 3: Machine Learning Workflow highlighting inputs and outputs

and a docking score.

The Morgan Fingerprint itself has two different parameters that significantly changes the fingerprint contents: depth and length. "Depth" typically refers to the radius parameter used in generating the fingerprints. Morgan fingerprints are circular fingerprints that capture local chemical environments around atoms in a molecule. The depth of the fingerprint determines how far from each atom the circular neighborhoods extend. By adjusting the depth parameter, you control the level of detail captured in the fingerprint. A larger radius captures more extended structural patterns and interactions, while a smaller radius focuses on local substructures.

The second parameter is "length" referring to the number of bits or dimensions used to represent the fingerprint. Each bit in the fingerprint corresponds to a specific substructure pattern or feature. The length of the fingerprint determines the level of granularity at which the molecular structure is encoded. In summary, the length of a Morgan fingerprint determines the dimensionality of the binary representation used to capture the molecular structure's features, and it plays a role in balancing representation power and computational considerations.

In order to identify the best fingerprint depth and size, we conducted a hyperparameter search of different fingerprint length and depth and measured the accuracy of the KNN model on a training/testing dataset partitioned in 70% training and 30% testing. The figure below shows the bright yellow square where fingerprint size is 128 and depth is 8, achieving an accuracy of 87%. Note that sub-optimal parameters for the Morgan Fingerprint can easily yield under 50% accuracy.

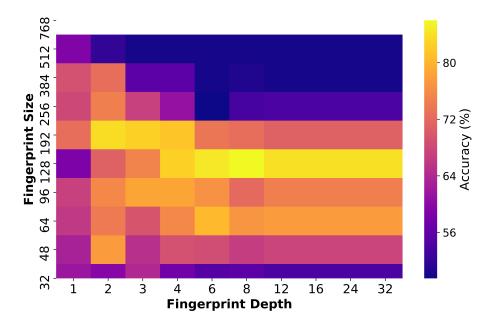


Figure 4: 2D-Heatmap showing KNN accuracy for fingerprint size vs depth

#### 5.4 Performance

#### 5.4.1 ParslDock vs Brute Force Docking

Putting it all together from the start (SMILES) to the end (top-4k ligands), we evaluated the time taken to run the entire ParslDockpipeline on two systems: an 8-core laptop and a 192-core server. Figure 5 shows the performance of the entire pipeline on two systems. The various stages are identified below in the different colors, from all the data transformations to PDF and PDBQT files, docking, machine learning building and testing, and final ligand selection of the best ligands identified. Most stages used Parsl to parallelize execution across the multicore systems in our testbed (the only exception was the building of the KNN, which was a significant part of the total pipeline). The speedup obtained on ParslDock compared to the brute force docking of the entire 4M ligands was 38X on the laptop and 37X on the server.

#### 5.4.2 Parsl Optimizations

Key to this work is the workflow graph generated by ParslDock. Each task consists of a molecule that is formatted and passed to a machine learning module for inference. This generates a bag-of-tasks graph, such a task graph is highly parallelizable. We use this scientific application to demonstrate the improved performance achieved by our optimizations.

ParslDock executed on the Mystic testbed mentioned previously. The dock-

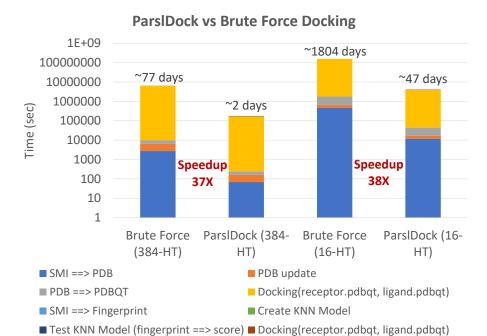


Figure 5: ParslDock and Brute Force Docking end-to-end pipeline execution time

ing simulation displayed Figure 6, Figure 7, and Figure 8 display runtime with varying numbers of workers and batch size. The simulation was run across 100k total molecules For serial, standard Parsl, and CDFK, the best runtimes are achieved with the largest batch size. This suggests that there is still work to be done to improve Parsl for fine-grained tasks.

Figure 6 shows the runtime of the docking simulation as a function of the batch size using serial code. Using a batch size of 1000 molecules the serial code achieves a runtime of around 35 seconds.

Figure 7 displays the runtime achieved by standard Parsl. Standard Parsl achieves its lowest runtime using 8 workers with a batch size of 1000, 11s. With standard Parsl using many workers or few workers returns a high runtime, 100 seconds.

Figure 8 shows the runtime achieved by CDFK Parsl. It scales linearly with the number of workers used, plateauing with hundreds of workers. CDFK Parsl with 128 workers and a batch size of 1000 simulates 100k molecules in 1.5 seconds. This runtime is 10x faster than standard Parsl's lowest runtime and 30x faster than serial.

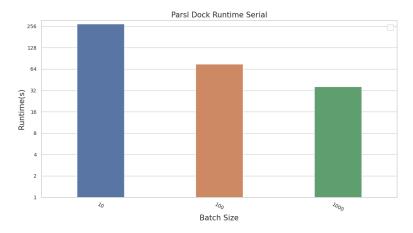


Figure 6: ParslDock runtime Serial

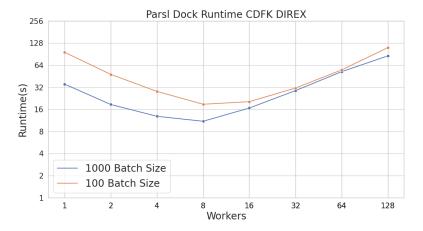


Figure 7: Standard Parsl

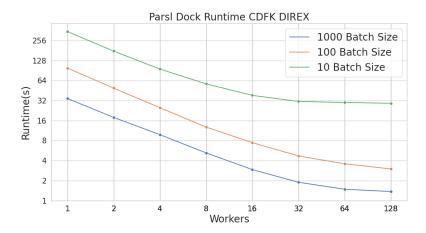


Figure 8: CDFK

## 6 Conclusions

Protein docking is a complex process that can be computationally expensive and time-consuming. ParslDock is a Python-powered automated pipeline that uses ML to accelerate the docking process and reduce compute costs by up to 38X. Our performance evaluation showed linear scalability from an 8-core laptop to a 192-core server. With further improvements, we believe we can bring down the computational requirements to the point that ParslDock will be tractable on a modern day personal computer.

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