

CS194-302

Joon Kim



Background

RNA is a good proxy measure for protein!

Classical (Bulk) RNA Seq: Systematic errors, (Pseudo) Alignments

↳ What can we say about the gene expression? Methods such as Library normalization, Dimensionality Reduction, Differential Expression.

Why Single Cell? Multiple cell measurements result in average.

↳ Single cell profiles recover heterogeneity! we don't know about the diversity b/w cells

Immunology readily adopted SC-omics due to immune cells being singular

Intrinsic Noise from biological data is due to stochastic processes

Week 2: Tumors

Immune System: protect the body from pathogens. — complement

innate → short term, fixed. adaptive → long term, needs learning

There are different types of immune cells. We are mainly interested in how these function together (systems immunology).

ex) CD4+ T cells have helper subsets, can be beneficial/deleterious.

- ↳ conduct *in vitro* experiments that control the environments that affect the "evolution" of the cells, and scatterplot markers.
(in lab)
- * *In vivo*, relationships between subsets are not very clean.
- ex) *In vitro* shows an XOR relation, but in nature, they can both exist.

Self vs Not-Self → checker for whether we should attack the cell

- Two bins of tumors: Hot(Lymphocyte infiltrating) / Cold(no Lymph. inf.)
- ↳ CD8 T cells can infiltrate(attack) hot tumor cells
 - ↳ CD4 T cells are not very helpful, sometimes even backfiring!
 - Tregs suppress other cells that can attack tumors → not good!

T cell exhaustion → bleeding edge, how to do it is unanswered

↑
Cancer cells try to trick attackers by sending "don't eat me" signals.

- ↳ CTLA4, PD1, CD47

Week 3: Batch Effects

- CAR-T Therapy: train patient's T cells to recognize tumors
- ↳ Blood tumors work well, solid tumors are in progress

Batch Effects: systematic (experimental) bias → how to overcome?

Curse of Dimensionality: All points are "equally" distant to each other

Principle Component Analysis: Linear transformation for dim. reduction

$X := N \text{ by } d$ matrix, covariance $\Sigma := X^T X \rightarrow$ not diagonal

↪ we need lin. transform P s.t. $(PX)^T (PX)$ is $\text{diag}(\lambda_1, \lambda_2, \dots)$.

Some confounding variables will have biological significance!

Harmony Algorithm: K-means, then shift data closer to centroids

Local Inverse Simpson's Index (LISI) → quantifies diversity of neighbors

↪ integration LISI: effective # of datasets (should \rightarrow # datasets)

↪ cell type LISI: effective # of cell types (should $\rightarrow \sim 1$)

Reciprocal PCA (rPCA): doesn't assume clusters from datasets!

↪ find "anchor" cells that are similar across datasets, then their difference vector is the batch effect.

1) Normalize dataset, select set of most common genes

2) Reciprocal Projection: PCA, then reciprocally superimpose datasets

- 3) Find anchors via Mutual Nearest Neighbors
- 4) Heuristically evaluate the quality of anchors (filtering, weighting)
- 5) Integration: pick a reference dataset, move others accordingly

Evaluation: how do we evaluate these methods?

Silhouette Scores: $a(i) :=$ mean distance in same batch,

$b(i) :=$ minimum mean distance to another batch,

$$s(i) := \frac{b(i) - a(i)}{\max\{a(i), b(i)\}}$$
 if $|B_i| > 1$, \emptyset if $|B_i| = 1$.

↪ we would want this score to decrease (close to \emptyset)

Week 4: Cell Trajectories

Metacells: group cells into a few representatives for strong signals

↪ helps with reducing the sparsity of the matrix.

MC2: Divide-and-Conquer, 2-sided Stability Score

↪ highly parallelizable, $O(N \log N)$ vs $O(N^2) \leftarrow (MC1)$

- how do we break up into subcomponents?

- how do we account for the fact that biology is not uniform?

Preliminary Phase: split cells into random piles, solve each one.

Metagroup Phase: partition metacells into metagroups, recurse.

Final Phase: treat outliers/rare cells separately

* also detects rare genes and separately processes them out of DaC.

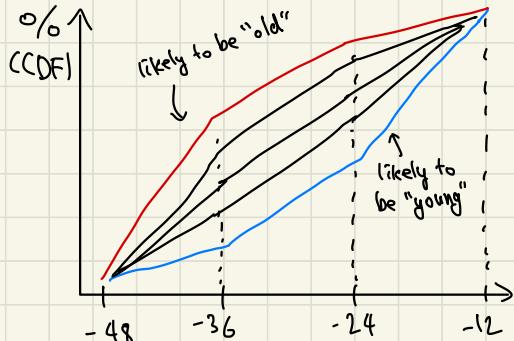
2-sided Stability Score: use iterative updates for accuracy.

Zman-seq: how do we analyze temporal processes in scRNA? → over course of day

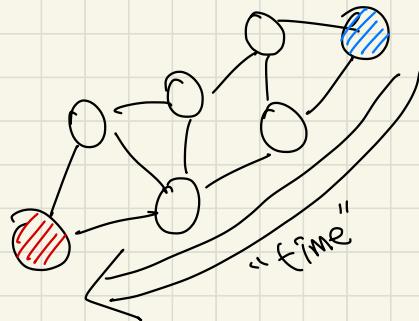
↳ add fluorescent pulse labels for temporal information.

↳ inject different colors at different timestamps to know when a particular cell entered the tumor!

cTET: approximate which metacell entered at what time by specifying a CDF of time entered (12, 24, 36, ... hrs) and calculating the AUC of each function → sense of time!

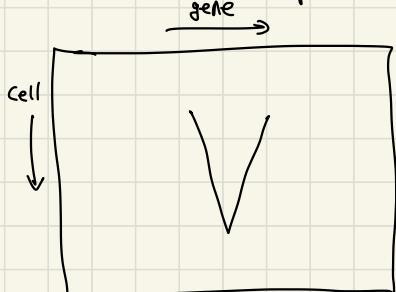


⇒

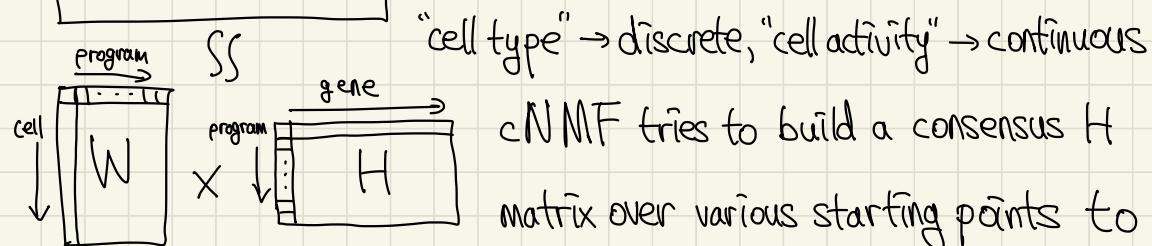


Week 5: (Consensus) Nonnegative Matrix Factorization

Bulk RNA-seq data → very large and sparse matrix V



(dropout is a factor)
traditional NMF is obfuscated by noise!
also, assignments are single & deterministic
linear analysis is efficient but not rigorous



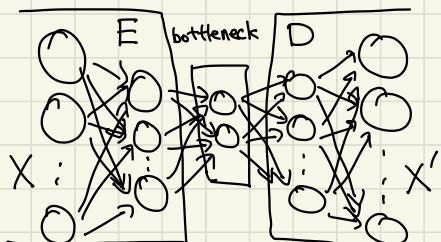
raise empirical accuracy. Programs need to be grouped intelligently!

Week 6: VAE

So far: Dimensionality Reduction → Clustering → Diff. Exp.

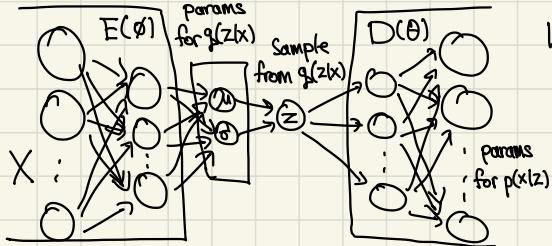
With VAE: Neural Networks → Clustering → Generative Exp.

Autoencoders: NN for dimensionality reduction; the NN tries



to minimize $\|x - x'\|$ while having a
"bottleneck" that learns a latent
representation of data

VAE: Probabilistic version of AE, predict a distribution on latent space



↳ this is a generative model, where we try to learn the joint distribution of x (data) and y (label).

$$* \text{Math: } \log p_\theta(x^{(i)}) = D_{\text{KL}}(q_{\theta^*}(z|x^{(i)}) || p_\theta(z|x^{(i)})) + \mathcal{L}(\theta, \phi; x^{(i)}),$$

$$\phi \rightarrow \begin{array}{c} z \\ \downarrow \\ N \end{array} \quad \mathcal{L}(\theta, \phi; x^{(i)}) = -D_{\text{KL}}(q_{\theta^*}(z|x^{(i)}) || p_\theta(z)) + E_{q_{\theta^*}(z|x^{(i)})} [p_\theta(x^{(i)}|z)]$$

how far are we away from the answer? ELBO

variational post. prior generative model
(encoding) (prior) (decoding)

scVI: z as latent cell representation, x as gene data

"Systems" Biology: Consider interactions between components!

Week 7: Multi-Modal Omics

(CD4 vs CD8 fate)

Today's Story: CITE-seq + total VI \rightarrow new bio discovery!

Flow Cytometry is powerful but is a manual procedure.

CITE-seq: RNA-seq + surface protein abundance quantification in sc.

↳ the readout is still sequencing, so a lot of caveats & noise

total VI: How can we explicitly incorporate protein data in modeling?

↳ "Joint" probability with a VAE! Proteins are modeled as mixture of Gaussians to better fit for foreground/background bimodality

* Gamma → Poisson mixture is just Negative Binomial, like scVI

Week 8: Cohort Studies

// Our presentation, refer to slides

Week 9: Spatial Omics I

(in-situ)

What if we were able to keep spatial information intact in sequencing?

Two flavors: imaging / sequencing

Imaging: single-cell resolution, but limited in # of genes

↳ Codex, MERFISH, ExSeq

Sequencing: transcriptome-wide measurement, but only near-single-cell

↳ Slide-Seq, (Spot Deconvolution)

Spot Deconvolution: "Separate" information. What is the proportion of cells under this specific bead?

TACCO: spatial deconvolution & categorical annotation methods

↳ "semi-unbalanced entropic optimal transport"

Optimal Transport: $\underset{\gamma \in \mathcal{P}(\text{pix}_a, \text{pix}_b)}{\operatorname{arg\min}} \sum_{a,b} \gamma_{a,b} M_{ab}$, s.t. $\gamma_{a,0} = p_a$, $\gamma_{0,b} = q_b$ (marginals)

↳ "convex" \Rightarrow LP!! but # of constraints is pretty large...

Entropic OT: regularize entropy of γ , add $\epsilon \cdot \sum_{a,b} \gamma_{ab} \log(\gamma_{ab})$

Semi-Unbalanced Entropic OT: if we don't know one of the marginal?

\Rightarrow add another term w.r.t. a "prior" \tilde{q} : $\lambda \cdot D_{KL}(q \parallel \tilde{q})$!

TACCO OT: "Objects" \rightarrow cell-like objects, "Categories" \rightarrow cell types

* OT doesn't explicitly use spatial data!

↳ TACCO incorporates both spatial & compositional annotation data

TACCO Spatial Framework: (boosters) \rightarrow comp. annotation \rightarrow deconvolution

Boosters: Platform Normalization ("change of basis"), Multicenter (k-means), Bisectioning (annotate, then subtract from data)

Object Splitting: derive several virtual observations for each one real

Week 10: Spatial Omics II

Physical tumors are 3-dimensional. How to analyze in 3D?

↳ Locations? Local communications? During homeostasis/disease?

Cellular Niches: Zones of tissue defined by mixture of cells & programs

2D → 3D: slicing, take 2D images at multiple places

↳ but how do we stitch these together?

CellCharter: Preprocess with scVI/scArches (VAE), then identify clusters incorporating neighbor information (Delaunay Triangulation!)

↳ Delaunay maximizes minimum angles → empty circumcircles!

Take n-neighbors, aggregate them for final cell representation.

Use EM algorithm on GMM for clustering. → # of clusters?

↳ do a parameter sweep and compare overlaps with $k' = k \pm 1$. (...?)

of clusters \approx # of subjects can lead to clustering based on subjects.

“ >> “ eventually leads to subclusters for subjects!

Week 11: scCRISPR Screens

Genetic knockout: originally very difficult & time-consuming
RNAi, ZFN, TALEN, ... CRISPR!

CRISPR: enables easier genome editing, modular & efficient

CRISPR + scRNA Seq? cannot read which guide is in which cell

↳ create a "barcode" mRNA ... but this breaks the virus!

↳ we can reverse the entire cassette to bypass this issue

Gene Regulatory Networks: nodes are regulators & targets, directed edges and types show the regulation

↳ Knockout of a TF can lead to many downstream effects

Double knockout experiments can give some evidence (hopefully)

↳ some technical / biological variations, some KOs are better than others

Linear Regression: $Y = X\beta + \epsilon$ naturally extends to $X \in \mathbb{R}^d$.

MIMOSCA: Y, X, β are all matrices, Y is (cells x expressions)

↳ uses elastic net regularization, $\lambda_1 \sum |\beta_j| + \lambda_2 \sum \beta_j^2$, L_1 & L_2 penalties.

↳ ridge helps with correlation, lasso helps with sparsity, general denoising

↳ use random permutation to simulate no associations, then compare

* t-tests/p-values don't work because of correlations!

→ find TF co-modules/gene programs, genetic expressions

Now we want to look at in vivo cells via CRISPR screening

What genes control T cell states in TME? Which TFs push cells?

Controlled model: 1) tumor with known antigen 2) T cells that recognize it

↳ How to choose which TF to KO? DE/DA to curate 180 TF-library

Week 12: Foundation Models

Motivation: How can we scale system biology better?

Challenges: 1) Knowledge is not shared amongst different datasets.

2) Each model is task-specific. 3) Poor out-of-distribution generalization

⇒ Can we build a "universal" model for sc biology?

Foundation Model: self-supervised feature extractor, tokenizes genes!

↳ Core philosophy: Pre-train universally, Fine-tune on demand.

* scRNA data is not sequence but a tablature, how to fit it in?

* some criticism on zero-shot performance being worse than specific models

scGPT: input of (expression values, gene tokens, condition tokens) for a cell

Nicheformer: FM for spatial omics