PCS Computational Science Project:

Unsupervised and supervised analysis of protein sequences

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We will start with the imports:

```
In [1]: %capture
%matplotlib inline
import numpy as np
import matplotlib.pyplot as plt

from sklearn.decomposition import PCA
from sklearn.model_selection import train_test_split
from sklearn.cluster import KMeans

plt.style.use('seaborn')
from matplotlib.pyplot import figure

figure(figsize=(8, 6), dpi=80)
```

Task 1

We will then read and one-hot encode the data.

```
In [2]: # read data as list of dictionary
        def read data(filename):
            with open(filename) as f:
                lines = f.readlines()
                data = []
                for line in lines:
                    if line[0] == ">":
                        properties = line[1:-1].split(" ")
                                   = properties[0]
                        functional = properties[1].split(" ")[1] == "true"
                        data.append({
                             "name"
                                         : name,
                            "functional" : functional,
                            "sequence" : ""
                        })
                    else:
                        data[-1]["sequence"] += line[:-1]
                return data
        # one-hot encode a single sequence
        def hot_encode_sequence(sequence):
            alphabet = ["A", "C", "D", "E", "F", "G", "H", "I", "K", "L", "M", "N", "P", "Q", "R", "S", "T",
            encoded sequence = []
            # encode each letter
            for letter in sequence:
                encoded_letter = [0]*(len(alphabet) - 1)
                letter idx = alphabet.index(letter)
                # if the letter is not the empty character
                if letter_idx < len(alphabet) - 1:</pre>
                    encoded_letter[letter_idx] = 1
                encoded_sequence += encoded_letter
            return encoded_sequence
        # test the one-hot encoder (see explaination below)
        print(hot_encode_sequence("C-W"))
```

```
# one-hot encode each sequence within the data
def hot_encode_data(data):
   for datum in data:
     datum["hot encoded sequence"] = hot encode sequence(datum["sequence"])
```

We tried to validate our *one-hote encoding* function on the simple sequence "C-W": This correctly gives us a 60-long vector (3*20) with only a 1 on the second place (corresponding to the first "C"), a one on the second-to last place (corresponding to the "W") with no one in between (corresponding to the "-").

```
In [3]: data_art = read data("MSA art.faa")
        hot encode data(data art)
        data nat = read data("MSA nat with annotation.faa")
        hot encode data(data nat)
In [4]: # print by hand to avoid a huge print out
        import json
        print_ = json.dumps(data_art[0], indent=4)
        print = print [:260] + "\t...\n ]\n}"
        print(print )
       {
           "name": "sequence 1",
           "functional": true,
           "sequence": "-----SLEELRKEIESIDREIVELIARRTYVAKTIAQIKRERGLPTTDESQEQRVMERAGSNAKQFD-VDANLVKAIFKLLIEL
       NKEEQRENR - - - "
           "hot encoded sequence": [
               0,
               Θ,
               0,
               Θ,
               Θ,
           ]
       }
```

We here review the encoding of the first sequence in the artificial data: we correctly read its sequence, name, that it is functional and then one-hot encoded its sequence.

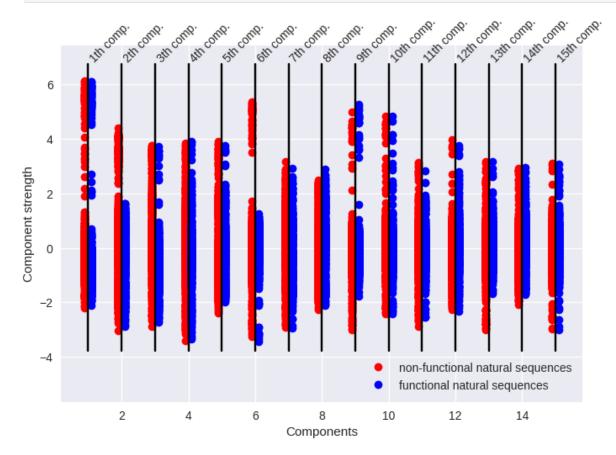
Task 2

for component in range(n components):

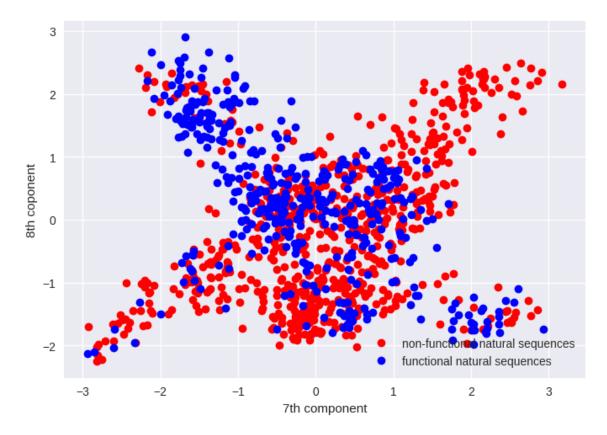
plt.plot(

```
We will no apply PCA to the one-hot encoded natural dataset to reduce its dimensionality.
In [5]: X nat = np.array([datum["hot encoded sequence"] for datum in data nat])
        X art = np.array([datum["hot encoded sequence"] for datum in data art])
        functionality nat = np.array([datum["functional"] for datum in data nat])
        functionality art = np.array([datum["functional"] for datum in data art])
In [6]: n components = 15
        model pca = PCA(n components=n components)
        eigen values nat = model pca.fit transform(X nat)
In [7]: functional eigen value nat
                                     = eigen values nat[
                                                                          functionality nat ]
        non functional eigen value nat = eigen values nat[np.logical not(functionality nat)]
        print(f"{ len(
                          functional eigen value nat) } functional sequences,",
              f"{ len(non functional eigen value nat) } non-functional sequences",
              f"for a total of { len(eigen values nat) } sequences.")
       423 functional sequences, 707 non-functional sequences for a total of 1130 sequences.
In [8]: Min, Max = 1.1*np.min(eigen_values_nat), 1.1*np.max(eigen_values_nat)
```

```
np.full(len(non_functional_eigen_value_nat), component + 0.9),
        non_functional_eigen_value_nat[:,component],
        "ro", label=("non-functional natural sequences" if component==0 else '_nolegend_'))
    plt.plot(
        np.full(len(functional_eigen_value_nat),
                                                     component + 1.1),
                                      [:,component],
        functional_eigen_value_nat
        "bo", label=("functional natural sequences" if component==0 else '_nolegend_'))
    plt.plot([component+1, component+1], [Min, Max], "k-")
    plt.text(component+1, Max, f"{component+1}th comp.",
            rotation=45,
            horizontalalignment="left", verticalalignment="bottom")
plt.ylim([Min*1.5, Max*1.1])
  = plt.xlabel("Components")
  = plt.ylabel("Component strength")
 = plt.legend(loc="lower right")
```



For some principal components (the 1st, 2nd and 6th for exemple) the functional and non-functional sequences occupy the same space, but some part of the space is reserved for one or the other (for exemple only non-fonctional sequence occupy the [2,4] space for the 2nd component, whereas both functional and non-functional sequences occupy the [-3,2] space for this component).



For other component (the 7th and 8th for exemple) the space seems fully shared. We would need to look at the higher dimensional data to see if in the 15-dimensional space they actually share the same space.

In the previous figure we see that in the 2D space of the 7th and 8th PCs, the functional and non-functional sequences are somewhat separated even though in they aren't separated in each 1D space.

We could expect that in the 15 dimentional PC space functional and non-functional sequence.

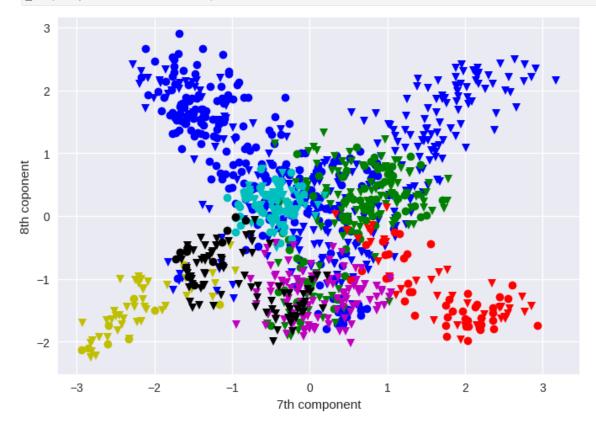
Task 3

We will now cluster the data using the K-mean method (apparently the subject ask us to cluster the one-hot encoded data and not the PC space data).

3.a - Cluster separation in PC space

We can visualize the clustered sequences in PC space as done in task 2:

```
In [11]: n1, n2 = 6, 7
         colors = ["b", "g", "r", "c", "m", "y", "k"]
         for cluster in range(n_cluster):
                                 = [cluster_idx == cluster for cluster idx in clustering]
             cluster mask
             X cluster
                                  = X nat[cluster mask]
             eigen_values_cluster = model_pca.transform(X_cluster)
             cluster functionality
                                                = functionality nat[cluster mask]
             eigen_values_cluster_functional = eigen_values_cluster[
                                                                                       cluster_functionality
             eigen_values_cluster_non_functional = eigen_values_cluster[np.logical_not(cluster_functionality)
             plt.plot(eigen values cluster functional
                                                        [:,n1], eigen_values_cluster_functional
             plt.plot(eigen_values_cluster_non_functional[:,n1], eigen_values_cluster_non_functional[:,n2], c
           = plt.xlabel(f"{n1+1}th component")
```



In the above figure we show the separation between the clusters (different colors) and between functional (circle marker) and non-functional (triangle marker) sequences.

We observe a partial separation in PC space of the different clusters, and somewhat of a separation between clusters of functional and non-functional sequences (which we will quantify later on).

3.b - Cluster separation between functional and non-functional sequences

After trying multiple number of clusters, and playing with the "cutoff" (we choose a cutoff equal to the global proportion of functional sequences) at which we choose if a cluster is a "primarly functional" cluster (majority of functional sequences) or a "primarly non-functional" cluster.

We can first define a function that categorizes between these two types clustes, and then check the accuracy of this function as a way of charectarizing if a sequence if functional or not:

```
In [12]: # functional proportion cutoff equal to the global proportion of functional sequence
         cutoff = len(functional_eigen_value_nat)/len(eigen_values_nat)
         # create a list of primarly functional clusters:
         primarly functional clusters = []
         for cluster in range(n_cluster):
             cluster mask = [cluster idx == cluster for cluster idx in clustering]
             cluster functionality = functionality nat[cluster mask]
             if np.mean(cluster functionality) > cutoff:
                 primarly functional clusters.append(cluster)
         # check if a cluster is in the list of primarly functional clusters
         def is functional cluster(cluster):
             return np.isin(cluster, primarly functional clusters)
         # check if a sequence is clustered withing a primarly functional cluster
         def is functional sequence(X):
             clustering = model cluster.predict(X)
             return is_functional_cluster(clustering)
```

```
In [33]: functional_functional, nonFunctional_functional = 0, 0
functional_nonFunctional, nonFunctional_nonFuctional = 0, 0
```

```
# count the true/false positive/negative rate
for cluster in range(n_cluster):
    cluster_mask = [cluster_idx == cluster for cluster_idx in clustering]
    cluster_functionality = functionality_nat[cluster_mask]
    if is_functional_cluster(cluster):
        functional_functional
                                += np.sum(cluster_functionality)
        nonFunctional_functional += len(cluster_functionality) - np.sum(cluster_functionality)
    else:
        functional nonFunctional += np.sum(cluster functionality)
        nonFunctional_nonFunctional += len(cluster_functionality) - np.sum(cluster_functionality)
print(f"{round(functional_functional)} functional sequence",
f"and {round(nonFunctional_functional)} non-functional sequences in cluster with more than {round(cu
f"corresponding to {round(functional functional/(functional functional + nonFunctional functional)*1
f"and {round(functional functional/len(functional eigen value nat)*100)}% of all functional sequence
print(f"{round(functional_nonFunctional)} functional sequence",
f"and {round(nonFunctional_nonFuctional)} non-functional sequences in cluster with less than {round(
f"corresponding to {round(nonFunctional nonFuctional/(functional nonFunctional + nonFunctional nonFu
f"and {round(nonFunctional nonFuctional/len(non functional eigen value nat)*100)}% of all non-function
```

400 functional sequence and 477 non-functional sequences in cluster with more than 37% of functional sequences, corresponding to 46% functional sequences in this group, and 95% of all functional sequences

23 functional sequence and 230 non-functional sequences in cluster with less than 37% of functional sequences, corresponding to 91% non-functional sequences in this group, and 33% of all non-functional sequences

After trying multiple number of clusters, and playing with the "cutoff" (we choose a cutoff equal to the global proportion of functional sequences) at which we choose if a cluster is a "primarly functional" cluster or a "primarly non-functional" cluster, we separated clusters in those two groups:

- a slight minority (46%) of sequences in "primarly functional clusters" are functional: this corresponds to *true* positives.
- a large majority (95%) of sequences in "primarly non-functional clusters" are non-functional: this corresponds to *true negatives*.
- a large majority (91%) of functional sequences end up in "primarly functional clusters".
- a minority (33%) of non-functional sequences end up in "primarly non-functional clusters".

We can conclude that functional and non-functional sequences are somewhat separated in the clusters, and thus are separated in PCs space.

3.c - Cluster separation between natural and artificial data

We will now do the same procedure, trying to separate natural and artificial data:

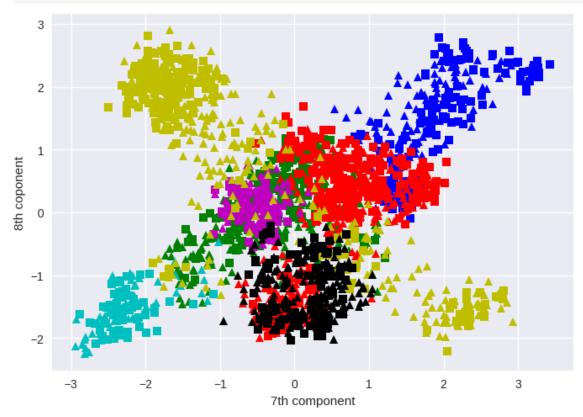
eigen_values_cluster_nat = model_pca.transform(X_nat_cluster)
eigen values cluster art = model pca.transform(X art cluster)

X art cluster = X art[cluster mask art]

```
proportion = len(X_nat_cluster)/(len(X_nat_cluster) + len(X_art_cluster))
if proportion > nat_art_cutoff:
    nat_nat += len(X_nat_cluster)
    art_nat += len(X_art_cluster)
else:
    art_art += len(X_art_cluster)
    nat_art += len(X_nat_cluster)

plt.plot(eigen_values_cluster_nat[:,n1], eigen_values_cluster_nat[:,n2], colors[cluster]+"^")
plt.plot(eigen_values_cluster_art[:,n1], eigen_values_cluster_art[:,n2], colors[cluster]+"s")

= plt.xlabel(f"{n1+1}th component")
= plt.ylabel(f"{n2+1}th coponent")
```



```
In [32]: print(f"{round(nat_nat)} natural sequence",
    f"and {round(art_nat)} artificial sequences in cluster with more than {round(nat_art_cutoff*100)}% of
    f"corresponding to {round(nat_nat/(nat_nat + art_nat)*100)}% natural sequences in this group,",
    f"and {round(nat_nat/len(X_nat)*100)}% of all natural sequences")

print(f"{round(nat_art)} natural sequence",
    f"and {round(art_art)} non-functional sequences in cluster with less than {round(cutoff*100)}% of natural f"corresponding to {round(art_art/(art_art + nat_art)*100)}% artificial sequences in this group,",
    f"and {round(art_art/len(X_art)*100)}% of all artificial sequences")
```

533 natural sequence and 373 artificial sequences in cluster with more than 53% of natural sequences, corresponding to 59% natural sequences in this group, and 47% of all natural sequences 597 natural sequence and 630 non-functional sequences in cluster with less than 37% of natural sequences, corresponding to 51% artificial sequences in this group, and 63% of all artificial sequences

Using the same way of separating between "primarly natural clusters" and "primarly artificial cluster" as previously with functional and non-functional sequences, we end up with poor to no separation between natural (triangle marker) and artificial (square marker) sequences; as the proportion of natural/artifical sequences does not significantly change after the clustering.

Task 4

We will now train various model that can classify between functional and non-functional proteins.

We first separate the natural dataset between a "training" an "testing" dataset:

```
In [17]: Y_nat, Y_art = functionality_nat, functionality_art
    X_train, X_test, Y_train, Y_test = train_test_split(X_nat, Y_nat, test_size=0.3, random_state=1)
```

4.a - Decision Tree

The first model we will train is a decision tree model similar to one that we used in a previous TD:

```
In [18]: %capture
         from sklearn.tree import DecisionTreeClassifier
         from sklearn.ensemble import AdaBoostClassifier
         # example stollen from the classifier TD
         weak classifier = DecisionTreeClassifier(max depth=1, random state=5)
                                                                                                            # h
         strong_classifier = AdaBoostClassifier(n_estimators=50, base_estimator=weak_classifier)
                                                                                                      # here wo
         model decisionTree = strong classifier.fit(X train, Y train)
In [19]: # function to meusure and print the true/false positive/negative rates
         def print_accuracy_matrix(test_function, X, Y):
             TP = test_function(X[Y], Y[Y])
             FN = 1 - TP #predicted negative while actually positive => false negative
             TN = test_function(X[Y==0], Y[Y==0])
             FP = 1 - TN #predicted positive while actually negative => false positive
             print(f"TP: {round(TP*100)}%, FP: {round(FP*100)}%")
             print(f"FN: {round(FN*100)}%, TN: {round(TN*100)}%")
         print("Natural data, test:")
         print accuracy matrix(model decisionTree.score, X test, Y test)
         print("\nNatural data, train:")
         print_accuracy_matrix(model_decisionTree.score, X_train, Y_train)
         print("\nArtificial data:")
         print accuracy matrix(model decisionTree.score, X art, Y art)
        Natural data, test:
        TP: 64%, FP: 17%
        FN: 36%, TN: 83%
        Natural data, train:
        TP: 81%, FP: 9%
        FN: 19%, TN: 91%
        Artificial data:
        TP: 74%, FP: 18%
        FN: 26%, TN: 82%
```

We can see that the true positive (TP) and true negative (TP) rates are higher for the data we trained on than the test data.

Interestingly we can observe that the model's TP is better on the artificial data than the natural data. We could assume that the artificial data has been generated by a generator trained to maximize such a model (as we will do later), thus artificial data could more distinguishable for our model as it was "generated to be differenciated".

4.b - Logistic regression

We will now move to a simple logistic regression model:

```
In [20]: %capture
    from sklearn.linear_model import LogisticRegression
    model_logic = LogisticRegression()
    model_logic.fit(X_train, Y_train)

In [21]: print("Natural data, test:")
    print_accuracy_matrix(model_logic.score, X_test, Y_test)

    print("\nNatural data, train:")
    print_accuracy_matrix(model_logic.score, X_train, Y_train)

    print("\nArtificial data:")
    print_accuracy_matrix(model_logic.score, X_art, Y_art)
```

```
Natural data, test:
TP: 65%, FP: 13%
FN: 35%, TN: 87%
Natural data, train:
TP: 100%, FP: 0%
FN: 0%, TN: 100%
Artificial data:
TP: 73%, FP: 18%
FN: 27%, TN: 82%
```

We obtain verry similar results to the decision tree model, and thus make largly the same observations.

4.c - Neural network

We will finally use a simple neural network based classifier provided by sklearn.

For that we will first need to one-hot encode the class (functional or non-functional) thus making it a vector of size 2. We will also split the natural data between a training and testing dataset as previously done:

```
Y nat 2d = np.array([[0,1] if f else [1,0] for f in functionality nat])
         Y_art_2d = np.array([[0,1] if f else [1,0] for f in functionality_art])
         X train 2d, X test 2d, Y train 2d, Y test 2d = train test split(X nat, Y nat 2d, test size=0.3, rando
In [23]: %capture
         from sklearn.neural network import MLPClassifier
         model nn = MLPClassifier(solver='lbfgs', hidden layer sizes=(50, 10), random state=1, alpha=1e-5)
         model nn.fit(X train 2d, Y train 2d)
In [24]: # we need a new function because the data is formated differently
         def print_accuracy_matrix_NN(test_function, X, Y):
             TP = test function(X[Y[:, 1] == 1], Y[Y[:, 1] == 1])
             FN = 1 - TP #predicted negative while actually positive => false negative
             TN = test_function(X[Y[:, 0] == 1], Y[Y[:, 0] == 1])
             FP = 1 - TN #predicted positive while actually negative => false positive
             print(f"TP: {round(TP*100)}%, FP: {round(FP*100)}%")
             print(f"FN: {round(FN*100)}%, TN: {round(TN*100)}%")
         # we need to define the accuracy function for the evaluation by hand
         def nn cost function(X, Y):
             prediction = model nn.predict(X)
             return np.mean(prediction[:, 0] == Y[:, 0])
         print("Natural data, test:")
         print accuracy matrix NN(nn cost function, X test 2d, Y test 2d)
         print("\nNatural data, train:")
         print_accuracy_matrix_NN(nn_cost_function, X_train_2d, Y_train_2d)
         print("\nArtificial data:")
         print_accuracy_matrix_NN(nn_cost_function, X_art, Y_art_2d)
        Natural data, test:
        TP: 62%, FP: 14%
        FN: 38%, TN: 86%
        Natural data, train:
        TP: 100%, FP: 0%
        FN: 0%, TN: 100%
        Artificial data:
        TP: 72%, FP: 15%
        FN: 28%, TN: 85%
```

The neural network-based model is significantly slower to train without having a huge advantage in discriminating between functional and non-functional proteins.

We want to train a model that generates sequences that are recognized by the one of the model we trained (we will use the logistic regression trained in section 4.b goind forward) as functional.

Idealy we could use this model as a fitness function for a neural network, but this isn't trivialy supported by sklearn, and it would be slow.

We thus choose to do the following work-around:

- We choose to train a multi-dimentional *gaussian mixture* model in PC space (to reduce the number of dimmension).
- We first train it to imitate the functional natural data distribution in PC space.
- We then iterate over the following algorithm so that it generates vectors in PC space that correspond to sequences that are considered to be functional by our *logistic regression* model:
 - 1. We generate a large number of vector in PC space (x time the number of natural data sequence).
 - 2. We transform them to sequences, and select the fraction (*y*) that has the highest likelyhood of being functional according to our *logistic regression* model.
 - 3. We go back to step 1, but this time training on a concatenanted dataset containing the functional natural data in PC space plus the highest scoring generated vector in PC space.
- Over the epochs (or iterations) we increase the fraction of the training dataset (*y*) that is composed of generated vector.

Additionaly, whenever we generate a vector in PC space and then transform it to a sequence, we "normalize" the sequence such that:

- Only up to one element can be 1 for each 20 number block corresponding to a letter.
- The element with the maximum value in the 20 number block gets assigned to 1 if it has a weight above 0.5, otherwise it gets assigned to 0.
- Every other elements in the block is assigned to 0.

The goal of this algorithm is that the initial trained distribution will try to match the natural data as much as possible, and then by this generation-selection loop we will "reinforce" the model to follow a distribution that maximizes the likelyhood of generated vector being functional according to our previously trained model.

```
In [25]: # function to round and pin between 0 and 1
         \# and to make sure that the sequence is valid (one or zero 1 per block of 20 corresponding to a lett
         def pin 01(X):
              # reshape as a list of list of one-hot encoded letter
             X \text{ reshaped} = \text{np.reshape}(X, (len(X), -1, 20))
             for X_ in X_reshaped:
                  for letter in X :
                      # find max in letter
                      idx = np.argmax(letter)
                      value = letter[idx]
                      # put all values to zero
                      letter = np.zeros_like(letter)
                      # if max is above 0.5 then put it to 1
                      if letter[idx] > 0.5:
                          letter[idx] = 1
              return np.reshape(X reshaped, (len(X), -1))
```

```
= pin 01(model pca.inverse transform(eigen gen))
          X_gen_fitness = model_logic.predict_proba(X_gen)[:, 1]
                                = int(len(functional eigen value nat)*train multiplier)
           fitest idx = np.argpartition(X gen fitness, -n select)[-n select:]
           eigen_train = np.concatenate((functional_eigen_value_nat, eigen_gen[fitest_idx]), axis=0)
           avg fitness = np.mean(X gen fitness)
           print(f"{i+1}th/{len(gen multipliers)} epoch: {round(avg fitness*100)}% average probability of formula fit is a second for the second fit is a second fit is 
1th/13 epoch: 55% average probability of functionality according to logistic regression
2th/13 epoch: 65% average probability of functionality according to logistic regression
3th/13 epoch: 69% average probability of functionality according to logistic regression
4th/13 epoch: 74% average probability of functionality according to logistic regression
5th/13 epoch: 78% average probability of functionality according to logistic regression
6th/13 epoch: 84% average probability of functionality according to logistic regression
7th/13 epoch: 84% average probability of functionality according to logistic regression
8th/13 epoch: 89% average probability of functionality according to logistic regression
9th/13 epoch: 89% average probability of functionality according to logistic regression
10th/13 epoch: 91% average probability of functionality according to logistic regression
11th/13 epoch: 91% average probability of functionality according to logistic regression
12th/13 epoch: 93% average probability of functionality according to logistic regression
13th/13 epoch: 93% average probability of functionality according to logistic regression
```

We obtained an average 93% predicted likelyhood that our generated sequences are functional, which is similar/higher than the true positive rate of this model. We thus hope that our generating distribution is trained to generate vector in PC space that correspond to functional sequences.

We will now generate a 1000 sequences and acess them using the methods developed in tasks 2-4:

Before moving on, we will check that we didn't overfit by checking that the generated data isn't just a copy of the natural data:

```
In [28]: is_natural = np.isin(X_gen, X_nat)
    existing_proportion = np.mean([np.any([np.array_equal(Y, X) for X in X_nat]) for Y in X_gen])
    print(f"{ round(existing_proportion*100) }% of the generated sequences existed in the natural data")
    0% of the generated sequences existed in the natural data
```

We see that none (or at least less than 1%) of our generated sequences are pre-existing in the natural data.

5.a - clustering

We will first acess the functionality of the generated sequences by checking if they are clustered within the "primarly functional" clusteres defined in task 3.

```
In [29]: X_gen_fitness = is_functional_sequence(X_gen)
avg_fitness = np.mean(X_gen_fitness)
print(f"{ round(avg_fitness*100) }% of the generated sequences are clustered within \"primarly functions"
```

99% of the generated sequences are clustered within "primarly functional" clusters

According to the clustering model 99% of the generated sequences are clustered within "primarly functional" clusters, thus suggesting that they could likely be functional.

5.b - Decision tree

We will now acess the functionality of these generated sequences using the decision tree trained in task 4.a:

```
In [30]: X_gen_fitness = model_decisionTree.predict(X_gen)
    avg_fitness = np.mean(X_gen_fitness)

print(f"{ round(avg_fitness*100) }% of the generated sequences are predicted to be functional by the
```

93% of the generated sequences are predicted to be functional by the decision tree model

According to the decision tree model 93% of the generated sequences are functional.

5.c - Neural netwrok

We will now acess the "functionality" of these generated sequences using the neural network trained in task 4.c.

Note that we don't acess the generated sequences using the logistic regression model as it was used in the training. We can however note that in training the logisitic regression model predicted up to a 93% rate of functionality in the generated sequences.

```
In [31]: X_gen_fitness = model_nn.predict(X_gen)[:, 1]
    avg_fitness = np.mean(X_gen_fitness)

print(f"{ round(avg_fitness*100) }% of the generated sequences are predicted to be functional accord.
```

95% of the generated sequences are predicted to be functional according to the neural network model According to our neural network based model 95% of the generated sequences are functional.

Conclusion

In task 2-4 we successfully built multiple independently trained tools to discriminate between functional and non-functional sequences, which where all capable of generalizing not only to the testing dataset (sub-portion of the natural sequences that was left out of the training dataset), but also to artificial sequences.

In task 5 we used one of these model (the *logistic regression* model) to iteratively train a probability distribution that successfully maximized this model's prediction that generated sequences were functional.

We then acessed the functionality of these sequences using the methods developed in tasks 2-4 and found that every method independily gave us a high likelyhood that the sequences were functional.

However, the training dataset for each of these method are the same and is generally quite a small sample (70% of a total of 1130 natural sequences) which thus prevents us from confidently saying that our generated sequences would actually be functional without more data, or even better actual experiments.