

Supplementary Material

1 SUPPLEMENTARY METHODS

1.1 TC recruitment

Under non-inflammatory conditions, it was assumed that TC entry and exit remain constant and TCs occupy a constant percentage (55%) of the total paracortical volume. The model represents a LN of 0.113-0.268 mm³, implying a LN mass of 0.18-0.44mg (based on collaborative unpublished measurements of murine popliteal LN mass versus volume). Reported lymphocyte recruitment for a popliteal LN of 1.15g is 4x10⁷ lymphocytes/hour and up to 40% of lymphocytes are B cells (Cahill et al., 1976; Habenicht et al., 2016; Battaglia et al., 2003). The model represents half a paracortex, therefore TC recruitment rate was estimated as 1950-9000 TCs/hour under non-antigenic conditions. Naive TC transit time through the LN (T_{res}) was estimated as 6-24 hours (Tomura et al., 2008).

1.2 Agents and agent migration

The agents were designated as members of the TC or DC class (Figure S1). Simulated DCs appear in the paracortex at a constant entry rate for 2 days, subsiding over the following 12 hours. Entry rate of DCs was scaled from counts of migrating DCs and initial TCs in a murine LN post-immunisation, with DCs totalling 4% of cells present (Acton et al., 2014). TC migration was assumed to follow a random walk with pauses. Previous models have described TC migration with Brownian motion, a random-walk with persistence, run and tumble, and Lévy walks amongst other methods, partly due to differing reports of *in-vivo* migration. (Celli et al., 2012; Bogle and Dunbar, 2008; Brown et al., 2018; Harris et al., 2012).

1.3 Agent interaction and signal integration

During TC-DC interaction, TCs remain stationary. Antigenic signal is presented by the DCs in the form of representative values of MHC I or MHC II, and decays with time (t) with the form :

$$MHC\text{I}(t) = MHC\text{I}_i(0.5)^{\frac{t}{MHC\text{I}t_{1/2}}} \quad (\text{S1})$$

$MHC\text{I/II}_i$ is initial MHC I or MHC II presented with estimated MHC half-lives $MHC\text{I}t_{1/2}$ and $MHC\text{II}t_{1/2}$. During cognate TC-DC interaction, CD4⁺/CD8⁺ TCs gain (S) at a rate proportional to the MHC I or MHC II signal respectively (Figure 2.C). Accumulated stimulation decays at a constant rate. Total change in TC stimulation is therefore given according to the first order rate equation:

$$\frac{dS}{dt} = K_s MHC\text{II}(t) - \lambda_S S(t) \quad (\text{S2})$$

Where K_s is a rate constant and λ_S is a decay factor. Accumulated simulation decays to a minimal value of S=1 to allow differentiation between cognate TCs that gain and lose simulation (S=1) and those that never gain simulation (S=0). Probability of cognate CD4⁺ activation (P_{a4+}) or cognate CD8⁺ activation (P_{a8+}) is calculated with a sigmoidal function given by:

$$P_{a4+} = \frac{1}{1 + e^{\frac{-S - Act\mu_4}{Actl_4}}} \quad (\text{S3})$$

Parameter $\text{Act}\mu_4$ determines the value of S required for 50% probability of activation and $\text{Act}\ell_4$ determines steepness of sigmoid inflection (Figure 2.D). Activation probability for CD8⁺ TCs (P_{a8+}) is determined with a sigmoid curve using a lower inflection point, ($\text{Act}\mu_8$), than for CD4⁺ TCs, unless the interacting agDC is licensed (see 2.4). Model parameters were estimated such that TC activation became apparent 8-15 hours post DC-arrival (Hugues et al., 2004; Mempel et al., 2004; Stoll et al., 2002). This method of signal integration and the subsequent progressive patterns of TC proliferation and differentiation is supported by *in-vivo* observations and modelling descriptions (Stoll et al., 2002; Germain and Stefanová, 1999; Rachmilewitz and Lanzavecchia, 2002; Lanzavecchia and Sallusto, 2002). It is assumed that co-stimulatory requirements are met as agDCs are highly efficient antigen-presenting cells.

Post-activation, proliferation is possible every 11 (CD4⁺) or 9 (CD8⁺) $\pm 1\text{hr}$ (De Boer et al., 2003; Foulds et al., 2002; Nelson et al., 2015; Tubo et al., 2013). Differentiation into effector or memory cells is possible after $>=4$ divisions, with differentiation probability determined with a second set of sigmoidal probability curves with midpoint $\text{Dif}\mu_{4+}$ and $\text{Dif}\mu_{8+}$ respectively (Miller et al., 2002; Linderman et al., 2010). Greater CD4⁺ TCs dependence on continued stimulation for differentiation than CD8⁺ TCs was implemented by using a higher minimum threshold of accumulated stimulation for CD4⁺ differentiation than for CD8⁺ TCs (Schrum et al., 2005; Wong and Pamer, 2001; Kaech and Ahmed, 2001; van Stipdonk et al., 2001; Shaulov and Murali-Krishna, 2008). Effectors TCs that underwent < 8 proliferations were less likely to differentiate into memory cells than those that underwent >8 proliferations. This was implemented by assigning differentiation ratios of dif_{early} and dif_{late} to the two different subsets ($<=8$ or >8 divisions).

During TC activation and proliferation, S1P₁r expression was estimated from several *in-silico* studies. Expression change was considered following TC migration velocity into areas of high S1P concentration, changes in TC egress from the LN and changes in S1P₁R expression relative to naive S1P₁r expression following TC activation and with subsequent proliferation (Figure 2.F) (Pham et al., 2008; Matloubian et al., 2004; Garris et al., 2014).

2 SUPPLEMENTARY TABLES AND FIGURES

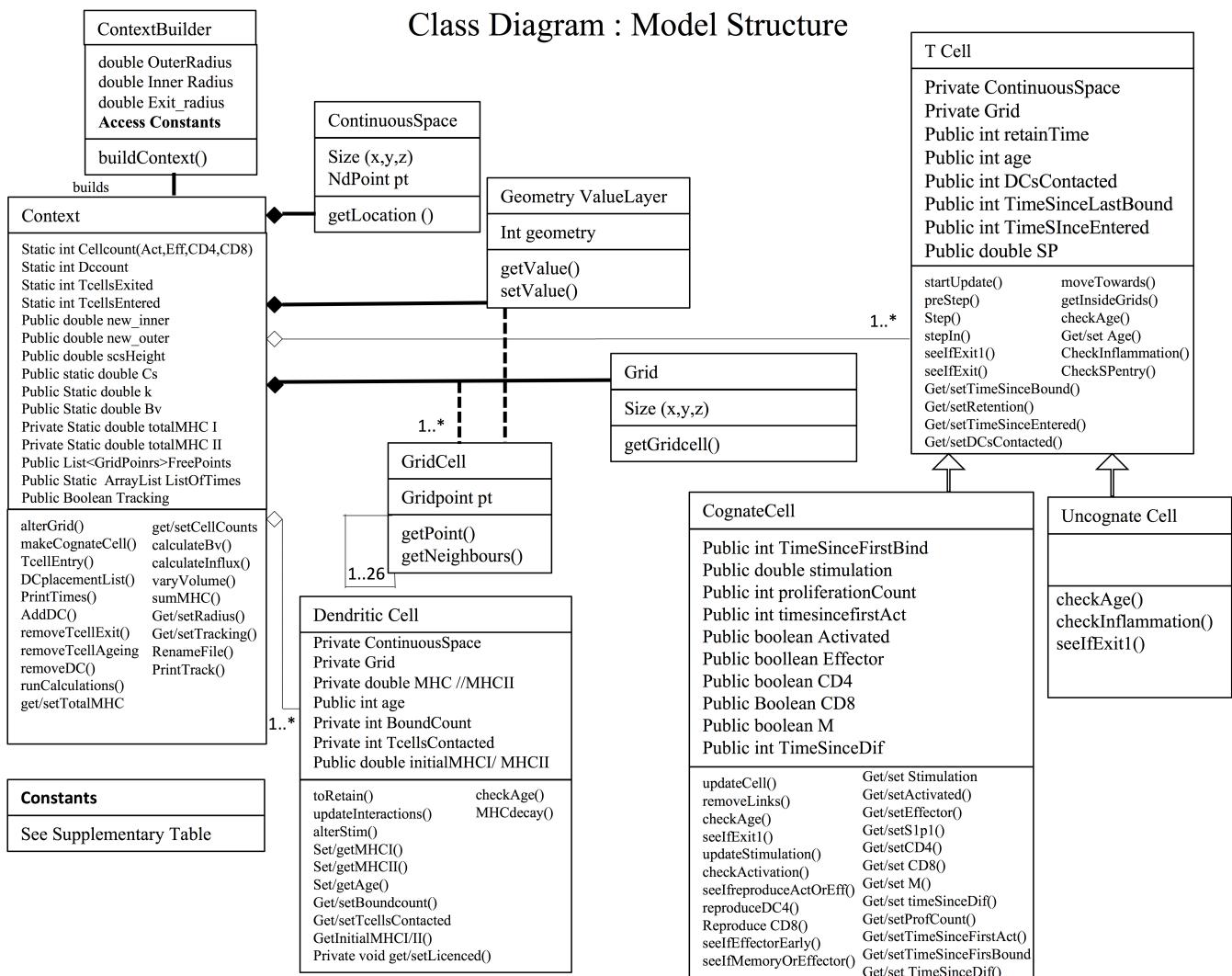


Figure S1. A class diagram displaying the underlying ABM structure. The model is constructed using instructions in the 'context builder' class. The entire modelling domain is described by the context class, and each compartment of the domain is described by the GridCell class. In a 3D simulation, each grid cell can be queried to identify the 26 neighbouring grids and how many agents they contain. The TC class is a template for the T cell object produced and is instantiated thousands of times to create T cells with the same variables but slightly different values. A subclass of cognate T cells extends the template to contains more methods and variables relating to interaction and proliferative response.

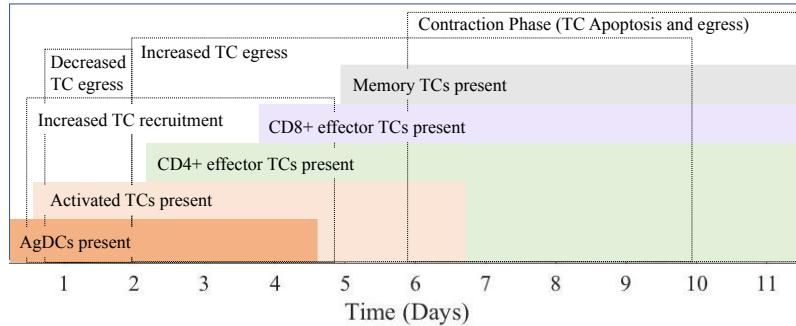


Figure S2. Captured phases of TC trafficking and response to AgDC stimuli. Changes in proliferation and differentiation continued after the initial stimulus was no longer present. TC recruitment and TC egress changes also accompanied the response.

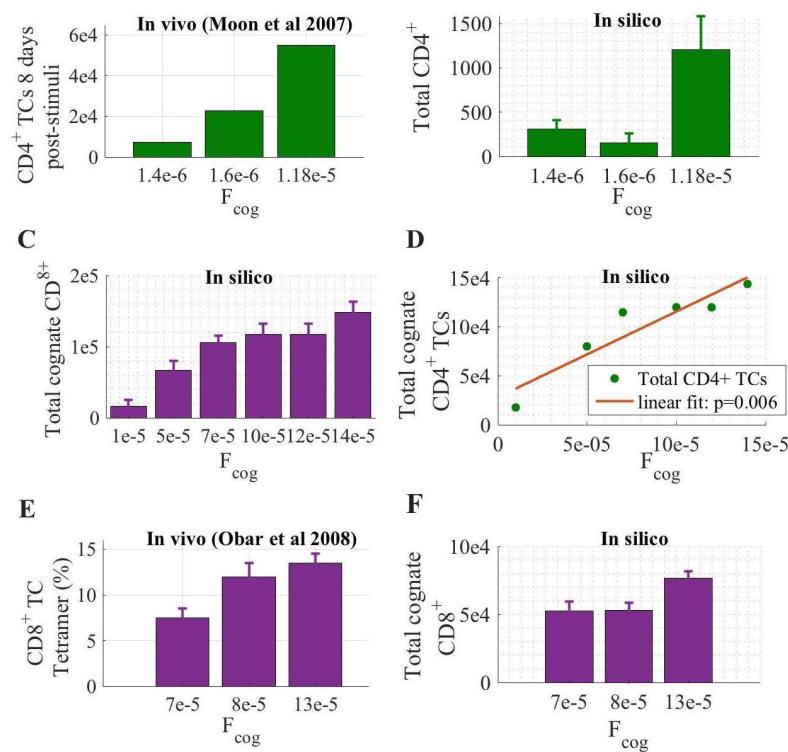


Figure S3. TC responses in-silico and in-vivo when proportion of cognate TCs was varied. A. CD4⁺ magnitude of response in dLNs of mice to injected antigen correlated to starting estimated frequency of cognate TCs in a sample of 1×10^7 TCs. B. Results in-silico showed an overall increase in response with increasing F_{cog} ($n=8$). Simulations using a wider range of F_{cog} values (C,D) confirmed no. of total cognate CD4⁺ TCs and CD8⁺ TCs increased linearly with F_{cog} . E. Mice were infected with VSV-M45 or VSV-ova with starting precursor CD8⁺ frequencies of 7×10^{-5} , 8×10^{-5} and 13×10^{-5} respectively. The peak number of resulting TCs as a percent of overall CD8⁺ TCs present is shown. F. Simulations using the same F_{cog} in-silico showed a similar increasing trend with similar increase rate.

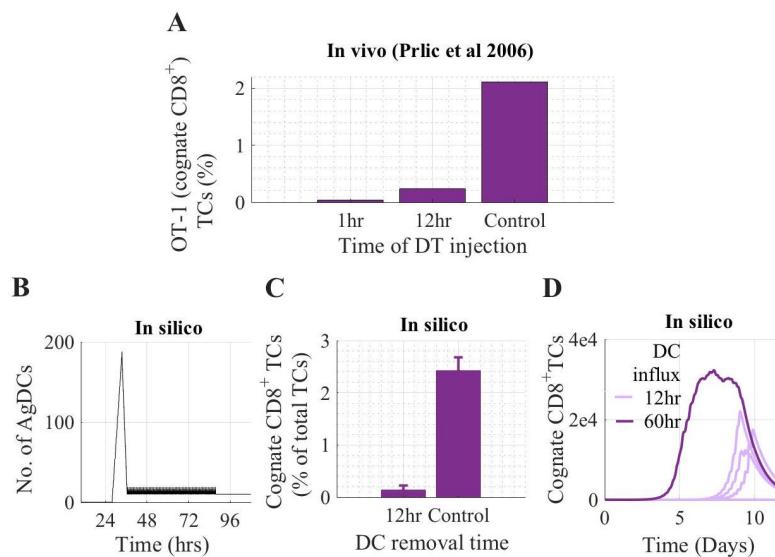


Figure S4. TC responses in-silico and in-vivo when a stimulus is abruptly abolished. A. Transgenic Rats were used that would eliminate specific injected agDCs when the rat was injected with DT within 12 hours. The rats were injected with OT-1 CD8⁺ TCs specific for agDCs that were subsequently injected. DT was then injected at 1h, 12 and 48hr(not-shown) later, curtailing the time that the agDCs would normally spend in the LN. Adapted from Prlic et al 2006. B. The simulated disrupted input stimuli achieved by curtailing the 60hr DC influx at 12+-2hr. C-D. The results of simulations (n=8). Mean (+SEM) CD8⁺ TCs were reduced 91% when the stimulus was curtailed at 12hrs compared to sustained entry for 60hrs. D. individual CD8+ TC responses varied by a factor of 10 in an all or nothing response manner.

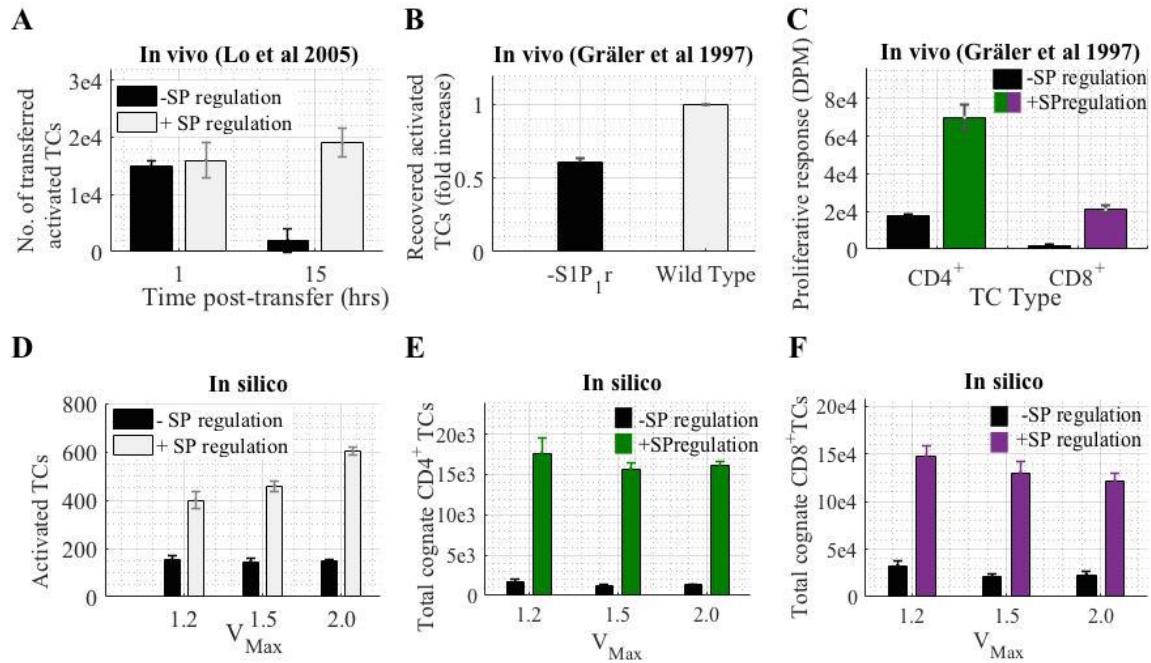


Figure S5. TC responses in-silico and in-vivo when S1P₁r down-regulation is inhibited. A. Pre-activated wildtype TCs (S1P₁r) and pre-activated TCs over-expressing S1P₁r (S1P₁r++) were transferred into mice and further entry of TCs was blocked. 15hrs later there was a 90% reduction in retention of S1P₁r++ activated TCs. Adapted from Lo et al 2005. B. The number of activated TCs in the LNs 24hours post-transfer dropped by 40% in transgenic mice with constitutive S1P₁r expression while (C) the proliferative CD4⁺ and CD8⁺ TC response decreased to 20% and 6% of that of the wild-type mice. Adapted from Gräler et al 1997. D-F. Simulation results (n=10) where S1P₁r down-regulation was prevented (-SP regulation), compared to baseline simulations (+SP regulation). Mean (+SEM) total number of activated TCs present was reduced 60%, 72% and 81% at V_{max}=1.2,1.5 and 2.0. E. The mean (+SEM) number of total CD8⁺ TCs, was diminished to 25,15 and 18% of control response at V_{Max}=1.2,1.5 and 2.0. F. CD4⁺ TCs were similarly diminished to 8-10% of control at all values of V_{max}.

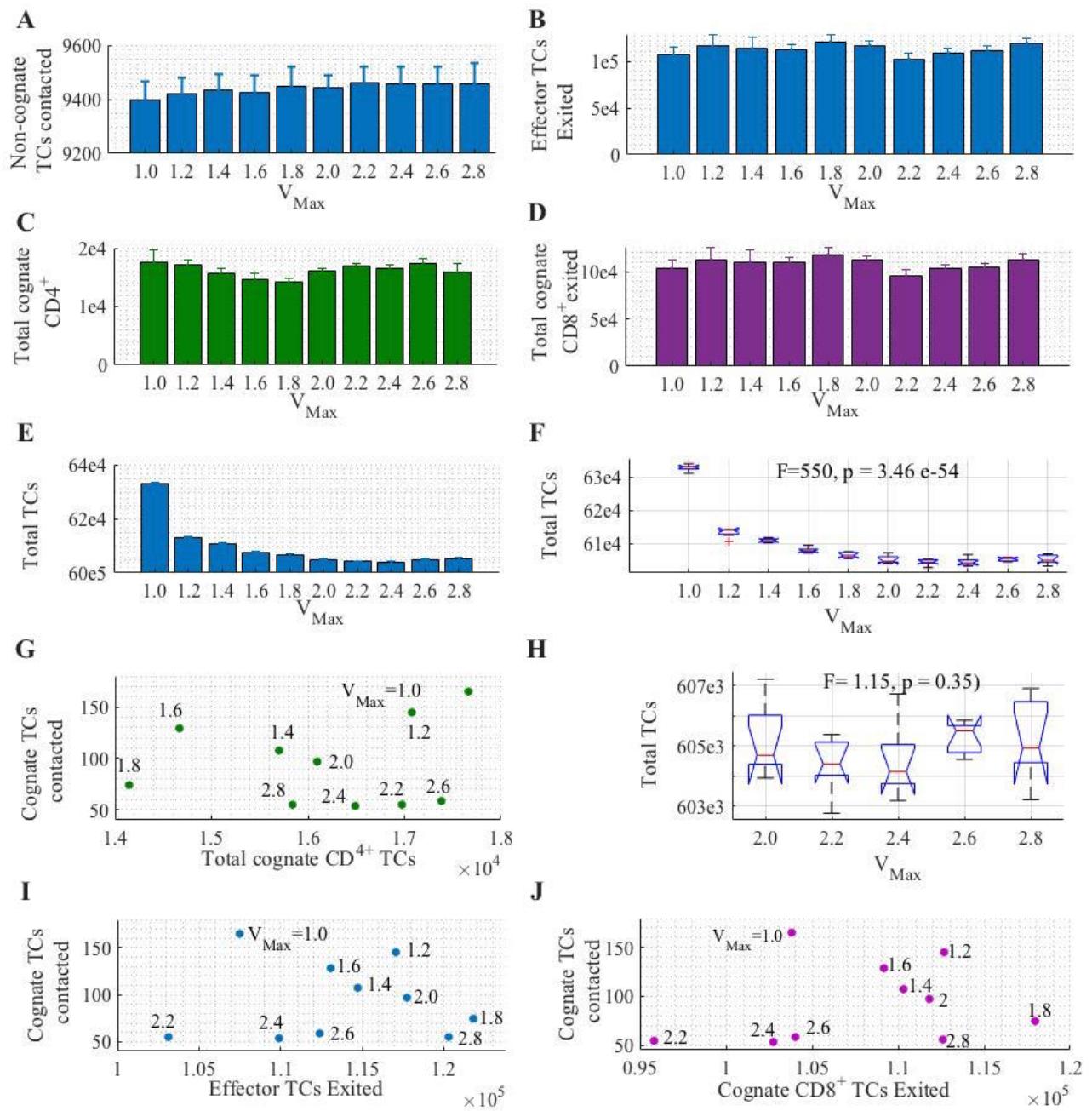


Figure S6. Additional data when V_{max} was varied with $T_{mid}=10e4$ TCs. A. The number of non-cognate TCs a DC contacted increased with LN swelling. B. No significant difference was apparent when effector TCs exited were plotted over time. C. Total cognate CD4⁺ TCs in the paracortex. D. Total CD8⁺ TCs that exited the paracortex. E,F. The total TCs present in the paracortex over the course of the simulation decreased as V_{max} increased, up to $V_{max}=2.0$ at which no further difference was observed (H). G, I, J. No correlation was observed between swelling, TC and DC contact and total cognate CD4⁺ TCs or Effector/cognate CD8⁺ TCs that exited by day 10.

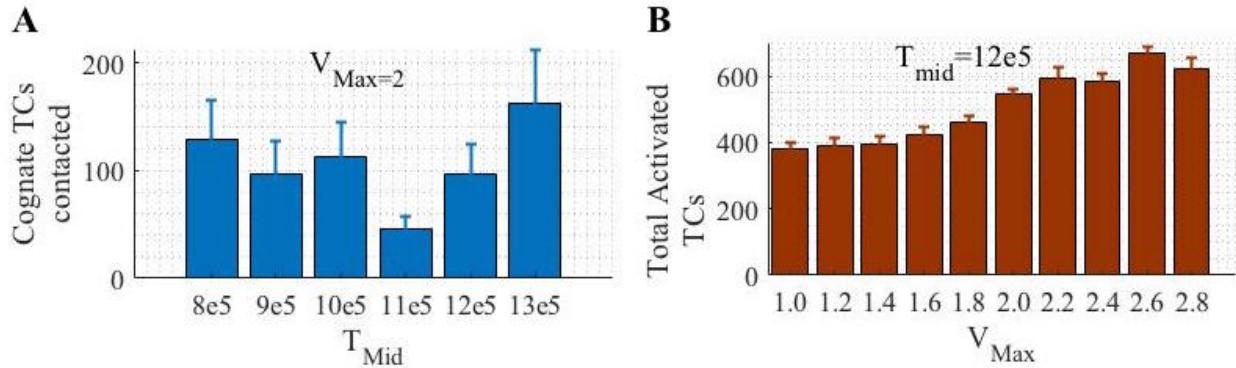


Figure S7. A. At $V_{max}=2.0$, no correlation between cognate TC and DC contacts and T_{mid} was observed. B. With a higher T_{mid} of 12e5, there remained a correlation between activated TCs and V_{Max} .

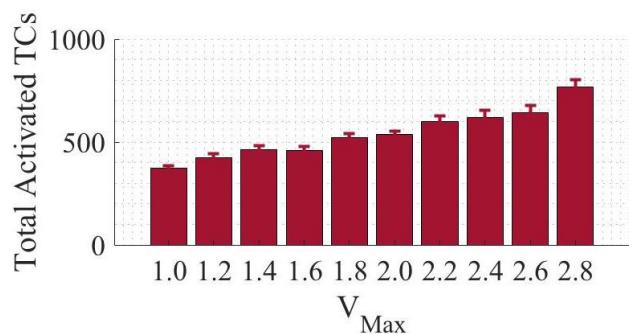


Figure S8. The activation of TCs at different maximal swelling when allowing TCs to return to HECs with a low probability increases with V_{max} ($p \cdot 10^{-5}$)

Table S1. Parameters that significantly influenced the number of effector TVs in the paracortex from day 3 to day 12 post-stimuli. There is a greater correlation between maximum CD8⁺ TC proliferation and effector TCs produced in the expanding paracortex than in the fixed volume paracortex. In an expanding paracortex there is also a negative correlation with TC recruitment, and maximal paracortical swelling in the second week of the simulation.

Parameter	Day 3	Day 4	Day 5	Day 6	Day 7	Day 8	Day 9	Day 10	Day 11	Day 12
Without Swelling										
Max_{P4+}							+		+	+
T_{short}					+					
Max_{NT}			+							
MHC_i					+					
F_{cog}	++	++	++	++	++	++	++	++		
ϕ_{DC}	++	++	++	++	++	++				
T_{DCin}	---	---	---	---						
SP_{early}	-									
With Swelling										
Max_{P8+}					+	++	++	++	++	++
Max_{P4+}					+					
T_{NC}								+	+	
T_{short}	+									
MHC_i								-		
P_{cog}	++	++	++	++						
ϕ_{DC}	++	++	+					-	-	
T_{DCin}	---	---	---	---	---	---	---			
SP_{early}	-									
R_F						-	--	--	--	--
V_{Max}	++		+	---	---	---	---	---	---	---
$0.05 > p > 0.001 = +/- \quad 0.001 > p > 10^{-6} = ++/- \quad 10e^{-6} > p > 0 = +++/- \quad -$										

Table S2. Parameters that significantly affected memory TCs number in the paracortex. Data is only shown from day 5 to day 12 post-stimuli, as they are not produced in the first few days. At day 5, V_{max} still showed some positive correlation with memory TCs present but by day 7 shows a negative correlation.

Parameter	Without Swelling				With Swelling			
	Day 5	Day 7	Day 9	Day 12	Day 5	Day 7	Day 9	Day 12
Max_{P8+}						++	++	+
$difflate$	++	++	++	++	++	++		+
γ			-		-			
F_{cog}	++	++	+		++			
ϕ_{DC}	++	++						
T_{DCin}	---	---			---	---		
R_F								
V_{Max}	N/A	N/A	N/A	N/A	+	-	---	---
T_{mid}		N/A	N/A	N/A	-			
$0.05 > p > 0.001 = +/- \quad 0.001 > p > 10^{-6} = ++/- \quad 10e^{-6} > p > 0 = +++/- \quad -$								

Table S3. Parameters that significantly affected the number of memory TCs exited in the paracortex from day 5 to day 12 post-stimuli.

Parameter	Without Swelling				With Swelling			
	Day 5	Day 7	Day 9	Day 12	Day 5	Day 7	Day 9	Day 12
TP_{4+}			+			+	+	
Max_{p8+}					-	--	--	--
dif_{early}	++				++			
dif_{late}	+++	+++	+++	+++	+++	+	++	+
T_{NC}							+	
K_S	+							
γ	--	--	-		--	--		
MHC_i						-	-	-
F_{cog}	+++	+++	++	++	+++			
ϕ_{DC}	--	--			--	--	--	--
T_{DCin}	--	--			--			
SP_{early}	--				--	-		
R_F					-	--	--	--
V_{Max}	N/A	N/A	N/A	(N/A)	--	--	--	--

0.05 > p > 0.001 = +/- 0.001 > p > 10^{-6} = ++/- - $10e^{-6} > p > 0$ = +++/- - -

Table S4. Parameters that were not varied in the global sensitivity analysis

Symbol	Parameter	Value	Reference
Model Geometry			
r^p	Initial paracortex radius	200 μm	
-	Entry radius	0.5 r^p	Mueller and Germain (2009); Kuka and Iannacone (2014)
-	Exit radius	0.07 r^p	
-	Sub-capsular Sinus Height	0.7 r^p	
-	Grid Size	6 μm	-
TC properties			
-	Initial occupation	55%	He (1985)
-	Ratio CD4 $^+$:CD8 $^+$	0.7:0.3	Mueller and Germain (2009); Kuka and Iannacone (2014)
-	TC naïve lifespan		Tough and Sprent (1995); Sprent and Tough (2001)
-	TC entry in afferent LVs to HEVs	0.1:0.9	Smith et al. (1970); Hall and Morris (1965)
Act_{4+}	Slope of CD4 $^+$ activation curve	-69.18	-
Act_{8+}	Slope of CD8 $^+$ activation curve	-80.71	-
Dif_{4+}	Slope of CD4 $^+$ differentiation curve	-17.26	-
Dif_{8+}	Slope of CD8 $^+$ differentiation curve	-13.58	-
β	Probability of movement	0.6	Park et al. (2010); Boscacci et al. (2010); Park et al. (2012); Girard et al. (2012); Miller et al. (2002)
P_e	Probability of egress	0.0126	-
γ	Max TCs per grid	2	-
T_{res}	TC residence time	24hrs	Catron et al. (2004); Tomura et al. (2008)
DC properties			
-	DC grid span	2 grids	Nitschké et al. (2012); Paharkova Vatchkova et al. (2004)
-	DC lifespan	2.5 days	Acton et al. (2014); Kamath et al. (2002); Bousso (2008)

Table S5. Parameters that were varied in the global sensitivity analysis.

Symbol	Parameter description	Default	Min	Max	Mean	SD	Distribution	Reference
TC response parameters								
Act μ_4	CD4* activation curve mean	120	70	230	-	-	Unif	Bajénoff et al. (2003); Yoon et al. (2007); Lawrence and Braciale (2004); Demotz et al. (1990); Lee et al (2002); Arens and Schoenberger (2010)
Act μ_8	CD8* activation curve mean	140	90	250	-	-	Unif	
Dif μ_4	CD4* differentiation curve mean	60	30	90	-	-	Unif	
Dif μ_8	CD8* differentiation curve mean	40	20	60	-	-	Unif	
TP ₄	Min. time between CD4* proliferations (hrs)	11	-	-	11	1.16	Norm	De Boer et al (2003); Nelson et al (2015); Foulds et al (2002); Tubo et al
TP ₈	Min. time between CD8* proliferations (hrs)	7	-	-	7	0.88	Norm	De Boer et al (2003); Foulds et al. (2002)
Max μ_8	Max proliferations CD8*	16	-	-	16	1.2	Norm	Butz and Bevan (1998); Murali-Krishna et al (1998); Busch et al. (1998); van Stipdonk et al. (2001)
Max μ_4	Max proliferations CD4*	10	-	-	10	1.2	Norm	Nelson et al. (2015); Tubo et al (2013); Foulds et al (2002)
Dif _{early}	Ratio of early differentiation into effector/memory TCs	0.01	0.001	0.02	0.01	-	Exp	William and Bevan (2004)
Dif _{late}	Ratio of late differentiation to effector /memory TCs	0.04	0.01	0.08	-	-	Unif	
TC interaction dynamics								
T _{NC}	Mean non-cognate T-DC interaction (min)	3.5	-	-	3.5	1	Norm	Bousso (2008); Miller et al (2004)
T _{Short}	Short cognate TC-DC interaction (min)	10.-15	-	-	10	3	Norm	Bousso (2008); Miller et al (2004); Mempel et al (2004)
T _{long}	Long cognate TC-DC interaction (min)	50.-70	-	-	50	12	Norm	von Andrian and Mackay (2000); Hugues et al. (2004); Mempel et al (2004); Stoll et al (2002)
T _{change}	Time TCs switch to long interactions (hrs)	8	-	-	8	1	Norm	
B _{Max}	Max TCs a DC can bind	3	1	5	-	-	Unif	
B _{Step}	Max TCs a DC can bind per step	15	4	20	-	-	Unif	Bousso and Robey (2003)
TC stimulation								
K _s	Stimulation gain coefficient	0.015	0.005	0.02	-	-	Unif	-
λ	TC stimulation decay factor	0.99	0.99545	0.9999	-	-	Norm	-
MHC _i	Initial MHC _i or MHCII	250	150	350	-	-	Norm	Cella et al. (1999); Kukutsch et al. (2000); Cella et al. (1997);
MHC _i _{1/2}	MHC _i half life (hrs)	19.7	-	-	19.7	6	Norm	Cella et al. (1999); Kukutsch et al.
MHCII _{1/2}	MHCII half life (hrs)	60	-	-	60	6	Norm	Cella et al. (1997); Baumgartner et al. (2010)
F _{cog}	Frequency of cognate TCs that enter	1.00E-04	5.00E-05	1.50E-04	-	-	Unif	Laouinin et al. (2000); Blattman et al. (2002); Nelson et al. (2015); Jenkins and Moon (2012)
ϕ_{DC}	Total DCs entering as % of initial TCs	0.04	0.02	0.06	-	-	Norm	Acton et al. (2014)
T _{DCin}	DC entry duration (days)	2.5	0.5	4.5	-	-	Unif	Acton et al. (2014)

Continued overleaf

Symbol	Parameter description	Default	Min	Max	Mean	SD	Distribution	Reference
	Sphingosine-1-phosphate receptor regulation							
SP _{entry}	S1P ₁ r expression post-entry	0.1	0.01	1	-	-	Unif	Pham et al. (2008), Matloubian et al. (2004), Lo et al. (2005)
SP _{act}	S1P ₁ r expression when activated	0.01	0.001	0.02	-	-	Unif	Pham et al. (2008), Matloubian et al. (2004), Lo et al. (2005), Garris et al. (2014)
SP _{early}	Effector S1P ₁ r expression (Proliferation>6)	0.8	0.3	1.3	-	-	Unif	Garris et al. (2014); Pham et al. (2008)
SP _{late}	Effector S1P ₁ r expression (Proliferation<=6)	0.4	0.01	1	-	-	Unif	
SP _{mem}	Memory S1P ₁ r expression	1	-	-	1	0.1	Norm	
SP _{IF}	S1P ₁ r on all TCs during inflammation	0.4	0.2	0.8	-	-	Unif	-
T _{Entry}	Time S1P ₁ r remains down-regulated post entry (min)	60	13	120	-	-	Unif	-
T _{Inflam}	Time to alter S1P ₁ r during inflammation (hrs)	4	1	7.5	-	-	Unif	-
	T cell recruitment							
RT1	Antigenic stim. threshold for TC recruitment increase (\sum MHCI)	2.00E+04	2.00E+04	1.00E+05	-	-	Unif	Hay and Hobbs (1977); Drayson and Smith (1981); Mackay et al. (1992); Webster et al. (2006)
RT2	Antigenic stim. threshold for TC recruitment increase (\sum MHCI)	4.00E+05	2.00E+05	2.00E+06	-	-	Unif	
R _F	Recruitment factor	3.00E-06	1.00E-06	4.00E-06	-	-	Unif	
	Paracortical expansion							
V _{Max}	Max fold-volume increase	1.00	2.00	2.50	-	-	Unif	-
<i>l</i>	Rate of volume change around m	7.00E-05	3.00E-05	1.00E-04	-	-	Unif	-
T _{mid}	No. of TCs present to reach 50% max volume	1.00E+05	9.00E+04	1.50E+05	-	-	Unif	-

2.1 UML diagrams

A series of UML diagrams that present the logic underlying the sub-methods called during the simulation (Supplementary Figure 9-26) are available at https://github.com/johnsara04/ABM_SF9-26

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