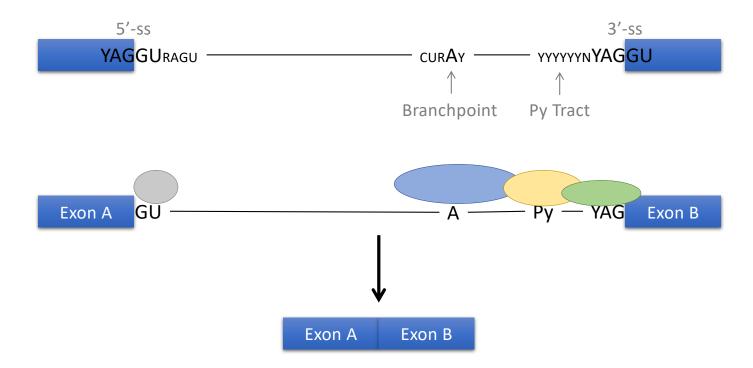
Splicing/Motif Mark

February 13, 2019

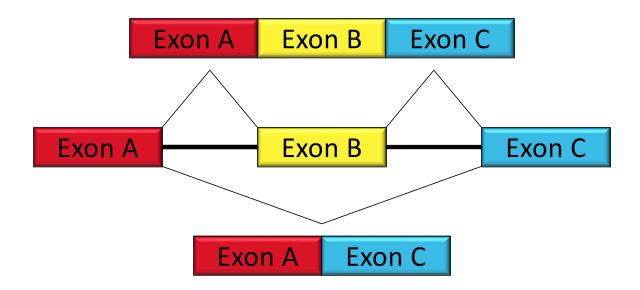
Observations

• MBNL1 and RBFOX1 are both regulators of alternative splicing

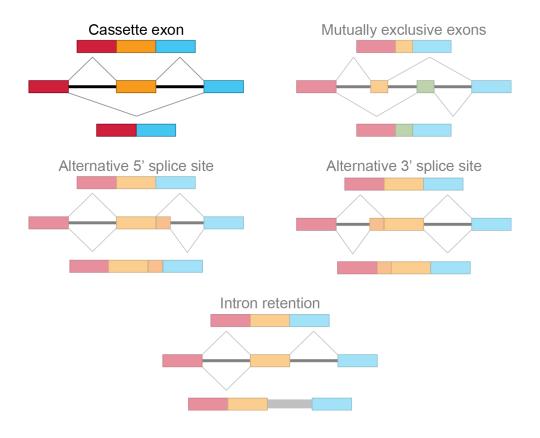
What is splicing?



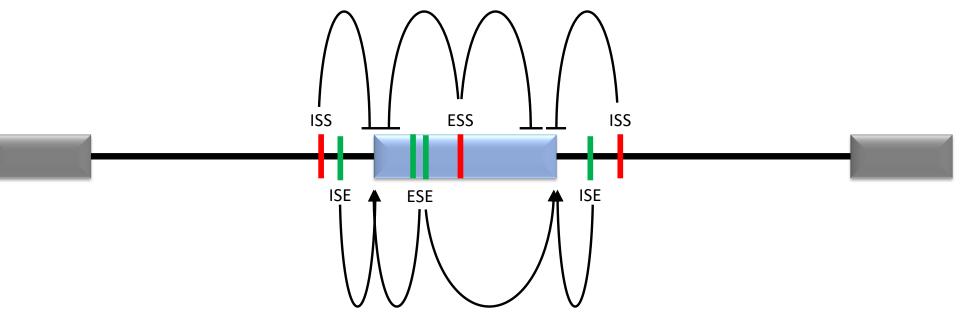
What is alternative splicing?



What is alternative splicing?

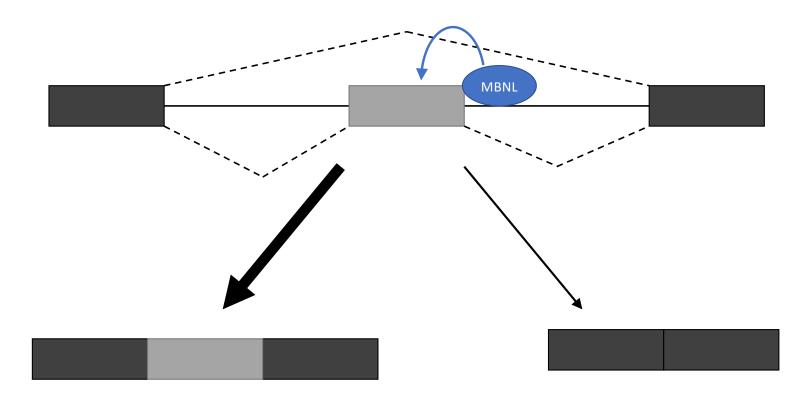


How is the splicing decision made?

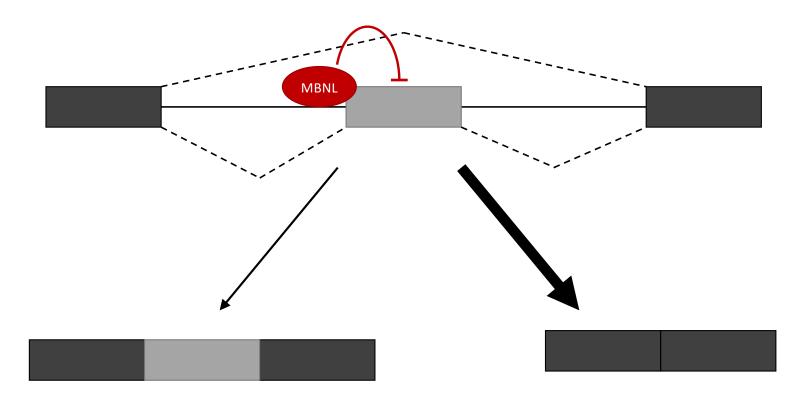


- ISS intron splicing silencer
- ISE intron splicing enhancer
- ESS exon splicing silencer
- ESE exon splicing enhancer

MBNL is a splicing factor that is both an activator and suppressor of exon inclusion for pre-mRNA splicing



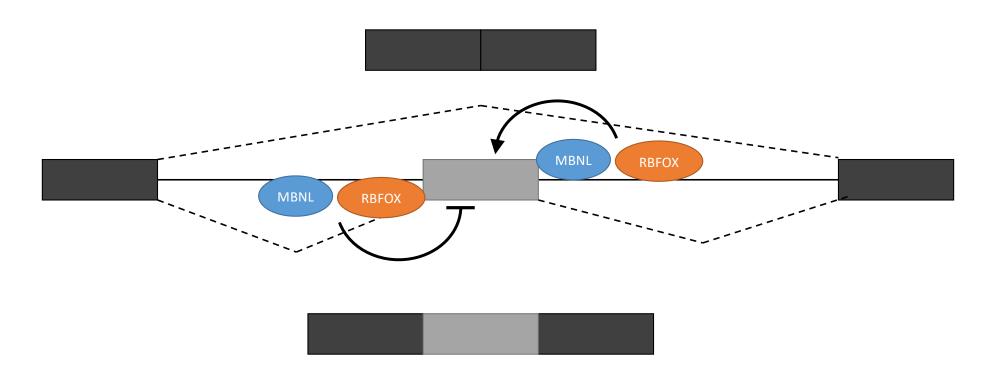
MBNL is a splicing factor that is both an activator and suppressor of exon inclusion for pre-mRNA splicing



Observations

- MBNL1 and RBFOX1 are both regulators of alternative splicing
- MBNL1 and RBFOX1 regulate splicing in similar manners

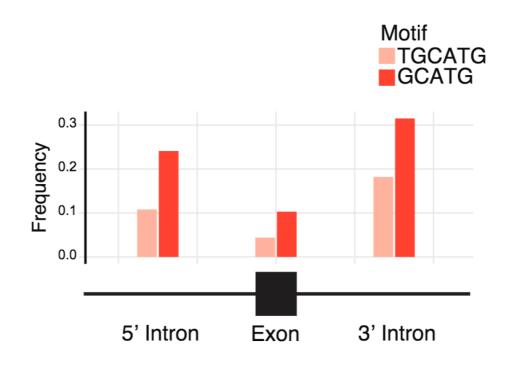
RBFOX1 regulates alternative splicing in a similar manner to MBNL1



Observations

- MBNL1 and RBFOX1 are both regulators of alternative splicing
- MBNL1 and RBFOX1 regulate splicing in similar manners
- MBNL1 and RBFOX1 have similar expression profiles
 - Both expressed in brain, heart, and skeletal muscle
 - Increased expression leads to alternative splicing changes critical for development
 - Depletions in expression have been associated with neurological disorders
- Some MBNL1 regulated splicing events have RBFOX1 binding sites

Analysis of 203 MBNL-regulated exons in mouse revealed the presence of RBFOX binding motifs



66% contain a GCAUG and 33% contain a UGCAUG RBFOX motif in the exon or within 250 nucleotides for the flanking exons

Research question

 How do MBNL1 and RBFOX1 work together to regulate splicing events in transcripts that are regulated by both?

Our goal

 Develop a Python script to plot protein binding motifs on an image of the an exon and flanking introns >INSR chr19:7149896-7151209 (reverse complement)

agaaa gaagtggctgagtcagttgtgatgtccacatgtagtcacgtttgacatcccagggccacctcagcaggccgtctct ggggagaattttctctgatttcttccccttcccttgctggacccctgcacctgctggggaagatgtagctcactcc gtctagcaagtgatgggagcgagtggtccagggtcaaagccagggtgcccttactcggacacatgtggcctccaag tgtcagagcccagtggtctgtctaatgaagttccctctgtcctcaaaggcgttggttttgtttccacagAAAAACC TCTTCAGGCACTGGTGCCGAGGACCCTAGgtatgactcacctgtgcgacccctggtgcctgctccgcgcagggccg gcggcgtgccaggcagatgcctcggagaacccaggggtttctgtggctttttgcatgcggcgggcagctgtgctgg agagcagatgcttcaccaattcagaaatccaatgccttcactctgaaatgaaatctgggcatgaatgtggggagaa tgcagccactgtttgctcactaaacatctctgcacctcccgcgtgcatttgcagaggtggggggtccccggag tctgagctccccgcggctgggtgccccgacccagcagctcctacaccatgaatggaggttgatctggaaacagaat attttcatgaaagggcgacagggtatgaacaaaagaacaccgtgtcgctcactgaattccacggaggagagtcagg ctttctttccttcttttc

>INSR chr19:7149896-7151209 (reverse complement)

agaaa gaagtggctgagtcagttgtgatgtccacatgtagtcacgtttgacatcccagggccacctcagcaggccgtctct gtctagcaagtgatgggagcgagtggtccagggtcaaagccagggtgccttactcggacacatgtggcctccaag tgtcagagcccagtggtctgtctaatgaagttccctctgtcctcaaaggcgttggttttgtttccacagAAAAACC TCTTCAGGCACTGGTGCCGAGGACCCTAGgtatgactcacctgtgcgacccctggtgcctgctccgcgcagggccg gcggcgtgccaggcagatgcctcggagaacccaggggtttctgtggctttttgcatgcggcgggcagctgtgctgg agagcagatgcttcaccaattcagaaatccaatgccttcactctgaaatgaaatctgggcatgaatgtggggagaa tgcagccactgtttgctcactaaacatctctgcacctcccgcgtgcatttgcagaggtggggggtccccggag tctgagctccccgcggctgggtgccccgacccagcagctcctacaccatgaatggaggttgatctggaaacagaat attttcatgaaagggcgacagggtatgaacaaaagaacaccgtgtcgctcactgaattccacggaggagagtcagg ctttctttccttcttttc

Activity

- Working in groups, discuss general strategy for the algorithm
 - What should the input look like?
 - What should the output look like?
 - What functions are you going to write?
 - How can you handle multiple motifs and multiple genes?
 - How are you going to identify motifs with ambiguity, i.e. YCGY?

Input

- FASTA file
 - How denote introns vs exons?
- Coordinates
 - UCSC genome browser
 - Need assembly info
 - How interface with UCSC?
- Other?

Output

- Vector based image
- To scale!
- Introns vs exons
- Denote multiple motifs
- Key?



>INSR chr19:7149896-7151209 (reverse complement)

agaaa gaagtggctgagtcagttgtgatgtccacatgtagtcacgtttgacatcccagggccacctcagcaggccgtctct ggggagaattttctctgatttcttccccttcccttgctggacccctgcacctgctggggaagatgtagctcactcc gtctagcaagtgatgggagcgagtggtccagggtcaaagccagggtgccttactcggacacatgtggcctccaag tgtcagagcccagtggtctgtctaatgaagttccctctgtcctcaaaggcgttggttttgtttccacagAAAAACC TCTTCAGGCACTGGTGCCGAGGACCCTAGgtatgactcacctgtgcgacccctggtgcctgctccgcgcagggccg gcggcgtgccaggcagatgcctcggagaacccaggggtttctgtggctttttgcatgcggcgggcagctgtgctgg agagcagatgcttcaccaattcagaaatccaatgccttcactctgaaatgaaatctgggcatgaatgtggggagaa tgcagccactgtttgctcactaaacatctctgcacctcccgcgtgcatttgcagaggtgggggtgggggtccccggag tctgagctccccgcggctgggtgccccgacccagcagctcctacaccatgaatggaggttgatctggaaacagaat attttcatgaaagggcgacagggtatgaacaaaagaacaccgtgtcgctcactgaattccacggaggagagtcagg ctttctttccttcttttc

Functions

- Parse FASTA
- Parse file with motifs
- Drawing function?

Activity

- Working in groups, discuss general strategy for the algorithm
 - What should the input look like?
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 - How are you going to identify motifs with "flexibility", i.e. YCGY?



conda install -c conda-forge pycairo

```
import cairo
surface = cairo.SVGSurface("plot.svg", width, height)
context = cairo.Context(surface)
context.set_line_width(1)
context.move to(50,25)
context.line to(intron1+exon+intron2,25)
context.stroke()
```

https://pycairo.readthedocs.io

https://cairographics.org/documentation/pycairo/3/

Your assignment: Motif Mark

- Create a repository in your Github profile called motif-mark
- Write a python script to visualize motifs on sequences
 - Minimum requirements:
 - Python3 compatible code
 - Use argparse
 - Input FASTA file and motifs file
 - Multiple sequences
 - Multiple motifs
 - · Ambiguous motif handling
 - svg output
 - Key/labeling

- Stretch:
 - Alternate inputs (coordinates)
 - · Option of output
 - Offers user customizable colors
 - Color palate options

 Turn in your code and a figure for the data Leslie will provide on Canvas