STANDARD OPERATING PROCEDURE

SOP # 1 Page: 1 of 3	Version 1.1 Date: 07/08/2017 Revise by: 07/08/2017	Prepared By: Dr Joyanta Modak Approved By: Dr Vijay Dhanasekaran
	1tevise by: 07/00/2017	

Laboratory of Viral Evolution, Department of Microbiology, Monash University

SOP 1: Virus RNA isolation using QIAamp Viral RNA mini kit

1 Introduction

This SOP describes the use of QIAamp Viral RNA mini kit (QIAGEN) to isolate RNA from sample containing viruses.

2. Scope

This procedure serves as a guide to laboratory personnel engaged in viral RNA purification using the QIAamp® Viral RNA Mini Kit by QIAGEN.

3. Safety

All lab workers must:

- a. Wear appropriate personal protective equipment (PPE) (Nitrile gloves, lab coat, closed-toed shoes, and safety glasses when specified by risk assessment) when carrying out the procedure.
- b. If procedure is performed on reassortant viruses, please wear an N95 mask for additional personal protection.
- c. Carry out the procedure in the appropriate setting, i.e. tissue culture room.
- d. Read and understand the recommendations in this SOP to ensure uniformity in practice.
- e. New staff or students should seek supervision from a senior lab co-worker during first attempt of the procedure.

4. Definition

Viral RNA purification: To extract purified viral RNA from samples plasma, serum, cell-free body fluids or cell-culture supernatants.

5. Procedures

5.1 Materials Needed

- a. Virus-infected cell-culture supernatant
- b. QIAamp® Viral RNA Kit (250) (QIAGEN Cat# 52906), which includes:
 - i) QIAamp® Mini spin columns
 - ii) Collection tubes (2 mL)
 - iii) Buffer AVL

STANDARD OPERATING PROCEDURE

SOP # 1 Page: 2 of 3	Version 1.1 Date: 07/08/2017 Revise by: 07/08/2017	Prepared By: Dr Joyanta Modak Approved By: Dr Vijay Dhanasekaran

Laboratory of Viral Evolution, Department of Microbiology, Monash University

- iv) Buffer AW1
- v) Buffer AW2
- vi) Buffer AVE
- vii) Carrier RNA (poly A)
- c. Ethanol (96 100%)
- d. 1.5 mL micro-centrifuge tubes (Axygen Cat# MCT-150-C)
- e. ART 200 and ART 1000XL aerosol barrier pipette tips (Molecular Bio-Products Cat# 2069-05-HR and 2179-05-HR)
- f. P100 and P1000 single-channel pipettes (Eppendorf Cat # 3111000.149 and 31111000.165)
- g. Micro-centrifuge
- h. Freezer -20°C
- i. Freezer -80°C

5.2 Reagent Preparation

To prepare carrier-RNA-containing Buffer AVL, carry out the following in accordance:

- a. Add 310 μ L of Buffer AVE to the tube containing 310 μ g lyophilized carrier RNA to obtain a solution of 1 μ g/ μ L.
- b. Dissolve the carrier RNA thoroughly, divide it into conveniently sized aliquots, and store it at -20°C. Do not freeze-thaw the aliquots of carrier RNA more than 3 times.
- c. For each sample, add 1 volume of carrier RNA-AVE to 100 volumes of Buffer AVL. (i.e. 5.6 μ L of carrier RNA-AVE to 0.56 mL of Buffer AVL for 1 sample)

To prepare Buffer AW1 and Buffer AW2, add the appropriate amount of ethanol (96-100%) as indicated on the bottle.

5.3 Purification of Viral RNA using Spin Columns

- a. Pipette 560 μ L of prepared Buffer AVL containing carrier RNA into a 1.5 mL microcentrifuge tube. (If the sample volume is larger than 140 μ L, increase the amount of Buffer AVL-carrier RNA proportionally.)
- b. Add 140 μL cell-culture supernatant to the Buffer AVL-carrier RNA in the micocentrifuge tube. Mix by pulse-vortexing for 15 s.

STANDARD OPERATING PROCEDURE

SOP # 1 Page: 3 of 3	Version 1.1 Date: 07/08/2017 Revise by: 07/08/2017	Prepared By: Dr Joyanta Modak Approved By: Dr Vijay Dhanasekaran
Page: 3 of 3	Revise by: 07/08/2017	

Laboratory of Viral Evolution, Department of Microbiology, Monash University

- c. Incubate at room temperature $(15 25^{\circ}C)$ for 10 min.
- d. Briefly centrifuge the tube to remove drops from the inside of the lid.
- e. Add 560 μ L of ethanol (96 100%) to the sample, and mix by pulse-vortexing for 15 s. After mixing, briefly centrifuge the tube to remove drops from inside the lid. (If the sample volume is greater than 140 μ L, increase the amount of ethanol proportionally.)
- f. Carefully apply 630 μ L of the solution from the previous step to the QIAamp Mini column (in a 2 mL collection tube) without wetting the rim. Close the cap, and centrifuge at 6000 x g (8000 rpm) for 1 min. Place the QIAamp Mini column into a clean 2 mL collection tube, and discard the tube containing the filtrate.
- g. Carefully open the QIAamp Mini column, and repeat the previous step.
- h. Carefully open the QIAamp Mini column, and add 500 μ L of Buffer AW1. Close the cap, and centrifuge at 6000 x g (8000 rpm) for 1 min. Place the QIAamp Mini column in a clean 2 mL collection tube (provided), and discard the tube containing the filtrate.
- i. Carefully open the QIAamp Mini colum, and add 500 μ L of Buffer AW2. Close the cap and centrifuge at full speed (20,000 x g; 14,000 rpm) for 3 min
- j. (Opional) Place the QIAamp Mini column in a new 2 mL collection tube (not provided), and discard the old collection tube with the filtrate. Centrifuge at full speed for 1 min.
- k. Place the QIAamp Mini column in a clean 1.5 mL microcentrifuge tube (not provided). Discard the old collecion tube containing the filtrate. Carefully open the QIAamp Mini column and add 60 μ L of Buffer AVE equilibrated to room temperature. Close the cap, and incubate at room temperature for 1 min. Centrifuge at 6000 x g (8000 rpm) for 1 min.
- 1. Store the purified viral RNA at -20°C or -70°C.

1.0 RECORDS

Nil

2.0 APPENDIXES

Nil

3.0 REFERENCES

QIAGEN, 2010. *QIAamp® Viral RNA Mini Handbook*. **Third Edition**.